

Single-molecule sequencing of the desiccation-tolerant grass *Oropetium thomaeum*

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Plant genomes, and eukaryotic genomes in general, are typically repetitive, polyploid and heterozygous, which complicates genome assembly¹. The short read lengths of early Sanger and current next-generation sequencing platforms hinder assembly through complex repeat regions, and many draft and reference genomes are fragmented, lacking skewed GC and repetitive intergenic sequences, which are gaining importance due to projects like the Encyclopedia of DNA Elements (ENCODE)². Here we report the whole-genome sequencing and assembly of the desiccation-tolerant grass *Oropetium thomaeum*. Using only single-molecule real-time sequencing, which generates long (>16 kilobases) reads with random errors, we assembled 99% (244 megabases) of the *Oropetium* genome into 625 contigs with an N50 length of 2.4 megabases. *Oropetium* is an example of a 'near-complete' draft genome which includes gapless coverage over gene space as well as intergenic sequences such as centromeres, telomeres, transposable elements and rRNA clusters that are typically unassembled in draft genomes. *Oropetium* has 28,466 protein-coding genes and 43% repeat sequences, yet with 30% more compact euchromatic regions it is the smallest known grass genome. The *Oropetium* genome demonstrates the utility of single-molecule real-time sequencing for assembling high-quality plant and other eukaryotic genomes, and serves as a valuable resource for the plant comparative genomics community.

The genomes of *Arabidopsis*³, rice⁴, poplar, grape and *Sorghum*⁵ were first sequenced using high-quality and reiterative Sanger-based approaches producing a series of 'gold standard' reference genomes. The advent of next-generation sequencing (NGS) technologies reduced costs of sequencing substantially, which has enabled sequencing of over 100 plant genomes¹. The quality of plant genome assemblies depends on genome size, ploidy, heterozygosity and sequence coverage, but most NGS-based genomes have on the order of tens of thousands of short contigs distributed in thousands of scaffolds. The short read lengths of NGS, inherent biases and non-random sequencing errors have resulted in highly fragmented draft genome assemblies that are not complete, which means they are missing biologically meaningful sequences including entire genes, regulatory regions, transposable elements, centromeres, telomeres and haplotype-specific structural variations. It is becoming clear from ENCODE projects that complete genomes are needed to better understand the importance of the non-coding regions of genomes².

More than 40% of calories consumed by humans are derived from grasses, and the grass family (Poaceae) is arguably the most important plant family with regard to global food security⁶. The size and complexity of most grass genomes has challenged progress in gene discovery

and comparative genomics, although draft genomes are now available for most agriculturally important grasses¹. The largest genome assemblies, such as maize (2,300 megabases (Mb))⁷, barley (5,100 Mb)⁸ and wheat (hexaploid, 17,000 Mb)⁹ are highly fragmented as a result of the inability of current sequencing technologies to span complex repeat regions. Near-finished reference genomes are available for rice⁴, *Sorghum*⁵ and *Brachypodium*¹⁰, but more high-quality grass genomes are needed for comparative genomics and gene discovery. Here we present the 'near-complete' draft genome of the grass *Oropetium thomaeum*, the first high-quality reference genome from the Chloridoideae subfamily. The draft genome is near complete because we were able to sequence through complex repeat regions that are unassembled in most draft genomes. *Oropetium* has the smallest known grass genome at 245 Mb and is also a resurrection plant that can survive the extreme water stress such as loss of >95% of cellular water (Fig. 1)¹¹.

Single-molecule real-time (SMRT) sequencing (Pacific Biosciences) produces long and unbiased sequences, which enables assembly of complex repeat structures and GC- and AT-rich regions that are often unassembled or highly fragmented in NGS-based draft genomes. We generated ~72× sequencing coverage of the *Oropetium* genome using 32 SMRT cells on the PacBio RS II platform (which is equivalent to <1 week of sequencing time and <US\$10,000 in reagents). The resulting sequence had a read N50 length of over 16 kilobases (kb), and there was 10× coverage of reads over 20 kb in length (Extended Data Fig. 1a). The raw reads were error-corrected using the hierarchical genome assembly process (HGAP), and the longest reads (>16 kb) were assembled using Celera assembler followed by two rounds of genome polishing using Quiver¹². The assembly contains 650 contigs spanning 99% (244 Mb) of the estimated 245 Mb genome size (Extended Data Fig. 1b) with a contig N50 length of 2.4 Mb (Extended Data Fig. 1c). The final assembly consists of 625 contigs after removal of the complete chloroplast genome, mitochondria-derived contigs and contaminants. The 35 largest contigs span half the genome, and the largest 107 contigs contain 90% of the sequence. The 135,324 base-pair (bp) chloroplast genome assembled into a single contig that includes both ~25 kb of inverted repeat regions which typically collapse into a single copy during assembly. The mitochondria genome was assembled into 20 partially overlapping circular chromosomes, which are the product of intramolecular recombination events that collectively span 1,100 kb.

The *Oropetium* genome has high contiguity for an uncurated draft plant genome. The average contig N50 length for all published plant genomes is 50 kb compared to 2.4 Mb for *Oropetium* (Extended Data Fig. 1d, e). After manual curation and data augmentation, only the *Arabidopsis* (TAIR10)¹³, rice (V7) and *Brachypodium* (V 2.1)¹⁰ genomes have longer contig N50 lengths. The accuracy rate is very

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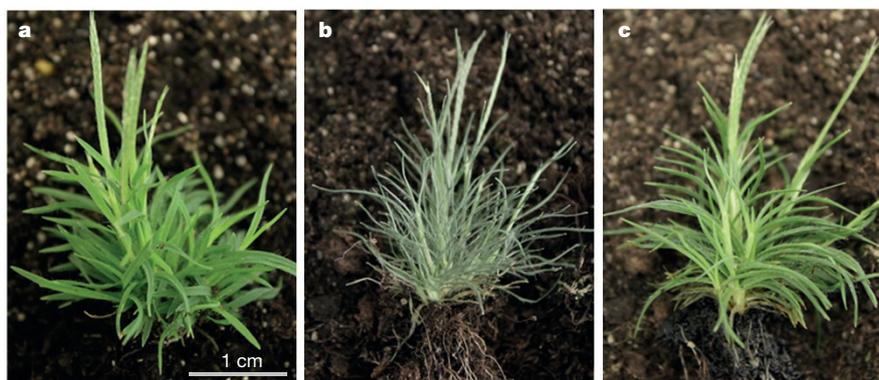


Figure 1 | Desiccation tolerance in the resurrection grass *Oropetium thomaeum*. **a**, Well watered. **b**, Desiccated (relative water content <5%) after 9 days of drought stress. **c**, Condition 24 h post-hydration (relative water content >70%).

high at 99.99995%, which is similar to Sanger-based approaches and higher than most NGS-based assemblies (Extended Data Fig. 1h). We plotted repeat density and GC content along the length of the contigs to identify factors causing contig breaks (Extended Data Fig. 1f, g). There is no correlation between repeat density and GC content at contig break points. This suggests that contig break points occur at the start of repeats or that most assembly breaks are caused by other factors, such as within-genome heterozygosity or haplotype-specific structural variation. To test this, we also tried ‘diploid-aware’ assemblers Falcon (<https://github.com/PacificBiosciences/falcon>) and MinHash Alignment Process (MHAP)¹⁴. These assemblies had similar metrics but were less contiguous overall (Extended Data Fig. 1i).

The completeness of the *Oropetium* genome allowed us to accurately survey its highly repetitive features that are often unassembled in most plant genomes. The *Oropetium* assembly captures all 18 telomeric arrays (Extended Data Table 1) with repeat number ranging from 40 to 900, suggesting that at least some are full length. Three of the nine centromeric satellites are completely assembled into large inverted repeats spanning 400 kb with a base monomer length of 155 bp, and higher order structures of dimers (310 bp), trimers (465 bp) and tetramers (620 bp; Fig. 2, Extended Data Fig. 2 and Supplementary Table 1). The remaining 40 centromeric sequences are incomplete centromere repeat fragments broken during assembly or solo repeats not associated with a larger centromere satellite. Nucleolus organizer regions contain tandem arrays of the 18S, 5.8S and 25S ribosomal RNA (rRNA) genes and typically span several megabase pairs with hundreds of nearly identical 10-kb arrays. Twenty-two full-length rRNA tandem arrays in six contigs are found in the *Oropetium* assembly (Extended Data Table 2). The largest tandem array contains five identical and one partial 9-kb repeats collectively spanning 51 kb; this is approaching the theoretical limit given the read-length distributions of our data. The remaining rRNA tandem repeats probably collapsed during read correction or genome assembly given their high sequence conservation.

Most repeats are incomplete, unassembled or highly collapsed in Illumina/454 NGS-based genomes, which has led to an underestimation and misclassification of repeat content in most plant genomes. Repetitive elements account for a surprisingly high proportion of the *Oropetium* genome (43%) compared to 21% in *Brachypodium*¹⁰, 35% in rice⁴, 54% in *Sorghum*⁵ and over 90% in wheat⁹ (Extended Data Table 3). Similar to these other genomes, the long terminal repeat (LTR) retrotransposons are the most abundant class and account for 35.6% of the *Oropetium* genome. We identified 3,247 intact LTRs in 358 families, which is similar to rice (3,663) and *Brachypodium* (2,162), but far less than *Sorghum* (17,022)¹⁵. Only ~2% of the repeats are unclassified, which reflects the completeness of individual repeat elements due to the long reads.

Genome size in the grasses varies by several orders of magnitude as a consequence of polyploidy and genome bloating due to repetitive DNA accumulation¹⁶. *Oropetium* has the smallest known genome among the

grasses¹⁷ at 90%, 60%, 50%, 30% and 10% the size of *Brachypodium*¹⁰, rice⁴, *Setaria*¹⁸, *Sorghum*⁵ and maize⁷, respectively. We found that *Oropetium* has a solo:intact LTR ratio >1, which is similar to small grass genomes like rice and *Brachypodium*, where proliferating LTRs are removed by illegitimate recombination, whereas large grass genomes like *Sorghum* and maize have solo:intact LTR ratios <1 (ref. 15). Despite its compact size, the *Oropetium* genome has a typical number of predicted protein coding genes at 28,446. A pan-cereal whole-genome duplication (WGD) event, called rho, occurred before the diversification of grasses^{5,19}. There appear to have been no further WGDs in the selected grass genomes, including *Oropetium*, since the shared rho event^{4,5}.

