Cytokeratin 20-negative Merkel cell carcinoma is infrequently associated with the Merkel cell polyomavirus

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Merkel cell carcinoma is a rare, highly aggressive cutaneous neuroendocrine carcinoma most commonly seen in sun-damaged skin. Histologically, the tumor consists of primitive round cells with fine chromatin and numerous mitoses. Immunohistochemical stains demonstrate expression of neuroendocrine markers. In addition, cytokeratin 20 (CK20) is expressed in ~95% of cases. In 2008, Merkel cell carcinoma was shown to be associated with a virus now known as Merkel cell polyomavirus in ~80% of cases. Prognostic and mechanistic differences between Merkel cell polyomavirus-positive and Merkel cell polyomavirus-negative Merkel cell carcinoma may exist. There has been the suggestion that CK20-negative Merkel cell carcinomas less frequently harbor Merkel cell polyomavirus, but a systematic investigation for Merkel cell polyomavirus incidence in CK20negative Merkel cell carcinoma has not been done. To test the hypothesis that Merkel cell polyomavirus is less frequently associated with CK20-negative Merkel cell carcinoma, we investigated 13 CK20-negative Merkel cell carcinomas from the files of the Cleveland Clinic and the University of Michigan for the virus. The presence or absence of Merkel cell polyomavirus was determined by quantitative PCR performed for Large T and small T antigens, with sequencing of PCR products to confirm the presence of Merkel cell polyomavirus. Ten of these (77%) were negative for Merkel cell polyomavirus and three (23%) were positive for Merkel cell polyomavirus. Merkel cell polyomavirus is less common in CK20-negative Merkel cell carcinoma. Larger series and clinical follow-up may help to determine whether CK20-negative Merkel cell carcinoma is mechanistically and prognostically unique.

Modern Pathology (2015) 28, 498-504; doi:10.1038/modpathol.2014.148; published online 14 November 2014

In 1875 Friedrich Merkel described the *Tastzellen*, or 'touch cells,' that now bear his name.¹ Nearly 100 years later, in 1972, Cyril Toker described trabecular carcinoma, named after its characteristic growth pattern, and favored a sweat gland origin for this tumor.² Later ultrastructural studies demonstrated dense core neurosecretory granules within the cells of 'trabecular carcinoma,' and subsequently the Merkel cell became a favored candidate for this malignancy's cell of origin.³ Recent evidence suggests that the cell of origin for Merkel cell carcinoma may be a Merkel cell precursor stem cell.⁴

Merkel cell carcinoma is among the most lethal cutaneous carcinomas, with significant mortality rate.⁵ Therefore, accurate distinction of Merkel cell carcinoma from histologically similar tumors is critical for prompt and appropriate management. Merkel cell carcinoma may display histologic overlap with basaloid or small cell cutaneous malignancies including basal cell carcinoma, adnexal carcinoma, lymphoma, and small-cell melanoma, although in most cases morphologic distinction is possible between Merkel cell carcinoma and these entities.⁶ However, Merkel cell carcinoma cannot be morphologically distinguished from cutaneous metastases of small cell carcinoma. Therefore, immunohistochemistry is critical for definitive diagnosis. Characteristically, Merkel cell carcinoma is positive for cytokeratins and neuroendocrine markers like synaptophysin, chromogranin, neurofilament, and neuron-specific enolase.^{6,7} Cytokeratin 20 has

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Received 8 July 2014; revised 2 September 2014; accepted 3 September 2014; published online 14 November 2014

become a standard marker for Merkel cell carcinoma and is positive in ~95% of cases, frequently with a classic paranuclear dot-like pattern of staining.⁷⁻⁹ Cytokeratin 20 expression is absent in most other small round cell malignancies including metastatic small cell lung carcinoma, as well as less-aggressive cutaneous malignancies such as basal cell carcinoma.^{6–8}

