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 Alter, A. A., Lee, S. L., Pourfar, M., and Dobkin, G., J. clin. Invest., 41, 1341 (1962).
 Alter, A. A., Lee, S. L., Pourfar, M., and Dobkin, G., Blood, 22, 165 (1963).
 Trubowitz, S., Kirman, D., and Masek, B., Lancet, ii, 486 (1962).
 Brandt, N. J., Freland, A., Mikkelsen, M., Nielsen, A., and Tolstrup, N., Lancet, ii, 700 (1963).
 Krone, W., Wolf, U., Goedde, H. W., and Baitsch, H., Lancet, ii, 590 (1964).
 Mellman, W. J., Oski, F. A., Tedesco, T. A., Maciera-Coelho, A., and Harris, H., Lancet, ii, 674 (1964).
 Epstein, C. J., Science, 163, 1078 (1969).
 Tischfield, J. A., Creagan, R. P., and Ruddle, F. H., Cytogenet. Cell Genet., 13, 167 (1974).
 Long, C., Chan, T., Levytska, V., Kusano, T., and Green, H., Biochem. Genet., 9, 283 (1973).
 Giblett, E. R., Genetic Markers in Human Blood, 466 (Blackwell, Oxford and Edinburgh, 1969).
 Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J., J. biol. Chem., 193, 265 (1951).
 Tarlov, A. R., Brewer, G. J., Carson, P. E., and Alving, A. S., Archs intern. Med., 109, 209 (1962).
 Nyhan, W. L., Pesek, J., Sweetman, L., Carpenter, D. G., and Carter, C. H., Pediat. Res., 1, 5 (1967).
 Gartler, S. M., Scott, R. C., Goldstein, J. L., Campbell, B., and Sparkes, R., Science, 172, 572 (1971).
 Silvers, D. N., Cox, R. P., Balis, E., and Dancis, J., New Engl. J. Med., 286, 390 (1972).
 de Bruyn, C. H. M. M., Oei, T. L., and ter Haar, B. G. A., Clin. Genet., 5, 449 (1974).

- 390 (1972).
 de Bruyn, C. H. M. M., Oei, T. L., and ter Haar, B. G. A., Clin. Genet., 5, 449 (1974).
 Sinet, P.M., Allard, D., Lejeune, J., and Jerome, H., C. r. hebd. Séanc. Acad. Sci. Paris, D278, 3267 (1974).
 Tan, Y. H., Schneider, E. L., Tischfield, J., Epstein, C. J., and Ruddle, F. H., Science, 186, 61 (1974).

Why didn't Gregor Mendel find linkage?

IT is quite often said that Mendel was very fortunate not to run into the complication of linkage during his experiments. He used seven genes and the pea has only seven chromosomes. Some have said that had he taken just one more, he would have had problems. This, however, is a gross oversimplification. The actual situation, most probably, is shown in Table 1. This shows that Mendel worked with three genes in chromosome 4, two genes in chromosome 1, and one gene in each of chromosome 5 and 7. It seems at first glance that, out of the 21 dihybrid combinations Mendel theoretically could have studied, no less than four (that is, a-i, v-fa, v-le, fa-le) ought to have resulted in linkages. As found, however, in hundreds of crosses and shown by the genetic map of the pea¹, a and i in chromosome 1 are so distantly located on the chromosome that no linkage is normally detected. The same is true for ν or le on the one hand. and fa on the other, in chromosome 4. This leaves v-le, which ought to have shown linkage.

Mendel, however, seems not to have published this particular combination and thus, presumably, never made the appropriate

Table 1 Relationship between modern genetic terminology and character pairs used by Mendel

Character pair used by	Alleles in	Located in
Mendel	modern terminology	chromosome
Seed colour,		
yellow-green	I-i	1
Seed coat and flowers,		
coloured-white	A-a	1
Mature pods,		
smooth expanded-wrinkled		
indented ¹	V_{-v}	4
Inflorescences,		
from leaf axils-umbellate in		
top of plant	Fa–fa	4
Plant height,	•	
>1m-around 0.5 m	Le–le	4
Unripe pods,		
green-yellow	Gp– gp	5
Mature seeds,	2 01	
smooth-wrinkled	R-r	7

cross to obtain both genes segregating simultaneously. It is therefore not so astonishing that Mendel did not run into the complication of linkage, although he did not avoid it by choosing one gene from each chromosome.

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Blixt, S., in Handbook of Genetics, 2 (edit. by King, R. C.), (Plenum, New York, 1974).

Defect of macrophage function in the antibody response to sheep erythrocytes in systemic Mycobacterium lepraemurium infection

THE need for macrophages for optimal antibody responses, both in vivo and in vitro, to certain antigens is now established^{1,2} and this paper describes experiments which have used this requirement to demonstrate a deficiency of macrophage function in mice experimentally infected with the rodent leprosy bacillus, Mycobacterium lepraemurium. This organism is an obligate intracellular dweller and is found particularly inside cells of the macrophage series although in terminal infection other cell types may be invaded. Systemically infected mice characteristically show macrophages overloaded with bacilli, ever increasing numbers of granulomata and increasing spleno- and hepatomegaly³. There is an increase in the phagocytic activity of the spleen and liver at an early stage of infection (I.N.B. and V.S.S., in preparation) indicative of alterations in macrophage function and we report here experiments which show that the in vivo and in vitro antibody response to sheep erythrocytes (SRBC) is depressed at later stages of infection. The results indicate a defect of macrophage function.

Young adult CBA female mice were injected intravenously with 10° M. lepraemurium freshly isolated from the heavily infected livers of mice infected 4-6 months previously (I.N.B. and H. N. Krenzien, in preparation). At 11-15 weeks after infection, when the present experiments were carried out, the mice appeared clinically healthy but at autopsy showed gross enlargement of liver and spleen. Ziehl-Neelsen stain revealed the presence of numerous mycobacteria in macrophages of various tissues, particularly liver, spleen and bone marrow. Uninfected mice of the same age and sex were used as controls. The in vivo antibody response was measured in mice injected intravenously with either 5×10° or 10° SRBC. Four days later, the number of antibody producing cells (PFC) present in the spleen of each mouse was determined using the method of Cunningham⁴. The in vitro response of spleen cells cultured for 4d according to the method of Marbrook⁵ was measured in the same way.

The results given in Table 1 show that the antibody response in vivo was reduced 11 weeks after infection and even more so at 14 weeks. This depression was more pronounced in animals injected with the smaller number of SRBC. It seemed greater when the results were expressed as PFC per 10 spleen cells because there was a 4-5-fold increase in the cellularity of the spleens from infected compared with normal mice. Serum antibody levels were also reduced in infected mice (results not shown). Spleen cells from infected animals cultured in vitro in the presence of SRBC failed to generate the expected number of PFC, particularly at 14 weeks.