

A small sample of the amine is allowed to react with 1:2:4-fluorodinitrobenzene and the dinitrophenyl-derivative is identified by reversed-phase chromatography on paper. In a typical experiment, fluorodinitrobenzene (0.15 μ l.) in 0.2 ml. of ethanol was added to 10 μ gm. of the amine or its hydrochloride in 0.1 ml. water, followed by 0.2 *N* sodium bicarbonate solution (0.1 ml.). After standing for 15 min. at room temperature, the solution was placed at 105° in a sealed tube for 2 hr. Excess fluorodinitrobenzene was removed by addition of a solution of glycine (0.1 ml. containing 300 μ gm.) and allowing the mixture to stand for 2 hr. at room temperature⁶. Ethanol was then removed in a vacuum desiccator, 0.15 ml. of 0.2 *N* sodium bicarbonate solution added and the dinitrophenyl-amines extracted into ether. The residue obtained on evaporation of the ether solution was dissolved in 5 μ l. ethanol and 1.7 μ l. of the solution applied to a paper chromatogram. Whatman No. 1 paper treated with Dow Corning silicone No. 1107⁷ was used and the chromatograms were developed with the more polar phase obtained on mixing methanol, chloroform and water (10:10:6 by volume). The chromatograms were run as recommended by Kritchewsky and Tiselius⁷ and allowed to develop for 16 hr., when the solvent front had run off the bottom of the sheet (35 cm. long). 2:4-Dinitroaniline (R_F , 0.60) had then travelled 26.25 cm. The values of R_A (defined as the ratio of the distance moved by the substance to the distance moved by 2:4-dinitroaniline) and of R_F for the dinitrophenyl-derivatives of the amines used are shown in Table 1.

Table 1. BEHAVIOUR OF DINITROPHENYL-AMINES ON PAPER CHROMATOGRAMS

$$R_A = \frac{\text{Distance moved by substance}}{\text{Distance moved by 2:4-dinitroaniline}}$$

Dinitrophenyl-derivative of	R_A	R_F
Ammonia	1.00	0.60
Methylamine	0.88	0.53
Dimethylamine	0.80	0.48
Ethylamine	0.67	0.40
isoPropylamine	0.47	0.28
Diethylamine	0.44	0.26
isoButylamine	0.33	0.20
isoAmylamine	0.20	0.12
isoHexylamine	0.12	0.07

By plotting the values of $R_M \left[= \log_{10} \left(\frac{1}{R_F} - 1 \right) \right]$

for the dinitrophenyl-derivatives of the primary and secondary amines against the number of methylene groups in the amines, two parallel straight lines are obtained for the two homologous series (cf. Bate-Smith and Westall⁸).

With the small spots applied to the chromatograms, all the dinitrophenyl-amines listed are separated from each other except the derivatives of isopropylamine and diethylamine. These two substances can be separated by prolonged development of the chromatogram. The spots can easily be eluted from the appropriate areas of the chromatograms with 3 ml. of ethanol. Measurement of the optical density of the resulting solutions at 345 $m\mu$ shows that the primary and secondary amines used have reacted almost quantitatively with 1:2:4-fluorodinitrobenzene. Ammonia gives only about 50 per cent of the expected yield and the high temperature used for the reaction is required to obtain this yield. The low solubility of the dinitrophenyl-derivative of methylamine in ethanol may also result in low recoveries in certain cases. Where an elongated spot due to

dinitrophenyl-methylamine will not interfere with other substances, the spots may be applied to the chromatogram in ethanol-acetone (1:1) instead of ethanol. The theoretical recovery of dinitrophenyl-methylamine is then obtained. The method therefore appears to have potentialities as a semi-quantitative tool.

Since the dinitrophenyl derivatives of the primary aliphatic amines show an absorption maximum at 345 $m\mu$, while the corresponding derivatives of secondary amines show a maximum at 365–370 $m\mu$, the measurement of the optical density of an eluate at two wave-lengths provides corroborative evidence as to whether the substance is derived from a primary or secondary amine.

I. M. LOCKHART

Sir William Dunn School of Pathology,
University of Oxford.

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- ¹ Sanger, F., and Thompson, E. O. P., *Biochem. J.*, **53**, 353 (1953).
² Lockhart, I. M., Abraham, E. P., and Newton, G. G. F., *Biochem. J.*, **61**, 534 (1955).
³ Bremner, J. M., and Kenten, R. H., *Biochem. J.*, **49**, 651 (1951).
⁴ Dougall, D., and Abraham, E. P., *Nature*, **176**, 256 (1955).
⁵ Schwyzer, R., *Acta Chem. Scand.*, **6**, 219 (1952).
⁶ Grassmann, W., Hormann, H., and Endres, H., *Ber. Dtsch. Chem. Ges.*, **86**, 1477 (1953).
⁷ Kritchewsky, T. H., and Tiselius, A., *Science*, **114**, 299 (1951).
⁸ Bate-Smith, E. C., and Westall, R. G., *Biochim. Biophys. Acta*, **4**, 427 (1950).

Erosion in Meanders

IN connexion with the communication by J. H. Horlock in *Nature* of November 26, p. 1034, I am writing to urge that the term 'meander' should be strictly limited to natural meanders, not to bends due to secondary flows. The term 'meander' is derived from the River Meander, which swings from bank to bank in an almost perfect pattern. The Central Board of Irrigation, India¹, many years ago defined "a meandering river as a river which follows a sinuous path due to natural physical causes, not imposed by external restraint". The experiments quoted were carried out under very unnatural conditions and hence produced highly unnatural results.

C. C. INGLIS
(Director)

Hydraulics Research Station,
Howbery Park,
Wallingford, Berks.

¹ See "The Behaviour and Control of Rivers and Canals (with the aid of Models)"; obtainable from Publications Branch, General Department, India House, Aldwych, W.C.2. 13s.

I AM grateful to Sir Claude Inglis for pointing out that the term 'meander' should be restricted to bends similar to natural meanders. I agree that the initial shapes of the bends used in the experiments described in my communication were not true meanders; but the point of the experiment was to demonstrate that the nature of the erosion in such bends would depend largely on the initial curvature at entry to the bend. Such artificial bends probably develop into natural meanders. Subsequent experiments on natural meanders of similar amplitude and wave-length show that erosion occurs on the banks on the outside of the bends, and not on one bank, as in the original experiments with artificial bends.

J. H. HORLOCK

Engineering Laboratory,
Cambridge.