Supplementary Methods

Antibodies
Primary and secondary antibodies used for immunostaining were as follows; mouse monoclonal antibodies against myc-tag (1:500; Biomol), Calbindin-28kD (1:1000; Swant), vimentin (1:800; Sigma), MHC (MF20) (1:20; Developmental Studies Hybridoma Bank), rabbit polyclonal antibodies against HA-epitope (1:500; Upstate), GFAP (1:1000; Chemicon), BLBP (1:1000; kindly provided by M. Watanabe), GLAST (1:1000; Covalab), DNER\(^{17}\) (1:500), Pax6 (1:1000; Chemicon), goat polyclonal antibody against Notch1 (1:100; Santa Cruz) and Alexa488 or Alexa568-conjugated anti-rabbit, anti-mouse or anti-goat secondary antibodies (1:400, Molecular Probes).

Golgi staining
Cerebella were dissected out at P18. They were immediately immersed into Golgi-rapid fixative solution, composed of 5% potassium dichromate, 5% chloral hydrate, 5% formalin and 1.25% glutaraldehyde, and kept at room temperature for 2 to 4 days with a change of solution. They were then briefly washed with 1.5% silver nitrate and kept in fresh 1.5% silver nitrate solution for 2 days with a change of solution. They were dehydrated through a graded ethanol series, embedded in polyester wax and cut coronally at 50-80 μm in thickness. Sections were mounted onto glass slides, coverslipped and observed under a LM.

BrdU incorporation
Nine-day-old mice were injected subcutaneously with 50 mg/kg of 5-bromo-2-deoxyuridine (Sigma). Three hours or two days after injection, they were cryoanesthetized and perfused with 4% paraformaldehyde. Cerebella were then dissected out and sagittal cryosectioned at 14 μm. Sections were permeabilized in 0.4% Triton X-100, treated with 2 M HCl for 30 min at 37°C and washed twice in 0.1 M sodium borate. They were immunostained with anti-BrdU antibody (Roche) with or without anti-BLBP antibody.

EGL explant culture
Reaggregate cultures of the EGL were performed as previously described by Kawaji et al. (2004). The EGLs from postnatal day 4 to 5 mice were dissected, and fractions containing small neurons were collected and transfected with pCA-EGFP. The transfected cells were centrifuged in an Eppendorf tube and incubated for 1 hour to form reaggregates. The resultant aggregates were cut into 300-400 μm pieces and placed on glass-bottom culture dishes (Asahi Techno Glass, Tokyo, Japan) coated with poly-D-lysine/laminin. Cultures were maintained in serum-free media at 37°C in 5% CO\(_2\) and analyzed after 4 days.