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Localization and Distribution of Tritiated **Histidine in Growing Mouse Incisor**

HISTIDINE has been reported as being the predominant basic amino-acid of developing human enamel¹. At present specific histochomical methods for in situ demonstration of histidine are not available². With the recent availability of tritiatod histidine, autoradiography appears to offer an adequate method for tracing its utilization in the growing incisors of mice.

Twenty-four female Brookhaven National Laboratory Swiss albino mice, 16 days of ago, were used in this work. The mice received a subcutancous injection of 1 µc. of tritiated histidine per gram of body-weight 1, 4, 8, 24 hr., 7 and 14 days prior to killing with other. Immediately after killing the lower jaws were romoved and fixed for 1 week in cold buffered formalin and washed for an additional 24 hr. in running tap water. Tissue samples were decalcified in a solution of 10 per cent 'Versene'. Paraffin blocks were prepared and sections cut at 5μ . Autoradiographs were prepared using Kodak ' NTB_3 ' liquid emulsion. The preparations were kept in a cold, dry atmosphere and exposed for 30 days. After proper exposure the auto-radiographs were developed, stained with hæmatoxylin and eosin, and grain counts were made. These were based on a unit area of 750 μ^2 .

Autoradiographic results showed that labelling of various incisor cells was intense after 1 hr. (20 grains). Histidine labelling was predominantly seen at the dentinal end of the cytoplasm of ameloblasts and the adjacent enamel matrix forming a band of reduced silver grains (Fig. 1). With increasing time the grain count over the cytoplasm of ameloblasts became diminished while additional grains appeared in the enamel matrix. As the grain count increased in the matrix the band of reduced silver grains became wider. At seven days the band appeared about 3-4 times wider (Fig. 2). At 4 hr. a marked decrease in labelling was observed in ameloblasts (9.6 grains). The labelling intensity of the enamel matrix adjacent to ameloblasts was maintained at 4 hr., but at 8 hr. the grain density was diminished. Enamel matrix grain distribution was altered with time so that at 7 days the grain density was higher at the widening band front (Fig. 2), and at 14 days the entire width of the enamel matrix was covered with grains. The total population of grains, however, was smaller than at earlier periods. Only a fow grains were observed in ameloblasts at this time (1.1 grains).

Odontoblasts (11.1 grains) and dentinal matrix (7.5 grains) were labelled most strongly at 1 hr.; however, the total grain density was much lower than that of the amoloblast (20 grains) and enamel matrix (62.4 grains). Changes in grain distribution with time were essentially similar to that described for the ameloblasts and enamel matrix, except for the reverse direction of the wave front of dentine proper. The pulp, which is composed of several cell types, revealed a broad distribution of grains. At the base of the incisor the ameloblasts and odontoblasts were weakly labelled.

The presence of a high concentration of histidine in the developing enamel revealed by biochemical analysis was confirmed autoradiographically¹. Histidine turn-over is more rapid in amcloblasts than in odontoblasts during matrix formation. Turn-over rate of these colls is quite rapid since labelled matrix was seen within 1 hr. Gould *et al.*³ stated that cortain tissues, such as liver, bone and periodontal membrane, exhibit a relatively rapid turn-over of

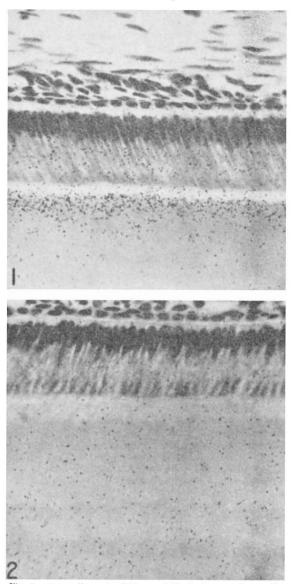


Fig. 1. Autoradiograph of the amcloblastic layer and enamel matrix of mouse incisor labelled with tritiated histidine 1 hr. following the administration of the isotope. (Oil immersion, $\times c.480$) Fig. 2. Autoradiograph of the amcloblastic layer and enamel matrix of mouse incisor labelled with tritiated histidine 7 days following the administration of the isotope. (Oil immersion, $\times c.480$)

collagen. The present studies reveal that the population of silver grains present over enamel and dentinal matrix was far less at 14 days than the number observed at earlier periods. This fact is taken to represent a fairly rapid turn-over of histidine in tho incisor matrix of the mouse.

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Eastoe, J. E., Nature, 187, 411 (1960).

- Eastoe, J. E., Nature, 187, 411 (1960).
 ^a Pearse, A. G. E., Histochemistry, Theoretical and Applied, second ed., 91 (Little, Brown and Co., Boston, 1960).
 ^a Gould, B. S., Manner, G., Goldman, H. M., and Stolman, J. M., Ann. New York Acad. Sci., 85, 385 (1960).