Lack of cytokeratin 20 expression has been described in $\sim 5\%$ of Merkel cell carcinoma.^{6,10-12} Diagnosis of cytokeratin 20-negative Merkel cell carcinoma typically involves immunohistochemical studies for cytokeratins other than cytokeratin 20 (including AE1/3, Cam5.2, or CK7) and neuroendocrine markers. The important differential diagnosis in this context is metastatic small cell lung carcinoma. Thyroid transcription factor-1 is a useful marker for small cell lung carcinoma; although it may be negative in 10-15% of cases and has been reported to be expressed in exceptional cases of Merkel cell carcinoma.^{7,8,13,14} Neurofilament protein expression is frequently observed in Merkel cell carcinoma but not in small cell lung carcinoma.^{7,15} Although it has been speculated that cytokeratin 20 negativity may be the result of de-differentiation,¹⁶ it is unclear whether the absence of cytokeratin 20 expression is an isolated finding in Merkel cell carcinoma, or whether these tumors are characterized by distinct molecular changes and prognostic differences relative to cytokeratin 20-positive Merkel cell carcinoma.

A breakthrough in understanding of the molecular pathogenesis of Merkel cell carcinoma came in 2008, when Merkel cell polyomavirus was shown to be present in a high percentage of Merkel cell carcinomas.¹⁷ Merkel cell polyomavirus may promote tumorigenesis via oncogenic actions of viral small T antigen and large T antigen, which are generated from a single gene via alternative splicing.⁵ Merkel cell polyomavirus associated with Merkel cell carcinoma displays tumor-specific mutations or deletions in large T antigen which render the virus replication deficient, a common characteristic of oncogenic viruses.^{5,18} Although numerous assays for Merkel cell polyomavirus detection have been reported, PCR targeting small T antigen and/or the 5' region of large T antigen is commonly used and highly sensitive.^{19–21}

Although present in the majority of Merkel cell carcinoma, Merkel cell polyomavirus has not been commonly observed in other malignancies. In particular, the presence of Merkel cell polyomavirus is highly specific for Merkel cell carcinoma relative to small cell lung carcinoma,^{12,22–24} and hence might be considered as a diagnostic marker in challenging cases including cytokeratin 20-negative Merkel cell carcinoma. However, studies examining the association between cytokeratin 20 negativity and the presence of Merkel cell polyomavirus in Merkel cell carcinoma have been limited to case reports or

incidental findings within larger case series. To our knowledge, there have been seven total reported cases in the literature of cytokeratin 20-negative cutaneous Merkel cell carcinoma assessed for Merkel cell polyomavirus status; of these, five were negative for Merkel cell polyomavirus.^{12,13,25,26} We hypothesized that Merkel cell polyomavirus is less frequently associated with cytokeratin 20-negative Merkel cell carcinomas.

Better understanding of the incidence of Merkel cell polyomavirus in cytokeratin 20-negative Merkel cell carcinoma has implications for the diagnostic utility of Merkel cell polyomavirus in this setting and may yield clues regarding the molecular pathogenesis of this subset of Merkel cell carcinomas. Therefore, we examined the incidence of Merkel cell polyomavirus by PCR in a cohort of 13 cytokeratin 20-negative Merkel cell carcinoma tumors.

Materials and methods

Case Selection

All studies were conducted with prior approval by the Institutional Review Board. The archives of the Cleveland Clinic and the University of Michigan were searched for cytokeratin 20-negative Merkel cell carcinomas. For inclusion in the study, the tumors had to have typical histologic features of Merkel cell carcinoma and compatible immunohistochemical profiles including positivity for cytokeratins coupled with positivity for at least one neuroendocrine marker. The medical charts were reviewed, or for cases received in consultation, contributors or treating physicians were contacted to establish that the tumors were clinically consistent with Merkel cell carcinoma and that there was no evidence of a neuroendocrine carcinoma for a different site.

