

NOTE

Cycloexpansamines A and B: spiroindolinone alkaloids from a marine isolate of *Penicillium* sp. (SF-5292)

Chiwook Lee^{1,2}, Jae Hak Sohn², Jae-Hyuk Jang³, Jong Seog Ahn³, Hyuncheol Oh¹, Jonas Baltrusaitis⁴, In Hyun Hwang⁵ and James B Gloer⁵

The Journal of Antibiotics (2015) 68, 715–718; doi:10.1038/ja.2015.56; published online 13 May 2015

Marine microorganisms are recognized as an important source of structurally diverse bioactive secondary metabolites.^{1,2} Particularly, secondary metabolites from marine fungal isolates have attracted significant interest, as many of them provide unique structural features and interesting biological properties.^{2–4} In our search for new metabolites from marine-derived fungi, we recently reported the isolation of an anti-inflammatory metabolite named penicillinolide A from extracts obtained from cultures of a marine-derived isolate of *Penicillium* sp. (SF-5292).⁵ As part of our continuing efforts to explore the chemistry of this isolate, studies of extracts obtained from larger-scale cultures of the fungus were undertaken. From this investigation, two minor metabolites, cycloexpansamines A (**1**) and B (**2**) were isolated (Figure 1) in addition to previously encountered penicillinolide A.

The fungal strain was cultured on 80 petri-dish plates (90 mm), each containing 20 ml of potato dextrose agar media (0.4% (w/v) potato starch, 2% (w/v) dextrose, 3% (w/v) NaCl, 1.5% (w/v) agar). Plates were individually inoculated with 2-ml seed cultures of the fungal strain. Plate cultures were incubated at 25 °C for a period of 14 days. Extraction of the agar media with EtOAc (2 × 500 ml) provided an organic phase, which was then concentrated *in vacuo* to yield 1.9 g of an extract. The EtOAc extract was subjected to C₁₈ flash column chromatography (5 × 40 cm), eluting with a stepwise gradient of 20, 40, 60, 80 and 100% (v/v) MeOH in H₂O (500 ml each). The fractions eluted at 80% MeOH (418 mg) were purified by semi-preparative reversed-phase HPLC eluting with a gradient from 40 to 100% MeOH in H₂O (0.1% formic acid) over 56 min to yield compound **1** (10.2 mg, *t_R* = 15.9 min). The fraction collected between 17 and 18 min in the above HPLC step was then subjected to further HPLC, eluting with a gradient from 40 to 65% MeOH in H₂O (0.1% formic acid) over 40 min to yield compound **2** (1.4 mg, *t_R* = 27.6 min).

Cycloexpansamine A (**1**) gave an [M+H]⁺ ion at *m/z* 450.2380 in the HRESI mass spectrum, indicating a molecular formula of C₂₆H₃₁N₃O₄, which was fully supported by the ¹H and ¹³C NMR data (Table 1). Analysis of its ¹³C NMR and DEPT spectra revealed the

presence of four methyl groups, two sp² methines, one sp³ methine and seven sp³ methylene units (including one bound to a heteroatom), as well as three carbonyl and four non-protonated aromatic/olefinic carbons. Because only six out of the 13 unsaturations were accounted for by these units, cycloexpansamine A (**1**) was required to contain seven rings. The structure of **1** could be fully defined by comprehensive analysis of 1D and 2D NMR data, including ¹H, ¹³C NMR, COSY, HSQC and HMBC experiments (Table 1). A literature search indicated that the planar structure of **1** was proposed for a metabolite of another unidentified *Penicillium* sp. in a recent US patent.⁶ However, ¹H and ¹³C NMR position assignments were not provided, nor were HRMS data and stereochemical assignments. Therefore, our attention turned into the comprehensive structure determination of **1**, including absolute configuration.

The relative configuration of compound **1** was determined by analysis of NOESY data (Figure 2). Since **1** consists of separated fused ring systems connected at a spiro-carbon atom, the two ring systems are virtually perpendicular to one another. NOESY correlations of H-10 with H₃-28 and H-23β, and of H-23β with NH-25 indicated that one of the methyl groups at C-13, the C1–C11 bond and the amide group (C-24) are β-oriented. This was also supported by the observation of NOESY correlations of H₃-28 with H-23β. On the other hand, NOESY correlations of H₃-29 with H-14; of H-14 with H-21α and H-23α; and of H-21α with H-23α suggested α-orientations for H-14, H-21α, H-23α and H₃-29. Therefore, the relative configuration of **1** was suggested as shown.

