

the latter 1.0, and these means differ significantly ( $t = 3.25$ ;  $P < 0.01$ ). This ratio is markedly affected by thyroid function<sup>2</sup> and our results may indicate that thyroid function is greater in Japanese than in British women. Since thyroid disease is reported to affect the incidence of breast cancer<sup>3,4</sup> the difference in 5 $\alpha$ /5 $\beta$  ratios between the populations may be relevant to the difference in incidence.

R. D. BULBROOK  
B. S. THOMAS

Imperial Cancer Research Fund,  
Lincoln's Inn Fields,  
London, W.C.2.

J. UTSUNOMIYA

Tokyo Medical and Dental University,  
1-3 Chomo, Yushima,  
Bunkyo-ku, Tokyo.

<sup>1</sup> Wynder, E. L., Bross, I. J., and Hirayama, T., *Cancer*, **13**, 559 (1960).

<sup>2</sup> Hellman, L., Bradlow, H. L., Zumoff, B., Fukushima, D. K., and Gallagher, T. F., *J. Clin. Endocrinol. and Metab.*, **19**, 936 (1959).

<sup>3</sup> Reper, R. W., *J. Michigan State Med. Soc.*, **51**, 1815 (1952).

<sup>4</sup> Loeser, A. A., *Brit. Med. J.*, **ii**, 1880 (1954).

### Effect of Norleucine on the Synthesis of Demethylchlortetracycline by *Streptomyces aureofaciens*

A NUMBER of recent reports have dealt with inhibition of methylation in *Streptomyces aureofaciens*. Strains of this organism which normally synthesize chlortetracycline can be directed toward the production of the 6-demethyl analogue by adding a number of compounds to the medium. Such compounds have included a variety of sulphonamides<sup>1,2</sup> and aminopterin<sup>3</sup> which act as inhibitors of folic acid mediated C6-methylation. In addition ethionine<sup>3-5</sup>, and to some extent D-methionine<sup>6</sup>, when added to the medium have also caused 6-demethylchlortetracycline production presumably by acting as antagonists of L-methionine, the normal source of the 6 methyl group<sup>6</sup>.

Norleucine, a methionine antagonist in *Escherichia coli*<sup>7-9</sup>, was tested against *S. aureofaciens* BC-41, a strain which produces only chlortetracycline and tetracycline. Using the paper chromatographic system previously described and a device to scan the chromatograms<sup>5</sup>, a small but consistent amount of the 6-demethyl analogue could be detected on addition of norleucine to the medium. This is illustrated in Table 1. Only the D-isomer of norleucine was found effective. The effect of norleucine could be fully reversed by methionine and to a much lesser extent by leucine and isoleucine. A similar effect has been noted in *E. coli*<sup>9</sup>.

Table 1. EFFECT OF NORLEUCINE ON BIOSYNTHESIS OF CHLORTETRACYCLINE AND 6-DEMETHYLCHLORTETRACYCLINE

DL-Norleucine (p.p.m.)	Chlortetracycline ( $\mu\text{g/ml.}$ )	6-Demethylchlortetracycline ( $\mu\text{g/ml.}$ )
0	6,340	0
350	4,605	175
500	3,030	120
1,000	1,390	70

A number of compounds related to norleucine were also tested for their ability to cause 6-demethylchlortetracycline biosynthesis. The unsaturated analogue of norleucine, *cis*-crotyl glycine, was reported by Skinner *et al.*<sup>10</sup> to be a potent methionine antagonist in *E. coli*. Through the courtesy of Dr. Skinner an authentic sample was obtained and tested in our system. It was without effect on growth or biosynthesis of tetracyclines. Similarly, norvaline<sup>7</sup> and its dehydro analogue, allyl glycine, were without effect in causing 6-demethylchlortetracycline biosynthesis, though both compounds did result in a decrease in growth and total tetracyclines production.

All methylation inhibitors thus far reported in the literature to be effective against *S. aureofaciens* cause a severe depression in total tetracyclines production, and, as

can be seen from Table 1, norleucine is no exception. Compared with the sulphonamides<sup>1,2</sup>, it causes the production of only a low proportion of demethylated to methylated antibiotic. For large-scale production of 6-demethylchlortetracycline, the use of mutant strains<sup>11</sup> remains the method of choice.