Immunohistochemistry

All of the immunohistochemical stains were reviewed. Most were performed at the time of initial diagnosis either by the referring institutions or at the Cleveland Clinic or the University of Michigan. A limited number of stains were performed at the Cleveland Clinic for the purposes of this study including cytokeratin 7 (DAKO, Carpinteria, CA, USA; clone OV-TL 12/30, dilution 1:40), neuron-specific enolase (DAKO; clone BBS/ NC/VI-H14, dilution 1:50), chromogranin A (DAKO; clone DAK-A3, dilution 1:100), and synaptophysin (BioGenex, Fremont, CA, USA; clone Snp88, prediluted) using the Ventana BenchMark XT or Bench-Mark Ultra autostainer (Ventana Medical Systems, Tucson, AZ, USA).

DNA Extraction and PCR

Five $4 \mu m$ tissue sections per specimen were deparaffinized and DNA was extracted using the

commercial microbead system (NucliSENS-easy-Mag, BioMírieux, Durham, NC, USA), following the manufacturer's instructions, as previously described.²³ DNA quantification was performed and diluted if necessary to achieve DNA concentrations between 3 and $50 \text{ ng}/\mu$ l. Qualitative PCR was performed for Merkel cell polyomavirus using previously reported primers sets including two (LT1 and MCVPS1) targeting the large T antigen and a third (LT3) predominantly targeting the small T antigen unique region (Table 1).¹⁹ Quantitative PCR for β -globin was used as the positive control to confirm the presence of amplifiable DNA. Negative controls included purified, distilled water and a sample of aortic valve tissue from a patient unaffected by Merkel cell carcinoma.

Quantitative PCR with LT1 and LT3 primer sets was performed using SybrGreen fluorescence (Life Technologies, Carlsbad, CA, USA). Threshold cycles were calculated for all samples. All samples were subsequently submitted for Sanger sequencing to confirm the presence of Merkel cell polyomavirus.

MCVPS1 quantitative PCR was performed using a TaqMan[®] probe (Life Technologies) for its high sensitivity and specificity.²⁷ The TaqMan[®] probe used was designed to specifically identify nucleotides 1104–1123 (5'-3' sequence GGACCCGA-TATACCTCCCGAAC) within the sequence amplified by the MCVPS1 primer set. All 14 samples were evaluated with the MCVPS1 primer set in conjunction with the MCVPS1 TaqMan[®] probe.

Studies evaluating the melting temperature of double-stranded PCR products were consistent with expected melting temperatures of LT1 and LT3 double primer sets and support the results of PCR and sequencing.

Results

Thirteen cases of Merkel cell carcinoma, occurring in 7 men and 6 women ranging in age from 56 to 93 years (mean 72), were identified out of ~380 cases in the combined archives. The tumors arose on the head and neck (n=7), upper extremities (n=3), lower extremities (n=2), and in an unspecified location (n=1). None of the patients had evidence of a neuroendocrine carcinoma from another potential primary. Patients treated at the authors' institutions all had appropriate clinical evaluations, including radiologic staging studies. For all cases seen in consultation with the exception of one, we were able to obtain additional clinical information. None of the consultation cases had evidence of a neuroendocrine carcinoma elsewhere. These patients had detailed clinical and radiologic evaluations. For the case in which additional information was unable to be obtained, the patient was a 93-year-old man with a lesion on the nose (see below).

Histologically, the tumors showed characteristic features of Merkel cell carcinoma, including trabecular or nodular proliferations of primitive round cells with high nuclear-to-cytoplasmic ratios, frequent mitoses, and a stippled ('salt and pepper') chromatin pattern. Nuclear molding was present at least focally in six of the cases (Figure 1). There were no morphologic findings to suggest basal cell carcinoma or adnexal carcinoma with aberrant neuroendocrine marker expression.

