

continuously to facilitate entry of the instillate into each lobar and segmental bronchus. Each lamb was then quickly ventilated by hand once with 100% oxygen and then ventilated with room air (to avoid the complications of oxygen toxicity) at 21° and 50% humidity using a small animal volume respirator (Harvard). The initial respirator settings were with a rate of 60/min, a volume of 5 ml/kg (based upon the estimated body weight), a positive end expiratory pressure (PEEP) of 4 cm water, and an inspiration to total cycle ratio of 0.4. An umbilical artery catheter was placed and advanced to the abdominal aorta just below the diaphragm for purposes of recording blood pressure and heart rate and for monitoring blood gases and blood pH. The blood pressure was monitored continuously. The blood gases and pH were determined immediately and thereafter at least every 30 min.

The body temperature of each lamb was monitored with a rectal thermometer. In pilot experiments, the body temperature rapidly dropped to quite low levels, so in subsequent studies we employed both a heating blanket (Gaymar) under each fetus as well as an overhead heat lamp. By adjusting these, we were able to maintain the rectal temperature in the range of 37–38°. No attempt was made to correct the lamb's metabolic acidosis when it occurred. The respirator rates and volumes were altered in both twins, however, depending on the responses of the lambs including the results of the blood gases.

After 2 hr of ventilation, each lamb that was still alive was sacrificed by cisternal injection of 2% Xylocaine. Immediately after the death of the animal (whether natural or induced) and while the lamb was still attached to the respirator the endotracheal tube was clamped at peak inspiration. At autopsy, the trachea was ligated and the chest was opened from the abdomen. The lungs and heart were carefully removed en bloc. The heart was removed and the entire lung was weighed. Portions of the lungs were removed and used as follows: right upper lobe for histology, histochemistry, and biochemical analysis (for phospholipid composition of surfactant fraction and residual fraction); right lower lobe for surface tension measurements; infracardiac lobe for lung water content; and the left lung for pressure-volume measurements.

PREPARATION OF NATURAL SURFACTANT

Fresh 2- to 3-day-old lamb lungs were perfused free of blood with cold 0.9% saline. The trachea was cannulated and the lungs were washed repeatedly with cold saline as reported previously (22). The turbid fluid washings were centrifuged at $200 \times g$ at 20° for 7 min to remove the cell debris. The supernatant was further centrifuged at $1000 \times g$ at 2° for 1 hr. The suspensions were pooled and again centrifuged at $100 \times g$ at 2° for 1 hr. The pellet obtained was suspended in an equal volume of saline and resedimented at $200 \times g$ at 2° for 10 min to remove the cell debris. This procedure was repeated until no yellow cellular layer was visible at the bottom. The resultant white layer was referred to as "natural surfactant." With this procedure it was possible to obtain as much as 250 mg surfactant lipid from a 2-day-old newborn lamb.

CHEMICAL COMPOSITION OF NATURAL SURFACTANT

In order to determine the purity and reproducibility of the material isolated, compositional studies were performed in five preparations from eight newborn lambs. For lipid analysis, an aliquot of surfactant was lyophilized and the lipids were extracted with chloroform-methanol (2:1, vol/vol) and washed by the method of Folch *et al.* (21). The total lipids were determined gravimetrically using a Cohn M-10 electrobalance. The phosphorus was determined by the method of Bartlett (6).

Individual phospholipid classes were quantitated by phosphorus analysis after separation by two-dimensional thin layer chromatography on silica gel H-coated plates. The plates were developed in the first dimension using CHCl_3 -methanol-ammonia (65:35:5), and in the second dimension using CHCl_3 -acetone-methanol-acetic acid- H_2O (5:2:1:1:0.5). The lipids on the plates were visu-

alized using 2',7'-dichlorofluorescein spray under UV light. The saturated phosphatidyl choline was determined by the method of Shimojo *et al.* (31). The neutral lipid classes, after separation by thin layer chromatography in ether petroleum ether-acetic acid, were estimated gravimetrically using a Cahn M-10 electrobalance. Protein was determined by the method of Lowry *et al.* (27) using crystallized bovine serum albumin, fraction V, as the standard.

PRESSURE-VOLUME MEASUREMENTS ON ISOLATED LEFT LUNG

A metal cannula connected to a polyethylene tube 10 mm in diameter was inserted into the left main stem bronchus via the trachea and ligated tightly just below the level of the carina. The lung was degassed in a vacuum jar and placed in a saline water bath at 37°. The polyethylene tube was connected to a constant infusion pump (Harvard) with a syringe of 200 ml and to a water manometer by a T tube. The lung was inflated with air over 7–17 min to 30 cm H_2O pressure in increments of 2.4 ml/min to avoid the stress relaxation phenomenon, and then was deflated in a similar fashion. The volume reached at 30 cm H_2O pressure was assumed to be the total lung capacity.

Pressure-volume curves of volume of gas per gram of lung tissue were derived from the pressure-volume measurements and weight using a computer plotter Wang model 600. Gas volumes were determined by subtracting the tissue volumes measured by displacement in the water manometer. The gas volume per g lung tissue on deflation was used for comparison of the pressure-volume characteristics of each lung.

