

Configuration of the C/D Ring Junction in Equilenin and Other Steroids

THE configuration of the C/D ring fusion in equilenin has been the subject of a number of communications¹⁻³. We consider that the arguments advanced for a *trans* ring junction leave much to be desired. Shoppee's¹ conclusion that the dehydrogenation of equilin to equilenin under mild conditions proves the steric identity of the C/D union in these two substances can no longer be accepted, since *isoequilin*, which is now known^{4,5} to be also *trans*, gives *isoequilenin* (which has the opposite C/D configuration to equilenin) under the same conditions⁶.

It is evident that the same uncertainty is present in any correlation based on the formation of equilenin (or *isoequilenin*) by reactions involving a centre adjacent to C₁₄². One would anticipate epimerization at C₁₄ in some of these experiments because of the possibility offered of relieving the strain associated with the *trans* C/D hydrindane system.

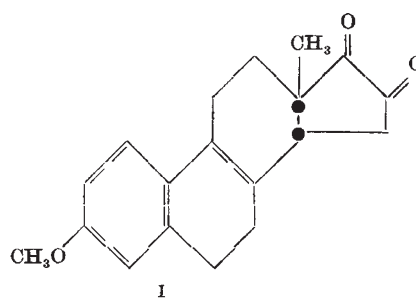
Another attempt to correlate the C/D configuration of equilenin with that of other natural steroids through a determination of the configuration at C₁₄ was based on the use of the method of molecular rotation differences³. Since this centre is adjacent to an unsaturated system which might give rise to considerable 'vicinal effect'⁷, the validity of this conclusion is questionable.

An unambiguous proof of the configuration of equilenin follows from the fact that Bachmann's⁸ α -7-methoxy-2-methyl-2-carboxy-1,2,3,4-tetrahydrophenanthrene-1-acetic acid (III) (which was converted to *isoequilenin*) is obtained⁵ by hydrogen peroxide cleavage of the α -diketone (I) to the diacid (II), followed by dehydrogenation. The diketone I is one of the products of the Diels-Alder reaction between 1-vinyl-6-methoxy-3,4-dihydronaphthalene and 3-methyl-cyclopentene-1,2-dione^{9,5}, and must therefore be *cis*.

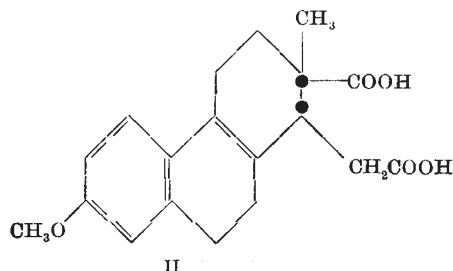
Dehydrogenation of II would not be expected to invert C₁ (C₁₄ sterol numbering), since the strain associated with a *trans* hydrindane system is no longer present. Dehydrogenation experiments in the doisylic acid series confirm this view¹⁰. The C/D ring junction is therefore *cis* in *isoequilenin* and *trans* in equilenin.

Compound II proved identical with a diacid prepared by Miescher *et al.*². This diacid, which is therefore unambiguously *cis*, gave after homologation and ring closure a substance which was *not dl-isoequilin*. *Isoequilin* is therefore *trans*, and its formation from equilin by acid isomerization cannot involve epimerization at C₁₄ (which would have produced a *cis* compound). This means that the C/D junction in *cestrone* must then also be *trans*, since it follows from the work of Pearlman and Wintersteiner¹¹ that this junction is the same as that of equilin.

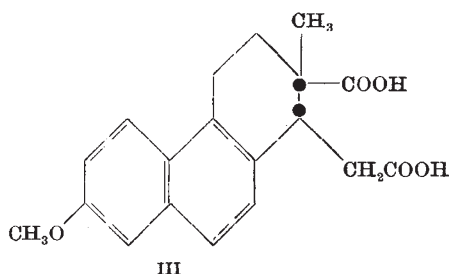
This derivation of the *trans* C/D junction in *cestrone* can be used as chemical evidence for the *trans* C/D union in the non-aromatic steroids, since cholesterol can be degraded to *cestrone* by a reaction which, although a pyrolysis, cannot involve C₁₄^{12,13}. This evidence is more convincing than that of Wieland and Dane¹⁴, which is open to serious criticism¹⁵, or that of Dimroth and Jonsson¹⁶, which logically demonstrates only the fact that vitamin D₂ has a C/D *trans* union, because the identity of the C/D junction in vitamin D₂ and ergosterol has not been convincingly demonstrated. It was possible that the *trans* C/D vitamin D₂ had resulted from the irradiation



I



II



III

of a *cis* C/D ergosterol. Such a change to the less stable configuration is known¹⁷ to take place when *isohydrocholesterol*, which is a C₅ α -compound, is transformed during irradiation to the C₅ β -configuration, which contains the less stable *cis*-decalin system.

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- ¹ Shoppee, *Nature*, **161**, 207 (1948).
- ² Rosenkranz, Djerassi, Kaufmann, Pataki and Romo, *Nature*, p. 814 of this issue.
- ³ Klyne, *Nature*, **161**, 434 (1948).
- ⁴ Heer, and Miescher, *Helv. Chim. Acta*, **31**, 1289 (1948).
- ⁵ Singh, G., *J. Amer. Chem. Soc.* (in the press).
- ⁶ Hirschmann, H., and Wintersteiner, *J. Biol. Chem.*, **126**, 737 (1938).
- ⁷ Barton, *J. Chem. Soc.*, 783 (1948).
- ⁸ Bachmann, Cole and Wilds, *J. Amer. Chem. Soc.*, **62**, 824 (1940).
- ⁹ Dane and Schmitt, *Ann.*, **537**, 246 (1939).
- ¹⁰ See, for example, Anner and Miescher, *Helv. Chim. Acta*, **30**, 1422 (1947).
- ¹¹ Pearlman and Wintersteiner, *J. Biol. Chem.*, **130**, 35 (1939); **132**, 605 (1940).
- ¹² Inhoffen and Zühlendorf, *Ber.*, **74**, 1911 (1941).
- ¹³ Wilds and Djerassi, *J. Amer. Chem. Soc.*, **68**, 2125 (1946).
- ¹⁴ Wieland and Dane, *Z. Physiol. Chem.*, **216**, 91 (1933).
- ¹⁵ Peak, *Nature*, **140**, 280 (1937).
- ¹⁶ Dimroth and Jonsson, *Ber.*, **74**, 5201 (1941).
- ¹⁷ Windaus and Zühlendorf, *Ann.*, **536**, 209 (1938).