J. J. GOODMAN  
MARY MATRISHIN

Lederle Laboratories Division,  
American Cyanamid Co.,  
Pearl River, New York.

<sup>1</sup> Goodman, J. J., and Matrishin, M., *J. Bact.*, **82**, 615 (1961).

<sup>2</sup> Perlman, D., *et al.*, *J. Amer. Chem. Soc.*, **83**, 4481 (1961).

<sup>3</sup> Niedleman, S. L., Bienstock, E., and Bennett, R. E., *Biochim. Biophys. Acta*, **71**, 199 (1963).

<sup>4</sup> Hendlin, D., *et al.*, *Biochim. Biophys. Acta*, **58**, 635 (1962).

<sup>5</sup> Goodman, J. J., and Miller, P. A., *Biotech. and Bioeng.*, **4**, 391 (1962).

<sup>6</sup> Miller, A. P., McCormick, J. R. D., and Doerschuk, A. P., *Science*, **123**, 1030 (1956).

<sup>7</sup> Harris, J. S., and Kohn, H. I., *J. Pharmacol. Exp. Therap.*, **73**, 383 (1941).

<sup>8</sup> Lampen, J. O., and Jones, M. J., *Arch. Biochem.*, **13**, 47 (1947).

<sup>9</sup> Harding, W. M., and Shive, W., *J. Biol. Chem.*, **174**, 743 (1948).

<sup>10</sup> Skinner, C. G., Edelson, J., and Shive, W., *J. Amer. Chem. Soc.*, **83**, 2281 (1961).

<sup>11</sup> McCormick, J. R. D., *et al.*, U.S. Patent 2,878,289 (1959).

### Extraction from Human Skin of Soluble Collagen Molecules containing only Beta Components

SOLUBLE collagens have been extracted from connective tissue with various media<sup>1,2</sup>. One type of molecular species consisting of  $\alpha$  components has been obtained by extracting with neutral salt solutions, and another type consisting of varying proportions of  $\alpha$  and  $\beta$  components has been obtained by extraction with buffered acid solutions. This communication describes the extraction of a different molecular species of collagen which is composed of  $\beta$  components only.

Specimens of human skin of the new-born were acquired as previously described<sup>3</sup>. The connective tissue was treated (repetitively) with each of the following in succession: 0.15 M, 0.45 M, 1.0 M, and 2.0 M sodium chloride; followed by 0.2 M citrate buffer (0.1 M citric acid and 0.1 M sodium citrate, pH 4.3). These extractions removed both the neutral salt-soluble and the citrate-soluble collagens. The resulting tissue paste was then repeatedly treated with 0.15 M citrate buffer (0.1 M citric acid and 0.05 M sodium citrate; reduced to pH 1.5 with hydrochloric acid) for 24 h. The yield decreased progressively with each successive extraction which was precipitated by dialysis with 0.01 M disodium hydrogen phosphate. The precipitate was collected and dissolved by dialysis against formate buffer, pH 3.75, ionic strength 0.15. All these procedures were done at 5° C.

Each collagen fraction from successive extractions with citrate buffer at pH 1.5 was heated to 40° C for 20 min and run in the ultracentrifuge at 35° C. The patterns exhibited increasing  $\beta$  content until finally  $\beta$  alone was seen after several extractions (Fig. 1). The ultracentrifuge patterns of this collagen, which was denatured in 2 M KSCN, were identical to those observed following heat denaturation. By this preparative procedure using extraction at low pH, molecules have been isolated that are made up of only  $\beta$  components. From these results, it may be concluded that the molecular weight of the undenatured collagen must be a whole-number multiple of the molecular weight of the  $\beta$  chain.

The undenatured collagen containing  $\beta$  chains had the same physical parameters which have been recorded previously<sup>4</sup> of citric acid soluble collagen prepared by extraction with citrate at pH 4.3 with no prior neutral salt extraction:  $S_{20,w}$ , 3.1; intrinsic viscosity, 13.5; optical rotation, -414°; and temperature of denaturation, 36° C. From the sedimentation and viscosity data, the molecular weight of the collagens obtained by the different extraction