LUNG WATER CONTENT

The lung tissue of the infracardiac lobe was removed and weighed by a Right-A-Weight balance. The lung tissue was placed in an oven at 120° for 24 hr, after which it was again weighed. The water content was calculated as follows:

$$\text{Lung water content (\%)} = \frac{(\text{lung wet weight}) - (\text{lung dry weight})}{(\text{lung wet weight})} \times 100$$

LUNG SURFACE TENSION

Three grams of lung tissue were taken from the right lower lobe and were prepared and used for surface tension measurements as reported previously (22). The surface activity of the saline extract was measured on a modified Wilhelmy balance. The surface area was cycled until a stable curve was recorded, and the highest and lowest tensions noted on the last cycle were recorded as γ maximum and γ minimum. The stability index (12) was also calculated.

HISTOLOGIC AND HISTOCHEMICAL TECHNIQUES

After the lungs were weighed, surgical scissors were used to cut a slice of lung, 5 mm thick, in a plane at right angles to the right upper lobe bronchus. The tissue was dropped into Elftman's dichromate-sublimate fixative, and was processed according to Elftman's recommendations (14–16), as described by Emmel and Cowdry (17).

Serial paraffin sections were cut at 6 μ and were stained both with routine histologic stains and with Sudan black and nuclear fast red, for localization of phospholipids. Sections of the whole lobe were studied under the light microscope at all degrees of magnification.

RESULTS

GENERAL COMMENTS

The emphasis of this report is on the clinical and physiologic effects of surfactant. Only brief mention will be made of the biochemical, histologic and histochemical findings, since they will be described in greater detail in subsequent reports.

The immaturity of all the lambs was obvious for a number of reasons: their low body weights; the sparseness of their hair; the fusion of their eyelids, and their soft nailbeds. At the time of delivery, all lambs were active and in good condition as evidenced by normal umbilical vein blood pH, pO₂, and pCO₂. Without exception, the "shake test" for surface activity was negative on the amniotic fluid and tracheal fluid of all the lambs. Furthermore, the amount of phosphatidyl choline in the surfactant fraction isolated from the tracheal fluid was very low (0.07 mg/100 ml). Thus, none of the lambs studied had more than a trace amount of surfactant in the tracheal and amniotic fluids at the time of delivery.

AMOUNT OF NATURAL SURFACTANT INSTILLED

The amounts of natural surfactant instilled into the lungs of each of the experimental lambs are shown in Table 1. The total lipids administered per kg body wt varied from 113–229 mg (mean 173) and the total lipids per g lung wet wt from 2.5–5.8 mg (mean 4.0). The total protein content contained in the surfactant administered varied from 40–76.9 mg (mean 56.5).

CHEMICAL COMPOSITION OF NATURAL SURFACTANT

The chemical composition of the natural surfactant used is shown in Table 2. These determinations were made on an aliquot of the material from eight newborn lambs 2–3 days of age. The average amount of total lipid was 18.5 mg/ml: 79% phospholipid and 54% saturated phosphatidyl choline. The average protein content was 4.1 mg/ml, giving a lipid protein ratio of approximately 4:1. Phosphatidyl choline comprised 84% of the total phospholipid, and saturated phosphatidyl choline (predominantly dipalmitoyl lecithin) accounted for 62% of the phospholipid.

EFFECTS OF SURFACTANT ON CLINICAL PARAMETERS

The administration of natural surfactant had a striking effect on the clinical course of the experimental lambs in contrast with the control twins that received saline or distilled water. As shown in Table 3, all of the lambs that received surfactant survived for the full 2 hr, whereas all of the control lambs died prematurely, usually before 60 min (average = 48 min).

Experimental lambs showed some spontaneous movement and responded promptly to painful stimuli, whereas control animals had no spontaneous movements with or without stimuli. On auscultation of the chest experimental lambs had predominantly vesicular breath sounds whereas control animals had bronchial breath sounds.

Figures 1, 2, and 3 show a plot of the arterial pH, pO₂, and pCO₂ against time for both the experimental and control lambs. All control lambs died with severe acidosis (pH below 7.0) and

most had hypercarbia and hypoxemia. In contrast, the experimental lambs had acceptable arterial blood pH, pO₂, and pCO₂, considering that they received no bicarbonate and were ventilated on room air.

An attempt was made to ventilate all the lambs in a similar fashion with similar volumes of air (5 ml/kg). In all pairs of twins, the peak ventilation pressure of the surfactant treated lamb was lower than the control (Table 3 and Figure 4); in many, it was approximately one-half the pressure used in the control lamb ($P < 0.001$). Using the formula:

$$\text{Effective compliance} = \frac{\text{peak lung volume}}{\text{peak inspiratory pressure} - \text{PEEP}}$$

the calculated effective compliance was strikingly different between the experimental and control lambs. Furthermore, the compliance of the lambs that received surfactant seemed to improve with time as shown in Figure 5.

AUTOPSY APPEARANCE OF LUNGS

The lungs of control animals were liver-like in appearance without exception. (Plate 1) The lungs of all lambs that received surfactant were aerated and their pleural surfaces looked like those of normal, full term breathing lambs. Where aeration was patchy the units that were either inflated or that were not, always followed the pattern of "pneumons" (26, 33).