Genome alignments between *Oropetium* and selected grass genomes are mostly one-to-one after exclusion of the alignments derived from the shared genome duplication events (Extended Data Fig. 3a–e). Overall, 75% of the *Oropetium* genome, or 89% of its gene space, is contained in conserved syntenic blocks when compared to other grasses. Genomic colinearity across grass genomes is extensive, with a high density of orthologous genes spanning much of the euchromatin (Fig. 3). Insertions of retrotransposons and non-collinear genes that originated elsewhere in the genome contribute greatly to the differences in the intergenic sequences in grasses²⁰.

The relative sizes of syntenic blocks in the grass genomes track closely with the overall genome size difference (Extended Data Fig. 3f).

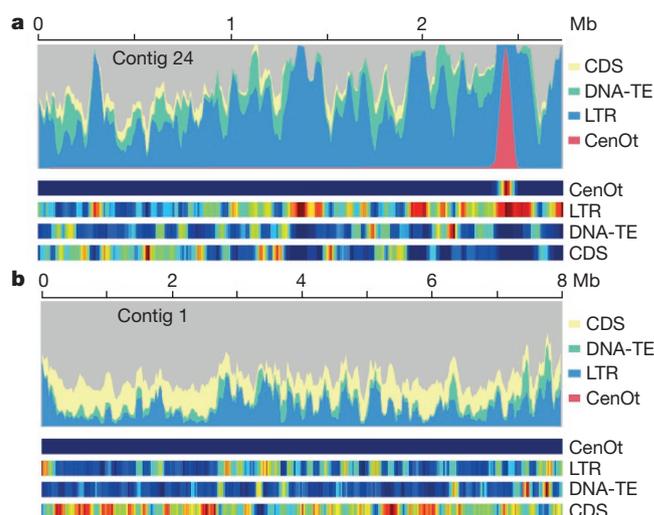


Figure 2 | SMRT sequencing enables contiguous sequencing over complex regions. The distributions of centromere-specific satellite DNA (CenOt), long terminal repeat retrotransposons (LTRs), DNA transposable elements (DNA-TE) and coding DNA sequences (CDS) are plotted. **a**, The gap-free assembly of a full-length centromeric array and the flanking highly repetitive pericentromeric region. **b**, The largest contig (7.8 Mb), which has a more typical distribution of elements.

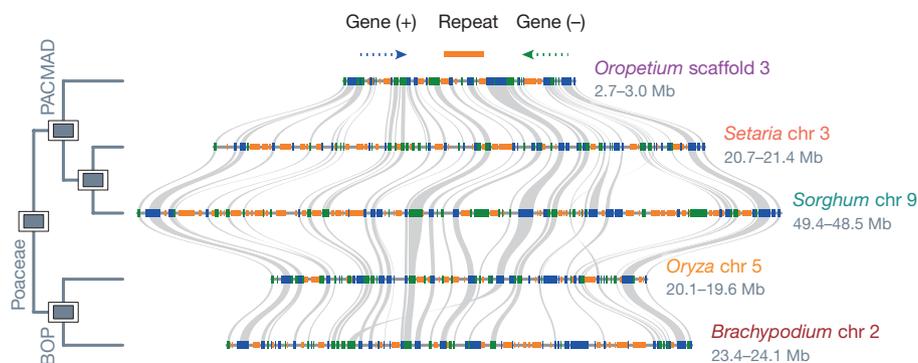


Figure 3 | Compact genome structure of *Oropetium*. *Oropetium*, part of the PACMAD clade, provides the first high-quality reference genome from the Chloridoideae subfamily—a large and diverse group of ~1,600 species that contains the orphan crops tef (*Eragrostis tef*) and finger millet (*Eleusine coracana*). Typical micro-colinearity patterns among genomic

regions from *Oropetium*, *Setaria*, *Sorghum*, *Oryza* and *Brachypodium* are shown. Rectangles show predicted gene models, and colours indicate relative orientations. Matching gene pairs are displayed as grey connections. chr, chromosome.

In contrast, the genomic span of coding sequences is similar across genes that are retained in orthologous locations, although coding features are slightly smaller in *Oropetium* (Extended Data Table 4). The relatively constant sizes of coding sequences among grass genomes confirm that genome size differences are indeed due to variations in the intergenic contents. It was thought that plants have a ‘one-way ticket to genome obesity’ due to the retention of proliferating transposable elements²¹. However, analysis of carnivorous plants *Utricularia gibba* (bladderwort, 82 Mb)²² and *Genlisea aurea* (corkscrew, 63.6 Mb)²³ provided evidence that almost all intergenic space can be purged. Small genomes also arise from a reduction in gene number as seen in the aquatic monocotyledon *Spirodela polyrhiza*, which has the fewest predicted protein coding genes at 19,623 (ref. 24). *Oropetium* seems to have reduced both its intergenic and intragenic sequence.

As the intergenic sequence in *Oropetium* is specifically reduced compared with other grasses (Extended Data Fig. 3f), we determined which sequence accounted for its smaller genome size by comparing highly syntenic regions of the larger 730 Mb *Sorghum* genome. To identify highly orthologous regions we looked for *Sorghum* genes (promoter, 5′UTR, exons, introns and 3′UTR) with an increased number of conserved noncoding sequences²⁵. We then analysed the top 48 *Sorghum* genes against their orthologous sequences in *Oropetium* and found that they were 38% (± 0.27 , 1 s.d.) larger in *Sorghum* (Extended Data Fig. 4a). The primary driver of gene-space expansion was highly unique ~1-kb intragenic sequences evenly spaced within the *Sorghum* genes. One explanation is that these evenly spaced highly unique sequences are degenerate remnants of transposons that have been partly purged from the *Sorghum* genome. *Oropetium* has a >1 solo:intact LTR ratio, consistent with active purging of transposons and complete loss of these regions. These results lend support to an emerging theory about the C-value paradox called the Genome Balance Hypothesis²⁶, which suggests that selection on gene networks and pericentromeric growth (centromere movement) is balanced by transposon proliferation and retention. Therefore, these evenly spaced highly unique sequences balance the 6:1 expansion of pericentromeric sequence in *Sorghum* as compared to *Oropetium* (Extended Data Fig. 4b).

Desiccation tolerance was a key adaptation that permitted the most recent common ancestor of terrestrial plants to survive on land. Desiccation tolerance is widespread in bryophytes and lichens but rare in flowering plants, although similar mechanisms have evolved in vascular plants for seed and pollen desiccation. Desiccation tolerance to survive prolonged drought evolved independently in diverse monocotyledon and eudicotyledon lineages, and is found in at least 300 species. Gene duplications have provided the raw material for evolutionary innovation across plants. Tandem duplicated genes are often involved in stress responses and are probably important for adaptive evolution in dynamically changing environments. *Oropetium* has 6,668 tandem duplicated genes in 2,326 clusters, which is a slightly higher number than in other

grasses, but a similar proportion (24% of genes). Tandem duplicated genes are enriched for gene ontology terms involved in response to abiotic stresses, gene regulation and cellular metabolism (Supplementary Table 2). In addition, *Oropetium* has 4,209 homeologous gene pairs retained from the rho WGD event, which are enriched for gene ontology terms related to gene regulation and stress responses such as transcription factor activity, nitrogen metabolism, response to abiotic stimulus, to salt stress and to oxygen-containing compounds (Supplementary Tables 3 and 4). Understanding the genomic mechanisms of extreme desiccation tolerance in resurrection plants such as *Oropetium* may provide targets for engineering drought and stress tolerance in crop plants.

Pacific Biosciences (PacBio) SMRT sequencing has been used to close gaps in the human genome²⁷, assemble complete bacterial genomes¹² and identify novel gene isoforms²⁸. Here we present a several hundred megabase plant genome, sequenced and assembled entirely by SMRT sequencing. The long SMRT reads produced a near-complete draft genome that captured three of nine complete centromeres, all of the telomeres and biologically relevant features of the *Oropetium* genome. The total time from extracted DNA to a complete assembly was less than one month, and costs for PacBio were comparable to an Illumina-based genome assembly. Our study demonstrates that SMRT sequencing enables a new level of genome assembly required for full ENCODE-type analysis of intergenic sequence, which is not currently possible with other NGS-based methods. The compactness of the *Oropetium* genome results from purging of both inter- and intragenic sequences, probably through small deletions during illegitimate recombination, as has been shown in other grasses. One hypothesis is that genome size is a function of cell size²⁹, and consistent with this, all small plant genomes sequenced to date including *Arabidopsis* (125 Mb), *Brachypodium* (272 Mb), *Selaginella* (100 Mb) *Spirodela* (158 Mb) and *Utricularia* (82 Mb) are plants of very small stature (Fig. 1). However, we provide evidence for the Genome Balance Hypothesis, which suggests that there is selective pressure on *Oropetium* to purge proliferating transposons in order to maintain expression balance of networked genes and spacing in centromeres. The complete assembly of complex and highly similar repeat sequences demonstrated here suggests that SMRT sequencing can be used to assemble large and polyploid plant and other eukaryotic genomes, assuming ample sequence coverage and computational resources. SMRT-sequencing-based assemblies provide an opportunity to determine how these regions play a role in genome architecture and dynamics.

Online Content Methods, along with any additional Extended Data display items and Source Data, are available in the online version of the paper; references unique to these sections appear only in the online paper.

