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D-Serine Augments NMDA-NR2B Receptor-Dependent Hippocampal Long-Term Depression and Spatial Reversal Learning

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The contributions of hippocampal long-term depression (LTD) to explicit learning and memory are poorly understood. Electrophysiological and behavioral studies examined the effects of modulating NMDA receptor-dependent LTD on spatial learning in the Morris water maze (MWM). The NMDA receptor co-agonist D-serine substantially enhanced NR2B-dependent LTD, but not long-term potentiation (LTP) or depotentiation, in hippocampal slices from adult wild type mice. Exogenous D-serine did not alter MWM acquisition, but substantially enhanced subsequent reversal learning of a novel target location and performance in a delayed-matching-to-place task. Conversely, an NR2B antagonist disrupted reversal learning and promoted perseveration. Endogenous synaptic D-serine likely saturates during LTP induction because exogenous D-serine rescued deficient LTP and MWM acquisition in Grin1^{D481N} mutant mice having a lower D-serine affinity. Thus, D-serine may enhance a form of hippocampal NR2B-dependent LTD that contributes to spatial reversal learning. By enhancing this form of synaptic plasticity, D-serine could improve cognitive flexibility in psychiatric disorders characterized by perseveration of aberrant ideation or behaviors.

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INTRODUCTION

In the hippocampus, activation of postsynaptic NMDA-type glutamate receptors by high-frequency stimulation triggers long-term potentiation (LTP) of glutamatergic neurotransmission, a form of synaptic plasticity thought to underlie some forms of explicit learning (O'Keefe, 1993; Martin *et al*, 2000; Lynch, 2004). In contrast, low-frequency stimulation can induce an NMDA-dependent long-term synaptic depression (LTD) (Dudek and Bear, 1992; Wagner and Alger, 1996; Kemp and Bashir, 2001). Behavioral experience can dynamically modulate the induction threshold for hippocampal LTD (Manahan-Vaughan and Braunewell, 1999; Kemp and Manahan-Vaughan, 2004) but, unlike LTP, the behavioral correlates of LTD modulation have not been extensively investigated.

The NMDA receptors that mediate most lasting forms of hippocampal plasticity are heteromeric protein complexes of different subunits (NR1_{a-h}, NR2A-D, NR3) that confer

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distinct biophysical properties (Monyer et al, 1994), and show specific patterns of expression during development and across the mature mammalian CNS (Dumas, 2005). In the mature hippocampus, functional receptors consist of two NR1 subunits associated with either two NR2A subunits, two NR2B subunits, or possibly one of each (Cull-Candy et al, 2001; Loftis and Janowsky, 2003). There is evidence to suggest that receptors with different NR2 subtypes mediate distinct forms of synaptic plasticity. Antagonists to NR2B-containing NMDA receptors were found to suppress specifically LTD in hippocampus (Liu et al, 2004) and cortex (Massey et al, 2004). More recent studies, however, have found both NR2B-independent LTD (Bartlett et al, 2007; Morishita et al, 2006) and NR2Bdependent LTP (Zhao et al, 2005; Bartlett et al, 2007; Fox et al, 2006). Also, NR2B antagonists have shown inconsistent and perplexing effects on hippocampus-dependent behaviors, inhibiting cued and contextual fear conditioning (Rodrigues et al, 2001) but not spatial learning (Guscott et al, 2003; Higgins et al, 2003).

The activation of NMDA receptors requires, in addition to concurrent binding of glutamate to NR2 and membrane depolarization, binding of glycine or D-serine to the 'glycine site' of NR1 (Johnson and Ascher, 1987; Clements and Westbrook, 2001). Increasing D-serine catabolism with D-amino acid oxidase (DAO) (Hayashi *et al*, 1997; Wake *et al*,

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2001) suppresses NMDA currents and blocks hippocampal LTP (Yang *et al*, 2003), suggesting that D-serine is the principal endogenous co-agonist in the hippocampus and could dynamically regulate LTP and LTD.

To examine further the role of LTD in spatial learning and memory, we identified a pharmacological method for specifically modulating the magnitude of NR2B-mediated LTD in adult mouse hippocampus. Performance in several spatial learning tasks was then assessed under conditions of both NR2B-dependent LTD enhancement and pharmacological blockade. We also employed the Grin1^{D481N/D481N} mutant mouse line (Kew *et al*, 2000) to examine the effects of exogenous D-serine on LTP and spatial learning under conditions of normal and limiting endogenous D-serine.

MATERIALS AND METHODS

Experimental Animals

Mice were housed, handled, and killed in accordance with institution and national guidelines for animal welfare (Province of Ontario Animals for Research Act, 1971 and the Canadian Council on Animal Care). C57BL/6J mice were purchased from JAX Research Systems (Bar Harbor, ME) or bred from JAX mice. Mice were housed in separately ventilated, filtered polycarbonate cages, 3-5 mice per cage, with sterile water, and rodent chow ad libitum. The animal colony is temperature $(20 \pm 1^{\circ}C)$ and humidity (50-60%) controlled, with a 12 h diurnal cycle (lights on from 0700 to 1900 h). Mice expressing the $Grin1^{D481N/D481N}$ mutation were derived from founders generously provided by Dr M Pauley-Evers, Hoffmann-La Roche Ltd. (Basel, Switzerland). The D481N mutation was confirmed in our Grin1 stock by sequencing the 450 bp fragment using an ABI PRISM 377 DNA Sequencer (Applied Biosystems Inc., Foster City, CA). These animals were backcrossed to the C57BL/6J strain for 11 generations before experiments. All animals used for behavioral testing and electrophysiology were 4-6 months of age. Mice that were tested in behavioral tests were not used for electrophysiological experiments. The Grin1^{D481N/D481N} mutation (aspartate to asparagine substitution at amino acid 481) is in the NR1 glycine-binding region and confers an approximately fivefold lower co-agonist affinity in the homozygous state (Kew et al, 2000).

To genotype the Grin1 line, tail clippings (0.5-1.0 cm) were taken and digested overnight in 600 ml TNES buffer (10 mM Tris-HCl, 400 mM NaCl, 100 mM EDTA, 0.6% SDS, pH 7.5) and 20 µl Proteinase K (Roche Diagnostics Canada, Laval, PQ). Genomic DNA was isolated by the addition of 170 µl 6.0 M sodium chloride, followed by ethanol precipitation and centrifugation. DNA was then dissolved in 20 mM Tris-HCl, pH 8. For PCR amplification of the Grin1 gene, the primers 5'-ATGTCAAGCCCACAATGAGT-3' and 5'-CCTGCTCCACGTCTACCTAA-3' were added to 50 ng genomic DNA, and the gene was amplified using Taq DNA polymerase (Fermentas Life Sciences Canada, Burlington ON). The 450 base pair (bp) product was then digested with the restriction endonuclease MlsI (Fermentas) and separated on a 1.5% agarose gel. Wild type fragments resisted digestion; heterozygous +/D481N fragments resulted in three bands of 450, 290, and 160 bp, while homozygous D481N/D481N animals generated 290 and 160 bp fragments.