LUNG WATER CONTENT

The lungs of both the experimental and control lambs (Table 3) remained wet (87–90%) and were in the range of values observed in fetal life (3). Thus, the administration of surfactant did not

Table 2. Chemical composition of natural surfactant

Total lipid	18.5 ± 5.3 mg/ml	
Protein	4.1 ± 1.2 mg/ml	
Lipid/protein ratio	4.0 ± 1.7	
Phospholipid	18.8 ± 3.6 μmol/ml	
Phosphatidyl choline	15.7 ± 2.2 μmol/ml	84.1 ± 4.1% of P
Saturated phosphatidyl choline	12.9 ± 1.4 μmol/ml	61.5 ± 7.8% of P
Phosphatidyl ethanolamine	1.0 ± 0.2 μmol/ml	4.6 ± 0.7% of P
Sphingomyelin	0.4 ± 0.05 μmol/ml	1.8 ± 0.3% of P
Phosphatidyl glycerol	0.9 ± 0.4 μmol/ml	4.4 ± 1.8% of P
Phosphatidyl inositol	0.09 ± 0.001 μmol/ml	0.4 ± 0.01% of P
Phosphatidyl serine	0.05 ± 0.04 μmol/ml	0.3 ± 0.2% of P

Table 1. Amount of natural surfactant instilled¹

Exp. no.	Body wt, kg	Lung wt, g	Total lipids, mg	Total PL, mg	Total Sat PC, mg	Total lipids/kg BW, mg	Total lipids/g LW, mg	Total protein, mg
452	1.7	82	375	322	153	221	4.6	53.7
453	2.0	92	282	266	118	141	3.1	70.1
454	1.7	90	277	252	118	163	3.1	76.9
458	1.8	73	272	245	115	151	3.7	54.6
459	1.2	54	136	113	54	113	2.5	40.0
460	1.7	58	306	254	122	180	5.3	75.0
461	2.0	89	275	208	104	138	3.1	48.0
462	1.2	61	275	208	104	229	4.5	48.0
465	1.5	65	265	202	107	177	4.1	48.8
466	1.2	46	265	191	113	221	5.8	50.4
Mean	1.6	71	273	226	111	173	4.0	56.6
± SE	0.1	5.2	18.4	17.6	7.7	12.6	0.3	4.0

¹ BW: body weight; LW: lung weight; PL: phospholipid; Sat PC: saturated phosphatidyl choline.

Table 3. *Effects of surfactant or diluent on clinical and autopsy parameters in premature twin lambs*

Exp no.	Body wt, kg	Treatment	Vent. pressure, cm H ₂ O	Survival time, min	Lung H ₂ O, %	Vol in ml/g at 7 cm pressure	Minimum surface tension, dynes/cm	Stability index
452								
A	2.1	Saline	33/8	97	88	0.1	29	0.5
B	1.7	Surfactant	30/7	120	86		13	1.1
453								
A	2.0	Surfactant	25/7	120	88	0.5	5	1.6
B	2.3	Saline	40/5	35	90	0.1	22	0.7
454								
A	1.7	Saline	28/5	34	87	0.1	31	0.4
B	1.7	Surfactant	17/6	120	89	1.1	4	1.7
458								
A	1.8	Surfactant	20/4	120	87	0.9	2	1.8
B	1.8	Saline	28/4	37	88	0.1	21	0.8
459								
A	1.3	Saline	35/5	49	87	0.1	34	0.4
B	1.2	Surfactant	20/5	120	88	1.3	11	1.2
460								
A	1.7	Surfactant ¹	23/5	120	88	1.1	8	1.4
B	1.4	Saline	28/4	91	89	0.1	25	0.7
461								
A	2.0	Water	36/4	27	89	0.1	32	0.5
B	2.0	Surfactant ¹	19/3	120	89	1.2	2	1.8
462								
A	1.2	Surfactant ¹	20/7	120	90	1.4	5	1.7
B	1.4	Water	33/5	25	89	0.1	35	0.4
C	1.1	Nil		0	89	0.1	36	0.4
465								
A	1.5	Water	29/5	23	89	0.1	34	0.4
B	1.5	Surfactant ¹	20/5	120	88	1.3	6	1.6
C	1.3	Nil		0	89	0	35	0.4
466								
A	1.2	Surfactant ¹	19/5	120	88	0.6	2	1.8
B	1.3	Water	37/5	42	87	0.1	35	0.5

¹ Surfactant in water.

seem to promote removal of lung fluid, although the histologic appearances suggested that fluid was redistributed from alveolar spaces to connective tissue and lymphatics.

PRESSURE-VOLUME CHARACTERISTICS OF LUNG

It was possible to measure the pressure-volume characteristics on the lungs of all lambs except 452B where an air leak occurred. As shown in Table 3, the volume of air (in milliliters per g wet lung tissue at 7 cm H₂O pressure on the deflation limb) was 2–10 times greater in the lambs that received surfactant ($P < 0.001$). The mean deflation limb values for the two groups of lambs are shown in Figure 6. The shape of the deflation pressure-volume curve at pressure = 0 in surfactant treated lambs suggests air trapping and/or reduced lung recoil.

LUNG SURFACE TENSION

Without exception, the lambs that received surfactant had a low minimum surface tension on the Wilhelmy balance and a high calculated stability index (Table 3, Fig. 7). In contrast, the lungs of control lambs had a high minimum surface tension and a low calculated stability index ($P < 0.001$). The values for the control lambs were similar to those obtained in infants who had died of hyaline membrane disease (5).