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1. Michael, T. P. & VanBuren, R. Progress, challenges and the future of crop genomes. *Curr. Opin. Plant Biol.* **24**, 71–81 (2015).
2. Kellis, M. *et al.* Defining functional DNA elements in the human genome. *Proc. Natl Acad. Sci. USA* **111**, 6131–6138 (2014).
3. The Arabidopsis Genome Initiative. Analysis of the genome sequence of the flowering plant *Arabidopsis thaliana*. *Nature* **408**, 796–815 (2000).
4. International Rice Genome Sequencing Project. The map-based sequence of the rice genome. *Nature* **436**, 793–800 (2005).
5. Paterson, A. H. *et al.* The *Sorghum bicolor* genome and the diversification of grasses. *Nature* **457**, 551–556 (2009).
6. Elert, E. Rice by the numbers: A good grain. *Nature* **514**, S50–S51 (2014).
7. Schnable, P. S. *et al.* The B73 maize genome: complexity, diversity, and dynamics. *Science* **326**, 1112–1115 (2009).
8. International Barley Genome Sequencing Consortium. A physical, genetic and functional sequence assembly of the barley genome. *Nature* **491**, 711–716 (2012).
9. International Wheat Genome Sequencing Consortium (IWGSC). A chromosome-based draft sequence of the hexaploid bread wheat (*Triticum aestivum*) genome. *Science* **345**, 1251788 (2014).
10. The International Brachypodium Initiative. Genome sequencing and analysis of the model grass *Brachypodium distachyon*. *Nature* **463**, 763–768 (2010).
11. Bartels, D. & Mattar, M. *Oropetium thomaeum*: A resurrection grass with a diploid genome. *Maydica* **47**, 185–192 (2002).
12. Chin, C.-S. *et al.* Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nature Methods* **10**, 563–569 (2013).
13. Lamesch, P. *et al.* The Arabidopsis Information Resource (TAIR): improved gene annotation and new tools. *Nucleic Acids Res.* **40**, D1202–D1210 (2012).
14. Berlin, K. *et al.* Assembling large genomes with single-molecule sequencing and locality-sensitive hashing. *Nature Biotechnol.* **33**, 623–630 (2015).
15. El Baidouri, M. & Panaud, O. Comparative genomic paleontology across plant kingdom reveals the dynamics of TE-driven genome evolution. *Genome Biol. Evol.* **5**, 954–965 (2013).
16. Michael, T. P. Plant genome size variation: bloating and purging DNA. *Brief. Funct. Genomic.* **13**, 308–317 (2014).
17. Jones, N. & Pašakinskienė, I. Genome conflict in the gramineae. *New Phytol.* **165**, 391–410 (2005).
18. Bennetzen, J. L. *et al.* Reference genome sequence of the model plant *Setaria*. *Nature Biotechnol.* **30**, 555–561 (2012).
19. Tang, H., Bowers, J. E., Wang, X. & Paterson, A. H. Angiosperm genome comparisons reveal early polyploidy in the monocot lineage. *Proc. Natl Acad. Sci. USA* **107**, 472–477 (2010).
20. Wicker, T., Buchmann, J. P. & Keller, B. Patching gaps in plant genomes results in gene movement and erosion of colinearity. *Genome Res.* **20**, 1229–1237 (2010).
21. Bennetzen, J. L. & Kellogg, E. A. Do plants have a one-way ticket to genomic obesity? *Plant Cell* **9**, 1509 (1997).
22. Ibarra-Laclette, E. *et al.* Architecture and evolution of a minute plant genome. *Nature* **498**, 94–98 (2013).
23. Leushkin, E. V. *et al.* The miniature genome of a carnivorous plant *Genlisea aurea* contains a low number of genes and short non-coding sequences. *BMC Genomics* **14**, 476 (2013).
24. Wang, W. *et al.* The *Spirodela polyrhiza* genome reveals insights into its neotenuous reduction fast growth and aquatic lifestyle. *Nature Commun.* **5**, 3311 (2014).
25. Lyons, E. & Freeling, M. How to usefully compare homologous plant genes and chromosomes as DNA sequences. *Plant J.* **53**, 661–673 (2008).
26. Freeling, M., Xu, J., Woodhouse, M. & Lisch, D. A solution to the C-value paradox and the function of junk DNA: the Genome Balance Hypothesis. *Mol. Plant* **8**, 899–910 (2015).
27. Chaisson, M. J. P. *et al.* Resolving the complexity of the human genome using single-molecule sequencing. *Nature* **517**, 608–611 (2015).
28. Au, K. F. *et al.* Characterization of the human ESC transcriptome by hybrid sequencing. *Proc. Natl Acad. Sci. USA* **110**, E4821–E4830 (2013).
29. Beaulieu, J. M., Leitch, I. J., Patel, S., Pendharkar, A. & Knight, C. A. Genome size is a strong predictor of cell size and stomatal density in angiosperms. *New Phytol.* **179**, 975–986 (2008).

Supplementary Information is available in the online version of the paper.

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Author Contributions R.V., D.Br., T.P.M. and T.C.M. designed and conceived research; D.Ba. and D.C. identified biological material, performed desiccation experiments and extracted DNA and RNA; R.V. prepared DNA for PacBio sequencing; T.P.M., R.V. and T.C.M. performed Illumina sequencing; K.S., R.H. and J.G. performed PacBio sequencing and assembly; B.T.H. and A.H. conducted the BioNano analysis. D.Br., R.V., T.P.M. and T.C.M. annotated genome features; E.L., M.F., D.Bu., R.V., D.Br., H.T., T.P.M., T.C.M. and P.P.E. analysed data; R.V., T.P.M. and T.C.M. wrote the paper. All authors read and approved the final manuscript.

Author Information The genome assembly and annotation have been deposited in CoGe under the accession code 25799 (<https://genomeevolution.org/CoGe/GenomeInfo.pl?gid=25799>), in the NCBI BioProject under PRJNA286116, and in GenBank under accession number LFFJQ0000000. Raw PacBio and Illumina reads are available at the Short Read Archive at NCBI under the aforementioned NCBI BioProject. Genome assembly and annotation are also available at <http://www.oropetium.org/>. Reprints and permissions information is available at www.nature.com/reprints. The authors declare no competing financial interests. Readers are welcome to comment on the online version of the paper. Correspondence and requests for materials should be addressed to T.P.M. (toddpmichael@gmail.com) or T.C.M. (tmockler@danforthcenter.org).



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METHODS

No statistical methods were used to predetermine sample size.

Plant material. *Oropetium thomaeum* is a compact resurrection plant that has the smallest known genome among the grasses, at 245 Mb and 9 chromosomes ($2n = 2x = 18$; $1C = 0.25 \text{ pg}$)¹⁷. We estimated the genome size to be 250 Mb by flow cytometry and 245 Mb by *k*-mer analysis (Extended Data Fig. 1b). *Oropetium thomaeum* plants were originally collected in Jodhpur, Rajasthan, India and propagated as previously described¹¹. *Oropetium* is a member of the Chloridoideae subfamily, a large and diverse group of roughly 1,600 species that contains the orphan crops *tef* (*Eragrostis tef*) and finger millet (*Eleusine coracana*) as well as some turf grasses (such as Bermuda grass, *Cynodon dactylon* and *Zoysia japonica*).

SMRT PacBio sequencing. Fifty micrograms of high-molecular-weight *Oropetium* gDNA was extracted using a modified nuclei preparation method³⁰ followed by an additional high-salt phenol–chloroform purification to minimize contamination. A 20-kb insert SMRTbell library was generated using a 15 kb lower-end size selection protocol on the BluePippin (Sage Science). Initial titration runs were performed to optimize loading on the SMRT Cell for maximum performance. The *Oropetium* genome was sequenced using 32 SMRT Cells with 4-h collections and P6-C4 chemistry on the PacBio RS II platform (Pacific Biosciences).

HGAP genome assembly. The *Oropetium* genome was assembled using the RS_HGAP_Assembly.3 protocol for assembly and Quiver for genome polishing in SMRT Analysis v2.3.0¹². This consisted of a three-step process involving (1) generation of preassembled reads with improved consensus accuracy; (2) assembly of the genome through overlap consensus accuracy using Celera; and (3) one round of genome polishing with Quiver. For HGAP, the following parameters were used: PreAssembler Filter v1 (minimum sub-read length = 3,000 bp, minimum polymerase read quality = 0.80, minimum polymerase read length = 3,000 bp); PreAssembler v2 (minimum seed length = 16,000 bp, number of seed read chunks = 6, alignment candidates per chunk = 10, total alignment candidates = 24, min coverage for correction = 6); AssembleUnitig v1 (target genome coverage = 30, overlap error rate = 0.06, minimum overlap = 40 bp and overlap *k*-mer = 14); and BLASR v1 mapping of reads for genome polishing with Quiver (max divergence percentage = 30, minimum anchor size = 12). A second round of genome polishing was performed using Quiver (SMRT Analysis v2.3.0) to further improve the site-specific consensus accuracy of the assembly. The following Quiver parameters were used for genome polishing: filtering (minimum sub-read length = 3,000 bp, minimum polymerase read quality = 0.80, minimum polymerase read length = 3,000 bp); mapping (maximum divergence percentage = 30, minimum anchor size = 12). Default parameters were otherwise employed for both HGAP assembly and Quiver protocols.

Falcon and MHAP assemblies. We also tested other assemblers to compare the PacBio HGAP assembly results (Extended Data Fig. 1i). Raw PacBio reads were error-corrected and assembled using Falcon and MHAP under default parameters. The Falcon and MHAP assemblies have lower contiguity than the HGAP assembly and have fewer assembled centromere and telomere sequences with a lower average length.