Electrophysiology

Mice were decapitated under halothane anesthesia, in accordance with institutional guidelines, and the head was submerged in partially frozen artificial cerebrospinal fluid (ACSF). The contents of this ACSF used for subsequent slice preparation, storage, and some electrophysiological studies contained (in mM): 124 NaCl, 26 NaHCO₃, 10 Glucose, 3.3 KCl, 1 NaHPO₄, 2 CaCl₂, 2 MgCl₂. Transverse slices from isolated hippocampi were cut on a Stoelting tissue chopper at 400 μm and stored in a tissue chamber aerated with 95% O_2 , 5% CO_2 at room temperature (21–23°C). Slices were stored for a minimum of 1.5h before experimentation. Individual slices were then transferred to a submerged superfusion chamber and secured with platinum wire. Slices were superfused at a rate of 3-4 ml/min for most experiments and at a slightly faster rate (5 ml/min) for the LTD and depotentiation studies using the glutamate uptake inhibitor TBOA (Figures 1a, b, g, and 5g). A bipolar stimulating electrode (FHC, Bowdoinham ME, USA), coupled to a physiological stimulator (Grass S88K) was placed in CA1 stratum radiatum. Field potential electrodes were pulled from 1.5 mm OD thin wall borosilicate glass (2- $5 M\Omega$, FHC) and advanced just below the slice surface of CA1 stratum radiatum. In some experiments (Figures 5e and f), a second electrode was placed in the CA1 stratum pyramidali for simultaneous fEPSP and population action potential (popAP) recordings. An Axopatch 1D amplifier (Axon Instruments, Foster City, CA) measured evoked excitatory postsynaptic field potentials (fEPSPs) in currentclamp mode, while an Axoclamp 2B amplifier was employed to measure popAPs. Slices were superfused with ACSF at $29.0 \pm 0.5^{\circ}$ C for $\approx 20-30$ min before baseline recording began. Data acquisition and analysis employed Clampex-9 software (Axon). Recordings of baseline and post-stimulus fEPSPs were filtered at 1 Hz low pass and 2 kHz high pass. Recordings of fEPSPs and popAPs during θ burst stimulation (TBS) were not low pass filtered. Only hippocampal slices showing maximum fEPSP amplitudes greater than 2 mV were used for experiments. To establish baseline (control) fEPSP slope, stimuli were delivered one every 30 s. LTP was evoked by high-frequency stimulation, consisting of either 100 Hz trains or of θ burst stimulation (50 ms, 100 Hz bursts delivered at 5 Hz) as indicated. Data is expressed as the slope of the fEPSP (over 20-60% of magnitude) vs time, normalized to a 20 min (40 sample) baseline. For figure clarity, individual fEPSPs were binned in groups of 3 or 4. For the LTD studies illustrated in Figure 1, the divalent ion composition of the ACSF was 1.0 mM MgCl₂ and 2.5 mM CaCl₂ because we found LTD to be more robust under these divalent conditions. Slices were preincubated for approximately 1 h before baseline recording to ensure complete re-equilibration of the extracellular cation concentration. We measured LTP and depotentiation under these same conditions for comparison. Other studies not directly comparable to the LTD experiments (Figures 2 and 5) were performed in our standard laboratory ACSF.

Behavioral Testing

Adult C57BL/6J, Grin1mutants and wild-type littermate mice were tested for spatial learning and memory in the

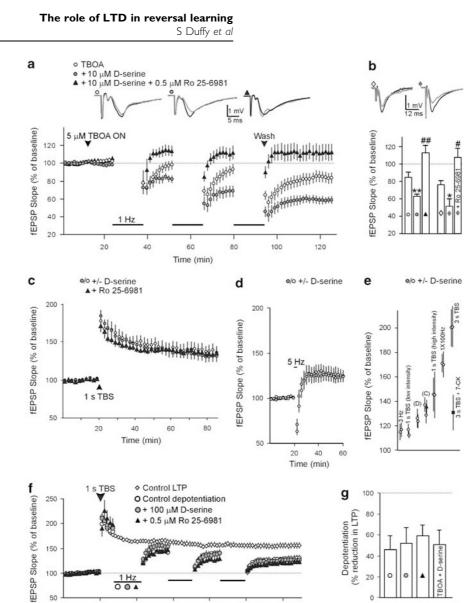


Figure I D-serine increased hippocampal LTD, but did not alter LTP or depotentiation. (a) In the presence of 5 µM TBOA, three, one Hz stimulus trains (900 pulses, fEPSPs at 40% of maximum) evoked a summating depression of the fEPSP slope (open circles). This depression was significantly increased when 10 μ M D-serine was co-applied with TBOA (closed circles), and completely inhibited by 0.5 μ M of the NR2B antagonist Ro 25-6981 (close triangles). Inset above: sample fEPSPs before the first I Hz train (black) and 30 min after the third (gray) for each condition. (b) Summary of average fEPSP amplitudes 25-30 min after the third 1 Hz train (TBOA vs TBOA + D-serine: **p < 0.02; TBOA + D-serine vs TBOA + D-serine + Ro 25-6981: $^{##}p < 0.0001$). Ro 25-6981 also inhibited the LTD induced by a single 1 Hz train in the presence of 30 µM TBOA and 100 µM D-serine. TBOA (open diamond) vs TBOA + D-serine (closed diamond): *p < 0.05; TBOA + D-serine vs TBOA + D-serine + Ro 25 6981: #p < 0.005. Inset above left: sample sweeps for 30 µM TBOA before and I h after I Hz stimulation. Inset above right: sample sweeps for 30 μM TBOA + 100 μM D-serine I h after I Hz stimulation. (c) In contrast, an even greater D-serine concentration (100 µM) did not alter the submaximal LTP induced by 1 s TBS (five bursts, each of six pulses at 100 Hz with fEPSP magnitude at 20% of maximum), nor was this LTP altered by Ro 25-6981. (d) D-serine did not alter the potentiation evoked by 3 min of 5 Hz at 40% of maximum fEPSP. (e) LTP was evoked by various stimulus protocols in the absence and presence of D-serine. Induction protocols are indicated above each group of points. LTP was measured from 50 to 60 min after induction except for 3 and 5 Hz experiments where it was measured 30-40 min post-stimulus. Data from (c) and (d) are also included as marked. Each point is the average of seven separate experiments, except for 100 Hz LTP (n = 5 for control and D-serine). The D-serine site antagonist 7-chlorokynurenic acid (7-CK, 10 µM) blocked LTP induced by 3 s TBS. (f) D-serine (100 µM), applied immediately following I s TBS (burst were four pulses at 100 Hz, evoked at 40% maximum fEPSP) did not alter depotentiation evoked by three, 1 Hz trains at 15 min intervals. (g) Summary of depotentiation experiments. Unlike LTD, depotentiation was not blocked by Ro 25-6981, nor was it altered in the presence of 0.5 μ M TBOA and $10 \,\mu\text{M}$ D-serine.

100

120

Morris water maze task. Approximately equal numbers of male and female mice were used for each behavioral task. A 1.17 m circular pool was filled with water $(26 \pm 1^{\circ}C)$ made opaque with white non-toxic paint. Distal visual cues were fixed to the room walls, approximately 1 m away from the pool edge, and four floor light sources of equal power provided uniform illumination to the pool and testing

50

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40

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Time (min)

80

room. A CCD camera suspended above the pool center was used for recording, and video output was digitized by a frame grabber (HVS Image, Twickenham, Middlesex, UK) controlled by the accompanying software (HVS Water 2020). Each experimental protocol began with a single day, four-trial stationary visible platform task. Mice were released in pseudo-random order from one of four

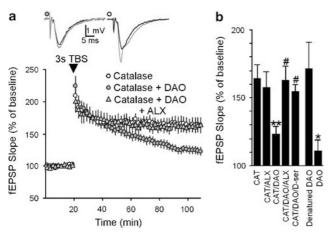


Figure 2 Endogenous D-serine is required to induce LTP. (a) LTP induced by 3 s TBS (15 bursts of six pulses at 100 Hz with fEPSP magnitude at 40% of maximum) at hippocampal Schaffer collateral-CA1 was reduced by preincubation of the D-serine catabolic enzyme D-amino acid oxidase (DAO) and catalase (CAT, shaded circles) relative to CAT alone (open circles). The glycine transport inhibitor ALX-5407 rescued LTP in the presence of catalase and DAO (shaded triangles). Plots are of average fEPSP slope normalized to baseline \pm SE. Individual fEPSP responses obtained every 30s were averaged in groups of 4 for clarity. Inset above: sample fEPSP sweeps recorded before (black) and 80 min after TBS (gray) for a slice treated with DAO and CAT (left) or CAT alone (right). (b) Summary bar graph of LTP amplitudes 80-90 min after induction by 3 s TBS for each treatment studied. These data also show rescue of LTP by D-serine in DAO/catalase-treated slices, and inhibition of the DAO mediated reduction by heat denaturation (symbols: CAT + DAO vs CAT: **p<0.004; CAT+DAO vs CAT+DAO+ALX: *p<0.006, CAT+ DAO + D-serine: p < 0.03; DAO vs denatured DAO: p < 0.04).

positions at the pool periphery. To escape, mice had to swim to a 10-cm-diameter platform that was elevated 0.5 cm above the water surface and demarcated with a 10 cm vertical pole. The edge of the platform was 30 cm from the pool wall in the visible and subsequent hidden platform task. The hidden platform task began 1 day later and lasted 4 or 5 days, each consisting of 4 trials with 1 h inter-trial intervals. The water level was adjusted so that the escape platform was submerged $\approx 1 \text{ cm}$ below the surface, and mice were allowed to search for the platform for 90 s. Mice that found the platform were left on it for 15s, while unsuccessful mice were assigned a 90 s latency and placed on the platform for 15 s. A single probe trial was performed 24 h following the final set of acquisition trials in which the platform was removed from the pool. Mice were released from the point furthest from the former platform location and the swim path was recorded for 60 s. To quantify spatial memory, we measured the time spent within an area three times the platform diameter centered at its former location. For some cohorts, the probe trial was followed by a reversal learning protocol where the hidden platform was moved to an opposite quadrant (SE to NW). Reversal learning entailed three additional days with 4 trials/day, similar to acquisition training. We also employed a delayed-matchingto-place (DMP) version of the Morris water maze (MWM) task consisting of a single day of visible platform training and 12 acquisition days where the hidden platform was moved to one of 12 predetermined locations at the beginning of each day. The edge of the platform at each position was either 25 or 40 cm from the wall. The methodology was similar to that described above for the stationary platform (reference memory) MWM task except that a larger 1.8-m-diameter pool was used to increase the possible distance between platform locations. Within a training day, the platform remained in the same position for four trials with an inter-trial interval of 45 min between all trials.