HISTOLOGIC AND HISTOCHEMICAL FINDINGS

The five serial sections from each right upper lobe were studied under the light microscope, and microphotographs were often taken at all levels of magnification using objective lenses of $\times 1$, $\times 10$, $\times 25$, $\times 40$, and $\times 100$. Observations were made, and results

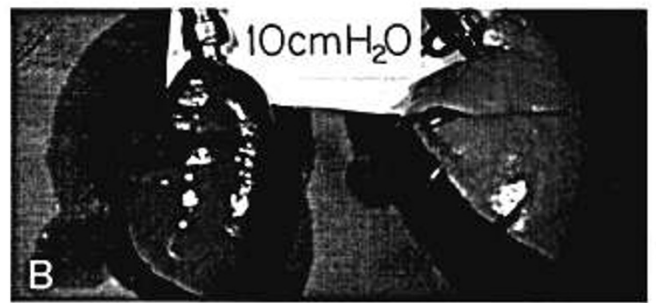
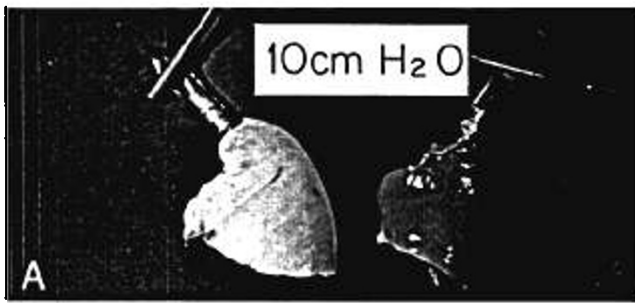
recorded both in words and on a 4-point scale without initial reference to whether or not the specimens were from a control or a surfactant-treated animal. The observed differences, however, were so uniformly striking that the diagnosis was quickly obvious, even though the section stained for phospholipid (which invariably showed large quantities of the instilled surfactant) was left to the last.

The histologic and histochemical findings will be reported in detail elsewhere. A few representative photomicrographs are shown in Plate 1. In contrast with the control lambs, those that received surfactant showed: good alveolar expansion, less obvious lung fluid, no hyaline membranes, and virtually no peripheral epithelial cell damage.

DISCUSSION

Our strikingly favorable results on the prevention of respiratory distress syndrome in premature lambs by the endotracheal instillation of natural surfactant prior to the first breath are consistent with the earlier observations of Enhörning and his collaborators (18–20).

In some respects, the studies of Enhörning and his colleagues were not comparable to ours. In their first report (20), the rabbit fetuses were dead at the time surfactant was instilled and the values given for the volume of air introduced into the lung were obtained on the inflation limb of the pressure-volume measurements rather than the standard way on the deflation limb. Furthermore, there was considerable overlap in the results they obtained in the controls as compared with those receiving surfactant. In their second report (18), the rabbit fetuses were delivered after the death of the mother and it took 3 min to cannulate each fetus.



#458 A B
SURFACTANT SALINE

#459 A B
SALINE SURFACTANT

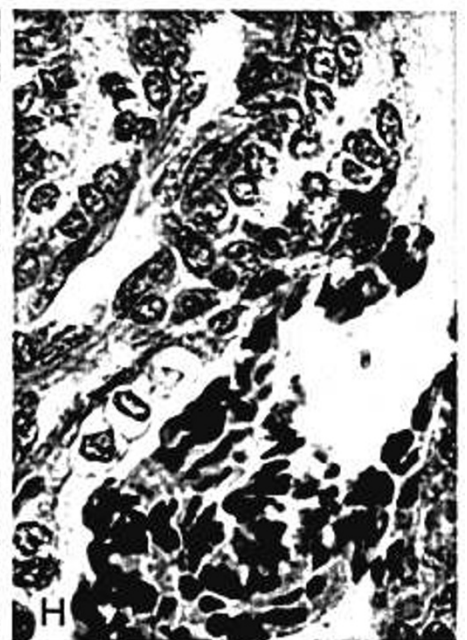
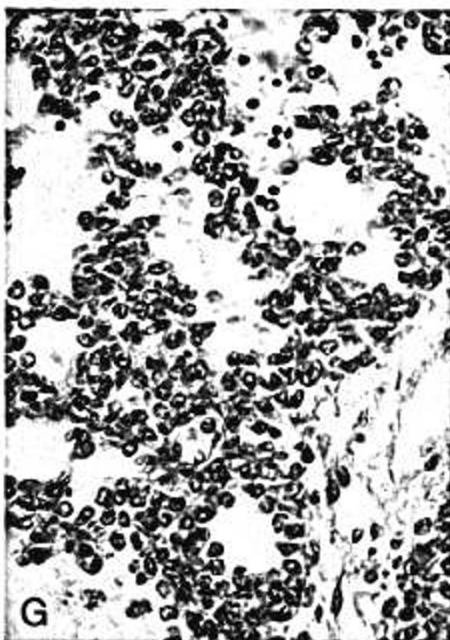
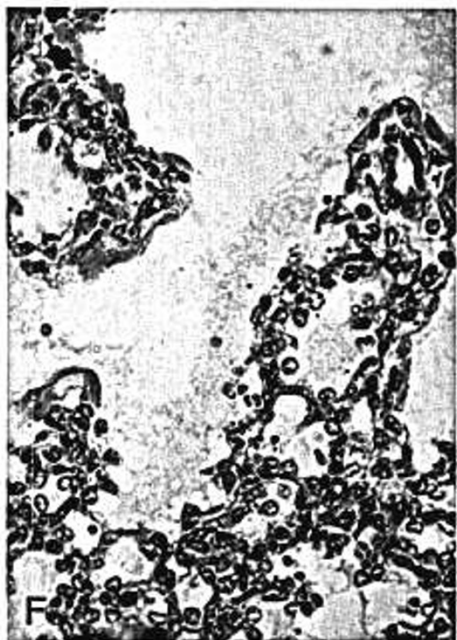
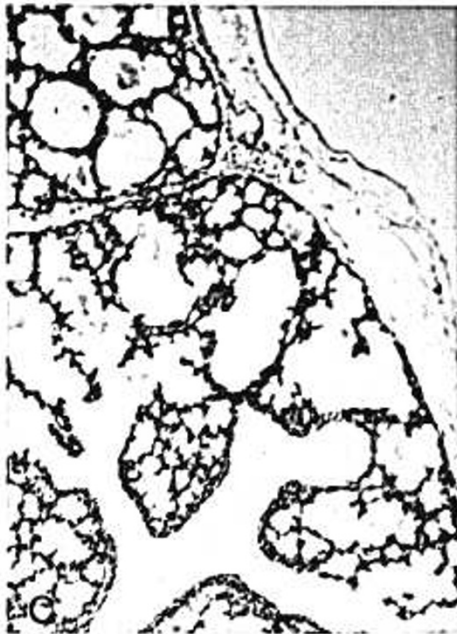


