

Detailed results of the investigations will be presented elsewhere.

Acknowledgment is made to the Council of Scientific and Industrial Research, India, for financial assistance.

S. C. AGARWALA  
A. KUMAR  
C. P. SHARMA

Department of Botany,  
University of Lucknow,  
Lucknow.

- <sup>1</sup> Hewitt, E. J., *Ann. Rep. Long Ashton Res. Sta.* (1948).  
<sup>2</sup> Smith, P. F., and Specht, A. W., *Plant Physiol.*, **28**, 371 (1953).  
<sup>3</sup> Crooke, W. M., *Ann. App. Biol.*, **43**, 465 (1955).  
<sup>4</sup> Wallace, J. M., *Dissertation Abstr.*, **17**, 2398 (1957).  
<sup>5</sup> Agarwala, S. C., and Kumar, A., *Proc. Forty-seventh Indian Sci. Congr.*, Pt. 3, 399 (1960).  
<sup>6</sup> Agarwala, S. C., and Kumar, A., *J. Indian Bot. Soc.* (in the press).  
<sup>7</sup> DeKock, P. C., *et al.*, *Plant. Physiol.*, **35**, 599 (1960).

## ENTOMOLOGY

### An Abnormal Form of Female Rat Flea, *Xenopsylla cheopis* Roths.

ROTHSCHILD, in 1911, separated the three species of fleas, *X. cheopis*, *X. astia* and *X. brasiliensis* of the genus *Xenopsylla* found on rats. *X. cheopis* is widely distributed in the tropics and is the principal plague flea in India. The three species (♀♀) mentioned here can be easily identified by the shape of the spermatheca. Until now, it has been reported that *X. cheopis* has a single spermatheca<sup>1-3</sup> (Fig. 1).

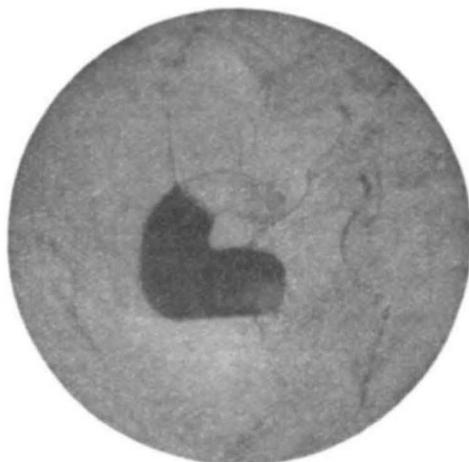


Fig. 1. ( $\times 132$ )

During a routine collection of rat fleas from Delhi City for conducting insecticide susceptibility tests, a single specimen of *X. cheopis* (♀♀) was observed which has the normal typical spermatheca and another spermatheca-like structure (Fig. 2). The normal spermatheca has all the characteristics of a typical specimen: (1) the tail is much longer and at the base it is slightly wider than at the head; (2) the terminal part of the tail is longer and narrows gradually; and (3) the lower margin of the head and tail-end are at about the same level (Fig. 2, A). The additional spermatheca-like structure (the tail-end of which is just close to the terminal part of the tail of the typical form (Fig. 2, B) is pear-shaped and has a small blunt tail, but no distinct head portion.

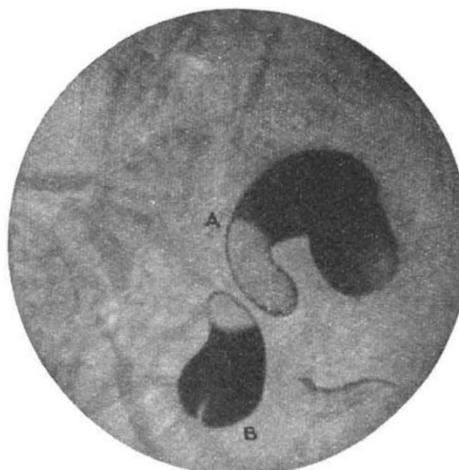


Fig. 2. ( $\times 132$ )

There has been no previous record in the literature, so far as we know, of the occurrence of a similar abnormality in *X. cheopis* (♀♀).

M. I. D. SHARMA  
G. C. JOSHI

Malaria Institute of India,  
Delhi.

- <sup>1</sup> Jordan, K., *A Handbook for the Identification of Insects of Medical Importance* by Smart, J. (printed by order of the Trustees of the British Museum, 1956).  
<sup>2</sup> Pollitzer, R., *Plague*. World Health Organization, Monograph Series No. 22 (World Health Organization, Palais des Nations, Geneva, 1954).  
<sup>3</sup> Roy, D. N., and Brown, A. W. A., *Entomology (Medical and Veterinary)* (Excelsior Press, Calcutta 12, 1954).

## MICROBIOLOGY

### Role of the Pentose Phosphate Pathway in *Pasteurella multocida*

It is acknowledged that in a large number of bacteria the main path for carbohydrate oxidation is the pentose phosphate cycle or other metabolic sequences not involving the Embden-Meyerhof pathway<sup>1</sup>. In the genus *Pasteurella*, work on *P. pestis* has shown that growing cells oxidize glucose through the pentose phosphate pathway<sup>2</sup>, but resting cells oxidize glucose through the combined action of glycolysis and the citric acid cycle<sup>3</sup>. In contrast with this metabolic duality, in *P. multocida* the pentose phosphate cycle is the main path for carbohydrate oxidation, both in growing and resting cells, and in regard to the citric acid cycle its principal role seems to be to supply carbon skeletons for the synthesis of amino-acids. These conclusions are borne out by experiments carried out with *P. multocida* (strain Beaufort No. 28 from the Institut Pasteur cultured aerobically for 4 hr. (growing cells) or 24 hr. (resting cells)) at 37° and pH 7.2 in a medium made of meat extract, 5 gm.; yeast extract (Difco), 1 gm.; peptone, 30 gm.; sodium chloride, 5 gm.; glucose, 20 gm.; agar, 25 gm. and water up to 1 litre. Our observations can be summarized as follows.

(a) Resting *P. multocida* oxidizes glucose, fructose, gluconate and ribose. The  $Q_{O_2}$  values ( $\mu$ l. oxygen consumption/mgm. dried cells/hr.) of the cells incubated with 5 mM substrate, in mM phosphate buffer pH 7.2 at 37°, were 24 with glucose or fructose, 18 with gluconate and 12 with ribose (control, 2.3-2.6).