To analyze performance in the Morris water maze, HVS Water 2020 software was used. Behavioral measures included latency to the platform, pathlength, thigmotaxis, swim speed, floating, % time within the target area, average proximity to the target area, and the number of platform crosses. Two independent observers that were blind to treatments, genotypes, and trial classified the swim strategies using Wintrack (program by Dr D Wolfer, www.dpwolfer.ch/wintrack). This program was used to classify search strategies into the following eight types: thigmotaxis (swimming along the wall of the pool), circling (tight circular swimming), floating (absence of swimming) activity), random (swimming over the entire pool surface), scanning (swimming in straight paths predominately restricted to a central area of the pool), chaining (circular swimming at roughly the same distance from the wall as the platform), incorrect focal searching (swimming that is restricted to and repetitive within a specific area of the pool where the platform is not located), correct focal search (swimming that is restricted to and repetitive within a specific area of the pool where the platform is located), and direct swim (swimming in a straight path directly to the platform) (Wolfer and Lipp, 2000; Janus, 2004). The strategies that do not involve searching for the platform (thigmotaxis, floating, and circling) were grouped under 'not searching', the strategies that did not require allocentric navigation (random, scanning, and chaining) were grouped under 'non-spatial strategy', and the strategies that did require formation of an allocentric cognitive map (correct and incorrect focal searches, direct swim) were grouped under 'spatial strategies'.

Drugs and Treatment Procedures

Salts used for ACSF, DAO, 7-chlorokynurenic acid, ALX-5407, Ro 25-6981, and β -cyclodextrin were purchased from Sigma-Aldrich Chemicals (St Louis, MO, USA). D-serine and DL-TBOA were purchased from Tocris (Ellisville, MO, USA). D-serine (600 mg/kg) was dissolved in 0.9% NaCl and injected subcutaneously 20 min before the first trial of each day, as described previously (Lipina *et al*, 2005). Ro 25-6981, a high affinity blocker of NR2B and mixed NR2A/B NMDA receptors (Chazot *et al*, 2002), was dissolved in 0.9% saline with 0.3% Tween and injected subcutaneously at 5 mg/kg 30 min before the first trial of each day (Chaperon *et al*, 2003). No drugs were given before the probe trials.

Statistical Analysis

For electrophysiological data, one-way analysis of variance (ANOVA) with *post hoc* least significant difference (LSD) tests were used. Data generated in the stationary platform (reference memory) and reversal MWM task for the Grin1^{D481N/D481N} mice were analyzed using a two-way repeated measures (RM) ANOVA with either drug treat-

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ment (saline and *D*-serine) or group (wild type, mutant, mutant + D-serine) as the between-subjects factor and days as the within-subjects factor. The DMP and the visible platform MWM task were analyzed using a two-way RM ANOVA with treatment or group as the between-subjects factor and trial as the within-subjects factor. To analyze the acquisition and reversal memory MWM task for the NR2B antagonist experiment, a three-way RM ANOVA was completed with drug treatment (vehicle, Ro 25-6981) as the between-subjects factor and days and trials as the within-subjects factors. To determine main effects within the individual days or across trials 2 and 3 on reversal days 8 and 9 (4), a two-way RM ANOVA with treatment as the between-subjects factor and trial as the within-subjects factor was used. For comparisons within individual days (Figures 3 and 6), individual trials (Figure 4), or probe trials, one way ANOVAs were used. Significant main or interaction effects were followed by a LSD post hoc test. Results were considered significant when p < 0.05. Analysis was performed using Statistica (StatSoft Inc.).

RESULTS

Endogenous D-Serine Enhanced LTD but not LTP or Depotentiation

To examine the role of LTD in hippocampus-dependent learning, we required a method for regulating specifically this form of plasticity. The obligatory NMDA receptor coagonist D-serine augmented NMDA currents evoked by single stimuli (Mothet *et al*, 2000; Chen *et al*, 2003), indicating that the D-serine concentration may not be sufficient to saturate the glycine site during low-frequency stimulation. Exogenous D-serine could, therefore, augment NMDA-dependent low-frequency-induced LTD. Similarly, if endogenous D-serine levels are also subsaturating during high-frequency stimulation, addition of exogenous D-serine would augment submaximal LTP.

LTD can be induced routinely at Schaeffer collateral-CA1 synapses in hippocampal slices from younger mice by lowfrequency stimulus trains alone. As they age, however, rats and mice become resistant to this protocol (Wagner and Alger, 1996). In adults, LTD can be induced by lowfrequency stimulation in the presence of glutamate transport inhibitors that facilitate stimulation of extra-synaptic NMDA-NR2B receptors (Massey et al, 2004). To measure LTD in mice that were aged matched to those used in subsequent behavioral analysis (4-6 months old), we applied 1 Hz stimulation during application of the glutamate transport inhibitor TBOA. In the presence of 5 µM TBOA, three 1 Hz trains of 15 min duration induced a gradual synaptic depression that was significantly augmented by 10 µM D-serine and blocked by 0.5 µM of the NMDA-NR2B subunit antagonist Ro 25-6981 (25-30 min after the third train, fEPSP slopes were TBOA: $84\pm6\%$ of baseline, n = 10; TBOA + D-serine: $62 \pm 3\%$, n = 9; TBOA + D-serine + Ro 256981: $112\pm9\%$, n=6 (main effect of treatment: $F_{(2,22)} = 13.73$, p = 0.0001; Figure 1a and b). In contrast, an even greater D-serine concentration (100 µM) did not significantly alter submaximal LTP induced by a variety of stimulus protocols designed to elicit LTP responses of varying magnitudes (including 3 Hz for 5 min with fEPSPs

at 40% of maximum; 5 Hz for 3 min with fEPSPs at 40% of maximum; low intensity 5 Hz θ burst stimulation with bursts consisting of three pulses at 100 Hz with fEPSPs at 20% of maximum; high intensity TBS with bursts consisting of six pulses at 100 Hz with fEPSPs at 20 or 40% of maximum; 3 s TBS with bursts of six pulses at 100 Hz, fEPSP at 40% of maximum, 100 Hz for 1 s with fEPSP at 40% of maximum; all performed in the presence of 2.5 mM CaCl₂ and 1 mM MgCl₂ except for the last two protocols listed performed in 2 mM CaCl₂ and 2 mM MgCl₂; Figure 1c-e). LTP induced by TBS was not blocked by Ro 25-6981 (Figure 1c). Furthermore, neither exogenous D-serine nor Ro 25-6981 altered the depotentiation of pre-established LTP using the same stimulation pattern $(3 \times 900 \text{ pulses at})$ 1 Hz every 15 min) employed to elicit LTD (Figure 1f and g). Similarly, addition of $5 \mu M$ TBOA and $10 \mu M$ D-serine did not alter depotentiation (LTP with post-tetanic TBOA and D-serine: $188 \pm 10\%$; LTP after 3×1 Hz stimulation: $144 \pm 11\%$, depotentiation = 50 ± 14%; Figure 1g). Thus, the effect of exogenous D-serine was specific for Ro 25-6981-sensitive (NMDA-NR2B-dependent) synaptic depression; D-serine had no effect on NMDA-NR2B-independent plasticity, even when induced by the identical pattern of synaptic stimulation (depotentiation).