Plate 1. Photographs A and B from experiments 458 and 459 show the typical appearances of the left lungs inflated to 10 cm H₂O pressure. The lungs of the surfactant treated lambs are well inflated whereas the lungs of the control lambs appear "liver-like." Photomicrographs C and D are adjacent sections of lung from surfactant-treated lamb 458A. In C, note the even expansion of the terminal bronchioles and alveoli and normal epithelium (Masson's stain; $\times 35$). In D, note the deposition of phospholipid (black) in the lumina of two adjacent pneumons (Sudan black and nuclear fast red stain; $\times 35$). Photomicrographs E, F, G, and H are from control lambs (survival times in parentheses) 466B (42), 452A (97), 461A (27), and 458B (37), respectively. In E, note poor peripheral expansion and dilatation of small airways proximal to sites of epithelial cell plugging in respiratory and terminal bronchioles (H and E stain; $\times 35$). In F, note typical hyaline membranes at points of division of bronchioles and alveolar ducts (H and E stain; $\times 140$). In G, note epithelial cells with pycnotic nuclei lying both in alveolar walls and free in the lumina. At the top is a clump of dying cells in the lumen of a bronchiole (H and E stain; $\times 140$). In H, note the clump of dying cells in the lumen of a bronchiole whose epithelium looks healthy above and less so below (H and E stain; $\times 350$). Damaged epithelial cells are present peripherally after 27 min and typical hyaline membranes after 90 min of respirator therapy on room air.

In our study, both the mother and the fetus were in good condition at the time of tracheal cannulation.

It would appear from rough calculation that we used considerably more surfactant in our studies than that reported by Enhörning *et al.* (18, 20). On an average we used 141.9 μg total phospholipid/g body wt compared to their value of 5.1 $\mu\text{g}/\text{g}$ body wt. Thus, our more favorable results may be in part due to the facts that our animals received larger amounts of surfactant and were artificially ventilated.

We were concerned over the possible adverse effects that the diluent might have on removal of alveolar and bronchial fluid since it is known that in dogs, endotracheally instilled saline disappears much more slowly from the lungs into the circulation than does plain water (29). In our small series, it did not seem to make a significant difference whether saline or distilled water was used as the diluent. Surfactant in saline or water seemed to produce less fluid microscopically in the alveoli and terminal bronchioles; however, both control and experimental lambs possessed very wet lungs (Table 3) similar to the fetal state (3). This

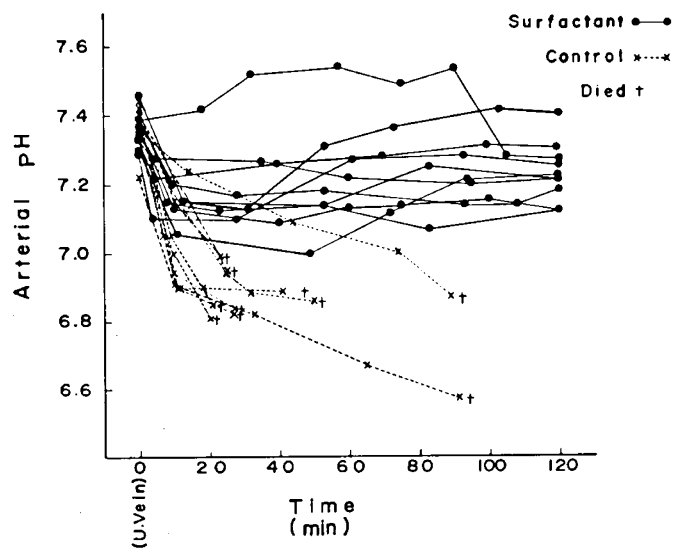


Fig. 1. Changes in arterial pH of surfactant-treated and control lambs. Note, no bicarbonate was given to either group.