All our cases were positive for at least one cytokeratin (Figure 2; Table 2); however, the most common pattern was diffuse cytoplasmic or

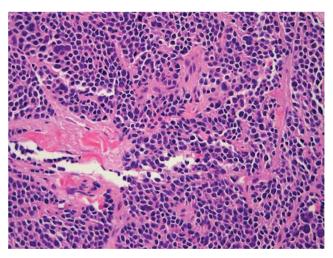


Figure 1 Histologic features of the cases. All of the cases of cytokeratin 20-negative Merkel cell carcinoma had typical histologic findings of Merkel cell carcinoma. The tumors were composed of cells with high nuclear-to-cytoplasmic ratios with a neuroendocrine chromatin pattern. Prominent nuclear molding was seen in six cases.

Table 1 Prin	mers used for MCP	vV detection (LT	1. LT3. MCVPS1) and control (β -globin)

Primer	NT location	Forward sequence	Reverse sequence	Probe	Amplicon size (bp)	Reference
LT1	1514–1953	TACAAGCACTCCACCAAAGC	TCCAATTACAGCTGGCCTCT	N/A	440	19
LT3	571-879	TTGTCTCGCCAGCATTGTAG	ATATAGGGGCCTCGTCAACC	N/A	308	19
MCVPS1	1071-1179	TCAGCGTCCCAGGCTTCAGA	TGGTGGTCTCCTCTCTGCTACTG	GGACCCGATAT ACCTCCCGAAC	109	19
β -Globin		ACACAACTGTGTTCACTAGC	CAACTTCATCCACGTTCACC	N/A	110	

Abbreviations: bp, base pair; NT, nucleotide.

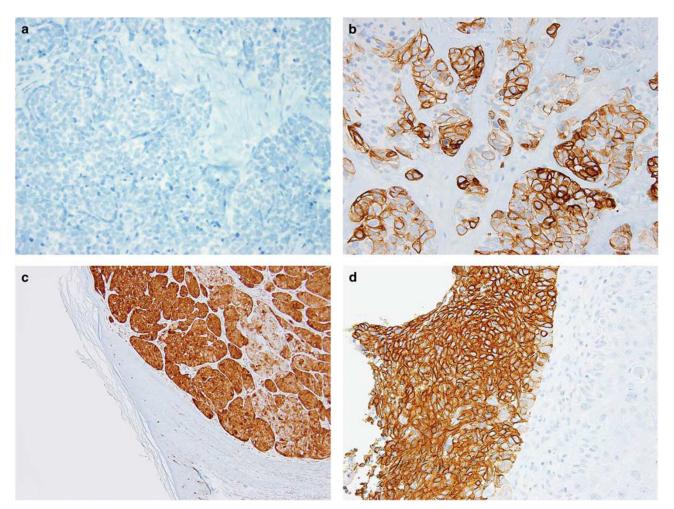


Figure 2 Summary of immunohistochemical stains. All specimens were negative for cytokeratin 20 (a); cytokeratin 7 was positive in a cytoplasmic, membranous pattern in two of our samples (b); all cases were positive for neuroendocrine markers such as neuron-specific enolase (c); other keratin immunohistochemical stains such as Cam5.2 usually had a diffuse cytoplasmic or membranous staining similar to stains for cytokeratin 7 rather than a perinuclear dot-like pattern (d).

membranous and not the usual paranuclear dot-like pattern. All had immunoreactivity for at least one neuroendocrine marker (Figure 2; Table 2). Synaptophysin was the most commonly positive neuroendocrine marker (6/8), while chromogranin was less sensitive (4/9). None of the cases tested were positive for thyroid transcription factor-1 (13/13). Dot-like immunoreactivity for neurofilament protein was seen in two of three consultation cases.

The one case in which there was no detailed clinical information had typical histologic features of Merkel cell carcinoma but also had a squamous cell carcinoma component that included overlying squamous cell carcinoma *in situ*. This case also exhibited dot-like immunoreactivity for neurofilament protein.