Construction of a genome map using the Irys system for contig anchoring and scaffolding. Genome mapping from BioNano Genomics³¹ was used to improve the assembly quality of the *Oropetium* genome with the eventual goal of producing a chromosome-scale assembly. High molecular weight genomic DNA was isolated from fresh *Oropetium* tissue using the following protocol outline. Three grams of leaves were collected from live *Oropetium thomaeum* plants and fixed with formaldehyde. After blending with a tissue homogenizer in isolation buffer, a filtration step and Triton-X washing treatment were performed. The nuclei were purified on percoll cushions. The nuclei were washed extensively and embedded in low melting agarose at different dilutions. Finally, the DNA plugs were treated with a lysis buffer containing detergent, proteinase K and β -mercaptoethanol (BME). In total, 53 Gb of data (>100 kb) were collected representing $\sim 200\times$ genome coverage with a molecule N50 length of 169 kb (Extended Data Fig. 5a). The size distribution was lower than expected and is probably a result of impurities during high-molecular-weight gDNA isolation that would cause shearing and inhibition of enzymes. Molecules were *de novo* assembled as previously described³². Two genome maps were assembled at different stringencies, map set 1 has 402 maps with an N50 length of 725 kb and spans 216 Mb (Extended Data Fig. 5b); the second genome map has 214 maps and an N50 of 1.674 Mb. Combining the genome maps with the PacBio assembly to produce a hybrid scaffold was performed sequentially with the two genome maps. The scaffolding merged 90 contigs producing an assembly of 46 primary scaffolds covering 94% of the sequence assembly with an N50 of 7.8 Mb; in total there are 535 scaffolds with an N50 of 7.1 Mb and total assembled size of 244 Mb.

Variant calling using Illumina data. WGS Illumina sequences from *Oropetium* gDNA were used to assess the error rate of the PacBio assembly and residual within-genome heterozygosity (Supplementary Table 5). Raw Illumina HiSeq data

from three different libraries of 570-bp insert, 1-kb insert and 3-kb insert sizes were trimmed for quality using Trimmomatic (v.0.32; ref. 33). Illumina sequence adaptors were removed, leading low quality (below quality 3) and N base pairs were trimmed, and reads were scanned using a 4-bp sliding window and trimmed when the average quality per base dropped below 30. Read pairs where both reads were ultimately of at least 36 bp in length following this quality control process were retained and used for subsequent analyses.

Quality trimmed data were aligned to our assembly using BWA mem (v. 0.7.12-r1039)³⁴. Duplicate alignments were marked using Picard tools v.1.104 MarkDuplicates (<http://broadinstitute.github.io/picard/>). Genome Analysis Toolkit (v.3.3.0)³⁵ IndelRealigner was used to perform local realignment around indels, followed by application of GATK HaplotypeCaller to call variants. Identified single nucleotide polymorphisms were filtered by depth, strand bias, mapping quality and read position. Identified indels were filtered by depth, strand bias and read position.

The native error rate of raw PacBio reads is in the range of 15–20%, raising the possibility that residual sequencing errors may be introduced into the final assembly of the *Oropetium* genome. Homozygous mismatches are classified as sequencing errors, and heterozygous mismatches indicate sites of heterozygosity. The accuracy rate is very high at 99.99995%, and a relatively high proportion of the errors (two-thirds) are small insertions or deletions (indels). The accuracy rate is similar to those obtained with WGS Sanger approaches^{5,36} and is higher than those reported for most NGS-based assemblies. The estimated residual within-genome heterozygosity for the *Oropetium* genome is very low at 0.087%, which probably contributed to the high contiguity of the assembly. This suggests that provided sufficient coverage, a PacBio SMRT-only approach can produce a high-quality complete plant genome.

Repeat annotation. To structurally annotate repeat sequences in the *Oropetium* genome, we began by discovering repetitive elements through application of the REPET v.2.2 packages TEde novo and TEannot³⁷. The TEde novo pipeline compares the genome with itself to identify and classify repeated genomic elements. All-by-all alignments were conducted with NCBI-BLAST+ using default TEde novo parameters. LTRharvest³⁸ was used for structural detection. During clustering, Grouper, Recon and Plier steps were invoked both with and without structural detection. Consensus building was performed using default parameters. During consensus detect features, repeat scout³⁹ was invoked, and Pfam26.0 HMM profiles⁴⁰ and Repbase (v18.08) nucleotide and amino acid databanks were used. Finally, consensus classification, filtering and clustering were performed using default parameters.

Output from the TEde novo pipeline was used as input to the TEannot pipeline. This pipeline mines the genome sequence using repeated sequences identified in the previous TEde novo pipeline to produce classified non-redundant consensus repeat sequences along with short simple repeats, which are exported to GFF3 format. First, a set of perfectly matching sequences from the TEde novo-output transposable elements (TE) library was selected by running a subset of the TEannot pipeline, producing a working reference TE library. This TE library was used in a full run of the TEannot pipeline. For alignment of the reference TE library, NCBI-BLAST+ was used, and blaster, repeat masker and censor steps were run both on the reference TE library and on randomized chunks. Filtering was applied using default parameters. Short simple repeats were identified using the crossmatch engine. Merging was performed using default parameters. For comparisons, Repbase (v18.08) nucleotide and amino acid databanks were used. Finally, filtering was applied using default parameters, and annotations were exported to GFF3 format.

To classify identified repeats, non-redundant consensus repeat sequences as output by TEanno were annotated via PASTEClassifier v1.0 <https://urgi.versailles.inra.fr/Tools/PASTEClassifier/README>. To classify these sequences, Repbase (v18.08)⁴¹ nucleotide and amino acid sequences were used, as were Pfam v26.0 (<http://pfam.xfam.org/>) HMM repeat profiles. Finally, identified LTRs were classified as Gypsy if homology or motif evidence existed for Gypsy and not for Copia, classified as Copia if the opposite were true, and otherwise classified as unknown.

Centromere and telomere identification. Centromeric repeats were identified using an approach outlined in ref. 42. Tandem repeat finder (TRF, Version 4.07b)⁴³ was used to find tandem repeats using the parameters '1 1 2 80 5 200 2000-d -h' in order to find high order repeats. The resulting 'dat file' was transformed into a GFF3 file, which was used to identify telomeric and centromeric repeats. To identify the centromeric repeats, the largest repeat arrays (period length X copy number) were identified and clustered. Clustered centromeric repeat regions were transformed into FASTA files and aligned using clustalX to identify array sequence composition and orientation. The base centromeric repeat was 155 bp dimers (310 bp), trimers (465 bp) and tetramers (620 bp) (Extended Data Fig. 2 and Supplementary Table 1). The three largest centromeric arrays (contigs 003, 028 and 064) were >400 kb and resolved into large inverted repeats, consistent with them being full length. The telomeric repeats were identified by searching

the ends of contigs for short (~7 bp) high copy number repeats; 18 telomeric repeat sequences with the monomer 'AAACCCCT' were identified (Extended Data Table 1). **Transcriptome assembly.** Total RNA was extracted from fresh, desiccated and 24-h post rehydration *Oropetium* leaf tissues with 2 biological replicates collected for each tissue. RNA-seq libraries were prepared from the total RNA and bar-coded using TruSeq RNA Sample Prep Kits (Illumina) according to the manufacturer's protocol. Raw Illumina RNA-seq data from the six libraries were trimmed for quality using Trimmomatic (v.0.32; ref. 33). Illumina sequence adaptors were removed, then leading low-quality (below quality 3) and N base pairs were trimmed and, finally, resulting trimmed reads were scanned using a 4-bp sliding window and cut when the average quality per base dropped below 30. Read pairs where both reads were ultimately of at least 36 base pairs in length following this quality control process were retained and used for subsequent analyses. Trinity (v.r20140717)⁴⁴ was used to assemble quality filtered data. Assembled transcripts were aligned to our genome sequence using NCBI blastn v.2.2.30+ with an *e*-value cut-off of 1×10^{-5} . Successfully aligned transcripts were clustered at 90% identity using CD-HIT (v. 4.5.4)⁴⁵, with representative sequences from each cluster retained and used to help parameterize gene calling. Eighty-seven per cent of the trimmed RNA-seq reads aligned to the *Oropetium* genome, suggesting that the genome is largely complete (Supplementary Table 5). Reads that failed to align may have been contaminants from other organisms.

Gene annotation. Maker v2.31.8⁴⁶ (<http://www.yandell-lab.org/software/maker.html>) was used to identify putative genes. Aligned and representative sequences from our transcriptome assembly were input to Maker as expressed sequence tag evidence. Rice and *Brachypodium* proteome sequences clustered at 90% identity using CD-HIT (v. 4.5.4)⁴⁵ with representative sequences from each cluster retained and input to Maker as multi-organism protein homology evidence. The *Oropetium* repeat database was input to Maker as a custom repeat library. SNAPhmm, Augustus, and GeneMarkHMM were invoked by Maker and were initially trained using rice and maize. Only genes for which the encoded protein was predicted to contain a complete open reading frame were retained.

On the basis of the gene annotations provided by Maker, cufflinks (v2.2.1)⁴⁷ was used to identify predicted genes without empirical expression evidence. Quality-trimmed data from all six RNA-seq libraries were input simultaneously to cufflinks, with results used to identify genes with and without expression.

Protein sequences from genes predicted by Maker were functionally annotated using NCBI blastp v.2.2.30+ versus the NCBI non-redundant refseq protein database (<http://www.ncbi.nlm.nih.gov/refseq/>), versus the UniProt database⁴⁸, and using InterProScan (v. 5.6-48.0)⁴⁹.