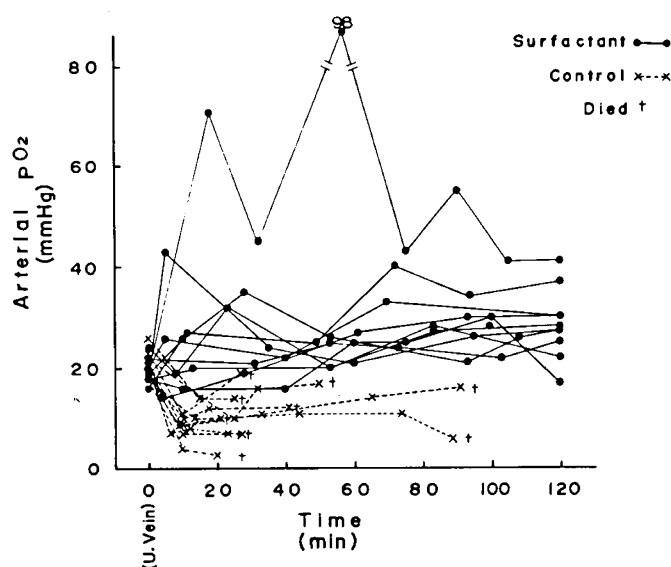


Fig. 2. Changes in arterial pO_2 on room air of surfactant-treated and control lambs.

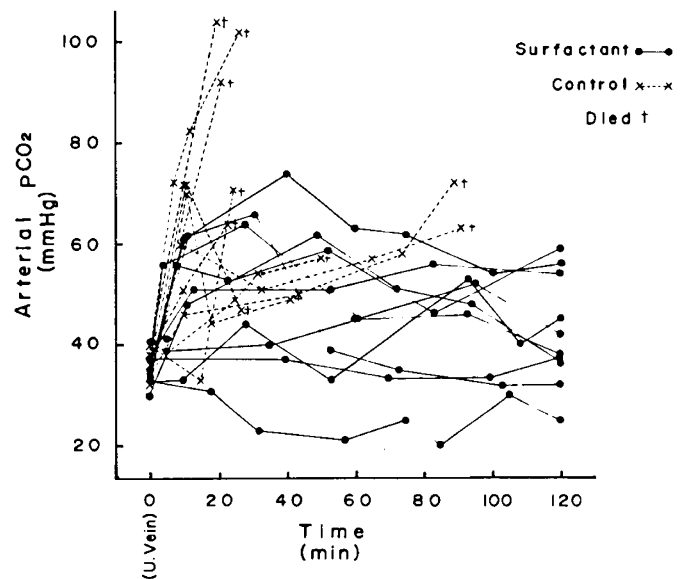


Fig. 3. Changes in arterial pCO_2 of surfactant-treated and control lambs.

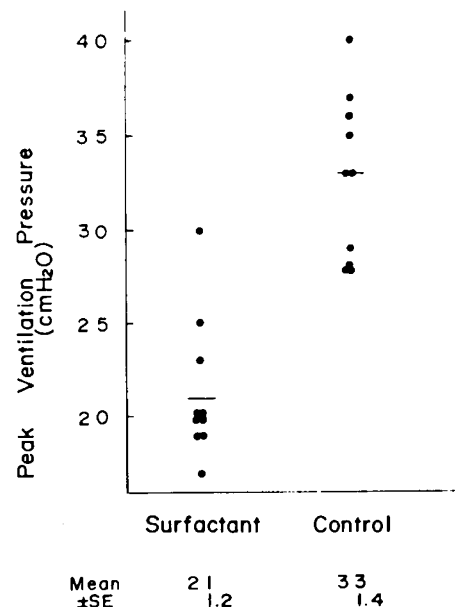


Fig. 4. Final peak ventilation pressure on respirator of surfactant-treated and control lambs (—: mean).

is not surprising since it is known that fetal lung fluid disappears slowly over a period of hours and days rather than minutes (4).

The prematurely delivered fetus has several additional features that puts it at a disadvantage in terms of the Starling equilibrium relative to fluid movement across a membrane: the colloid osmotic pressure of the plasma is known (28) to be low (12–16 torr in several of our experiments); the calculated pulmonary vascular resistance is higher than in adults (28); and most human premature infants have rather large left to right ductal shunts that can in turn produce pulmonary congestion and edema (32).

We are convinced that our immature lambs would not have survived without assisted ventilation. Very immature human infants also need such assisted ventilation frequently for long periods of time. Animal studies suggest the importance of PEEP and the need to keep the peak inspiratory pressure low (34). This was done in our studies. It may take some time, however, before the optimal mode of ventilation has been determined (7).

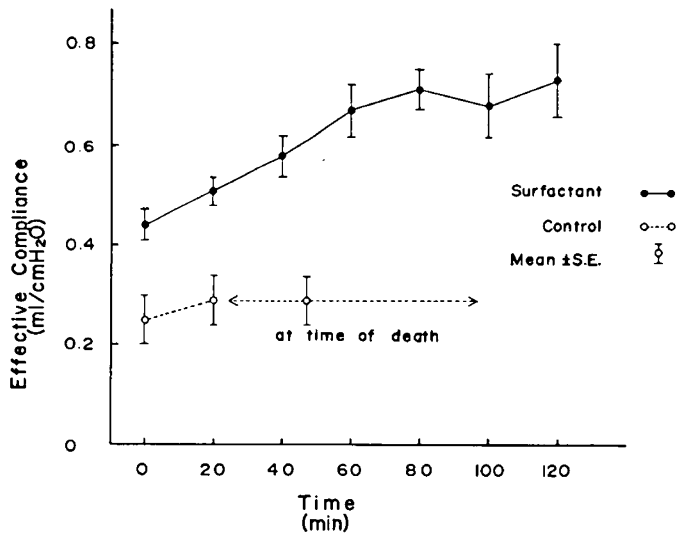


Fig. 5. Changes in effective lung compliance of surfactant-treated and control lambs.