The specimens were analyzed by PCR for MCVPS1, LT1, LT3, and β -globin. Ten cases (77%) were negative for Merkel cell polyomavirus. Three (23%) were positive for Merkel cell polyomavirus (Figure 3). Of these, two were positive for all three

primer sets tested, including MCVPS1 and LT1 (targeting large T antigen) and LT3 (predominantly targeting small T antigen). The remaining case was positive for MCVPS1 and LT3 (Table 3). PCR products for LT1 and LT3 were further analyzed by Sanger sequencing, which demonstrated >98% similarity to the Merkel cell polyomavirus genome (NC_010277.1) for all PCR amplicons (Figure 4). Case 4 displayed a C1923T mutation resulting in a premature stop codon in large T antigen, consistent with a tumor-specific truncating mutation.

Discussion

Merkel cell carcinoma is a primary cutaneous neuroendocrine carcinoma that typically occurs in older, fair-skinned patients. Merkel cell carcinoma is aggressive with metastatic rates of 36–55% and overall 5-year mortality between 31 and 64%.^{28–30} The diagnosis of Merkel cell carcinoma is confirmed

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with immunohistochemical stains for distinction from other small cell malignancies, including metastatic small cell lung carcinoma.^{6–8} Particularly useful is cytokeratin 20, which is positive in ~95% of Merkel cell carcinomas but negative in most small cell lung carcinomas.^{6–8}

Increased insight into the pathogenesis of Merkel cell carcinoma was gained in 2008 with the discovery that Merkel cell polyomavirus was genomically integrated in a significant subset of Merkel cell carcinoma.¹⁷ Merkel cell polyomavirus appears to be a common constituent of normal microfauna on the skin of healthy individuals.^{5,31} However, evidence suggests that Merkel cell polyomavirus detected in Merkel cell carcinoma has an oncogenic role, rather than incidental detection of background infection by wild-type virus. Merkel cell polyomavirus in Merkel cell carcinoma displays clonal integration, high viral copy number, and tumorspecific mutations of large T antigen resulting in incapacity to replicate.^{5,17,18,32} Furthermore, *in vitro* studies support a role for Merkel cell polyomavirus T antigens, especially small T antigen, in transformation and cell survival.⁵

Table 2. Summary of immunohistochemical stains

Stain	Positive/Total
Cytokeratins	
Cytokeratin 20	0/13
Cytokeratin-7	2/12
AE1/AE3	11/11
Cam 5.2	2/4
Any cytokeratin	13/13
Neuroendocrine markers	
NSE	2/3
Synaptophysin	7/9
Chromogranin	4/10
CD56	3/4
Neurofilament	2/3
Any neuroendocrine marker	13/13
Other	
TTF-1	0/12

Abbreviations: CK20, cytokeratin 20; CK7, cytokeratin 7; NSE, neuron-specific enolase; TTF-1, thyroid transcription factor 1.

Most studies have reported that a significant minority of Merkel cell carcinoma lack detectable Merkel cell polyomavirus,⁵ raising the question of how Merkel cell polyomavirus-negative tumors might differ from Merkel cell polyomavirus-positive tumors. Reports have been mixed regarding immunohistochemical and clinical differences between Merkel cell polyomavirus-positive and polyomavirus-negative Merkel cell tumors.³¹ Merkel cell carcinoma with squamous elements or coexisting squamous cell carcinoma is often Merkel cell polyomavirus negative.^{12,23,25,33–36} Recent findings indicate that Merkel cell polyomavirusnegative tumors may be associated with RB1 inactivating mutations³⁷ and (in a subset) PIK3CA activating mutations.³⁸ Hence, although additional study is needed to clarify differences between Merkel cell polyomavirus-positive and Merkel cell polyomavirus-negative tumors, current data suggest that molecular differences exist that may have implications for immune-based or targeted therapy.