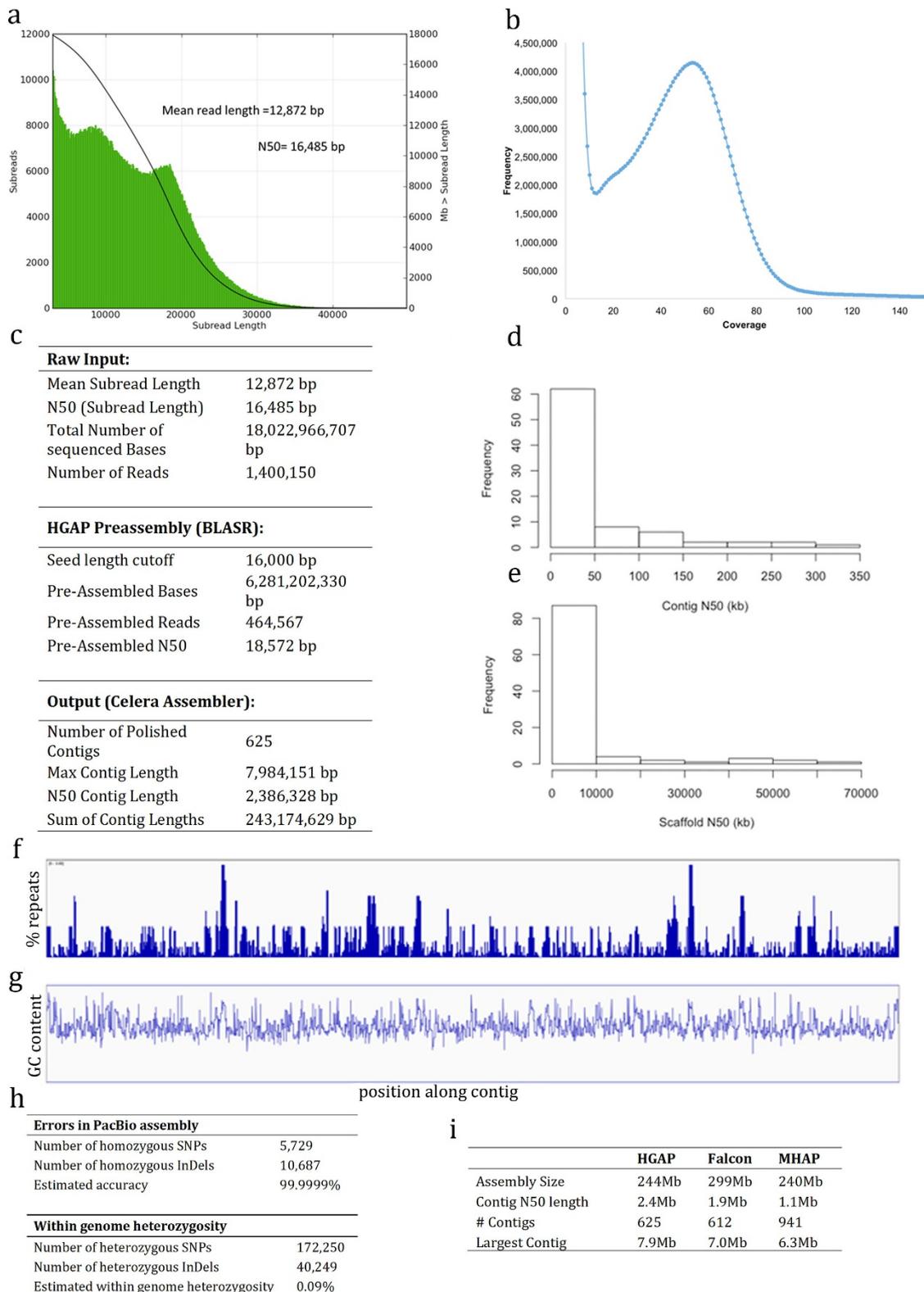
Finally, Maker-predicted genes were pruned based on a Maker-defined annotation edit distanced (AED) score that measures distance between the predicted gene and the evidence input to Maker, non-redundant (NR) annotation, Uniprot annotation, InterProScan annotation and expression level as output by cufflinks. Genes were removed that had no alignment evidence (AED = 1), no sequence match to either the NR or Uniprot databases, no InterProScan predicted domains and no expression evidence in our RNA-seq data.

Synteny and comparative genomics. Genome data sets from *Setaria*, *Sorghum*, rice and *Brachypodium* were downloaded from Phytozome (version 9.1) and subject to pairwise genome alignments against the *Oropetium* genome. For each pairwise alignment, the coding sequences of predicted gene models are compared to each other using adaptive seeds⁵⁰. Our synteny search pipeline defines syntenic blocks by chaining the large-scale alignment tool (LAST) hits with a distance cut-off of 20 genes apart, also requiring at least four gene pairs per syntenic block. The syntenic blocks were further screened using QUOTA-ALIGN⁵¹ to retain one-to-one blocks and to exclude weak blocks derived from shared ancient duplications. The resulting dot plots were visually inspected to confirm the structural similarity of the *Oropetium* genome in relation to other genomes (Extended Data Fig. 3a–e).

Pairwise genomic alignments, described above, combined with OrthoMCL⁵² analyses filtered to one-to-one hits were used to identify orthologous gene clusters between *Oropetium* and *Sorghum*, rice, *Vitis* and *Arabidopsis*. The complete *Oropetium*–*Arabidopsis* orthologue list was then filtered to focus on genes with functional data in the STRING v9.1 global *Arabidopsis* protein interaction network⁵³. Gene expression patterns and duplicated genes (tandem and whole-genome duplicates) were mapped onto this network using Cytoscape v3.1.1⁵⁴

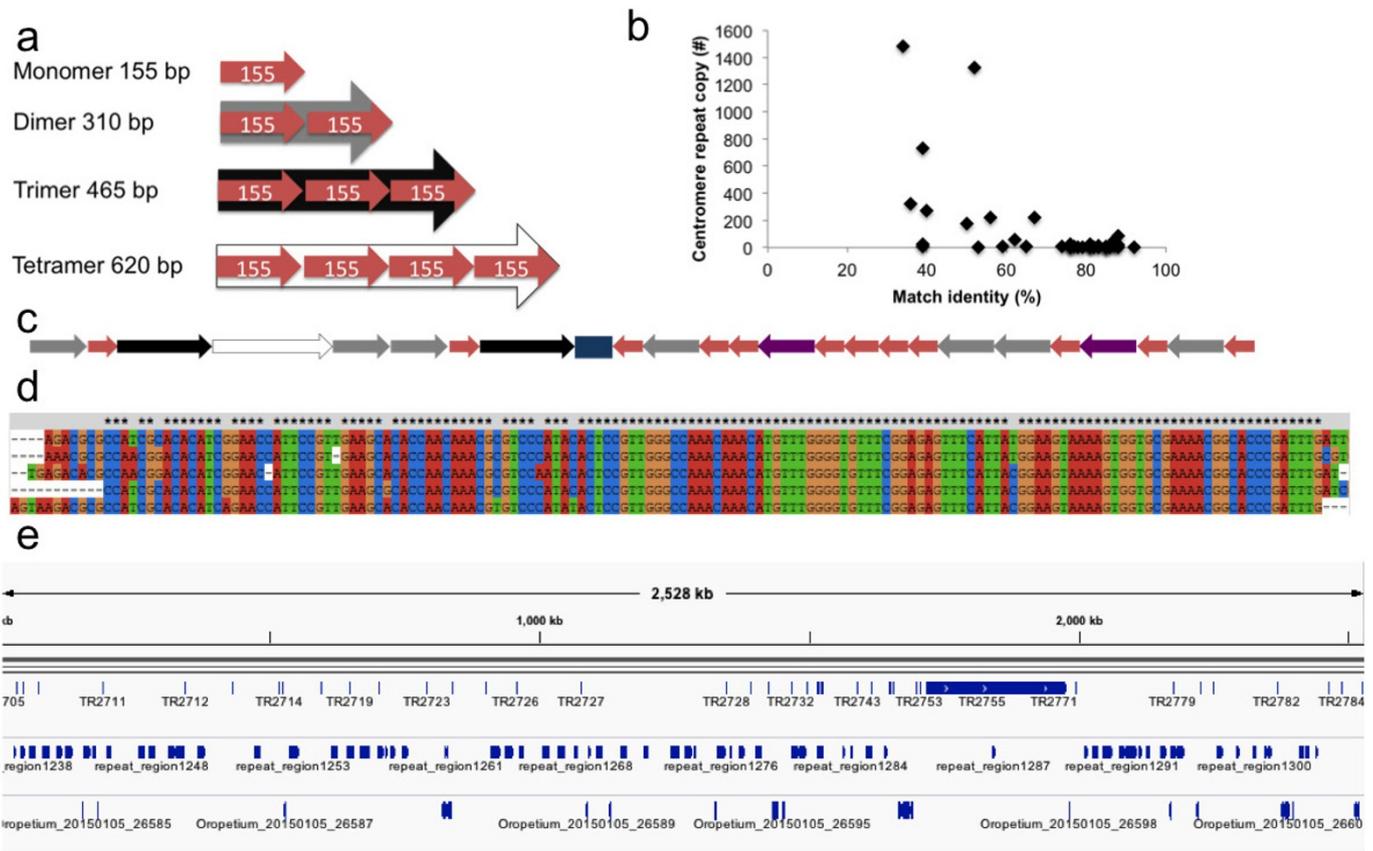
to identify clusters of co-expressed and interacting duplicate genes, respectively (Extended Data Fig. 6). Various network statistics were calculated using NetworkAnalyzer⁵⁵, including average number of neighbours (that is, protein interactions) and total number of isolated nodes (that is, without known interactors). **Constructing a gene interaction network.** We constructed a gene interaction network for *Oropetium* on the basis of orthologous relationships with *Arabidopsis* genes with validated interactions and expression data yielding a network with 4,421 nodes (gene products) with 36,918 edges (interactions). This network encompasses most metabolic pathways including photosynthesis, core anabolic and catabolic processes and stress response pathways (Extended Data Fig. 6).

