

Table 1. MILK-LEVELS OF OESTROGENS PRIOR TO AND AFTER SINGLE INTRAMUSCULAR INJECTION OF 50 MG OESTRADIOL-17 β

Day No.	ng per 50 ml.*			
	Ether soluble		Ether insoluble	
	Oestrone	Oestradiol	Oestrone	Oestradiol
1	1	3	1	6
2	2	3	2	8
3	2	5	<1	7
4	2	10	<1	8
5	Injection			
	Evening†	4	30 \pm 2.3	353 \pm 14
6	Morning	2	13	208
	Evening	2	8	146
7	Morning	—	7	114
	Evening	—	7	92
12	—	—	2	21
20	—	—	<1	6
21	—	—	<1	9

* Oestradiol calculated as oestradiol-17 β . Means of duplicate estimates uncorrected for losses due to the analytical procedure. The recoveries of free oestrogens (1- μ g level) added to intact milk or taken through hydrolysis in the presence of milk extracts (cf. method) amounted to 40–50 per cent.

† The milk was collected 8 h after injection. The values of the ether-insoluble oestrogens represent the means (\pm S.E.) of 11 estimates.

demonstrated in the first milk sample drawn after the injection of the β -epimer. A slight increase was recorded of the milk levels of ether non-soluble oestrone. Oestrone is another metabolite of oestradiol-17 β previously described in cattle¹.

In the milk produced during the first 48 h after injection the net increase of oestradiol recovered after hydrolysis of the postulated derivative insoluble in ether may be calculated to about 150 μ g or roughly 0.3 per cent of the dose. Since the recoveries of free oestradiol-17 β similarly treated were not better than 50 per cent the true value was presumably at least twice as high.

It was shown that the concentrations of free oestradiol in the blood on the day of injection rose to levels comparable with those found in the milk of the derivative insoluble in ether. Tentatively, no appreciable increase in the blood levels of oestrogen conjugates could be demonstrated. The possibility should thus be considered that the mammary gland itself might be the site of the apparent conjugation of the oestrogens being eliminated with the milk.

This work was supported by grants from the Agricultural Research Council of Norway.

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¹ Velle, W., thesis, Oslo (1958).

² Gorski, J., and Erb, R. E., *Endocrinol.*, **64**, 707 (1959).

³ Münch, U., *Milchwissenschaft*, **9**, 150 (1954).

⁴ Pope, G. S., and Roy, J. H. B., *Biochem. J.*, **53**, 427 (1953).

⁵ Edwards, R. W. H., Kellie, A. E., and Wade, A. P., *Mem. Soc. Endocrinol.*, **2**, 53 (1953).

⁶ Brown, J. B., *Biochem. J.*, **60**, 185 (1955).

⁷ Ittrich, G., *Z. physiol. Chem.*, **312**, 1 (1958).

⁸ Lunaas, T. (to be published).

Microbial Transformation of *n*-Octane into Dicarboxylic Acids

In 1960 Kester and Foster¹ demonstrated the conversion of certain liquid hydrocarbons to the corresponding alkanedioic acids in cultures of a Gram-positive bacterium. *n*-Alkanes possessing between 10 and 14 carbon atoms were shown to undergo this process of di-terminal oxidation. Evidence has been obtained^{2,3} that *n*-decanoic and 10-hydroxydecanoic acids are intermediates in the oxidation of *n*-decane to *n*-decanedioic acid.

In our experiments we have used a Gram-negative bacterium, tentatively identified as a *Pseudomonas*, isolated from the soil of a grass lawn which had been previously exposed to commercial petrol. The organism has been grown in a medium containing mineral salts and *n*-octane. Continuous ether extraction of acidified culture solution gives a mixture of acids, of which two have been identified. Suberic acid (*n*-octanedioic acid) has been isolated from the mixture in a pure crystalline

form and characterized on the basis of its chromatographic properties, melting point and mixed melting point with authentic material, elementary analysis, neutralization equivalent and infra-red spectrum. Adipic acid (*n*-hexanedioic acid) is present in smaller amounts and has been identified by partition chromatography of the free acid on paper and on silica-gel columns and further by vapour phase chromatography of the dimethyl ester.

In the light of the work of Kester and Foster, our investigation enhances the view that di-terminal oxidation is a significant aspect of *n*-alkane metabolism.

We thank Prof. A. A. Eddy for his interest in this work and Dr. J. W. Foster for providing us with unpublished information. For financial support we thank the Department of Scientific and Industrial Research (D. S. R.) and H. E. H. the Nizam's Sahibzadas Surf-e-Khas Trust, Hyderabad (M. Y. A. K.).

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¹ Kester, A. S., and Foster, J. W., *Bact. Proc.*, 168 (1960).

² Kester, A. S., *Dissertation Abstr.*, **22**, 3350 (1962).

³ Foster, J. W., in *The Oxygenases*, edit. by Hayaishi, O. (Academic Press, New York) (in the press).

Stimulation of Oxytetracycline Formation by *N*-Acetyl Derivatives of Certain Amino-acids

SINCE acetate can serve as an effective precursor to the formation of the ring structure of the tetracycline (TC) molecule¹, it was of interest to determine the role of certain *N*-acetylated amino-acids in formation of oxytetracycline (OTC). This communication summarizes several observations on the stimulation of antibiotic formation through the inclusion of certain *N*-acetylated amino-acids in *Streptomyces rimosus* (NRRL 2234) fermentations.

The procedures used for all shake flask fermentations, preparation of inocula, antibiotic assays and determinations of growth were similar to those previously described². Composition of the basal medium follows (g/l.): amino-acid, 0.5 (as organic nitrogen); glucose, 10.0 (autoclaved separately); NaCl, 5.0; K₂HPO₄, 2.0; MgSO₄·7H₂O, 1.0; CaCl₂·2H₂O, 0.4; (NH₄)₂HPO₄, 0.2; FeSO₄·7H₂O, 0.02, and ZnSO₄·7H₂O, 0.01.

Table 1. EFFECT OF *N*-ACETYL DERIVATIVES OF CERTAIN AMINO-ACIDS ON GROWTH AND ANTIBIOTIC PRODUCTION BY *Streptomyces rimosus*

Amino-acid source	Final pH	TC activity (μ g/ml.)	Dry cell weight	TC activity (mg/g cells)
None	7.3	13	47	26.8
DL-Alanine	8.0	47	322	14.6
<i>N</i> -Acetyl-DL-alanine	8.3	183	258	71.0
Glycine	8.4	42	225	18.6
<i>N</i> -Acetyl-glycine	8.2	199	257	77.4
DL-Methionine	7.3	28	234	12.0
<i>N</i> -Acetyl-DL-methionine	8.1	30	69	43.5
DL-Leucine	7.2	12	65	18.9
<i>N</i> -Acetyl-DL-leucine	8.0	38	237	16.0
DL-Tryptophan	7.0	12	57	20.2
<i>N</i> -Acetyl-DL-tryptophan	7.1	14	53	26.4
DL-Valine	7.1	70	248	28.2
<i>N</i> -Acetyl-DL-valine	7.3	72	226	31.8
L-Glutamic acid	9.1	63	289	21.8
<i>N</i> -Acetyl-L-glutamic acid	8.9	83	317	26.1

The results in Table 1 show that the most striking stimulation of antibiotic production was noted in media containing *N*-acetyl-DL-alanine or *N*-acetyl-glycine. Antibiotic yields per unit of dry-cell weight were increased 4- to 5-fold above those observed with the parent amino-acids. *N*-Acetyl-DL-methionine was not effectively utilized for growth but resulted in comparable titres of antibiotic activity per unit volume of broth as that found