The absolute configuration of **1** was proposed by analysis of its electronic circular dichroism (ECD) spectrum and by comparison to its calculated spectrum (Supplementary Information). The Cotton effect at 250–350 nm is considered to be related to the configuration at the spirogenic center in ECD spectra of spiro-oxindole alkaloids.^{7–9} The ECD spectrum of **1** (Supplementary Figure S1) showed a positive Cotton effect in the spiro-oxindole absorbance region around 270 nm, which suggested the 1S configuration for **1**, and this assignment is in agreement with the configurations previously established for related

¹Department of Pharmacy, College of Pharmacy, Wonkwang University, Iksan, Korea; ²Department of Bio-food Materials, College of Medical and Life Sciences, Silla University, Busan, Korea; ³Chemical Biology Research Center, Korea Research Institute of Bioscience and Biotechnology (KRIBB), Cheongwon, Korea; ⁴PhotoCatalytic Synthesis Group, University of Twente, Enschede, The Netherlands and ⁵Department of Chemistry, University of Iowa, Iowa City, IA, USA
Correspondence: Professor H Oh, Department of Pharmacy, College of Pharmacy, Wonkwang University, Iksan, 570-749, Korea.
E-mail: hoh@wonkwang.ac.kr

Received 28 February 2015; revised 3 April 2015; accepted 13 April 2015; published online 13 May 2015

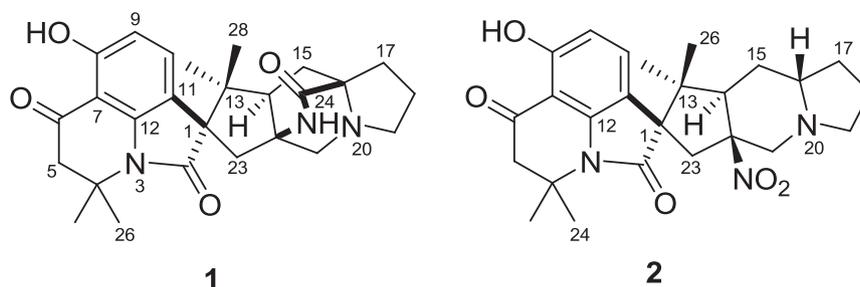


Figure 1 Structures of cycloexpansamines A (**1**) and B (**2**).

Table 1 NMR spectroscopic data for cycloexpansamine A (**1**) in pyridine-*d*₅

No.	δ_C^a	δ_H , mult. (J in Hz) ^b	Key NOESY ^c	HMBC (H → C#)
1	64.9	—	—	—
2	183.0	—	—	—
4	57.2	—	—	—
5	51.6	2.67, d (16.5) 2.99, d (16.5)	—	4, 6, 7, 26, 27
6	197.5	—	—	—
7	104.9	—	—	—
8	160.0	—	—	—
9	108.8	6.60, d (8.4)	—	6, 7, 8, 11
10	134.3	7.26, d (8.4)	H-23 β , H-28,	1, 8, 12
11	119.2	—	—	—
12	147.9	—	—	—
13	47.8	—	—	—
14	49.3	3.39, t (8.4)	H-15 α , H-21 α , H-29,	13, 15, 21, 22, 28, 29
15	32.7	(α) 1.90, m (β) 1.78, m	H-14 H-28	14, 16, 17, 22, 24
16	67.3	—	—	—
17	27.8	3.01, m 1.51, m	—	16, 24 15, 16, 24
18	24.9	1.86–1.99, m	—	17, 19
19	54.0	3.06, m 2.28, dd (17.2, 8.4)	—	16 18, 21
21	64.0	(α) 3.75, d (8.4) (β) 2.49, d (8.4)	H-23 α —	14, 15, 16, 22, 23 14, 19, 22
22	63.5	—	—	—
23	40.4	(β) 2.47, d (15.0) (α) 2.38, d (15.0)	H-10, H-25, H-28 H-21 α	2, 11 1, 2, 13, 21
24	174.0	—	—	—
25	—	9.06, s	H-23 β	14, 16, 22, 23
26	23.9	1.43, s	—	4, 5, 27
27	26.8	1.77, s	—	4, 5, 26
28	21.3	1.12, s	H-10, H-15 β , H-23 β	1, 13, 14, 29
29	23.3	0.84, s	H-14	1, 13, 14, 28

^aRecorded at 100 MHz.

