

to push the mucous film forward against its tendency to escape with the exhaled stream of water. Small particles and unicellular algæ have been found associated with the micro-gillrakers, although not in the quantities one might expect. The final collection and concentration of food is probably achieved by the pharyngeal apparatus⁴.

I have found micro-gillrakers in *T. nigra*, *T. mossambica*, *T. esculenta*, *T. variabilis* Blgr., *T. zillii* Gervais, and *T. melanopleura*. The shape of the micro-rakers and the teeth is identical in all these species, but in *T. variabilis*, *T. zillii* and *T. melanopleura* there are usually more teeth (10-16; Goose records 18 in *T. melanopleura* and *T. nilotica*). Adult *T. zillii* and *T. melanopleura* feed mainly on higher plants, but phytoplankton has been recorded as a subsidiary food in the latter³. Micro-rakers were not found in *T. grahami*, Blgr., a small species confined to the strongly alkaline waters of Lake Magadi, Kenya. They are however present in at least one species of *Haplochromis*, *H. bloyeti* (Sauvage)—again identical to those of *T. nigra*. In the cyprinid *Labeo cylindricus* Peters, an algal grazer, micro-rakers are absent. In this fish the gillrakers are very close set (about 60 on the lower part of the anterior arch) and probably provide a sufficiently fine filtration surface.

Micro-gillrakers may hold some systematic value (*vide* the greater numbers of teeth in certain species). They may however be of more importance at generic levels, particularly where specialization has been towards a piscivorous or molluscivorous habit. A rather simpler type of micro-raker is found in species of *Citharinus* (Characidae), another filter feeder⁵.

PETER J. WHITEHEAD

Ministry of Forest Development,
Game and Fisheries,
Nairobi, Kenya.

¹ Fish, G. R., *Nature*, **167**, 900 (1951).

² Vaas, K. F., and Hofstede, A. E., *Contr. Int. Fish Res. Sta. Bogor*, **1**, 1 (1952).

³ Fish, G. R., *Uganda J.*, **19** (1), 85 (1955).

⁴ Greenwood, P. H., *Nature*, **172**, 207 (1953).

⁵ Gosse, J.-P., *Soc. Roy. Zool. Belge*, **86**, 303 (1955).

Infection of Cattle with *Cysticercus bovis* by the Injection of Onchospheres

In 1907 Dévé¹ reported the successful infection of a rabbit by the subcutaneous injection of *Echinococcus granulosus* eggs which produced a single fertile hydatid cyst. Leonard and Leonard², studying the resistance of rabbits to the larvae of *Tenia pisiformis*, infected these animals by the injection of hatched eggs into the mesenteric vein. Fedeli Avanzi³ reported that the intraperitoneal injection of hydatid sand into rabbits failed to produce any infection, but Batham⁴ successfully infected mice by the intraperitoneal injection of *E. granulosus* eggs.

In the annual report of the Kenya Veterinary Department for 1944⁵ Mann is reported as having hatched *T. saginata* eggs in acid pepsin and pancreatin solutions, but the onchospheres failed to infect calves when injected intravenously or subcutaneously.

It has been found by Froyd⁶ that adult cattle in Kenya are apparently resistant to infection with *T. saginata* eggs when these are given *per os*. A similar experience has been reported by Urquhart⁷. In order to discover whether adults were totally resistant to infection, parenteral routes were tried, since work on other parasites had shown that there is probably more than one phase of resistance to parasitic infection^{2, 8, 9}.

Table 1. RESULTS OF INJECTION OF HATCHED *T. saginata* ONCHOSPHERES IN ADULT CATTLE

Route of Infection	Artificial Infection		Result doubtful
	Successful	Unsuccessful	
Naturally preinfected cattle			
Intravenous	0	2	—
Subcutaneous	1	2	—
Subcutaneous and intramuscular	3	2	—
Cattle not preinfected			
Intravenous	0	0	2, of which 1 died 11 days after injection
Subcutaneous	1	1	—
Subcutaneous and intramuscular	5	5	—
Total	10	12	2

To date, 24 adult cattle have been injected intravenously, subcutaneously and/or intramuscularly with onchospheres of *T. saginata*, hatched by Silverman's technique¹⁰. Some of these beasts were found to be naturally preinfected by *C. bovis*.

The results of these injections are shown in Table 1.

Thus in ten animals cysts were found at the injection sites. One other, doubtful, beast injected intravenously had two small cysts in the intramuscular fascia of a hindleg, but their histological appearance was not sufficiently conclusive to allow a definite diagnosis of *C. bovis* infection.

In two cases viable cysts were demonstrated at the injection site. In several others macroscopical evidence of cysts was noted, proved histologically to be indistinguishable from cestode tissues and in some cases showing scolices, thus leading to a definite diagnosis of successful parenteral infection.

Batches of eggs vary in the viability of the onchospheres which they produce. Seven of the negative cases above belonged to one batch which, though infective for calves, failed to infect any adult bovine. Excluding that group our series gave 58.8 per cent infection instead of 41.7 per cent all inclusive.

A fuller report on the above experiments will be published elsewhere.

This letter is published with the permission of the Director of Veterinary Services, Kenya.

G. FROYD
M. C. ROUND

Veterinary Research Laboratory,
Kabete,
Kenya.
July 1.

¹ Dévé, F., *C. R. Soc. Biol.*, **63**, 332 (1907).

² Leonard, A. B., and Leonard, A. E., *J. Parasit.*, **27**, 375 (1941).

³ Fedeli Avanzi, C., *Ann. Fac. Med. vet. Pisa*, **5**, 152 (1952); abstract in *Helminth. Abstr.*, **21**, 235 (1952).

⁴ Batham, E. J., *N.Z. Vet. J.*, **5**, 74 (1957).

⁵ Kenya Vet. Dep. Ann. Rep. 1944, 15 (1946).

⁶ Froyd, G. (unpublished results, 1958).

⁷ Urquhart, G. M. (private communication, 1959).

⁸ Chandler, A. C., *Amer. J. Hyg.*, **22**, 157 (1935).

⁹ Sarles, M. P., and Taliaferro, W. H., *J. Infect. Dis.*, **59**, 207 (1935).

¹⁰ Silverman, P. H., *Ann. Trop. Med. Parasit.*, **48**, 207 (1954).

Conjugation in *Spirostomum*

ANY report of conjugation in *Spirostomum* is of special interest because of its extreme rarity. To our knowledge the last report was by Bishop³ nearly thirty-six years ago in *S. ambiguum*. Earlier, Stein¹ and Balbiani² had observed the process in the same species. In no case was it noticed as a regular phenomenon (as in *Paramecium* and other ciliates) affecting considerable numbers of animals in a population. Nor were any of these authors able to present a full account of the conjugation process, since it was encountered by them very infrequently. As Bishop says, 'There