To show that SC-CA1 LTP was in fact dependent on endogenous D-serine, we confirmed and extended previous results by examining LTP under conditions of reduced endogenous D-serine. Extracellular D-serine was reduced by addition of DAO. Catalase (CAT) was also added in some trials because neurotoxic H_2O_2 is generated by DAOmediated catabolism of D-serine. Slices were preincubated for 1.0-1.5 h in either 0.2 U/ml DAO plus 260 U/ml catalase, catalase alone, 0.2 U DAO alone, or 0.2 U (by weight) DAO inactivated by 1-2h boiling in water. In some trials using DAO plus CAT, the glycine transport blocker ALX-5407 (1µM) was co-applied 30 min before LTP induction; in others, 1 mM D-serine was added 4.5 min earlier. Basal synaptic strength, determined by the relation between presynaptic fiber volley amplitude and postsynaptic fEPSP slope (IO), did not differ between slices preincubated in either active or denatured DAO (IO in active DAO was 103% of the IO in denatured DAO, data from 6 slices / group). The LTP induced by 3s of TBS was significantly reduced by DAO (plus CAT) relative to CAT alone, and rescued by application of the glycine transport inhibitor ALX-5407 (LTP 85-90 min post-TBS, CAT: $164 \pm 11\%$, n = 8; CAT + DAO: $123 \pm 6\%$, n = 9; CAT + DAO + ALX-5407: 163 ± 10 , n = 6; main effect of treatment: $F_{(4, 26)} = 3.82$, p = 0.014; Figure 2a and b). This deficient LTP was also rescued by application of D-serine (157+12, n=6) and was not observed if DAO was first heat-inactivated (DAO alone: $110\pm9\%$; n=6; denatured DAO: 164 ± 21 , n=7; effect of treatment: $F_{(1,10)} = 6.09$, p = 0.033; Figure 2b).

In sum, elevating extracellular D-serine augmented LTD but was without effect on sub-maximal LTP. Furthermore, D-serine appears to be the dominant physiologic co-agonist in the hippocampus. The synaptic D-serine concentration is probably saturating at the subsynaptic NMDA-NR1 D-serine binding sites that initiate LTP and depotentiation (Liu *et al*, 2004) but not at the NMDA receptors that mediate LTD. This conclusion is further supported by subsequent results presented in Figures 5 and 6.

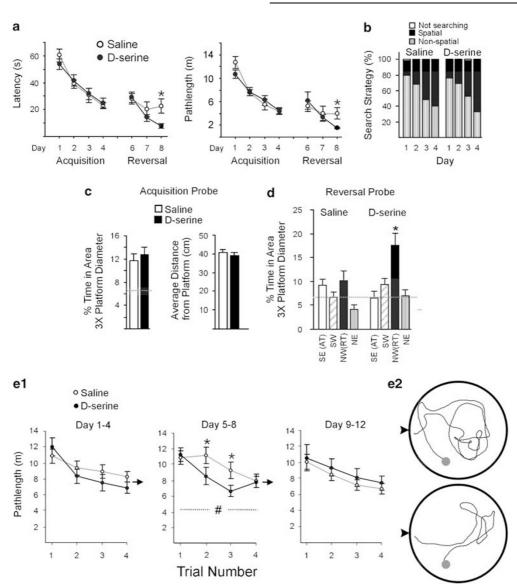


Figure 3 D-serine specifically enhanced spatial reversal learning in wild type mice. (a) The effects of D-serine on spatial learning in the stationary hidden platform version of the Morris water maze. Subcutaneous injection of D-serine (600 mg/kg) did not alter the initial acquisition of a hidden platform location as measured by both latency (left panel) and pathlength (right panel). Left: latencies to reach a hidden platform over a 4 day, 4 trials/day acquisition training period did not differ between D-serine-treated (closed circles) and saline-treated wild-type mice (open circles). In contrast, when the platform was moved to the opposite quadrant (reversal), D-serine-injected mice showed reduced latencies to find the hidden platform on the third day (*p<0.04) Right: similarly, the swim pathlengths from release point to platform were not altered by D-serine during acquisition training, but were significantly reduced on the third day of the reversal training (*p < 0.03). (b) The search strategies used over the 4 training days did not differ between saline-treated and D-serine-treated wildtype mice. Each bar shows the proportion of swims relative to the daily total (N mice × 4 trials/day) that were classified as not searching (floating, circling, and thigmotaxis, white area), Spatial searching (focal searches and direct swims, black area), or non-spatial searching (random, scanning, and chaining, vertical stripes). (c) The acquisition probe trial revealed that D-serine did not alter memory for the acquisition platform location. Left: the % of time spent in an area defined by three times the platform diameter (centered on its former location) did not differ significantly between mice injected with saline and mice injected with D-serine. The dashed line is the 'chance level', corresponding to the ratio of the 3 x diameter platform area to the total pool area (or 6.6%). Right: the average distance from the AT did not differ in the acquisition probe. (d) D-serine improved memory for the reversal platform location. In the reversal probe trial, mice injected with D-serine spent significantly more time in the area three times the platform diameter, centered around the reversal target (NW-RT), than did saline-treated controls (*p < 0.03). (e) Preinjection with D-serine increased the rate of acquisition of a novel platform location in the DMP version of the Morris water maze. (e1) The average pathlengths for each of the four daily trials over days 1–4, 5–8, and 9–12 are shown for saline-injected mice (open circles) and mice injected with 600 mg/kg D-serine (closed circles) before the first trial of each day. Pathlengths were significantly reduced during days 5-8 ($^{\#}p$ < 0.04 is the main effect across trials 2–4). (e2) Sample swim paths from day 6, trial 2, are shown at right for a saline-injected mouse (top) and a D-serineinjected mouse (bottom).

D-Serine Injection Improved Spatial Reversal Learning, but not Initial Acquisition, in Wild-Type Mice

In light of these effects on hippocampal synaptic plasticity, we examined if exogenous D-serine altered hippocampus-

dependent memory. To this end, we employed our colony of Grin1^{D481N/D481N} mice (Kew *et al*, 2000). The Grin1 mice have been backcrossed to the C57BL6/J strain for 11 generations, so wild-type Grin1^{+/+} are genetically comparable to those C57BL6/J mice used for electrophysiology in Figures 1 and 2.



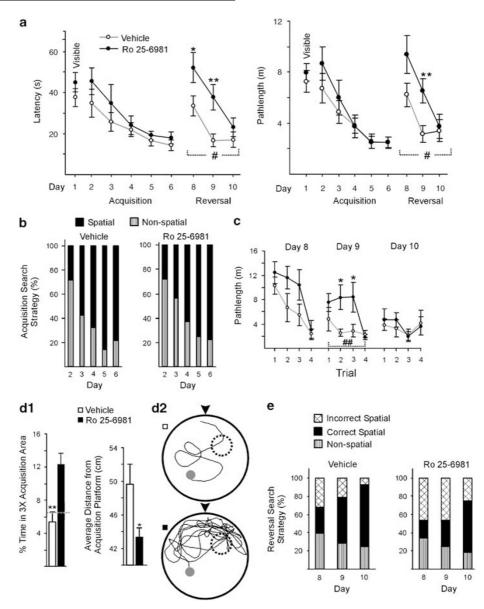


Figure 4 The NR2B antagonist Ro 25-6981 disrupts reversal learning and enhances perseveration in the Morris water maze. (a) The latency to find a platform (left panel) and pathlength (right panel) for wild type mice preinjected with vehicle (control, open circles) or 5 mg/kg Ro 25-6981 (closed circles). Neither the latency nor pathlength to find a visible platform was significantly different between treatment groups (day I, average latency for four trials). The latency to find a submerged hidden platform (acquisition) decreased for both groups over the 5 days of training (days 2–6, 4 trials/day), but neither latencies nor pathlengths were significantly different between treatment groups overall or on any day. When the hidden platform was moved to an opposite quadrant (reversal, days 8–10), mice preinjected with Ro 25-6981 showed significantly longer average escape latencies ([#]p < 0.005 is the main effect of drug over days, *p<0.05, **p<0.01 is main effect within days). (b) The search strategies employed by the treatment groups during acquisition training. (c) the average pathlengths to the new (reversal) platform location is plotted for each trial (1–4) on each reversal training day ($^{\#p} > 0.01$ for main effect by RM ANOVA; *p < 0.05, post hoc LSD test). (d1) Mice treated with Ro 25-6981 showed perseveration for the AT during reversal trials 2 and 3 on days 8 and 9. Left panel: the average time spent in the 3 × area of the former (acquisition) location for trials 2 and 3 on days 8 and 9. Mice injected with Ro 25-6981 spent significantly more time in the 3 × diameter area of the old acquisition location (**p<0.0002). Dashed line marks the chance time (6.6%). Right panel: average distance from the old acquisition location was significantly shorter during the platform search in Ro 25-6981-treated mice (*p < 0.04). (d2) Sample swim paths from trial 2, day 9, demonstrating the tendency of Ro 25-6981-injected mice to swim in the old acquisition platform location (demarcated with dashed circle) before locating the reversal platform (gray disk). (e) Search strategies during the reversal trials. The total fraction of spatial and non-spatial swims (area with vertical stripes vs black area plus hatched area) did not differ, but Ro 25-6981-treated mice showed a greater propensity for focal searches in an incorrect acquisition platform area during reversal days.

