

Group	Riboflavin in diet ( $\mu\text{gm./gm.}$ )	Serum riboflavin ( $\mu\text{gm./ml.}$ ) $\pm s$	Riboflavin in eggs ( $\mu\text{gm./gm.}$ ) $\pm s$
1	1.56	0.285 $\pm$ 0.03	2.31 $\pm$ 0.13
2	3.60	0.778 $\pm$ 0.07	4.18 $\pm$ 0.22
3	5.55	0.880 $\pm$ 0.07	4.92 $\pm$ 0.22

Fluorimetric assays were carried out on the diets, blood sera and eggs by the method of Kodicek and Wang<sup>2</sup>. The accompanying table shows the results obtained.

It will be seen from these results that the serum riboflavin is substantially increased by supplementing the diet and that the blood-levels are directly related to egg content. An increase in the diet above 3.6  $\mu\text{gm./gm.}$  did not result in a significant increase in serum level; this would appear to be in accordance with the observation of Petersen *et al.*<sup>3</sup> that further increase in hatchability from the inclusion of riboflavin in the diet is not attained beyond this dietary level for battery-kept birds.

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<sup>1</sup> Lepovsky, S., Taylor, L. W., Jukes, T. H., and Almquist, H. J., *Hilgardia*, 11, 559 (1938).

<sup>2</sup> Engel, R. W., Phillips, P. H., and Halpin, J. G., *Poult. Sci.*, 19, 135 (1940).

<sup>3</sup> Petersen, C. F., Lampman, C. E., and Stamborg, O. E., *Poult. Sci.*, 28, 187 (1947).

<sup>4</sup> Common, R. H., Rutledge, W. A., and Bolton, W., *J. Endocrinol.*, 5, 121 (1947).

<sup>5</sup> Kodicek, E., and Wang, Y. L., *Biochem. J.*, 44, 340 (1949).

### Production of Specific Group N Precipitating Sera for *Streptococcus cremoris* by 'Absorption' of Non-specific Antigens

DUE in part to the difficulty of preparing a specific group N precipitating serum for *Streptococcus cremoris*, the boundaries and constituents of serological group N have been so far incompletely determined. Although, by means of serological methods using interfacial precipitin tests with hydrochloric acid extracts, *Str. cremoris* has been grouped with *Streptococcus lactis* by Shattock and Mattick<sup>1</sup>, and with *Str. lactis* and with *Streptococcus diacetylactis* by Swartling<sup>2</sup>, final confirmation of its serological identity with group N must depend on the preparation of a specific group serum for *Str. cremoris* itself.

Early attempts in this laboratory to produce such a serum in rabbits, using as vaccines living and variously heat- and chemically-treated bacterial cells, yielded only non-specific products, cross-reactions occurring most commonly and intensively with hydrochloric acid extracts of the streptococci of groups K and L. Absorption of these non-specific sera with suspensions of K and L streptococci completely removed the non-specific antibodies and only partially impaired the group N reactions. In order to produce a powerful serum specific in the first instance, it was clearly necessary to inactivate the non-specific antigens in the *cremoris* cells constituting the vaccine before the immunization process was begun. By 'absorbing' *Str. cremoris* cells with sera of groups K and L, these antigens were eliminated or rendered non-antigenic, and specific group N sera were obtained forthwith. Such sera have been prepared

for both freeze-dried and freshly isolated strains of *Str. cremoris*.

The details of this work will be published elsewhere.

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<sup>1</sup> Shattock, P. M. F., and Mattick, A. T. R., *J. Hyg. Camb.*, 43, 173 (1943).

<sup>2</sup> Swartling, P. F., *J. Dairy Res.*, 18, 256 (1951).

### Grana-like Structures of *Synechococcus cedorum*

DURING photosynthetic studies in this laboratory, the desire for an organism possessing a simple internal structure prompted us to investigate the blue-green alga. It has occasionally been reported that these organisms contain grana or chloroplasts; but the most widely accepted opinion is that all their pigments are uniformly distributed throughout the cytoplasm<sup>1</sup>.

A pure culture of a unicellular blue-green alga, *Synechococcus cedorum*, was grown in an inorganic medium, and one-day old cells were used for investigating the pigment distribution. After the cells were harvested by centrifuging and washed with water, they were broken by grinding with alumina<sup>2</sup>. The mixture was diluted with water and centrifuged at 2,000 g for 10 min. to remove unbroken cells, cell debris and alumina. A blue-green supernatant having a strong Tyndall effect and slight fluorescence was obtained. This supernatant was then centrifuged in a refrigerated Spinco ultracentrifuge for 30 min. at 36,000 g. A clear blue supernatant above a very minute green sediment was obtained. The absorption spectra of the intact cells and of the several fractions were determined from 3500 A. to 7500 A. with a Cary recording spectrophotometer. The curves of the aqueous solutions or suspensions are shown in Fig. 1.

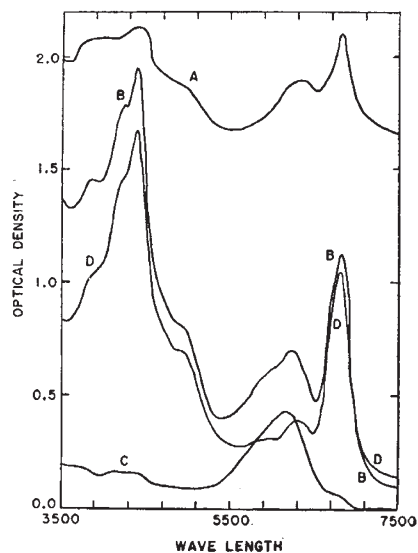


Fig. 1. Absorption spectra of *Synechococcus cedorum*. (A) Whole cells; (B) supernatant (2,000 g) from alumina-treated cells; (C) supernatant (36,000 g) after ultracentrifugation; (D) re-suspended particles obtained from ultracentrifugation