

DNA likely to match the quality of that from the skeleton found at Mezmaiskaya Cave². We recommend that any additional Neanderthal destined for destructive analysis should be carefully selected, taking into account its integrated thermal history.

We calculated the likely deterioration of DNA for a series of Holocene fossils and at Pleistocene sites^{1–8} for which we had reliable regional palaeoclimatic reconstructions^{9–11}. We assumed that depurination is the principal pathway of DNA decomposition in most environments¹² (activation energy, $E_a = 127 \text{ kJ mol}^{-1}$) and have taken account of altitude and the integrated regional palaeotemperature data in the calculations. For each sample, the expected deterioration in DNA by depurination alone is expressed in terms of its 'thermal age', which is the number of years required, at a constant 10 °C, to produce the degradation calculated from its thermal history.

Figure 1 shows that bones with thermal ages greater than that of the Feldhofer Neanderthal¹ (17,000 years at 10 °C) failed to yield DNA. The material from Mezmaiskaya², although this is at a lower latitude than Feldhofer¹, was cooler because of its altitude (1.3 km; ref. 2) and so has a lower thermal age. High altitude is a common factor in low-latitude sites that have yielded ancient DNA^{6,7} and is anticipated from thermal age analysis. The failure to amplify DNA samples in the polymerase chain reaction from specimens from two sites whose thermal ages are both younger than 17,000 years at 10 °C demonstrates that thermal age is not the only factor that preserves DNA quality for successful amplification. Nevertheless, our analysis indicates that the original Feldhofer Neanderthal DNA sequence¹ represents the current technical limit for retrieval.

When we subjected other Neanderthal sites from northwestern Europe to the same analysis, only nine (of 39) cave sites are thermally younger than Feldhofer, and none strikingly so (see table in supplementary information). Despite the importance of gaining further insight into Neanderthal genetics, our results reveal that careful consideration is necessary before embarking on the destructive analysis of remains from all but a handful of sites.

Colin I. Smith*, Andrew T. Chamberlain†, Michael S. Riley‡, Alan Cooper§, Chris B. Stringer||, Matthew J. Collins*

*Fossil Fuels and Environmental Geochemistry (Postgraduate Institute), NRG, Drummond Building, University of Newcastle, Newcastle upon Tyne NE1 7RU, UK
e-mail m.collins@ncl.ac.uk

†Department of Archaeology, University of Sheffield, Northgate House, Sheffield S1 4ET, UK

‡School of Earth Sciences, University of Birmingham, Edgbaston, Birmingham B15 2TT, UK

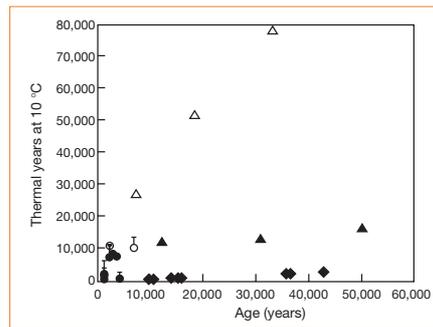


Figure 1 The success of amplifying ancient DNA by polymerase chain reaction^{1–8} is related to its thermal age — the thermal age of the original Neanderthal DNA amplification¹ represents the limit using current techniques. Filled symbols represent sites where ancient DNA has survived, clear symbols sites where it has not: diamonds, permafrost sites; triangles, cave sites; and circles, open sites. Thermal ages of open sites are calculated using mean annual air temperatures, with the upper range representing the effect of temperature fluctuation (estimated from monthly averages). The relation between air temperature and permafrost surface temperature is difficult to predict¹³, but these problems are largely offset because the rate of depurination is apparently weakly temperature-sensitive in frozen samples¹⁴. Methodological details are available from the authors.

