Tt

At present, only the racemic forms of laudanosine and of tetrahydropapaverine have been employed in preparing compound 20. Work on the resolution of laudanosine is now in progress, and in due course we hope to investigate the properties of the bis quaternary ammonium salts prepared from the stereoisomers.

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<sup>1</sup> Taylor, E. P., and Collier, H. O. J., Nature, 165, 602 (1950).

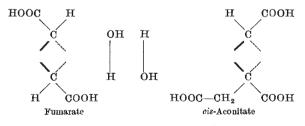
<sup>2</sup> Craig, L. E., and Tarbell, D. S., J. Amer. Chem. Soc., 70, 2783 (1948). <sup>3</sup> Paton, W. D. M., and Zaimis, E. J., Brit. J. Pharmacol., 4, 381 (1949).

## Specificity of the Enzyme Aconitase

BECAUSE the enzyme named 'aconitase' catalyses the conversion of cis-aconitate both to citrate and to isocitrate (and the reverse reactions), it has been thought<sup>2</sup> that it may be a mixture of two enzymes, though Buchanan and Anfinsen<sup>3</sup> found no evidence of this.

As in the case of hydration of fumarate to malate<sup>4</sup>, it is necessary to assume that H<sub>2</sub>O is added exclusively to one member of the double bond of cisaconitate, in order to explain the formation of optically active products. This necessitates, in both cases, the assumption of a 'three-point attachment' of the substrate to the enzyme<sup>5</sup>. The formation of citrate or isocitrate depends only on the direction of addition of H<sub>2</sub>O.

Now it is not necessary to assume that there is any preferred direction of addition of H<sub>2</sub>O to fumarate (relative to the asymmetric enzyme), since addition in either direction (see formulæ) yields the same



isomer of malate. This makes it likely that with cis-aconitate also there is no preferred direction and, thence, that citrate and isocitrate may be formed by the action of a single enzyme. It is of interest that Martius and Lynen<sup>6</sup> have shown that the initial rates of formation of citrate and isocitrate are approximately equal. The occurrence of the reverse reactions follows from the requirement of catalytic reversibility.

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- <sup>1</sup> Martius, C., and Knoop, F., Z. physiol. Chem., 246, 1 (1937). <sup>2</sup> Jacobsohn, K. P., and Tapadinhas, J., Enzymologia, 5, 388 (1938). <sup>3</sup> Buchanan, J. M., and Anfinsen, C. B., J. Biol. Chem., 180, 47 (1949).
- <sup>4</sup> Krebs, H. A., Harvey Lectures, Series xliv, 165 (1950).
- <sup>6</sup> Ogston, A. G., *Nature*, **162**, 963 (1948). <sup>6</sup> Martius, C., and Lynen, F., "Advances in Enzymology", **10**, 190 (1950).

## Formation of Hydroxylamine from Amino-Compounds in Áqueous Solution by Irradiation with X-Rays

In recent studies of the effects of ionizing radiations on aqueous solutions of compounds containing amino-groups such as, for example, amino-acids<sup>1</sup>, adenine<sup>2</sup>, nucleic acids<sup>2,3</sup>, it has been shown that, among other changes, ammonia is formed. In the case of the nucleic acids, for example, the ammonia arises from the constituent purine and pyrimidine bases as a result of a deamination process involving the radicals OH and HO2, which are formed in the primary radiation process according to<sup>4</sup>:

$$H_{0} \longrightarrow H + OH$$

which in the presence of oxygen is followed by :  $H + O_2 \rightarrow HO_2$ .

It may be expected that an attack by these radicals  
on the primary amino-groups 
$$(-NH_2)$$
 in the bases  
adenine, guanine and cytosine, and also in other  
amino-compounds, may, in addition, give rise to  
some hydroxylamine. By irradiation with X-rays

some vs of aqueous solutions of these bases, and also of their corresponding nucleosides and nucleotides, we have established that this process does, in fact, take place to some extent.

The presence of hydroxylamine was established by well-known colour reactions and also by a paperchromatographic method which was developed for this purpose. Quantitative determinations of the yields, for example, from adenine, show that doses of the order of 100,000 r. produce a concentration corresponding to about  $1 \times 10^{-6}$  moles/litre of hydroxylamine. However, only very small amounts are formed on irradiation of the nucleic acids. That the amino-group is the source of hydroxylamine is again demonstrated by the absence of this substance in irradiated solutions of, for example, hypoxanthine, uracil and uridine, in which cases no amino-groups are present.

Other compounds containing the primary aminogrouping have also been examined, and it has been found that hydroxylamine is produced in the irradiation of aqueous solutions of certain amino-acids (for example, alanine, lysine) and of certain amines (for example, ethylamine, histamine). In general, the yields of hydroxylamine depend markedly upon the pH of the irradiated solutions.

These observations may be of biological significance in view of the possible formation of hydroxylamine in vivo from amino-compounds present in tissues, as it is well known that hydroxylamine is a strong cell poison and a very effective catalase inhibitor.

On the other hand, we found that irradiation of ammonia does not yield hydroxylamine. Irradiation of ammonium salt solutions with X-rays results predominantly in an oxidation to nitrite, a reaction which depends markedly on the hydroxyl ion concontration and which is also enhanced by the presence of molecular oxygen. This oxidation process is of interest in view of the corresponding process occurring in plants and in nitrifying bacteria.

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King's College, University of Durham, Newcastle upon Tyne. Feb. 27.

- <sup>1</sup> Stein, G., and Weiss, J., Nature, 162, 814 (1948); J. Chem. Soc., 3256 (1949). Dale, W. M., and Davis, J. V., Nature, 163, 64 (1949). Dale, W. M., Davis, J. V., and Gilbert, C. W., Biochem. J., 45, 93 (1949).

- <sup>a</sup> Scholes, G., and Weiss, J., Nature, 166, 640 (1950).
  <sup>s</sup> Scholes, G., Stein, G., and Weiss, J., Nature, 164, 709 (1949).
  <sup>4</sup> Weiss, J., Nature, 153, 748 (1944); Trans. Farad. Soc., 43, 314 (1947).