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NATURE

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- ¹ Lucké, B., J. Exp. Med., 68, 457 (1938); Ann. N.Y. Acad. Sci., 54, 1093 (1952).
- ² Duryee, W. R., Ann. N.Y. Acad. Sci., 63, 1280 (1956).
- ³ Rose, S. M., Ann. N.Y. Acad. Sci., **54**, 1110 (1952). ⁴ Tweedell, K. S., Cancer Res., **15**, 410 (1955).
- Lucké, B., J. Exp. Med., 70, 269 (1939).
- Voungner, J. S., Proc. Soc. Exp. Biol. Med., 85, 202 (1954).
- ⁷ Bodian, D., Virology, 4, 575 (1956).

Acylase Activity of Ascites Tumour Cells on Dichloroacetyl Derivatives of Amino-acids

While investigating the specificity of acylase activity of soil bacteria on acyl derivatives of aminoacids1-2, we noted that the acetone powder of Ehrlich ascites carcinoma cells was able to hydrolyse dichloroacetyl-DL-aspartic acid but not (or only to a small extent) the dichloroacetyl derivatives of some aminoacids such as DL-glutamic scid, DL-serine, DL-

methionine and DL-phenylalanine4.

We have now investigated the acylase activity of lyophilized powder of ascites tumour cells such as Ehrlich ascites carcinoma, sarcoma 180, leukæmia SN 36, ascites hepatoma MH 134 and MH 129P in C3H mice, and AH 7974, AH 130, and Yoshida sarcoma in rats on dichloroacetyl derivatives of the following twelve amino-acids: DL-aspartic and DLglutamic acids, glycine, DL-alanine, DL-valine, DLleucine, pr-phenylalanine, pr-tryptophan, pr-serine, DL-threonine, DL-methionine, and E-N-benzoyl-DLlysine.

Lyophilized powder of ascites tumour cells was prepared as follows: intraperitoneally transplanted ascites tumour cells were aspirated at the fifth day after transplantation, centrifuged at 1,000 r.p.m. for 10 min., washed three times with phosphate buffer solution, and then lyophilized. It was confirmed that the cells contained almost 85-95 per cent tumour cells.

Five mgm. of lyophilized powder of ascites tumour cells were added to 1 c.c. of neutralized 0.1 M substrate and I c.c. of tris buffer at pH 8.2, and the mixture was incubated at 37° C. for 2 or 18 hr. in the presence of toluene, and then titrated by Grassmann-Heyde's method.

The results obtained in the experiments carried out, under comparable conditions, on the acylase activity of lyophilized powder of eight ascites tumours are presented in Table 1. Ehrlich carcinoma, sarcoma 180 and leukæmia SH 36 were all able to hydrolyse dichloroacetyl-DL-aspartic acid but not (or only to a small extent) the dichloroacetyl derivatives of the other eleven amino-acids. It is of interest that MH 134, MH 129P, AH 7974, AH 130, and Yoshida sarcoma differed from Ehrlich carcinoma, sarcoma 180, and leukæmia SN 36 in failing to hydrolyse dichloroacetyl-DL-aspartic acid. Incidentally, MH 134, MH 129P, AH 7974 and AH 130 were hepatoma. The low acylase activities of AH 7974 and AH 130 seem worth noting when considered together with the fact that the rat liver could hydrolyse dichloroacetyl-DL-aspartic acid5.

Thus, our experiment showed that there is a significant difference among ascites tumours in their capability of hydrolysing dichloroacetyl derivatives of amino-acids, and that acylase activity may be available as a tool for studying biochemical specificity of living material1-3.

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- Kameda, Y., et al., Nature, 169, 1016 (1952); 170, 888 (1952); 181, 1225 (1958); 182, 453 (1958).
 Kameda, Y., et al., Yakugaku Zasshi, 78, 202, 748, 754, 759, 763, 765, 767, 769 (1958); 80, 362 (1960).
- Kameda, Y., et al., Chem. Pharm. Bull., 6, 321, 394, 395, 441 (1958);
 7, 702, 785, 787, 789 (1959).
- Kameda, Y., Toyoura, E., Kimura, Y., Matsui, K., Omori, T., and Takeuchi, K., Yakugaku Zasshi, 77, 1351 (1957).
 Kameda, Y., et al., Chem. Pharm. Bull. (in the press).

BIOLOGY

Chloride Regulation in Triops

LITTLE physiological work on the branchiopods has been reported because of their relative scarcity and the unpredictability of supplies of animals. Among the freshwater species Krogh¹ reports on some pre-liminary experiments with Branchipus and Triops (Apus), Panikkar² gives some figures for the blood concentration of Chirocephalus, and Fritzche³ gives a blood concentration for Daphnia.

An opportunity arose for me to use a few specimens of Triops cancriformis (Bose) bred in laboratory conditions by Mr. A. R. Longhurst. The level of

Table 1. ACYLASE ACTIVITY OF ASCITES TUMOUR CELLS TOWARD N-DICHLOROACETYL AMINO-ACIDS

		rescentage of hydrolysed substrate for 2 and 18 nr. at 37° C.															
	Ehrlich carcinoma		Sarcoma		Leukæmia					100 D	177 =0-1				Yoshida		
	carcii		180		SN~36		MH~134		MH~129P		AH 7974		AH	AH 130		sarcoma	
Substrate	2	18	2	18	2	18	2	18	2	18	2	18	2	18	2	18	
Dichloroacetyl-DL-aspartic acid	27	56	28	55	22	52	0	7	0	3	1	3	2	3	1	2	
Dichloroacetyl-DL-glutamic acid	0	1	0	0	0	1	1	0	Ŏ	Ō	ō	2	2	2	õ	ō	
Dichloroacetylglycine	0	3	2	3	0	1	1	3	ō	1	2	3	ō	1	ĭ	2	
Dichloroacetyl-DL-alanine	1	4	1	4	0	2	1	5	0	1	2	2	2	2	ī	4	
Dichloroacetyl-DL-valine	0	2	0	3	0	0	0	2	1	3	ī	9	ī	2	1	î	
Dichloroacetyl-DL-leucine	0	2	0	3	1	2	1	4	0	3	2	2	ī	ñ	ō	2	
Dichloroacetyl-DL-phenylalanine	0	2	0	4	1	1	0	1	1	1	0	1	2	6	2	3	
Dichloroacetyl-DL-tryptophan	0	1	0	1	0	0	1	0	0	2	0	1	1	5	2	3	
Dichloroacetyl-DL-serine	0	2	1	2	0	1	0	2	0	1	2	3	1	3	ō	2	
Dichloroacetyl-DL-threonine	2	2	0	2	0	1	0	Ō	0	0	0	1	2	2	Ō	1	
Dichloroacetyl-DL-methionine	0	3	1	2	0	3	0	4	0	0	2	1	1	2	1	2	
$a-N$ -Dichloroacetyl- ε - N -benzoyl-												-	-			-	
DL-lysine	0	2	0	1	1	1	0	Ð	0	0	1	9	1	0	0	1	

The digests consist of 5 mgm. of lyophilized sample, 1 c.c. of neutralized 0·1 M substrate, and 1 c.c. of tris buffer at pH 8·2 at 37° C. Hydrolytic activity was measured by Grassmann-Heyde's alcoholic titration method.