^bRecorded at 400 MHz.

^cRecorded at 600 MHz.

compounds containing spiro-oxindole units, such as synthetic *ent*-(+)-paraherquamide B¹⁰ and (+)-versicolamide B.⁷ In an effort to provide independent support for this assignment, the ECD spectrum of an energy-minimized conformer of **1** (generated using Spartan '10, Wavefunction, Inc., CA, USA) was simulated using time-dependent density functional theory (TDDFT) calculations, and

compared with the experimental data for **1**. The experimental ECD spectrum of **1** matched closely with the calculated ECD spectrum. Therefore, on the basis of both empirical considerations and TDDFT calculations, the absolute configuration of **1** was assigned as shown.

Cycloexpansamine B (**2**) was assigned the molecular formula C₂₅H₃₁N₃O₅ (11 unsaturations) on the basis of high-resolution electrospray ionisation mass spectrometry (HRESIMS) and NMR data (Table 2). Analysis of its ¹³C NMR and DEPT spectra revealed the presence of four methyl groups, two sp² methines, two sp³ methines and seven sp³ methylene units, as well as two carbonyl and four non-protonated aromatic or olefinic sp² carbons. Analysis of the ¹H and ¹³C NMR data suggested that the overall structure of **2** is similar to that of **1**. The main difference was evident in the absence of one carbonyl signal in the ¹³C NMR spectrum, and changes in the chemical shifts corresponding to the C-15, C-16 (sp³ methine instead of sp³ quaternary carbon), and C-22 positions of **1**. These observations suggested that **2** differs from **1** by the absence of the bridged amide ring system and associated structural modification around this functionality. The planar structure of **2** was finally inferred by considering this information together with a comprehensive analysis of 2D NMR data as well as close comparisons of the data with those reported for related metabolites, cyclopiamides^{11,12} and citrinalins.¹³

The relative configuration of **2** was determined by analysis of the NOESY spectrum (Figure 2) and by analogy to cycloexpansamine A (**1**). As in the case of **1**, NOESY correlations of H₃-26 with H-10 and H-23 β indicated that one of the methyl groups at C-13, the C1–C11 bond and H-23 β were β -oriented. On the other hand, NOESY correlations of H-14 with H-17 α , H₃-27 and H-21 α suggested α -orientations for H-14, H-17 α and H-21 α . This assignment in turn suggested β -orientation for the nitro group at C-22. Finally, NOESY correlations of H-15 β with H-16, and of H-15 α with H-14, H-17 α and H₃-27 suggested β -orientation of H-16. Therefore, the relative configuration of **2** was proposed as shown. In addition, by analogy to the recently proposed biogenesis of nitro group containing prenylated indole alkaloids from bicyclo[2.2.2]diazaoctane-containing metabolites (discussed below), the absolute configuration of **2** was proposed to be analogous to that of **1**.

Cycloexpansamine A (**1**) is a novel heptacyclic spiroindolinone alkaloid consisting of a 4,5-dihydro-1*H*-pyrrolo[3,2,1-*ij*]quinoline-2,6-dione ring system and an amide-bridged cyclopenta[*f*]indolizidine ring system connected via a spiro-carbon atom. Compound **1** is structurally related to the family of prenylated indole alkaloids containing the bicyclo[2.2.2]diazaoctane ring system as a core structure.⁷ This family of natural products, including the brevianamides, paraherquamides, stephacidin, asperparalines, marcfortines, notoamides, malbrancheamides, avrainvillamide and sclerotiamide, has been reported mainly from various fungi of the genera *Aspergillus* and *Penicillium*,¹⁰ and several biological activities, including