30. Zhang, H.-B., Zhao, X., Ding, X., Paterson, A. H. & Wing, R. A. Preparation of megabase-size DNA from plant nuclei. *Plant J.* **7**, 175–184 (1995).
31. Lam, E. T. *et al.* Genome mapping on nanochannel arrays for structural variation analysis and sequence assembly. *Nature Biotechnol.* **30**, 771–776 (2012).
32. Cao, H. *et al.* Rapid detection of structural variation in a human genome using nanochannel-based genome mapping technology. *GigaScience* **3**, 34 (2014).
33. Bolger, A. M., Lohse, M. & Usadel, B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* **30**, 2114–2120 (2014).
34. Li, H. & Durbin, R. Fast and accurate short read alignment with Burrows–Wheeler transform. *Bioinformatics* **25**, 1754–1760 (2009).
35. McKenna, A. *et al.* The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res.* **20**, 1297–1303 (2010).
36. Ming, R. *et al.* The draft genome of the transgenic tropical fruit tree papaya (*Carica papaya* Linnaeus). *Nature* **452**, 991–996 (2008).
37. Flutre, T., Duprat, E., Feuillet, C. & Quesneville, H. Considering transposable element diversification in de novo annotation approaches. *PLoS ONE* **6**, e16526 (2011).
38. Ellinghaus, D., Kurtz, S. & Willhoeft, U. LTRharvest, an efficient and flexible software for de novo detection of LTR retrotransposons. *BMC Bioinformatics* **9**, 18 (2008).
39. Price, A. L., Jones, N. C. & Pevzner, P. A. De novo identification of repeat families in large genomes. *Bioinformatics* **21**, i351–i358 (2005).
40. Finn, R. D. *et al.* Pfam: the protein families database. *Nucleic Acids Res.* **42**, D222–D230 (2014).
41. Jurka, J. *et al.* Repbase Update, a database of eukaryotic repetitive elements. *Cytogenet. Genome Res.* **110**, 462–467 (2005).
42. Melters, D. P. *et al.* Comparative analysis of tandem repeats from hundreds of species reveals unique insights into centromere evolution. *Genome Biol.* **14**, R10 (2013).
43. Benson, G. Tandem repeats finder: a program to analyze DNA sequences. *Nucleic Acids Res.* **27**, 573 (1999).
44. Grabherr, M. G. *et al.* Full-length transcriptome assembly from RNA-Seq data without a reference genome. *Nature Biotechnol.* **29**, 644–652 (2011).
45. Huang, Y., Niu, B., Gao, Y., Fu, L. & Li, W. CD-HIT Suite: a web server for clustering and comparing biological sequences. *Bioinformatics* **26**, 680–682 (2010).
46. Cantarel, B. L. *et al.* MAKER: an easy-to-use annotation pipeline designed for emerging model organism genomes. *Genome Res.* **18**, 188–196 (2008).
47. Trapnell, C. *et al.* Differential gene and transcript expression analysis of RNA-seq experiments with TopHat and Cufflinks. *Nature Protocols* **7**, 562–578 (2012).
48. Wu, C. H. *et al.* The Universal Protein Resource (UniProt): an expanding universe of protein information. *Nucleic Acids Res.* **34**, D187–D191 (2006).
49. Quevillon, E. *et al.* InterProScan: protein domains identifier. *Nucleic Acids Res.* **33**, W116–W120 (2005).
50. Kielbasa, S. M., Wan, R., Sato, K., Horton, P. & Frith, M. C. Adaptive seeds tame genomic sequence comparison. *Genome Res.* **21**, 487–493 (2011).
51. Tang, H. *et al.* Screening synteny blocks in pairwise genome comparisons through integer programming. *BMC Bioinformatics* **12**, 102 (2011).
52. Li, L., Stoeckert, C. J. & Roos, D. S. OrthoMCL: identification of ortholog groups for eukaryotic genomes. *Genome Res.* **13**, 2178–2189 (2003).
53. Franceschini, A. *et al.* STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res.* **41**, D808–D815 (2013).
54. Saito, R. *et al.* A travel guide to Cytoscape plugins. *Nature Methods* **9**, 1069–1076 (2012).
55. Doncheva, N. T., Assenov, Y., Domingues, F. S. & Albrecht, M. Topological analysis and interactive visualization of biological networks and protein structures. *Nature Protocols* **7**, 670–685 (2012).



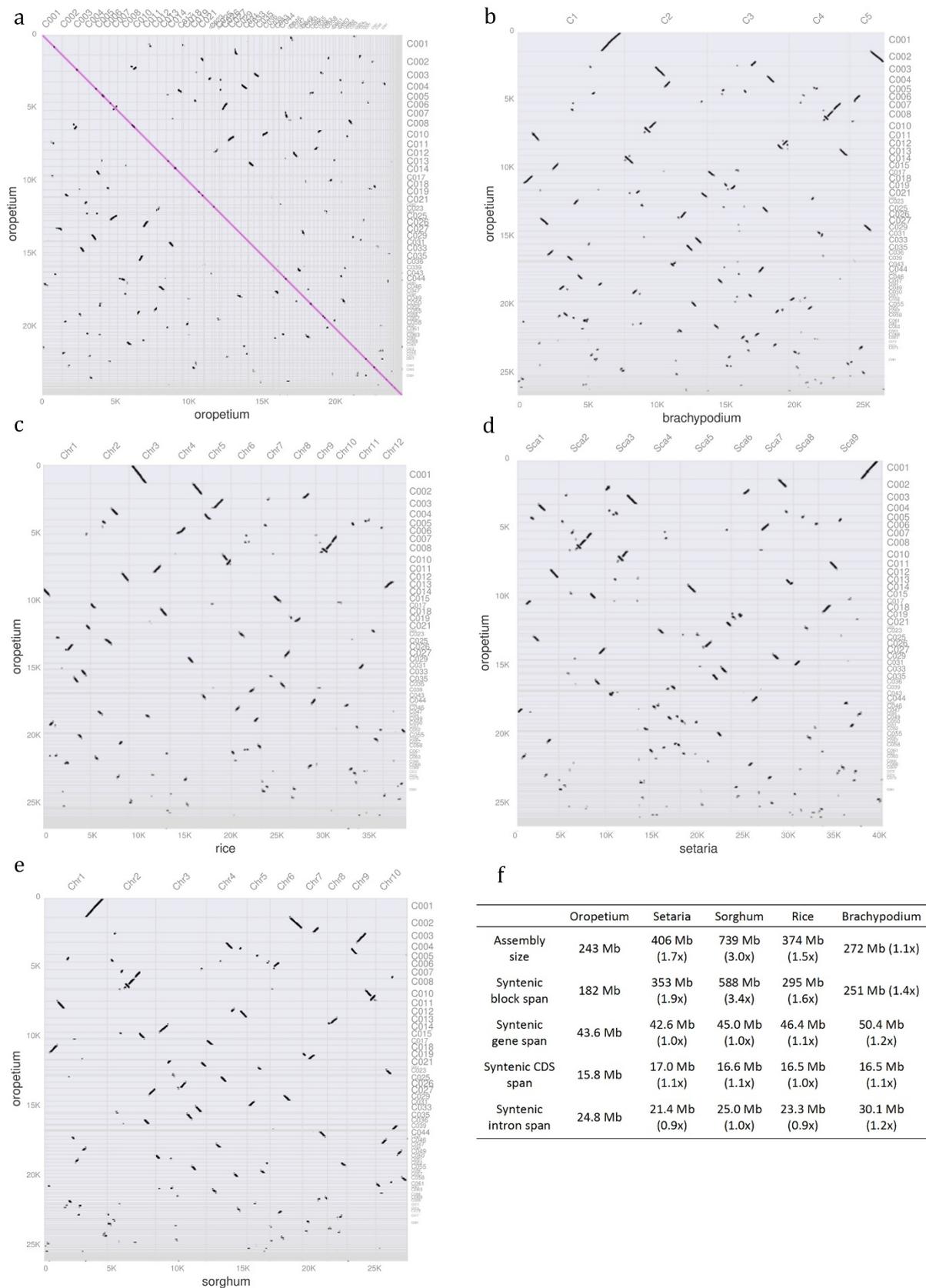
Extended Data Figure 1 | Summary of the *Oropetium* genome assembly statistics. **a**, Histogram of length distribution of raw P6C4 chemistry PacBio reads. The mean read length of the raw reads is 12,872 bp, and the N50 is 16,485 bp. **b**, Genome size estimation using *k*-mer distribution. *K*-mer distribution of unassembled *Oropetium* Illumina WGS reads. *K*-mer frequency displays a unimodal curve indicating a low rate of heterozygosity in the *Oropetium* genome. Frequency distribution suggests a genome size of ~245 Mb, consistent with flow-cytometry-based estimations. **c**, SMRT sequencing raw read, preassembly and assembly statistics. **d**, **e**, The distribution of the contig N50 length (**d**) and scaffold

N50 length (**e**) of all published plant genomes is plotted. The average contig N50 length for published plant genomes is ~50 kb compared to 2.4 Mb for *Oropetium*. **f**, **g**, Repeat density (as a function of percentage repeats) (**f**) and GC content (**g**) are plotted at a scaled position along each contig. Each contig was divided into 5,000 sliding windows with each window representing 0.02% of the contig length and the averages of each scaled sliding window are plotted. Repeat content and GC content do not vary at the ends of contigs. **h**, Estimated accuracy of SMRT PacBio assembly and within-genome heterozygosity. **i**, Comparison of HGAP Falcon and MHAP PacBio assemblers.



Extended Data Figure 2 | PacBio sequencing and assembly completely resolves the *Oropetium* centromeres. **a**, The *Oropetium* centromere repeat base is 155 bp (red arrow), whereas they are also found in dimer (310 bp, grey arrow), trimer (465 bp, black arrow) and tetramer (620 bp, white arrow) form. **b**, As the copy number of a repeat increases, the match

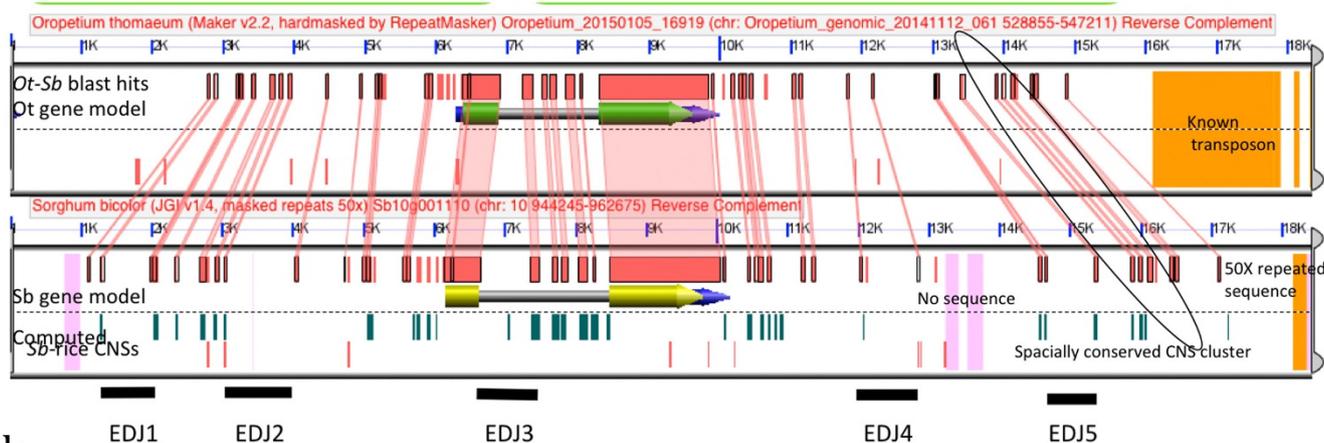
identity between monomers in the repeat decreases. **c**, The inverted repeat structure of the entire centromere on contig028 with a 60 kb spacer (blue box); arrows are as in **a**. **d**, Consensus 155 bp centromere monomer. **e**, Integrated genome browser view of centromere repeat, LTRs and predicted genes on contig028.



