difficult to decide which suprarenals are in the more physiological condition.

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Physiological Laboratory. Cambridge. Jan. 16.

¹ von Euler, U. S., Ergeb. Physiol., 46, 261 (1950).

² Holton, P., Nature, 163, 217 (1949).
⁸ Holton, P., J. Physiol., 108, 525 (1949).
⁴ von Euler, U. S., Nature, 163, 642 (1949).

Zinc Content of the Genital Organs of the Rat

IN a recent review, Vallee and Altschule¹ have suggested that though many workers have estimated zinc in living tissues, the methods used have often been inaccurate. We have estimated the zinc content of certain tissues of the Collip hooded rat by the dithizone method of Vallee and Gibson². Mature animals were used which were fed on a diet of fox chow containing about 75 $\mu gm.$ zinc per gm., supplemented with carrot and cabbage. Most tissues contained about 15-30 µgm. zinc per gm.; but certain male genital organs showed wide deviations from this range. The literature contains few references to the zinc content of the genital system, though Bertrand and Vladesco³ reported high results in herring's and horse's testis, pig's seminal vesicle, rat's epididymis and in the prostate of the bull and of man. Kadota⁴ mentioned briefly that zinc could be detected by a relatively insensitive histochemical method in the rabbit prostate. The figures given by Leiner and Leiner⁵ for rat's testis and by Eggleton⁶ for human testis were within the usual range for soft mammalian tissues.

The accompanying table shows the results we have obtained with the genital organs of the rat, compared with a selected group of other tissues of the rat. The amount of zinc in the seminal vesicle contents was at the lower limit of accuracy of the method of estimation, and some of the zinc found may have been due

ZINC CONTENT OF ORGANS OF THE RAT

Organ	No. of samples	Zinc content of or standard de (µgm./gm. wet weight)	
Seminal vesicle contents Seminal vesicle tissue Posterior prostate Ventral prostate Epididymis Coagulating gland Testis	3 5 11 9 8 6 7	$\begin{array}{rrrrr} 2\cdot15\pm & 0\cdot48\\ 22\cdot1\pm & 2\cdot64\\ 180\cdot0\pm & 45\cdot5\\ 18\cdot7\pm & 3\cdot24\\ 46\cdot2\pm & 4\cdot60\\ 25\cdot0\pm & 8\cdot47\\ 28\cdot9\pm & 1\cdot84\end{array}$	$\begin{array}{c} 0.42 \pm 0.09 \\ 1.98 \pm 0.30 \\ 15.2 \pm 3.62 \\ 1.49 \pm 0.43 \\ 4.36 \pm 0.50 \\ 2.49 \pm 0.81 \\ 2.68 \pm 0.23 \end{array}$
Uterus Ovaries	6 5	$ \begin{array}{r} 14 \cdot 4 \ \pm \ 3 \cdot 23 \\ 20 \cdot 3 \ \pm \ 1 \cdot 49 \end{array} $	$ \begin{array}{r} 1 \cdot 47 \pm 0 \cdot 33 \\ 1 \cdot 51 \pm 0 \cdot 23 \end{array} $
Brain Liver Lung Submaxillary	5 5 8	$\begin{array}{r} 14.8 \pm 1.32 \\ 30.3 \pm 1.57 \\ 18.7 \pm 1.61 \end{array}$	$\begin{array}{c} 1 \cdot 06 \pm 0 \cdot 08 \\ 2 \cdot 23 \pm 0 \cdot 12 \\ 1 \cdot 59 \pm 0 \cdot 16 \end{array}$
gland Pancreas Duodenum Diaphragm Tibia	5 6 5 5 5	$\begin{array}{r} 16.5 \pm 0.96 \\ 23.3 \pm 1.01 \\ 23.9 \pm 1.10 \\ 24.9 \pm 2.51 \\ 233.0 \pm 22.0 \end{array}$	$\begin{array}{c} 1 \cdot 17 \pm 0 \cdot 06 \\ 1 \cdot 28 \pm 0 \cdot 16 \\ 1 \cdot 93 \pm 0 \cdot 28 \\ 2 \cdot 17 \pm 0 \cdot 08 \\ 0 \cdot 39 \pm 0 \cdot 05 \end{array}$

to contamination by vesicular tissue juice. There is no doubt, however, that this material contained much less zinc than any other tissue examined. The posterior prostate, on the other hand, contained much more zinc than any other tissue except bone, and, when results were calculated as zinc per gm. ash, the posterior prostate was by far the richest source of zinc in the body. The epididymis also contained a considerable amount of zinc; but the female genital organs gave no striking results.

It is noteworthy that the ventral prostate of the rat contained comparatively little zinc. The fact that the ventral and posterior prostates are very different tissues does not appear to be widely recognized. though they differ not only in gross and histological appearance but also in citric acid and fructose content⁷ and in acid and alkaline phosphatase activity⁸.

By the courtesy of Prof. Lyman Duff, of Montreal, and Prof. W. L. Robinson, of Toronto, we have been able to examine a number of specimens of human prostate taken at operation and at autopsy. The results, which will be reported elsewhere, were very variable; but most of the apparently normal glandular tissue contained more than 100 µgm. zinc per gm., and one value of $307 \ \mu gm$. per gm. was observed. Four rabbit prostates contained 173, 233, 270 and 358 µgm. zinc per gm. It seems likely, therefore, that an exceptionally high zinc content is typical of the prostate gland.

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Atomic Energy Project, Chalk River, Ontario. Dec. 21.

- Vallee, B. L., and Altschule, M. D., Physiol. Rev., 29, 370 (1949).
 Vallee, B. L., and Gibson, J. G., J. Biol. Chem., 176, 435 (1948).
 Bertrand, G., and Vladesco, R., C.R. Acad. Sci., Paris, 173, 176 (1921).
- ⁴ Kadota, I., J. Lab. Clin. Med., 35, 568 (1950).

- ⁶ Kanota, I., J. Lab. Clin. Meta., 35, 508 (1907).
 ⁵ Leiner, M., and Leiner, G., Naturwiss., 29, 763 (1941).
 ⁶ Eggleton, W. G. E., Biochem. J., 34, 991 (1940).
 ⁷ Humphrey, G. F., and Mann, T., Nature, 161, 352 (1948).
 ⁸ Mawson, C. A., and Clayton. B. P. (unpublished).

Detection of Enzymes by the Agar-Plate Method and its Application to Paper Chromatography

In previous publications¹⁻⁴ from this laboratory, the use of the agar-plate method for the detection of enzymes has been described. This technique has now been adapted to the location of enzymes on paper. Preliminary investigations have shown that enzymes can be readily identified on the chromatogram by means of this technique.

The filter paper chromatographic experiments were carried out on paper strips (Whatman No. 1, 40 cm. imes12 cm.) by a procedure which resembles in general that developed by Consden, Gordon and Martin⁵. The paper carrying the enzyme near its upper end is hung from a glass trough containing aqueous alcohol or aqueous acetone (50 per cent) as solvent, the whole system being kept in a rectangular-sided glass jar saturated with the vapour of the solvent. After a suitable time of running (5-18 hr.), the paper is removed and the solvent front marked; the paper is allowed to dry at room temperature. The paper chromatogram is then placed on the surface of a thin layer of agar (2 per cent) containing the substrate. After a few hours (2-8 hr.)