We found no differences in basal synaptic transmission or LTP in hippocampal slices from C57BL/6J mice and Grin1⁺ mice (LTP at 60 min post-tetanus, C57BL6/J: $179\pm8\%$ of baseline, n=8; Grin1^{+/+}: $179\pm11\%$, n=9). In contrast, the mutant littermates (Grin1^{D481N/D481N}) express an NR1 subunit with 4-5-fold lower D-serine affinity (Kew et al,

2000). The Grin1 mutant line, therefore, provides a positive control for our subsequent experiments. Furthermore, we used these mutants to support our conclusions regarding the specific enhancement of LTD by exogenous D-serine.

We first measured spatial learning in wild-type (Grin1^{+/+}) mice using the reference memory version of

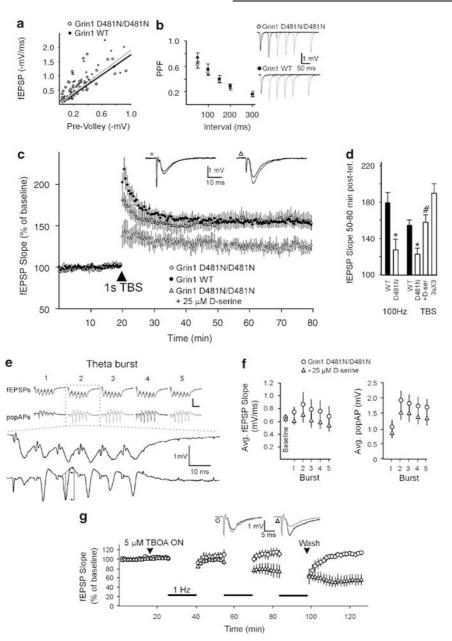


Figure 5 Exogenous D-serine rescued deficient LTP in hippocampal slices from Grin1 mutant (D481N/D481N) mice. (a) Plot of presynaptic fiber volley amplitude against fEPSP slope for mutant (Grin I mutants, open circles) and wild type (closed circles) revealed no difference in basal synaptic strength. (b) A plot of PPF ((fEPSP2 slope/fEPSP1 slope)-1) vs inter-stimulus interval revealed no difference between mutant (open circles) and wild type mice (closed circles). Inset to the right, sample fEPSPs in response to paired pulses of 50, 100, 150, 200, and 300 ms. (c) Hippocampal slices from Grin1 mutant mice exhibited diminished LTP induced by 1 s TBS (five bursts of six pulses at 100 Hz with fEPSP magnitude at 40% of maximum, open circles) relative to slices from wild type mice (closed circles). This deficient LTP was rescued by addition of exogenous D-serine (25 µM, open triangles). Inset above: sample sweeps recorded 60 min after LTP induction in slices from Grin1 mutant mice in the absence and presence of 25 µM D-serine. (d) Summary of experiments from (c). Graph also shows that slices from Grin I mutant mice displayed robust, long-lasting LTP in response to more intense stimulation (3 s TBS repeated three times at 5 min intervals, labeled as $3s \times 3$). Wild type vs mutant: *p < 0.02; mut vs mut + D-serine: *p < 0.01). (e and f) D-serine did not alter the neuronal depolarization evoked by TBS in slices from Grin I mutant mice. (e) Upper 2 traces: typical sample sweeps of simultaneously recorded fEPSP trains and popAP trains during the five 100 Hz heta bursts. Note that the intervals between bursts have been omitted. Lower two sweeps: the second burst on an expanded time scale showing the method used to quantify popAP amplitude (arrows). (f) Average intraburst fEPSP slopes (left panel) and popAP amplitudes (right panel) from Grin I mutant slices during TBS. The average intraburst fEPSP slope in the presence of D-serine (open triangles) was not significantly different from the average fEPSPs recorded in drug-free slices (open circles). Similarly, D-serine treatment had no effect on average intraburst popAP amplitude. (g) LTD could not be elicited by 3, I Hz stimulus trains in slices from Grin I mutant mice in the absence of exogenous D-serine (open circles). Inset above: sample sweeps from drug-free slices (open circle) and in the presence of D-serine (open triangles).

the Morris water maze, a test that depends on intact hippocampal function (Morris et al, 1986). Subcutaneous injection of D-serine (600 mg/kg) 20 min before the first of four daily trials did not augment the acquisition of a hidden platform location over a 4-day 'acquisition' training period as indicated by both the latency to find the platform (Figure 3a, left panel) and the pathlength (Figure 3a, right panel; average of 23 saline-injected and 19 D-serine-injected

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mice). Moreover, D-serine did not alter the navigational search strategy employed by wild type mice (Figure 3b). Spatial memory, as measured 24 h later (day 5) during a 60 s probe trial by the fractional time spent within an area three times the platform diameter centered over the former location $(3 \times \text{diameter area})$ was also not significantly different from saline-injected controls (Figure 3c, left panel, p = NS). Furthermore, a second measure of target memory, the average distance from the target, did not differ (Figure 3c, right panel). D-serine had no significant effects on swim speed (Control: 17.9 ± 0.4 cm/s; D-serine-injected: 18.3 ± 0.4 cm/s) or on the propensity for the non-searching behaviors of floating and thigmotaxis (p = NS). Both experimental groups, however, clearly learned the specific target platform location as evidenced by a significant preference for the $3 \times$ target location compared to the averaged analogous non-target locations (WT + saline: $F_{(1,44)} = 6.26$, p = 0.016; WT + D-serine: $F_{(1,36)} = 12.87$, p = 0.001).

We then sought to find a behavioral correlate to the specific alterations in LTD observed in vitro, reasoning that synaptic depression may contribute to the extinction or reversal of previously formed memories. Indeed, in contrast to acquisition of the first target, D-serine markedly enhanced the rate at which these mice learned the position of a new hidden target in a subsequent reversal learning task (Figure 3a, right side of each panel) as revealed by both significantly reduced latency and pathlength on the third day of training (effect of treatment: latency: $F_{(1, 21)} = 4.89$, p = 0.038; pathlength: F_(1, 21) = 6.23, p = 0.021, n = 12 salinetreated, and n = 11 D-serine-treated mice). Furthermore, the memory for this new reversal target position was substantially improved. Compared to saline-treated mice, D-serineinjected mice spent significantly more time in the $3 \times$ platform diameter area (RT) during a 60 s probe trial 24 h following the third reversal training day (effect of treatment: $F_{(1,21)} = 5.69$, p = 0.027; Figure 3d). The % time in RT was significantly greater in both groups compared to the averaged analogous area of the two non-targeted platforms (WT + saline: $F_{(1, 22)} = 4.37$, p = 0.048; WT + D-serine: $F_{(1, 20)} = 14.04$, p = 0.0013). When comparing the time spent in RT to that in the former acquisition target (AT), salinetreated mice did not show a preference $(F_{(1,22)} = 0.13)$, p = 0.772), indicating that their learning was not specific for

RT over AT, whereas D-serine-injected mice showed a significant preference for RT over AT ($F_{(1,20)} = 16.08$, p = 0.0007; Figure 3d).

We next examined the effects of D-serine during a more rigorous reversal task, a DMP version of the Morris water maze, where the location of the hidden platform was changed daily to one of 12 preassigned positions (Figure 3e). Mice were given 4 trials/day and the pathlength of each trial was averaged in three blocks (days 1-4, 5-8, 9-12) (Nakazawa et al, 2003). Over the first 4 days of training, the trial-to-trial pathlengths were significantly reduced for both saline-treated mice (n = 16) and D-serinetreated mice (n = 13) (main effect of trials on days 1-4: $F_{(3,81)} = 7.15$, p = 0.00025). The D-serine-injected group, however, did not reach significance compared to salineinjected controls on the non-random trials 2-4 (main effect of treatment for pathlength: $F_{(1, 27)} = 3.86$, p = 0.060). Over the next 4 days (5-8), a clear difference in pathlength was observed, with D-serine-injected mice showing a significantly greater decrease over the non-random trials 2-4 (main effect of treatment for pathlength: $F_{(1,27)} = 5.18$, p = 0.031, Figures 3e1 and e2). D-serine injection had no significant effect on swim speed, floating, or thigmotaxis (p = NS). With further training (days 9–12), saline-injected mice demonstrated similar pathlengths to the D-serine group.