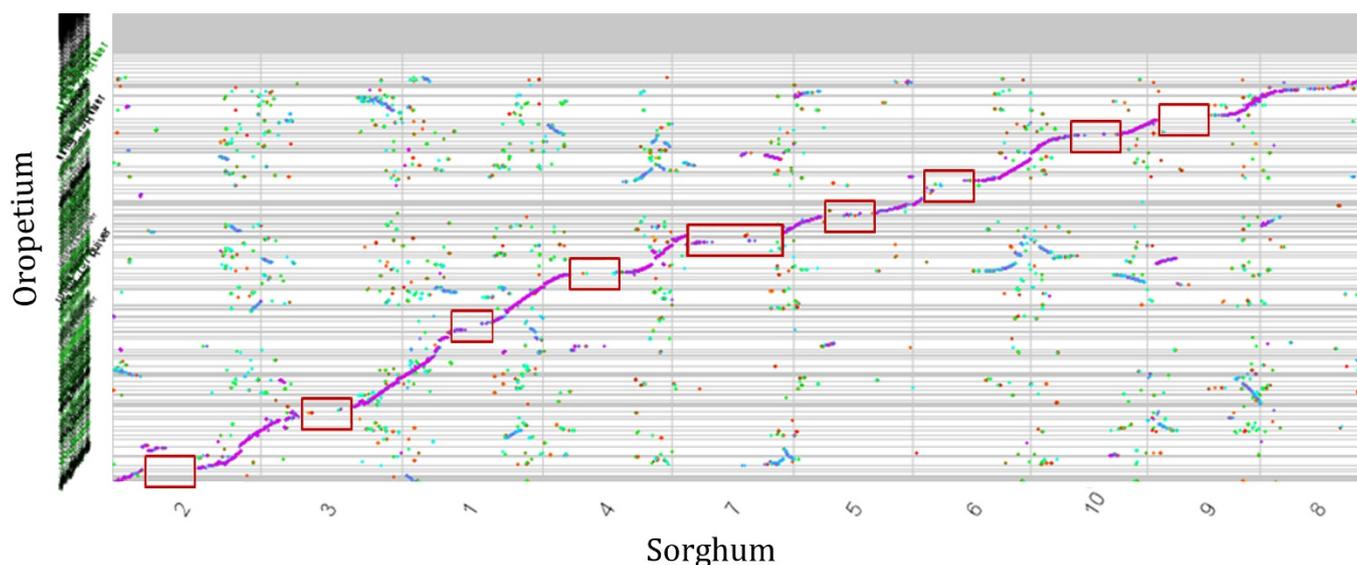
Extended Data Figure 3 | Macrocenty patterns and comparative genomics between the grasses. a–e, Macrocenty of *Oropetium* versus *Oropetium* (a); *Oropetium* versus *Brachypodium* (b); *Oropetium* versus rice (c); *Oropetium* versus *Setaria* (d); and *Oropetium* versus *Sorghum* (e). **f,** Genome compaction in *Oropetium* compared to related grass genomes.

Syntenic block span is based on regions that show conserved syntenly across all five genomes. Syntenic gene and coding DNA sequences span is based on 13,683 genes that are retained as genes in orthologous locations across all five genomes. The ratio compared to *Oropetium* is given in brackets.

a



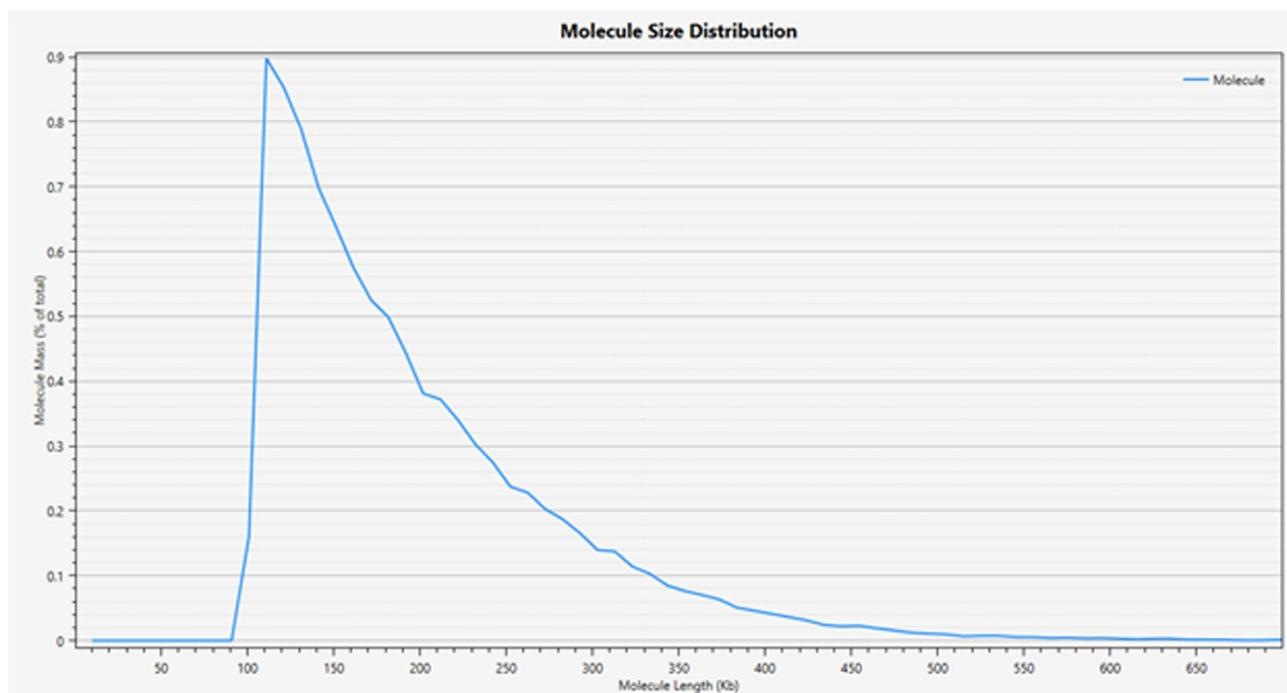
b



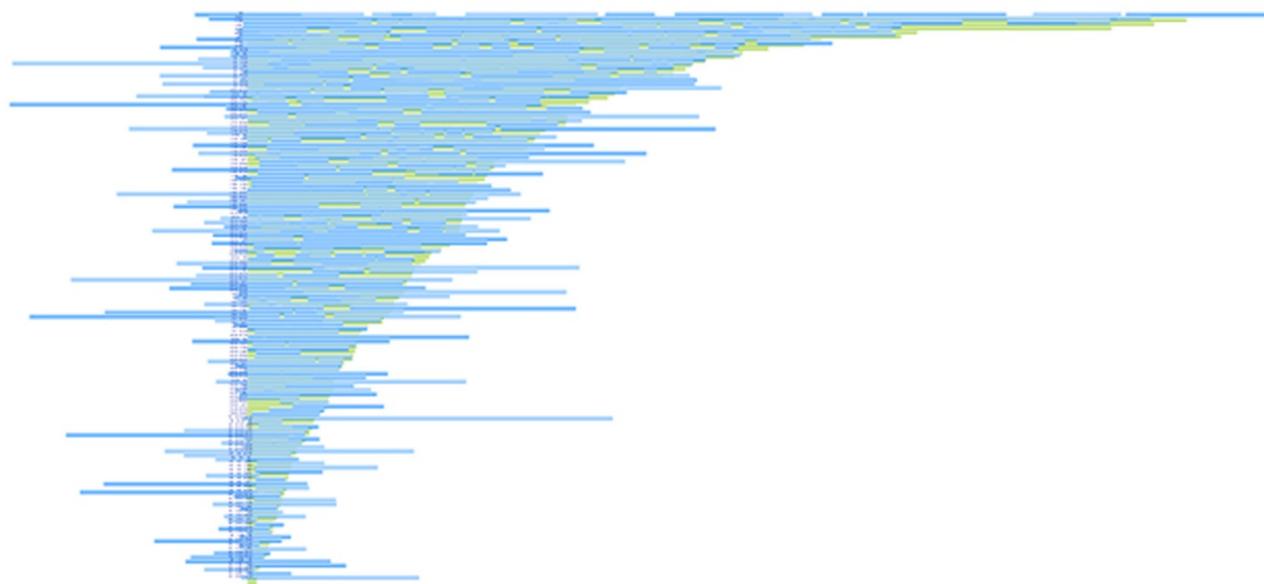
Extended Data Figure 4 | Expansion of intragenic and pericentric regions in *Sorghum* compared to *Oropetium*. **a**, A GEvo sequence similarity graphic of an *Oropetium* gene (upper) and its orthologous *Sorghum* gene (lower). Blast hits (high-scoring segment pairs) are denoted by red rectangles, and syntenic hits are connected by a red line. The green rectangles on the model line of *Sorghum* are conserved noncoding sequences (CNS) computed between *Sorghum* and rice; the expanse of CNS coverage defines 'gene space'. Within the oval are three CNS that may be spatially constrained. The expanded interspersed sequences are

annotated at the bottom in black. **b**, Pericentric region expansion in *Sorghum* compared to *Oropetium*. A syntenic dot plot of the *Sorghum* and *Oropetium* genomes is plotted. *Oropetium* contigs are ordered based on synteny with *Sorghum*. Hits are coloured based on K_s divergence, with purple blocks corresponding to 1:1 orthologous regions and other colours corresponding to retained genes from the rho and sigma WGDs. Pericentric regions in *Sorghum* have few syntenic matches to *Oropetium*, suggesting that much of the expansion occurred in pericentric regions.