It is unclear what role Merkel cell polyomavirus plays in the rare subset of Merkel cell carcinoma that are cytokeratin 20 negative. We undertook a study to examine series of cytokeratin 20-negative Merkel cell carcinoma to further explore the prevalence of

 Table 3 MCPyV positivity as detected by LT1 and LT3 (PCR, sequencing) and MCVPS1 (Taqman[®] probe-based PCR)

Case	MCVPS1	LT1	LT3
1	_	_	_
2	_	_	_
3	_	_	_
4	+	+	+
5	_	_	_
6	-	_	_
7	-	_	_
8	—	_	_
9	_	_	_
10	+	+	+
11	—	_	_
12	_	_	_
13	+	_	+
MKL-1	+	+	+

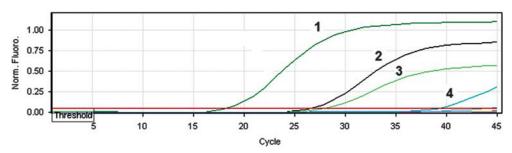


Figure 3 PCR melting curve analysis for Merkel cell polyomavirus. PCR melting curve for Merkel cell polyomavirus LT3: 1— Amplification of the Merkel cell polyomavirus cell line in the positive control cell line. 2, 3, and 4 demonstrate the amplification of the MCV in patients 4, 10, and 13, respectively. All of the other specimens were negative for Merkel cell polyomavirus.

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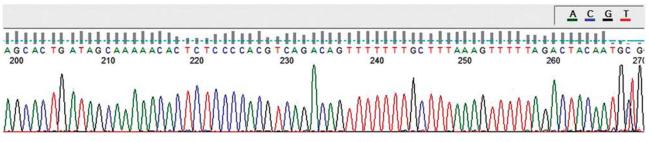


Figure 4 Example of Sanger sequencing for a Merkel cell polyomavirus-positive case. For the Merkel cell polyomavirus-positive cases, Sanger sequencing demonstrated >98% similarity to the Merkel cell polyomavirus genome (NC_010277.1) for all PCR amplicons.

Merkel cell polyomavirus in this subset of Merkel cell carcinoma. Our PCR results suggest that Merkel cell polyomavirus is uncommon in cytokeratin 20negative Merkel cell carcinoma, as 10/13 (77%) were negative for the virus. This finding agrees with the few previous reports of cytokeratin 20-negative Merkel cell carcinoma evaluated for Merkel cell polyomavirus, the majority of which were Merkel cell polyomavirus negative.^{12,13,25,26}

These results have diagnostic implications. Lack of cytokeratin 20 expression may make definitive diagnosis of Merkel cell carcinoma challenging, and additional markers may be desirable in this context. Detection of Merkel cell polyomavirus either by immunohistochemistry or by molecular means has been proposed as a diagnostic test for Merkel cell carcinoma, which may aid in distinction from other carcinomas, including metastatic small cell carcinoma.^{12,22-24} Our results suggest that Merkel cell polyomavirus detection is insufficiently sensitive to be useful in the diagnosis of cytokeratin 20-negative Merkel cell carcinoma. Other markers such as thyroid transcription factor-1 and neurofilament may be more useful in this context.7,15,26

In summary, our data suggest that cytokeratin 20negative Merkel cell carcinoma is more likely to lack detectable Merkel cell polyomavirus. Further investigations will address whether Merkel cell polyomavirus-negative/cytokeratin 20-negative Merkel cell carcinoma displays genetic changes similar to other Merkel cell polyomavirus-negative Merkel cell carcinoma such as *RB1* mutations, or whether these tumors possess a unique spectrum of mutations and hence represent a distinct molecular subclass of Merkel cell carcinoma.

Conclusions

We have shown through three primer sets that cytokeratin 20-negative Merkel cell carcinoma is associated with a low incidence of Merkel cell polyomavirus positivity. Further study of cytokeratin 20-negative, Merkel cell polyomavirus-negative Merkel cell carcinoma may prove helpful in elucidating genetic changes and prognosis of this subset of Merkel cell carcinoma.

Disclosure/conflict of interest

The authors declare no conflict of interest.

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