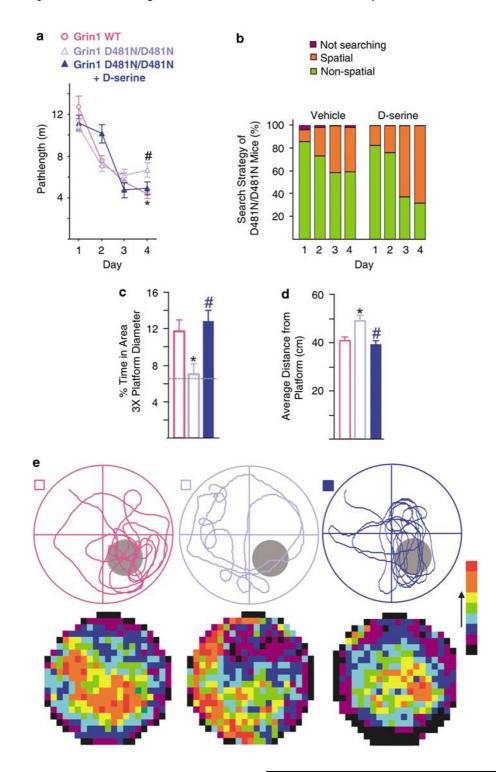
An NR2B-Antagonist Disrupts Spatial Reversal Learning and Promotes Perseveration

In the present study, hippocampal LTD in adult mice was blocked by Ro 25-6981 (Figure 1a), a highly specific antagonist for the NR1-NR2B subtype of the NMDA receptor (Figure 1a). We examined the effects of systemic injection of this drug on spatial acquisition and reversal learning in the MWM task. Preinjection of Ro 25-6981 had no significant effect on either latency or pathlength to a visible platform on day 1, or on the acquisition of a hidden platform location over 5 training days with 4 trials/day (main effect of treatment on acquisition latency: $F_{(1,15)} = 1.61$, p = 0.224; on pathlength: $F_{(1,15)} = 2.06$, p = 0.171; n = 9 vehicle-treated, and n = 8 Ro 25-6981-treated mice; Figure 4a). There was no change in the search strategy employed (spatial *vs* non-spatial) over the course of

Figure 6 Deficient spatial learning and memory in Grin I mutant mice was rescued by injection of D-serine (600 mg/kg). (a) Pathlengths during hidden platform searches for saline-injected wild type Grin I mice (open circles) and Grin I mutant mice injected with either saline (open triangles) or D-serine (closed triangles). Saline-injected Grin I mutant mice demonstrated poor acquisition of platform location during training, as indicated by a significantly longer pathlength on day 4 compared to wild-type littermates (*p < 0.006). This deficit was significantly reduced by injection of D-serine (#p < 0.05). (b) The search strategies used over the 4 training days for saline-treated and D-serine-treated mice. Not searching (floating, circling, and thigmotaxis, purple), non-spatial searching (random, scanning, and chaining, green) or spatial searching (focal searches and direct swims, orange). Non-spatial strategies predominant for both saline- and D-serine-injected groups on the first two days, but are gradually replaced by spatial strategies. D-serine increased the proportion of mutant mice using spatial strategies. (c) D-serine improved memory for the platform location during the probe trial. Time spent in an area three times the platform diameter was significantly lower in saline-injected mutant mice compared to both wild type mice (*p < 0.02) and Grin I mutants that were injected with D-serine during training ($^{\#}p < 0.01$). The dashed line is the chance level (6.6%). (d) The average proximity to the target was also significantly reduced in Grin I mutants relative to wild type mice (*p < 0.005) and D-serine-treated mutants ($\frac{a}{p} < 0.001$). (e) Upper panels: typical swim paths during the probe trial for a saline-injected wild type mouse (left), a mutant mouse injected with saline (middle) and a mutant injected with D-serine (right). The gray disk in the lower right quadrant demarcates the 3 × diameter area surrounding the former platform location. Lower panels: the spatial probability map for all animals in each group during the probe trial. Each map has a spatial resolution of approximately 5.8 cm (each pixel maps a pool area 5.8 \times 5.8 cm). The bright green color includes the chance probability level. The highest probability (orange-red) for wild type and D-serine-treated mutants lies over the former platform location (gray disc, top row). The equivalent area for the untreated mutants is at chance (green). A second area of high probability overlies the release point (NW periphery) in all three maps.

acquisition training (Figure 4b). Unlike acquisition training, Ro 25-6981-treated mice demonstrated significant deficits in acquisition memory as revealed by no preference for the target over the mean analogous non-target areas ($F_{(1,14)} = 0.04$, p = 0.844). In contrast, vehicle-treated mice showed a highly significant preference for the target ($F_{(1,14)} = 17.91$, p = 0.0008; data not shown).

When the platform location was switched to the opposite quadrant, mice injected with Ro 25-6981 showed a clear deficiency in initial performance during reversal trials relative to vehicle-injected controls (main effect of drug treatment over three reversal days: latency: $F_{(1, 15)} = 12.21$, p = 0.0033; pathlength: $F_{(1, 15)} = 11.06$, p = 0.0046; Figure 4a, reversal trials). This deficiency was significant within days on both the first (day 8) and second (day 9) reversal days (main effect of treatment for day 8 latency: $F_{(1, 15)} = 4.75$, p = 0.0457; day 9 latency: $F_{(1, 15)} = 10.19$, p = 0.006; day 9 pathlength: $F_{(1, 15)} = 9.20$, p = 0.0084; Figure 4a and c). Detailed analysis of swim paths on these days revealed that trials 2 and 3 on day 9 were the most deficient trials for both



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latency and pathlength (effect of treatment for latency: day 9, trial 2: $F_{(1,15)} = 8.79$, p = 0.0097; trial 3: $F_{(1,15)} = 5.18$, p = 0.038; pathlength: day 9, trial 2: $F_{(1,15)} = 8.20$, p = 0.0118; trial 3: $F_{(1,15)} = 4.67$, p = 0.0474; Figure 4c).

The Ro 25-6981-injected mice showed significant perseveration for the original (acquisition) platform location on trials 2 and 3 of days 8 and 9. That is, Ro 25-6981-treated mice spent significantly more time in the $3 \times$ diameter area of the acquisition platform location than did vehicleinjected controls (main effect of treatment: $F_{(1,15)} = 26.09$, p = 0.00016; Figure 4d1, left), had a shorter average distance to the AT (main effect of treatment: $F_{(1,15)} = 5.16$, p = 0.0384; Figure 4d1, right), had a reduced latency to the first AT crossing (vehicle: 45.56 ± 8.90 s; Ro 25-6981: 21.00 ± 5.40 s; main effect of treatment: $F_{(1, 15)} = 5.22$, p = 0.0373), and a great number of $3 \times$ diameter AT crossings (vehicle: 1.42 ± 0.35 ; Ro 25-6981: 4.84 ± 0.73 ; main effect of treatment: $F_{(1,15)} = 19.20$, p = 0.00054). This perseveration was also indicated by the prevalence of focal but incorrect searches relative to vehicle-treated mice (Figure 4e).

Finally, a reversal probe trial revealed a significant memory impairment in Ro 25-6981-treated mice compared to vehicle-treated mice, as indicated by reduced time in the $3 \times$ diameter reversal platform area (vehicle: 14.88 ± 1.14%; Ro 25-6981: $9.41 \pm 1.99\%$; effect of treatment: $F_{(1, 14)} = 5.67$, p = 0.0321), increased average distance from the reversal platform (Vehicle: 41.88 ± 1.17 cm; Ro 25-6981: $48.50 \pm$ 2.40 cm; effect of treatment: $F_{(1, 14)} = 6.16$, p = 0.0264), and by reduced number of crossings (vehicle: 6.38 ± 0.32 ; Ro 25-6981: 4.38 ± 0.78 ; effect of treatment: $F_{(1, 14)} = 5.65$, p = 0.0325). Also, Ro 25-6881-treated mice showed no preference in % time spent within the reversal target area compared to either the averaged unbiased non-target areas $(F_{(1,14)} = 1.50, p = 0.242)$ or the AT $(F_{(1,14)} = 0.31, p = 0.31)$ p = 0.584). In contrast, vehicle-treated mice demonstrated a significant preference for the reversal target over both the averaged unbiased analogous targets ($F_{(1,14)} = 6.81$, p = 0.021) and the AT (F_(1,14) = 4.92, p = 0.0435; data not shown). While Ro 25-6981 had significant effects on learning and memory, preinjection did not alter swim speed, floating, or thigmotaxis on visible, acquisition, or reversal trials (p = NS). In sum, a drug that blocked LTD also interfered with learning and led to a perseveration behavior.