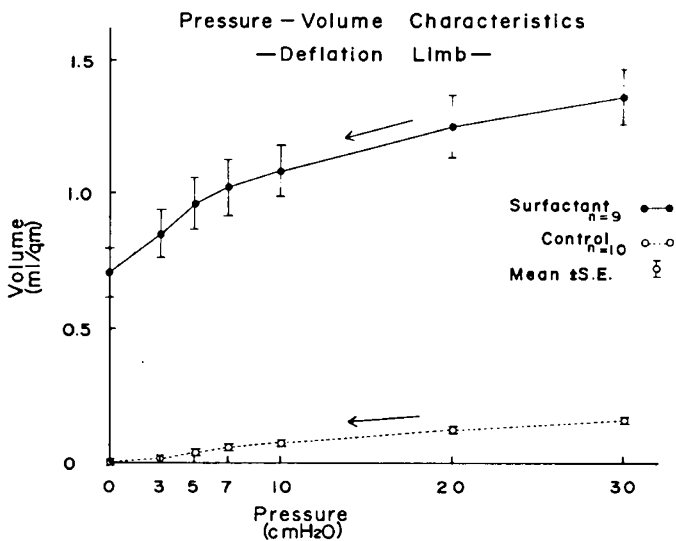


Fig. 6. Changes in pressure-volume characteristics on deflation limb of surfactant-treated and control lambs.

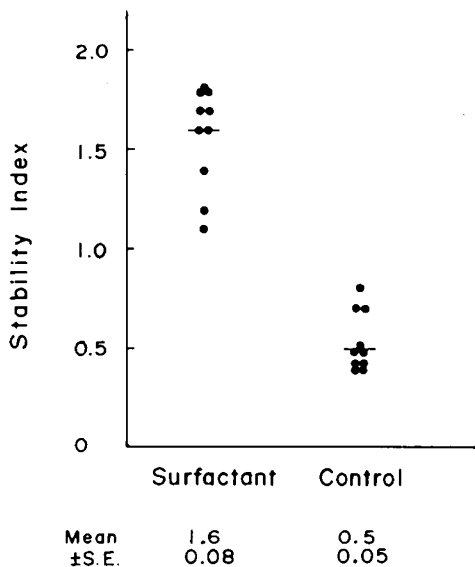


Fig. 7. Calculated stability index values from lung surface tension measurements of surfactant-treated and control lambs.

As stated earlier, the synthesis and secretion of surfactant by the cells of the fetal lung have been well studied in animals and humans (1, 3, 8, 22-24, 33). It is also known that a large amount of surfactant is secreted into the alveolar space of the newborn after the onset of respiration (1, 3, 23, 24). Since the lung fluid of our fetal lambs contained only trace amounts of surfactant, and since the control lambs died so quickly, it can be assumed that our lambs were very immature and possessed very little releasable surfactant.

In our study each set of twin lambs was treated in a similar fashion except that one received natural surfactant and the other received diluent (saline or water). Except for the variation in the amount of surfactant that the experimental lambs received (Table 1), ours is a controlled study. Further investigations are still indicated however to answer such questions as: what is the optimal amount of surfactant that should be used; how long is a single dose of surfactant effective; how can surfactant best be administered after the onset of breathing; and is natural surfactant antigenic?

