Curve A: $(\text{Ph}_3\text{MeAs})_3(\text{Fe}(\text{CNS})_6)_3$ in *o*-dichlorobenzene ($4.0 \mu\text{gm. Fe/ml.}$) + 70 equivalents of $\text{Ph}_3\text{MeAsCNS}$. 0.5 cm. cell.
 Curve B: $(\text{Ph}_3\text{MeAs})_3(\text{Fe}(\text{CNS})_6)_3$ in *o*-dichlorobenzene ($4.0 \mu\text{gm. Fe/ml.}$). 0.5 cm. cell.
 Curve C: $(\text{Ph}_3\text{MeAs})_3(\text{Fe}(\text{CNS})_6)_3$ in *o*-dichlorobenzene ($0.5 \mu\text{gm. Fe/ml.}$). 4.0 cm. cell.

plex anions such as FeCl_4^- , BiI_4^- , $\text{Co}(\text{CNS})_4^-$, and dissociation of the above type is probably a general phenomenon.

The dissociation of complex salts in water, as pointed out by Bjerrum³, is complicated by the competition of water molecules for co-ordination positions of the metal, so that complexity constants are not measures of the affinity of the metal for a particular ligand, but of the change in free energy when a solvent molecule is replaced by the ligand. The determination of dissociation constants in solvents such as *o*-dichlorobenzene or carbon tetrachloride should give a very close evaluation of the true affinities of metals and ligands, as well as the energy of dissociation, since these solvent molecules can be attached by only the weakest of bonds.

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Identity of Lespedin with Kämpferitrin

FROM the leaves of *Lespedeza cyrtobotrya* Miquel, a deciduous shrub of the natural family Leguminosæ, M. Hasegawa and I isolated a kämpferol glycoside which we named lespedin^{1,2}, its structure being shown to be kämpferol 3,7-dirhamnoside. It has also been isolated from *Celastrus orbiculatus* Thunberg (family Celastraceæ) by M. Kanao and M. Shimokoriyama³.

When crystallized from water, lespedin had melting point 195° with or without water of crystallization ($3\frac{1}{2}\text{H}_2\text{O}$), whereas from ethanol it melted at 234° . It thus appeared to be different from kämpferitrin, melting point $201\text{--}203^\circ$ ⁴.

Having learnt from Dr. E. C. Bate-Smith that the kämpferitrin employed in his chromatographic studies was obtained from the original collection of the late Prof. A. G. Perkin (through the courtesy of Dr. E. J. Cross of the Department of Colour Chemistry, University of Leeds), I sent a specimen of lespedin to Dr. Bate-Smith in order to have the mixed melting point of the two glycosides tested. The results he has communicated to me are as follows:

	Softening point	Melting point
Kämpferitrin	$190\text{--}192^\circ$	194°
Mixture	$190\text{--}194^\circ$	195°
Lespedin	$190\text{--}194^\circ$	195°

R_F values in butanol-acetic acid-water (4:1:5) and phenol-water are for both 0.71. The colours given in ultra-violet light and in ammonia are also identical.

I am satisfied that the identity of lespedin with kämpferitrin has been thus established, and the name 'lespedin' becomes, therefore, redundant.

I wish to thank Dr. T. Swain, who carried out the determinations, and Dr. E. C. Bate-Smith, both of the Low Temperature Research Station, Cambridge, for their kindness.

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Cholinesterase Activity in the Secretions of the Pancreas of the Dog and Parotid of the Pig

It has been recognized for some time that the pancreas of the dog contains large amounts of pseudo-cholinesterase¹, and the parotid gland of the pig is known to have a much higher cholinesterase activity than other digestive glands in this species². Ginsberg *et al.*³, using a biological method, found that dog's pancreatic juice possessed at least fifteen times the enzyme activity of the blood, but this observation does not so far seem to have been confirmed.

In view of this work and other experiments which will be reported separately, dog's pancreatic juice, secreted in response to different stimuli, was examined for the presence of pseudo-cholinesterase. The animals were anaesthetized with chloralose before cannulation of the accessory pancreatic duct. Extracts of the mucosa of the small intestine and pilocarpine were injected intravenously to stimulate secretion. Pseudo-cholinesterase in the juice and in water homogenates of the gland was estimated manometrically at pH 7.4 with 0.06 M acetylcholine perchlorate (B.D.H.) as the substrate. The unit of cholinesterase is defined as the amount of enzyme which liberated $1 \mu\text{l.}$ carbon dioxide/min.