§Departments of Biological Anthropology and Zoology, University of Oxford, Oxford OX2 6QS, UK
||Department of Palaeontology, The Natural History Museum, London SW7 5BD, UK

1. Krings, M. *et al.* *Cell* **90**, 19–30 (1997).
2. Ovchinnikov, I. *et al.* *Nature* **404**, 490–493 (2000).
3. Cooper, A. *et al.* *Science* **277**, 5329–5332 (1997).
4. Leonard, J. A., Wayne, R. K. & Cooper, A. *Proc. Natl Acad. Sci. USA* **97**, 1651–1654 (2000).
5. Höss, M., Pääbo, S. & Vereshchagin, N. K. *Nature* **370**, 333 (1994).
6. Cooper, A. *et al.* *Nature* **381**, 484 (1996).
7. Fleischer, R. C., Olson, S. L., James, H. F. & Cooper, A. C. *Auk* **117**, 1055–1060 (2000).
8. Colson, I. B., Bailey, J. F., Vercauteren, M. & Sykes, B. C. *Ancient Biomol.* **1**, 109–117 (1997).
9. Vandenbergh, J., Coope, R. & Kasse, K. *J. Quat. Sci.* **13**, 361–366 (1998).
10. Guiot, J. *et al.* *Palaeogeogr. Palaeoclimatol. Palaeoecol.* **103**, 73–93 (1993).
11. Elias, S. A. *Quat. Res.* **53**, 229–235 (2000).
12. Lindahl, T. & Nyberg, B. *Biochemistry* **11**, 3610–3618 (1972).
13. Zhang, T., Osterkamp, T. E. & Stamnes, K. *Permafrost Periglacial Process.* **8**, 45–67 (1997).
14. Osborne, M. R. & Phillips, D. H. *Chem. Res. Toxicol.* **13**, 257–261 (2000).

Supplementary information is available on Nature's website at <http://www.nature.com> or as paper copy from the London editorial office of Nature.

Ovchinnikov *et al.* reply — Smith *et al.* have shown an interesting correlation between the thermal age of Pleistocene–Holocene fossils and ancient DNA retrieval and advocate using this criterion to assess the merit of subjecting ancient bones to destructive analysis, particularly any scientifically valuable bones of Neanderthals and anatomically pre-modern humans. The post-mortem DNA quality is, of course, dependent on the surrounding temperature, but — as the authors point out — many other important factors influence fossil DNA preservation, such as air and soil humidity, soil pH, phosphorus content of the soil, average tempera-

ture in different earth layers, and microbial-mediated decay, which also have to be taken into account.

The complex interaction of several factors is illustrated by differential preservation of skeletons and bones from the same site, and even between different parts of the same skeleton or mummy. In addition, some specimens recovered from permafrost or glacial conditions after present-day thawing have shown variable degrees of mitochondrial DNA preservation (for example, in the Tyrolean Ice Man¹) or they contained no recoverable mitochondrial DNA (as discovered in the Altai Princess and Warrior mummies; I.V.O. and W.G., manuscript in preparation), illustrating that low temperatures alone are not sufficient to preserve all specimens.

Relying too heavily on temperature alone could lead to important specimens being excluded from DNA analysis. The Feldhofer Neanderthal is an outlier based on this model, which would predict that the possibility of recovering fossil DNA from this specimen would be very remote. Obtaining a sample of the Feldhofer Neanderthal for destructive analysis would presumably have been even more difficult if too much significance had been placed on such a model at the time the work was carried out.

Another example for which the thermal age may not be an accurate guide to the state of preservation is the Neanderthal recovered from the Marillac Cave in France. As this has a thermal age of 30,539 years, the model would predict that there should be very little chance of fossil DNA remaining. However, the collagen yield is comparable to that of the Mezmaiskaya Neanderthal, which had very good DNA preservation^{2,3}.

In conclusion, we agree that there is a real correlation between DNA survival and thermal history, but the accuracy to which the survival of DNA can be predicted is limited. Important specimens could be overlooked if too much emphasis is placed on the thermal age alone.

Igor V. Ovchinnikov*†, Anders Götherström§, Galina P. Romanovall, Vitaliy M. Kharitonov¶, Kerstin Lidén§, William Goodwin*

*Human Identification Centre, University of Glasgow, Glasgow G12 8QQ, UK
e-mail: w.goodwin@formed.gla.ac.uk

†Institute of Gerontology, Moscow 129226, Russia

‡Department of Medicine, Columbia University, New York, New York 10032, USA

§Archaeological Research Laboratory, Stockholm University, 106 91 Stockholm, Sweden

||Institute of Archaeology, Moscow 117036, Russia

¶Institute and Museum of Anthropology, Moscow State University, Moscow 103009, Russia

1. Handt, O. *et al.* *Science* **264**, 1775–1778 (1994).
2. Bocherens, H. *et al.* *J. Hum. Evol.* **20**, 481–492 (1991).
3. Ovchinnikov, I. V. *et al.* *Nature* **404**, 490–493 (2000).