

Cortisone and Hydrocortisone in Cerebrospinal Fluid

ALTHOUGH numerous measurements have been made of the concentration of adrenocortical steroids in various body fluids, particularly since the development of paper chromatographic methods, no data are available concerning cerebrospinal fluid.

Cerebrospinal fluid was obtained by lumbar puncture from patients in a neurological unit in the course of air encephalographic studies. Each sample was extracted within twenty-four hours of withdrawal by a modification of the method of Nelson and Samuels¹. The sample was shaken three times with 1.5 volumes of chloroform. The combined chloroform extracts were taken to dryness. The residue was dissolved in 50 ml. of 70 per cent ethanol, and washed three times with 25 ml. of hexane. The ethanolic solution was taken to dryness, and the residue dissolved in 0.1 ml. of ethanol for chromatography. Paper chromatography was carried out using the benzene-formamide system of Zaffaroni, Burton and Keutmann², using appropriate standards. The chromatograms were run for 60 hr. Provisional identification and semi-quantitative comparison with standards were based on: (1) running properties of unknown compounds, compared with standards added to cerebrospinal fluid and extracted in a similar manner; added cortisone and hydrocortisone run more rapidly in extracts of cerebrospinal fluid than in the standard solutions in ethanol; (2) ultra-violet absorption³ at 254 m μ ; (3) reaction with a tetrazolium salt⁴—blue tetrazolium was used; (4) soda fluorescence⁵.

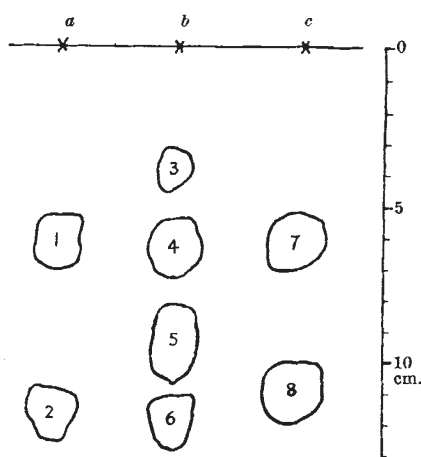
Recoveries of cortisone and hydrocortisone added to cerebrospinal fluid exceeded 75 per cent. There was no conversion of either compound to the other during extraction.

Examination of seven single samples of cerebrospinal fluid (each of 50–100 ml.) revealed in five samples doubtful traces of substances exhibiting soda fluorescence, and tentatively identified as cortisone and hydrocortisone by their running properties. As well as the substances exhibiting soda fluorescence, a spot (R_F value relative to cortisone = 0.85) was occasionally found, detectable by ultra-violet absorption only.

Two pooled extracts, each representing 500 ml. of cerebrospinal fluid, were now run. Both showed the same pattern (see diagram). Substances provisionally identified as cortisone and hydrocortisone were found in concentrations of 0.1–0.2 μ gm. and 0.2–0.4 μ gm. per 100 ml. respectively. Absolute identification by means of crystallization and melting-point determinations was not carried out because of the small amounts detected. For the same reason, running properties of the acetyl derivatives and the infra-red and ultra-violet absorption spectra were not investigated.

The provisional identification of cortisone in cerebrospinal fluid is of importance in view of the failure of some workers to find cortisone in normal human plasma^{1,6}. Morris and Williams⁷, on the other hand, identified both cortisone and hydrocortisone (in addition to other steroids) in human plasma, in the same ratio (1:2) as we have found in cerebrospinal fluid, although the concentrations were twenty-five times greater in plasma.

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Tracing of soda fluorescence on a chromatogram run for 60 hr. at room temperature in the benzene-formamide system. (a) Extract of standards added to 50 ml. cerebrospinal fluid; (1) 2 μ gm. hydrocortisone; (2) 2 μ gm. cortisone. (b) Pooled extract from 500 ml. cerebrospinal fluid: (3) yellow fluorescence, indicating trace of unidentified substance; R_F value relative to cortisone, 0.3; (4) hydrocortisone (1.0–2.0 μ gm.); (5) blue fluorescence, with similar running properties to substance X (see ref. 5); (6) cortisone (0.5–1.0 μ gm.). (c) Standards in ethanol solution: (7) 2 μ gm. hydrocortisone; (8) 2 μ gm. cortisone.

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¹ Nelson, D. H., and Samuels, L. T., *J. Clin. Endocrinol.*, **12**, 519 (1952).

² Zaffaroni, A., Burton, R. B. and Keutmann, E. H., *Science*, **111**, 6 (1950).

³ Haines, W., and Drake, M. A., *Fed. Proc.*, **9**, 180 (1950).

⁴ Burton, R. B., Zaffaroni, A., and Keutmann, E. H., *J. Biol. Chem.*, **188**, 763 (1951).

⁵ Bush, I. E., *Biochem. J.*, **50**, 370 (1952).

⁶ Bayliss, R. I. S., and Steinbeck, A. W., *Biochem. J.*, **54**, 523 (1953).

⁷ Morris, C. J. O. R., and Williams, D. C., *Biochem. J.*, **54**, 470 (1953).

Canine Distemper Virus Complex

Plurality of the Virus. Some cross-immunity between canine distemper and human influenza viruses¹ has been an intriguing problem for some time, and the striking immunological relationship between canine distemper and poliomyelitis has also been reported². It was thought that canine distemper virus represented one etiological entity³ *Tarpeia canis*⁴, which is usually referred to as Carre or Laidlaw–Dunkin virus. It has been suggested that the disease commonly referred to as ‘hard pad’ is produced by a different virus⁵. This was questioned^{6–8}, because ‘hard pad’ virus in the hands of other workers was found to be immunologically indistinguishable from Carre–Laidlaw–Dunkin virus^{7,8}, although cause of infection differed in the ferret⁷; it is noteworthy that after as few as five consecutive passages a