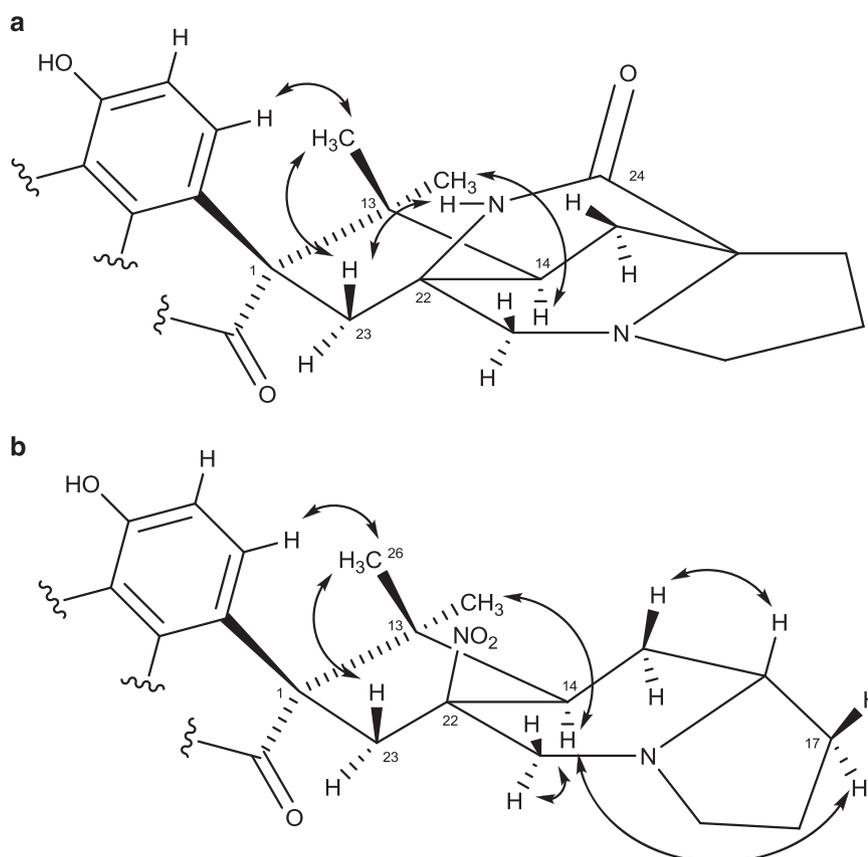


Figure 2 Key NOESY correlations (a) for the amide-bridged cyclopenta[*f*]indolizidine portion of cycloexpansamine A (**1**), and (b) for the cyclopenta[*f*]indolizidine portion of cycloexpansamine B (**2**).

insecticidal, antitumor, anthelmintic, calmodulin inhibitory and antibacterial properties, are displayed by members of this family.⁷ Because of their complex structures and diverse bioactivities, studies of synthetic and biosynthetic pathways leading to these alkaloids has become an area of significant interest.^{13–16} In line with this, it is noteworthy that cycloexpansamine A (**1**) represents a rare example of prenylated indole alkaloids with the anti-relative configuration between the C13–C14 and C16–N24 bonds in the [2.2.2]diazaoctane core system.^{7,17} In addition, cycloexpansamine A (**1**) is the only member of this family that contains a 4,5-dihydro-1*H*-pyrrolo[3,2,1-*ij*]quinoline-2,6-dione ring system derived from condensation of the indole ring and a prenyl group.

Cycloexpansamine B (**2**) is closely related to cyclopiamines A¹¹ and B,^{11,12} which are the only precedent secondary metabolites possessing a 4,5-dihydro-1*H*-pyrrolo[3,2,1-*ij*]quinoline-2,6-dione ring system. The relative configurations of cyclopiamine B was originally established by single-crystal X-ray crystallography.¹¹ In the course of the structure elucidation of cyclopiamine B (a diastereomer of cyclopiamine A), the desmethylated analog of cyclopiamine B, which possesses the same planar structure as that of **2**, was prepared by treatment with aluminum chloride in nitrobenzene.¹⁸ Recently, *O*-desmethylcyclopiamine B has been asymmetrically synthesized in the course of total synthesis of cyclopiamine B possessing absolute configuration of 1*R*, 14*R*, 16*R* and 22*S*.⁹ The structure of **2** differs from *O*-desmethylcyclopiamine B only in the configurations at stereogenic centers. To confirm the diastereomeric relationship between *O*-desmethylcyclopiamine B and **2**, the NMR data of **2** in CDCl₃ were compared with those reported for the asymmetrically synthesized *O*-desmethylcyclopiamine B in the same