a

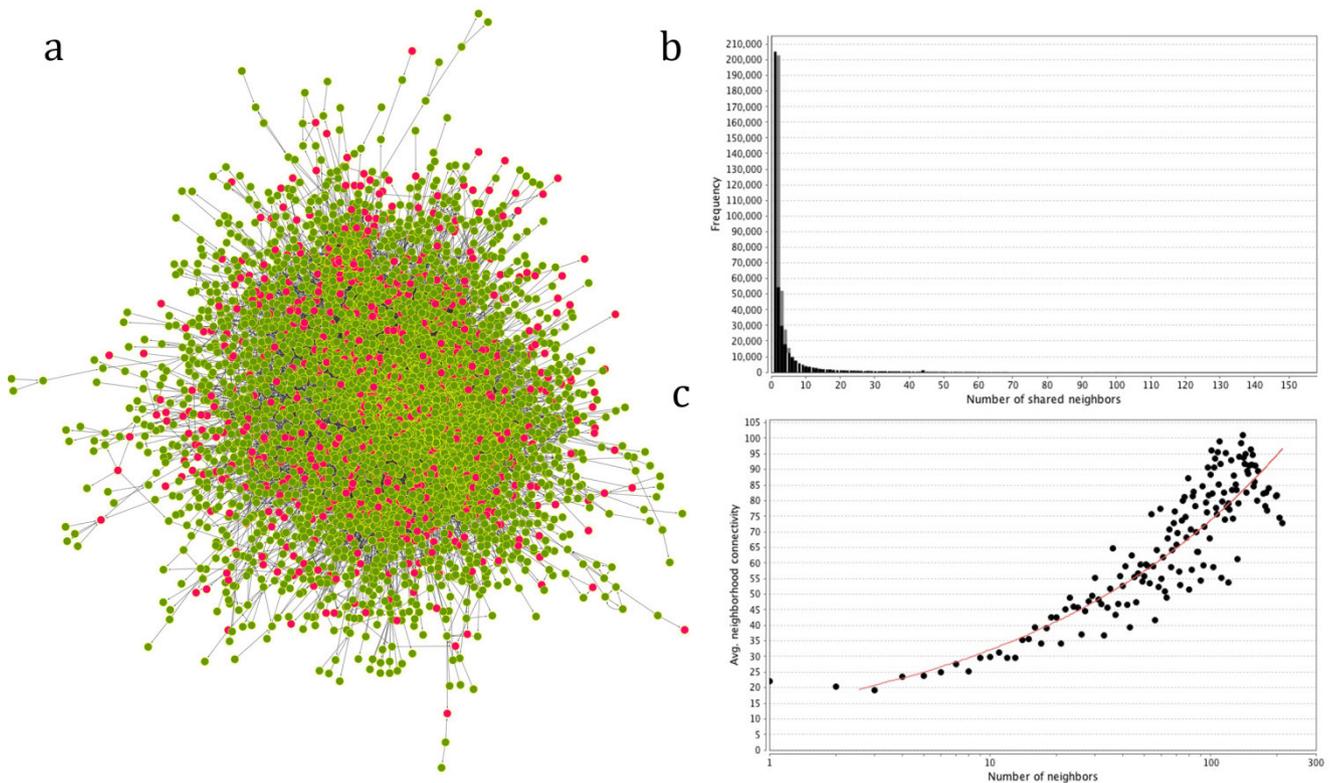


b



Extended Data Figure 5 | Assembly improvement using a BioNano-based genome map from the Irys system. **a**, Distribution of molecule size for raw single-molecule genome mapping data. Size of single molecules in nanochannel arrays is plotted. **b**, Integration of the genome map with

the genome assembly. Overlap between the PacBio-based contigs and the genome map. Each line shows a single PacBio contig in green; genome maps are shown in light blue.



Extended Data Figure 6 | Network statistics for tandem duplicated genes. a, Tandem duplicated genes in the metabolic network are shown in pink. b, Distribution of shared neighbours. c, The average number of neighbours.

Extended Data Table 1 | Telomere repeat (AAACCCT) locations and organization in the *Oropetium* genome

Contig Name	Contig Size (bp)	Start of telomeric array	End of telomeric array	Size of centromeric array (bp)	Teleomeric repeat sequence	Position of Telomere on contig	Number of telomeric repeats
Oropetium_genomic_143	99,304	1	6,446	6,445	AACCCTA	start	910.1
Oropetium_genomic_058	1,564,795	1,560,696	1,564,795	4,099	AGGGTTT	end	580.9
Oropetium_genomic_552	22,498	18,622	22,498	3,876	GTTTAGG	end	562.9
Oropetium_genomic_043	1,920,679	1	3,643	3,642	CCCTAAA	start	515.7
Oropetium_genomic_050	1,822,802	1	3,223	3,222	CCTAAAC	start	453.3
Oropetium_genomic_125	248,855	1	3,182	3,181	AAACCCT	start	452.4
Oropetium_genomic_027	2,706,558	2	2,092	2,090	AAACCCT	start	301.9
Oropetium_genomic_169	56,172	54,243	56,170	1,927	TTAGGGT	end	279.7
Oropetium_genomic_103	526,141	524,277	526,139	1,862	GTTTAGG	end	265.9
Oropetium_genomic_124	262,476	260,617	262,476	1,859	TTAGGGG	end	264
Oropetium_genomic_090	736,395	1	1,601	1,600	CCTAAAC	start	227.4
Oropetium_genomic_010	4,141,579	4,140,107	4,141,579	1,472	TTAGGGT	end	208.7
Oropetium_genomic_076	1,024,162	1	1,169	1,168	CCCTAAA	start	166.1
Oropetium_genomic_493	25,796	24,869	25,795	926	GGGTTTA	end	129.9
Oropetium_genomic_136	153,270	152,446	153,270	824	GTTTAGG	end	119.1
Oropetium_genomic_155	63,826	63,040	63,826	786	TTAGGGG	end	110.4
Oropetium_genomic_019	3,122,409	1	347	346	AAACCCT	start	48
Oropetium_genomic_149	80,145	1	294	293	AAACCCT	start	40.4

Extended Data Table 2 | rRNA tandem array locations and organization in the *Oropetium* genome

Contig Name	Contig Size (bp)	Start of NOR array (bp)	End of NOR array (bp)	Size of NOR tandem array (bp)	Position of NOR on contig	Number of NOR tandem repeats
Oropetium_genomic_182	51,716	1	51,716	51,716	spans contig	5.7
Oropetium_genomic_265	38,885	1	38,885	38,885	spans contig	4.3
Oropetium_genomic_168	56,772	1	56,772	56,772	spans contig	6.3
Oropetium_genomic_192	48,530	1	42,860	42,860	start	4.7
Oropetium_genomic_214	44,298	31,977	44,298	12,321	start	1.3
Oropetium_genomic_539	23,633	1	20,975	20,975	start	2.3

Extended Data Table 3 | Repeat annotation of the *Oropetium* genome

Repeat Class	Number of Elements	Percent Base Pairs Covered
Retrotransposon	214,698	35.60%
Long terminal repeat (LTR)	107,010	25.50%
Gypsy (RLG)	83,872	21.80%
Copia (RLC)	18,223	36.90%
Penelope (RPX)	1,548	0.15%
Unknown LTR (RLX)	3,367	0.44%
LINE (RIL)	17,399	1.90%
SINE (RSX)	2,735	0.07%
DIRS (RYD)	5,098	3.00%
Unknown retrotransposon (RXX)	82,456	7.50%
DNA transposon	69,217	8.50%
Maverick (DMX)	68	0.01%
TIR (DTX)	41,930	6.60%
Unknown DNA transposon (DXX)	27,219	1.90%
No category	7,902	1.00%
Total	291,817	43.80%

Extended Data Table 4 | Comparisons of repeats and coding features in the monocotyledons

Common name	Species name	chr. #	genome size (Mb)	repeat #	Gene #	Transcript statistics				Exon statistics			Intron statistics		
						Avg. Length	Median Length	Avg. Num Exons	Median Num Exons	Count	Avg. Length	Median Length	Count	Avg. Length	Median Length
Greater duckweed	<i>Spirodela polyrhiza</i>	20	150	23	19,519	4,718	3,015	5.22	3	101,867	222	129	82,368	757	202
Oropedium	<i>Oropetium thomaeum</i>	9	250	43	28,446	2,729	1,928	4.55	3	129,421	210	126	100,975	446	168
brachy	<i>Brachypodium distachyon</i>	5	272	21	42,868	3,819	3,128	5.38	4	154,738	254	137	120,380	402	142
rice	<i>Oryza sativa</i>	12	403	35	66,338	3,191	2,701	4.4	3	238,247	331	162	177,497	389	166
setaria	<i>Setaria italica</i>	9	510	40	29,448	3,299	2,563	4.96	3	134,802	261	137	106,488	436	145
sorghum	<i>Sorghum bicolor</i>	10	818	62	40,599	2,745	2,189	4.74	3	160,151	252	140	122,497	326	133
corn	<i>Zea mays</i>	10	2,300	85	63,540	4,236	2,747	4.6	3	203,643	238	133	149,177	670	154