

ment appears long after physiological mechanisms begin to operate. The synthesis of needed enzymes and the oxidation of carbon provide the energy and intermediates that are necessary for germ tube formation.

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Effect of *Pseudomonas tabaci* on the Metabolism of Starch in Tobacco Leaves

In the host tissues infected by various plant pathogens, including viruses, bacteria and fungi, as a rule starch accumulates around the infection area¹⁻³. By contrast, the toxin-induced chlorotic halo formed in White Burley tobacco leaves infected by *Pseudomonas tabaci* (Wolf and Foster) Stevens fails to show any starch reaction (Fig. 1). The low starch content of the affected area was also shown by quantitative chemical determination⁴ (Table 1). The assays were made 7 days after needle inoculation.

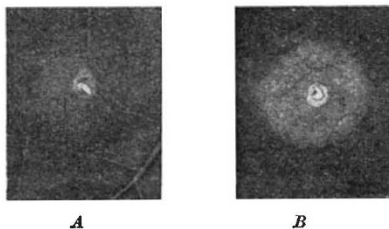


Fig. 1. Parts of a tobacco leaf after needle-inoculation with *Pseudomonas tabaci*. A, Untreated; B, stained with iodine

The decrease in starch content could also be induced by the injection of bacteria-free culture filtrate (Table 1). Czapek-Dox nutrient medium incubated with *Ps. tabaci* for 48 h was used. Treatments were carried out on intact plants by injecting one half of the leaves with the culture filtrate and the other with Czapek-Dox nutrient as control. Measurements were made 3 days after injection.

Table 1. EFFECT OF *Pseudomonas tabaci* ON THE STARCH CONTENT AND PHOSPHORYLASE ACTIVITY OF TOBACCO LEAVES

Treatment of leaves	Starch content* (mg/g fresh weight)	Phosphorylase activity*	
		Synthesized amylose (mg/g fresh weight/1 h)	PO ₄ ³⁻ liberated (mg/g fresh weight/1 h)
Control †	2.6	4.5	1.8
Needle inoculated	1.5	1.2	0.6
Control ‡	5.2	4.4	1.6
Injected with culture filtrate	2.3	2.3	0.9

* Values from representative experiments.

† Unaffected areas of the same leaf.

‡ Half-leaf injected with Czapek-Dox nutrient.

To elucidate the possible mechanism responsible for decrease in starch content the activity of the two major enzyme systems involved in starch metabolism was measured. A negligible and equal amylase activity was found in the extracts from control and culture filtrate treated leaves. The activity of phosphorylase, however, exhibited striking differences. The extent of amylose synthesis from glucose-1-phosphate was measured as follows: 500 mg tissue was extracted in the cold in 2.5 ml. acetate buffer pH 6.0. The reaction mixture contained

1 ml. crude extract (without centrifugation), 10⁻² M sodium fluoride, 0.2 ml. 1 per cent dextrin as primer, and 1.7 × 10⁻² M glucose-1-phosphate in 2.3 ml. final volume. After incubation at 30° C for 30 min the reaction was stopped by adding equal amounts of 10 per cent trichloroacetic acid (controls were treated with trichloroacetic acid at zero time). The starch (amylose) content was determined colorimetrically in centrifuged samples by adding 0.2 ml. of an iodine solution containing 60 mg iodine/100 ml. A standard curve was prepared from soluble starch. Alternatively the orthophosphate set free during starch synthesis was measured⁵.

As shown in Table 1 phosphorylase activity, as compared with the controls, was markedly less both in the halo areas of the infected leaves and in the tissues treated with culture filtrates.

The *Ps. tabaci* strain used in these experiments does not hydrolyse starch. The culture filtrate added to phosphorylase-containing plant extracts *in vitro* did not diminish the activity of phosphorylase. Therefore, it can be stated that the decrease in phosphorylase activity is due to an indirect effect of *Ps. tabaci* on the metabolism of the host. This effect is specific because the activity of some other enzymes, for example, glucose-6-phosphate dehydrogenase, increases in the tissues surrounding the infection area. The parallelism between the decrease in starch content and inhibition of phosphorylase activity suggests a causal relationship.

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Optimum Leaf Area Index in the Potato Crop

In his review of the effect of competition for light on crop growth Donald¹ notes the existence of but two published accounts presenting evidence to support the generally adopted conception of an 'optimum leaf area index' as the ratio of leaf area to ground covered at which the balance of photosynthesis over respiration is at a maximum value. For kale crops in southern England the figure lies between 3 and 5.4 (ref. 2). In Australia, subterranean clover was found to reach its highest net assimilation rate when the leaf area index lay between 4 and 5 (ref. 3). More recently, Stern and Donald⁴ have shown that the optimum leaf area index for clover in Australia varies with the amount of light present, particularly the amount which penetrates to the lower leaves, rising from 3.5 at a radiation-level of 50 g cal/cm²/day to a nearly constant 5 above 150 g cal/cm²/day. Donald¹ calls for more evidence for other crops and different regions.

In an experiment on the effect of applying four levels of a 12 per cent N : 12 per cent P₂O₅ : 18 per cent K₂O fertilizer to a crop of Majestic potatoes at Boghall, Midlothian, in 1961, measurements were made of leaf area and the dry matter of leaves, stems, roots and tubers at four dates each approximately three weeks apart. The plants which received 2.5 and 5 cwt. per acre rates of fertilizer (Fig. 1) show a steady rise in leaf area index to 4.4 and 5.3 respectively at the third sampling date, declining thereafter to 3.9 and 4.9. The plants given the 7.5 and 10 cwt. per acre treatments rose swiftly through the level of leaf area index 5 to reach maxima of 7.0 and 8.3, afterwards declining to 5.5.