REFERENCES AND NOTES

- Adams, F. H., Fujiwara, T., Emmanouilides, G. C., and Riih , N.: Lung phospholipids of human fetuses and infants with and without hyaline membrane disease. *J. Pediat.*, 77: 833 (1970).
- Adams, F. H., Fujiwara, T., Emmanouilides, G. C., and Scudder, A.: Surface properties and lipids from lungs of infants with hyaline membrane disease. *J. Pediat.*, 66: 357 (1965).
- Adams, F. H., Fujiwara, T., and Latta, H.: "Alveolar" and whole lung phospholipids of premature newborn lambs. *Biol. Neonat.*, 217: 198 (1971).
- Adams, F. H., Yanagisawa, M., Kuzela, D., and Martinek, H.: The disappearance of fetal lung fluid following birth. *J. Pediat.*, 78: 837 (1971).
- Avery, M. E., and Mead, J.: Surface properties in relation to atelectasis and hyaline membrane disease. *Amer. J. Dis. Child.*, 97: 517 (1959).
- Bartlett, G. R.: Phosphorus assay in column chromatography. *J. Biol. Chem.*, 234: 466 (1959).
- Belenky, D. A., Orr, R. J., Woodrum, D. E., and Hodson, W. A.: Is continuous transpulmonary pressure better than conventional respiratory management of hyaline membrane disease? A controlled study. *Pediatrics*, 58: 800 (1976).
- Brumley, G. W., Chernick, V., Hodson, A., Normand, C., Penner, A., and Avery, M. E.: Correlation of mechanical stability, morphology, pulmonary surfactant and phospholipid content in the developing lamb lung. *J. Clin. Invest.*, 46: 863 (1967).
- Brumley, G. W., Hodson, W. A., and Avery, M. E.: Lung phospholipids and surface tension correlations. *Pediatrics*, 40: 13 (1967).
- Chu, J., Clements, J., Cotton, E., Klaus, M., Sweet, A., Thomas, M., and Tooley, W.: The hypoperfusion syndrome. *Pediatrics*, 35: 733 (1965).
- Chu, J., Clements, J. A., Cotton, E. K., Klaus, M. H., Sweet, A. Y., and Tooley, W. H.: Neonatal pulmonary ischemia. *Pediatrics*, 40: 709 (1967).
- Clements, J. A., Hustead, R. F., Johnson, R. P., and Grietz, I.: Pulmonary surface tension and alveolar stability. *J. Appl. Physiol.*, 16: 444 (1961).
- Clements, J. A., Platzker, A. C., et al.: Assessment of the risk of the respiratory distress syndrome by a rapid test for surfactant in amniotic fluid. *New Engl. J. Med.*, 286: 1077 (1972).
- Elftman, H.: Controlled chromatation. *J. Histochem. Cytochem.*, 2: 18 (1954).
- Elftman, H.: Phospholipid fixation by dichromate-sublimate. *Stain Technol.*, 32: 29 (1957).
- Elftman, H.: Effects of fixation in lipid histochemistry. *J. Histochem. Cytochem.*, 6: 317 (1958).
- Emmel, V., and Cowdry, E.: *Laboratory Technique in Biology and Medicine*, Ed. 4, p. 108, (Williams & Wilkins, Baltimore, 1964).
- Enh rning, G., Grossman, G., and Robertson, B.: Tracheal deposition of surfactant before the first breath. *Amer. Rev. Resp. Dis.*, 107: 921 (1973).
- Enh rning, G., Grossman, G., and Robertson, B.: Pharyngeal deposition of surfactant in the premature rabbit fetus. *Biol. Neonate*, 22: 126 (1973).
- Enh rning, G., and Robertson, B.: Lung expansion in the premature rabbit fetus after tracheal deposition of surfactant. *Pediatrics*, 50: 58 (1972).
- Folch, J., Lees, M., and Sloane-Stanley, G. H.: A simple method for the isolation and purification of total lipids from animal tissue. *J. Biol. Chem.*, 226: 497 (1959).
- Fujiwara, T., Adams, F. H., Sipos, S., and Salawy, A. E.: "Alveolar" and whole lung phospholipids of the developing fetal lamb. *Amer. J. Physiol.*, 215: 375 (1968).
- Fujiwara, T., Adams, F. H., Sipos, S., and Salawy, A. E.: "Alveolar" and whole lung phospholipids of newborn lambs. *Proc. Soc. Exp. Biol. Med.*, 127: 962 (1968).
- Gluck, L., Montoyama, E. K., Smits, H. L., and Kulovick, M. V.: The biochemical development of surface activity in mammalian lung. *Pediat. Res.*, 1: 237 (1967).
- Ikegami, M., Hesterberg, T., Nozaki, M., and Adams, F. H.: Restoration of lung-pressure-volume characteristics with surfactant: Comparison of nebulization versus instillation and natural versus synthetic surfactant. *Pediat. Res.*, 11: 178 (1977).
- Lauweryns, J. M., and Rosan, R. C.: The unit lobule: A revised concept of the

- neonatal lung. Proceedings of the Second European Congress of Perinatal Medicine, London, 1970, p. 259 (Karger, Basel, 1971).
27. Lowry, O. H., Rosebrough, N. J., Farr, A. L., and Randall, R. J.: Protein measurement with the Folin phenol reagent. *J. Biol. Chem.*, *193*: 265 (1951).
 28. Meschia, G.: Colloidal osmotic pressures of fetal and maternal plasmas of sheep and goats. *Amer. J. Physiol.*, *181*: 1 (1955).
 29. Pearce M. L.: Sodium recovery from normal and edematous lungs studied by indicator dilution curves. *Circ. Res.*, *24*: 815 (1969).
 30. Shannon, D. C., Kazemi, H., Merrill, E. W., Smith, K. A., and Wong, P. S.: Restoration of volume-pressure curves with a lecithin fog. *J. Appl. Physiol.*, *28*: 470 (1970).
 31. Shimojo, T., Abe, M., and Ohta, M.: A method for determination of saturated phosphatidyl choline. *J. Lipid Res.*, *15*: 525 (1974).
 32. Thibeault, D. W., Emmanouilides, G. C., Nelson, R. J., Lachman, R. S., Rosengart, R. M. and Oh, W.: Patent ductus arteriosus complicating the respiratory distress syndrome in preterm infants. *J. Pediat.*, *86*: 120 (1975).
 33. Towers, B.: The fetal and neonatal lung. In: N. S. Assali: *The Biology of Gestation, Vol. II: The Fetus and Neonate*, p. 189 (Academic Press, New York, 1968).
 34. Webb, H., and Tierney, D. F.: Experimental pulmonary edema due to intermittent positive pressure ventilation with high inflation pressures: Protection by positive end-expiratory pressure. *Amer. Rev. Resp. Dis.*, *110*: 556 (1974).
 35. The authors are indebted for help and advice and wish to thank the following: Donald F. Tierney, Bertrand Shapiro, Goran Enhörning, Bengt Robertson, Alex Sevanian, Takafumi Nagatomo, Tom Hesterberg, and John Zeller.
 36. This research was supported by funds from the USPHS and the National Cystic Fibrosis Research Foundation.
 37. Requests for reprints should be addressed to: Dr. Forrest H. Adams, Department of Pediatrics, Division of Cardiology, University of California School of Medicine, The Center for the Health Sciences, Los Angeles, CA 90024 (USA).
 38. Received for publication September 9, 1977.
 39. Accepted for publication October 26, 1977.