${\rm D}\mbox{-serine}$ Rescued LTP and Spatial Learning in $Grin1^{D481N/D481N}$ Mutant Mice

To support further the hypothesis that the lack of effect of D-serine on TBS-evoked LTP and spatial acquisition was due to NR1 saturation (Figures 1c, 2a and b), and to determine whether our D-serine treatment protocols can enhance deficient LTP and learning, we repeated the electrophysiological and behavior tests in Grin1^{D481N/D481N} mutants. Synaptic strength, as indicated by the prevolley amplitude to fEPSP slope relation, was not altered by the Grin1 mutation (Figure 5a), indicating that basal AMPA-mediated neurotransmission was unchanged. Similarly, paired-pulse facilitation (PPF), a presynaptic form of short-term potentiation that may regulate LTP by controlling the level of postsynaptic depolarization, was unaltered (Figure 5b, n = 9 slices). Compared to wild-type littermates, Grin1^{D481N/D481N}

mutant mice expressed a major LTP deficit in response to 1 s of TBS that was rescued by addition of exogenous D-serine (at 1 h post-TBS, wild-type LTP: $154\pm6\%$, n=9; Grin1^{D481N/D481N} LTP: $123\pm7\%$, n=7; LTP in Grin1^{D481N/D481N} + 25 μ M D-serine: $158\pm8\%$, n=9; effect of group: $F_{(2,22)} = 6.40$, p = 0.006; Figure 5c and d). This LTP deficit and rescue by exogenous D-serine may be explained by the lower NR1 D-serine affinity conferred by the D481N mutation. In contrast to wild-type mice, exogenous synaptic D-serine may be subsaturating at CA3-CA1 synapses due to the expression of the low-affinity mutant NR1, leading to aberrantly reduced NMDA receptor activation that is restored by exogenous D-serine.

Activation of NMDA receptors also depend on depolarization from AMPA-mediated EPSPs and action potentials, so it is possible that p-serine rescued LTP by enhancing neuronal depolarization. To examine this possibility, we measured simultaneous fEPSPs and population action potentials (popAPs) during TBS (Figure 5e). Application of p-serine (25μ M) did not significantly alter the average intraburst fEPSP slope or the average intraburst popAP recorded during individual theta frequency bursts (average of seven TBS responses from seven drug-free slices and nine responses from nine p-serine-treated slices; Figure 5f). Consequently, the rescue of LTP in Grin1 mutants by p-serine is most likely due to enhanced NMDA receptor activation independent of membrane potential.

Finally, we showed that the Grin1 mutation also reduced LTD. Slices from these mice were resistant to LTD induction using the stimulation protocol employed previously in wild-type mice $(110\pm5\%)$ of baseline, n=6) unless the exogenous D-serine $(25\,\mu\text{M})$ was added $(58\pm11\%)$ of baseline, n=5, p<0.005, Figure 5g). Grin1^{D481N/D481N} mutant mice also demonstrated a

significant deficit in memory for the hidden platform location compared to wild type littermates (Figure 6a; average of 23 wild type, 24 saline-injected mutant mice, and 16 D-serine injected mutant mice). Subcutaneous injection of 600 mg/kg D-serine improved acquisition relative to saline-treated mutants by the fourth training day (group \times days interaction for pathlength: $F_{(6, 180)} = 3.83$, p = 0.0013; effect of group for pathlength on training day 4: $F_{(2, 60)} = 4.46$, p = 0.0156; main effect of group for average distance from target: $F_{(2, 60)} = 9.22$, p = 0.00032; group \times days interaction for average distance; $F_{(6, 180)} = 2.83$, p = 0.012; effect of group for average distance on day 4: $F_{(2,60)} = 9.50$, p = 0.0003). Post hoc LSD tests revealed a significant effect between wild type and mutant mice for pathlength (day 4: p = 0.0055), average distance from target (day 4: p = 0.00025), and between saline-treated and D-serine-treated mutants for pathlength (day 4: p = 0.050), and average distance from the target (day 4: p = 0.001).

Injection of D-serine also altered the prevailing swim strategy (Figure 6b). During the early training days, Grin1 mutants predominantly employed non-spatial search strategies (chaining, random swimming, and scanning). With training, such search strategies diminished and spatial or focal strategies emerged. D-serine increased the propensity for spatial searching, the increased efficiency of which could account for the observed improvement in performance.

Saline-treated Grin1 mutants performed very poorly in the subsequent probe trial relative to wild-type and D-serine-treated littermates, as measured by time spent in the $3 \times$ diameter area of the former target (effect of group: $F_{(2, 59)} = 5.17$, p = 0.0085; Figure 6c), by the average distance to the platform (effect of group: $F_{(2,59)} = 7.79$, p = 0.001; Figure 6d), by the latency to the first crossing (wild type: 11.22 ± 1.77 ; mutants: 23.22 ± 6.02 ; D-serine-injected mutants: 9.78 \pm 2.12; effect of group F_(2, 59) = 3.25, p = 0.0456), and by number of crossings over the $3 \times$ diameter area (wild type: 6.04 ± 0.50 passes, n = 23; mutants: 3.13 ± 0.47 passes n = 23; D-serine-injected mutants: 6.36 \pm 0.83, n = 16; effect of group: $F_{(2,59)} = 9.82$, p = 0.0002). Post hoc analysis revealed significant differences between wild type and mutants in time in the $3 \times$ diameter area (p = 0.011), average distance from platform (p = 0.0026), latency to first crossing (p = 0.035), and number of crossings over the $3 \times$ diameter area (p = 0.0004), and between saline-treated and D-serine-treated mutants in time spent (p = 0.0062), average distance (p = 0.0008), latency (p = 0.0325), and number of crossings (p = 0.00035). There were no significant differences in these parameters between wild type and D-serine-treated mutants, indicating that D-serine rescued the spatial learning and memory deficit in the Grin1 mutants. These effects are demonstrated by the spatial probability map derived from the swim paths recorded during the probe trial (Figure 6e, lower panels). Both wildtype mice and D-serine-treated mutants showed significant preferences for the target over the averaged analogous nontarget locations (wild type: $F_{(1,44)} = 6.26$, p = 0.016; mutant + D-serine: $F_{(1, 30)} = 4.39$, p = 0.045), while the saline-treated mutants showed no preference ($F_{(1, 44)} = 2.18$, p = 0.153).

To verify that the Grin1 mutation and D-serine did not affect motor co-ordination, vision, or motivation, groups were trained for four trials in a visible platform MWM task. Even though this test was run in a larger (1.8 m) pool to exacerbate any differences, neither latency nor pathlength differed between wild types, saline-treated wild types, and saline-treated mutants (main effect of group for latency: $F_{(2, 42)} = 1.34$, p = 0.273; pathlength: $F_{(2, 42)} = 1.31$, p = 0.282). In contrast, there was a strong main effect of trials (latency: $F_{(3, 126)} = 37.23$, $p = 2.7 \times 10^{-17}$; pathlength: $F_{(3, 126)} = 28.31$, $p = 4.6 \times 10^{-14}$), indicating that learning in the groups occurred across trials. Average swimming speed did not differ between mutant and wild type mice as measured on the day 4 of hidden platform training (wild type: 16.9 ± 0.5 cm/s; mutants: 15.5 ± 0.7 cm/s). Finally, we used a 'probe trial' following a 12 day delayed-matching to place training session (as in Figure 3c) to demonstrate that neither wild type nor mutant mice had an inherent bias for quadrant location (main effect of % time spent in each quadrant: $F_{(3, 39)} = 0.21$, p = 0.888).

In sum, these results indicate that exogenous D-serine can rescue spatial learning, but only when endogenous D-serine is limiting, in this case by a reduced NR1 affinity.

DISCUSSION

Exogenous D-serine selectively enhanced NMDA-NR2B receptor-dependent LTD at hippocampal CA3-CA1 synapses *in vitro*, and improved reversal learning in hippocampus-dependent behavioral tasks. Conversely, the NR2B-selective antagonist Ro 25-6981 disrupted both

hippocampal LTD and reversal learning with a concomitant increase in perseveration. These results point to an

The role of LTD in reversal learning

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increase in perseveration. These results point to an important role for NMDA-NR2B receptor-mediated plasticity in hippocampus-dependent reversal learning, and suggest that D-serine may improve cognitive flexibility by enhancing this process.

Specific Regulation of NMDA-NR2B-Dependent LTD by Extracellular D-Serine

The most parsimonious explanation for the specific enhancement of NMDA-NR2B-dependent LTD by exogenous D-serine is that normal endogenous D-serine levels are subsaturating in the proximity of NR2B-expressing receptors, but are near saturation at those NMDA receptors mediating the induction of LTP and depotentiation. D-serine is clearly the dominant physiological co-agonist for activation of NMDA receptors linked to LTP, as evidenced by the inhibitory effect of the catabolic enzyme *D*-amino acid oxidase and rescue by D-serine (Yang et al, 2003). Furthermore, exogenous D-serine rescued LTP and LTD in Grin1 mutants expressing a lower NMDA-NR1 D-serine affinity, indicating that exogenous D-serine application substantially increased the intra-synaptic D-serine concentration. That this treatment had no effect on submaximal LTP in wild-type mice indicates that *D*-serine is normally not the limiting factor for NMDA receptor activation during tetanic stimulation (Billups and Attwell, 2003).

Alternatively, D-serine is most likely subsaturating around NMDA-NR2B receptors that mediate LTD, so that exogenous D-serine can increase receptor activity. Receptors expressing NR2B subunits have a co-agonist affinity that is actually higher than those expressing only NR2A (Priestley et al, 1995), so any difference in the degree of saturation must be determined by extracellular concentration gradients. Astrocytic processes containing D-serine are directly opposed to the synapses and so are in close proximity to subsynaptic NMDA receptors (Schell et al, 1997). In the mature hippocampus, however, NR2B-containing receptors are thought to be primarily extra-synaptic (Clark and Cull-Candy, 2002) and so may be exposed to lower D-serine concentrations emanating from a relatively distal source. Alternatively, differential saturation could reflect a dynamic coupling of *D*-serine release to the frequency of synaptic activation, with more D-serine released by LTP-inducing high-frequency stimulation. Aside from synthesizing and storing D-serine (Schell et al, 1995, 1997; Mothet et al, 2000; Wolosker et al, 1999), perisynaptic astroglial processes express a variety of neurotransmitter receptors and ion channels, and so are well positioned to respond in a graded manner to transmitters and ions (Vesce *et al*, 1999; Scemes, 2000; Somjen, 2002). However, depotentiation was not altered by exogenous D-serine, indicating that synaptic D-serine levels are sufficient to saturate these NMDA-NR2B antagonist-insensitive receptors at low frequencies (1 Hz).

The Role of NR2B-Dependent Synaptic Plasticity in Hippocampus-Dependent Reversal Learning

The precise neurocellular mechanisms underlying hippocampus-dependent spatial learning are still debated, but activation of hippocampal NMDA receptors and the

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induction of multiple forms of NMDA-dependent synaptic plasticity appear necessary (Martin et al, 2000; Lynch, 2004). Our results indicate that the NMDA-NR2B subtype of NMDA receptor mediates a form of hippocampal synaptic depression that may contribute to spatial reversal learning and oppose behavioral perseveration. D-serine augmented NR2B-dependent hippocampal LTD and improved reversal learning in both a reversal version of a reference memory task, and in a hippocampus-dependent version of the delayed-matching-to place task (Steele and Morris, 1999). Conversely, blockade of NMDA-NR2B receptors inhibited both LTD and reversal learning, and significantly increased the time mice spent within the previous acquisition target. This behavior is referred to as perseveration, and significantly, is also observed after partial damage to the hippocampus or transection of the fibria-fornix (Whishaw and Tomie, 1997). Thus, while systemic injection of D-serine and Ro 25-6981 will, respectively, augment and suppress NMDA-NR2B receptor activity throughout the brain, it is likely that actions within the hippocampus account for the deficit in reversal learning and increased perseveration.

Evidence for a unique role of NMDA-NR2B receptors in synaptic plasticity and learning has been somewhat contradictory, possibly due to species-dependent differences and changes in receptor subtypes during development. Hippocampal overexpression of the NR2B subunit was shown to enhance learning in the MWM task in adult mice (Tang et al, 1999). Consistent with these transgenic studies, we observed that pharmacologic downregulation of NR2B receptor function disrupted spatial memory in wild type mice. In contrast, NR2B antagonists had no effect on spatial memory in rats (Guscott et al, 2003; Higgins et al, 2003), suggesting that encoding of spatial memories may be more dependent on other NMDA receptor isoforms in mice.

There is also considerable ambiguity regarding the differential contributions of NR2A- and NR2B-containing receptors to hippocampal LTP and LTD. The idea that NR2A and NR2B subtypes have strictly delineated roles in the induction of LTP and LTD respectively is very likely an oversimplification. NR2B receptors have been shown to participate in LTP induction at young mouse CA3-CA1 synapses (Berberich et al, 2005, 2007), as well as in other brain regions (Rodrigues et al, 2001; Weitlauf et al, 2005; Zhao et al, 2005). The NR2B-antagonist Ro 25-6981 blocked LTD in adult rats (Liu et al, 2004; Massey et al, 2004) but, conversely, did not block hippocampal LTD in very young rats in vitro (Bartlett et al, 2007; Morishita et al, 2006). Curiously, however, this drug inhibited LTD in young rats in vivo (Fox et al, 2006). A putative NMDA-NR2A-specific antagonist also blocked LTD (Fox et al, 2006) but has also been shown to inhibit NR1-NR2B receptors (Weitlauf et al, 2005; Frizelle et al, 2006). It should be emphasized that the subcellular distribution of NR2A and NR2B receptors changes during maturation of the hippocampus (Dumas, 2005), so direct comparisons between young rats or mice and the adult mice employed in our study are not possible. Furthermore, the different LTD induction protocols employed may engage different signaling pathways that could account for some of these discrepancies. The LTD induction method employed here (see Massey et al, 2004) induces a form of LTD that requires Ro 25-6981-sensitive NR2B receptors. Likewise, the θ burst protocol that closely mimics

the endogenous hippocampal firing pattern (O'Keefe, 1993) induces LTP that is NR2B receptor-independent. The strongest evidence that an NR2B-dependent LTD is involved in reversal learning in vivo is the opposing effects of D-serine and the NMDA-NR2B antagonist on both reversal learning and in vitro LTD. Disruption of reversal learning was concomitant with perseveration. In light of previous results showing that the same NMDA-NR2B antagonist promotes perseveration in a serial reaction task (Higgins et al, 2003), we suggest that a major function of NR2Bmediated LTD is to augment behavioral flexibility, independent of the specific cognitive requirements of the task.

How specifically could LTD and NMDA-NR2B receptors facilitate behavioral flexibility? By weakening certain synapses in response to changing environmental conditions, LTD could enhance the signal-to-noise, dynamic range, and plasticity of hippocampal circuits. Indeed, the propensity for LTD has been shown to change during exploration and learning. In rats, the threshold for LTD was lowered by exposure to a simple but novel environment, and when the positions of salient cues within that environment were changed (Manahan-Vaughan and Braunewell, 1999; Kemp and Manahan-Vaughan, 2004). Changing the location of a hidden platform may be analogous, requiring a refinement or reconfiguration of the previously formed spatial map. Similarly, calcineurin knockout mice show both diminished LTD and poor performance in the delayed-matching-to-sample task (Zeng et al, 2001), while environmental enrichment, the most studied form of cognitive enhancement, increases the propensity for LTP, LTD, and depotentiation/dedepression (Duffy et al, 2001; Abraham et al, 2006; Artola et al, 2006). Modulation of LTD threshold has also been associated with cognitive dysfunction; for example, chronic stress reduces the induction threshold for LTD and leads to cognitive impairments (Xu et al, 1997; Yang et al, 2005). However, our results, and those cited, indicate that LTD is not necessarily detrimental to explicit learning. Rather, whether LTD promotes or inhibits learning depends on the context of induction. Specific enhancement of LTD by D-serine would increase the dynamic range of hippocampal synapses, and so long as LTD and LTP induction occur in appropriate contexts, learning may be improved.

D-Serine in Psychiatric Disease

Recent evidence points to NMDA-receptor hypofunction mediated by low D-serine as a specific causative factor in some behavioral endotypes of schizophrenia (Mohn et al, 1999; Goff and Coyle, 2001; Chumakov et al, 2002). Treatments that can augment the synaptic concentrations of *D*-serine or glycine are, therefore, possible therapies (Javitt, 2004). Indeed, rodents treated with NMDA receptor antagonist show behavioral endotypes of schizophrenia that can be partially ameliorated by D-serine (Andersen and Pouzet, 2004; Lipina et al, 2005). The role of D-serine as a cognitive enhancer in the normal brain is not as well documented. Here we show that D-serine does not improve acquisition of spatial tasks in wild-type mice, but allows acquired information to be utilized more effectively under changing conditions. These findings underscore the potential of *D*-serine in the treatment of a range of behavioral

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DISCLOSURE/CONFLICT OF INTEREST

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