solvent. Although a comprehensive comparison was hampered by the limited availability of **2**, clear differences in proton shift values for prominent peaks, including the aromatic and methyl proton signals were observed (**2**: aromatic ¹H signals at δ_H 7.41 (br d, *J*=7.0, 1H) and 6.57 (br d, *J*=7.0 Hz, 1H); methyl ¹H signals at δ_H 1.77 (s, 3H), 1.46 (s, 3H), 0.97 (s, 3H) and 0.80 (s, 3H); *O*-desmethylcyclopiamine B: aromatic ¹H signals at δ_H 7.20 (br d, *J*=8.3, 1H) and 6.48 (br d, *J*=8.3 Hz, 1H); methyl ¹H signals at δ_H 1.73 (s, 3H), 1.41 (s, 3H), 1.03 (s, 3H) and 0.89 (s, 3H)). These data support the conclusion that **2** is a diastereomer of asymmetrically synthesized *O*-desmethylcyclopiamine B, possessing inverted configurations at C-1, C-14 and C-16 compared to the respective positions in *O*-desmethylcyclopiamine B. It is also noteworthy that encountering fungal metabolites possessing a nitro group is unusual. Cyclopiamines A and B from *Penicillium cyclopium*,¹¹ cyclopiamin B from *Aspergillus caespitosus*¹² and citrinalins A, B¹³ and 17-hydroxycitrinalin B¹⁸ from *P. citrinum* are rare examples of precedent fungal metabolites bearing such functionality. It has been suggested that cyclopiamines and citrinalins biogenetically arise from a bicyclo[2.2.2]diazaoctane precursor *via* hydrolysis of the amide bridge, decarboxylation and amino group oxidation to the nitro group. However, such a precursor was unknown until a recent report of the co-isolation of nitro group containing 17-hydroxycitrinalin B and a bicyclo[2, 2, 2]diazaoctane-containing precursor.¹⁸ Thus, the isolation of cycloexpansamines A and B is an additional example of co-isolation of biogenetically related metabolites, and this supports the existence of a common bicyclo[2.2.2]diazaoctane-containing biogenetic precursor to nitro group containing prenylated indole alkaloids.

Table 2 NMR spectroscopic data for cycloexpansamine B (**2**) in pyridine-*d*₅

No.	δ_c^a	δ_H , mult. (<i>J</i> in Hz) ^b	Key NOESY ^c	HMBC (<i>H</i> → <i>C</i> #)
1	61.8	—	—	—
2	182.3	—	—	—
4	57.0	—	—	—
5	51.1	2.66, d (16.5) 2.99, d (16.5)	—	4, 6, 7, 24 4, 6, 24, 25
6	197.0	—	—	—
7	104.7	—	—	—
8	159.7	—	—	—
9	108.9	6.68, d (8.3)	—	7, 8, 11
10	134.1	7.28, d (8.3)	H-23 β , H-26	1, 8, 12
11	118.6	—	—	—
12	147.5	—	—	—
13	47.4	—	—	—
14	47.5	3.15, m	H-15 α , H-17 α , H-21 α , H-23 α , H-27	15, 21
15	22.4	(β) 2.95, m (α) 1.61, m	H-16, H-26 H-14, H-17 α	13, 16, 17, 22 13, 16, 22
16	58.8	3.16, dd (9.9, 4.3)	H-15 β	13, 15, 21
17	27.5	(α) 1.69, m (β) 1.63, m	H-14, H-15 α H-15 β	18 18
18	22.6	(β) 1.81, m (α) 1.50, m	H-19 β H-19 α	17 —
19	54.3	(β) 2.70, m (α) 2.65, m	H-18 β , H-21 β H-18 α	16 18, 21
21	60.5	(β) 3.22, d (11.0) (α) 2.98, d (11.0)	H-19 β H-14	14, 16, 22 16, 19
22	95.9	—	—	—
23	43.7	(β) 3.08, d (15.8) (α) 2.67, d (15.8)	H-26 H-14	1, 2, 11, 13, 22 1, 2, 13, 22
24	23.7	1.39, s	—	4, 5, 25
25	26.5	1.74, s	—	4, 5, 24
26	22.9	0.94, s	H-10, H-15 β , H-23 β	1, 13, 27
27	23.4	0.96, s	H-14, H-15 α	1, 14, 26

^aRecorded at 150 MHz.^bRecorded at 600 MHz.^cRecorded at 600 MHz.

Cycloexpansamine A (**1**) moderately inhibited the activity of protein tyrosine phosphatase 1B,^{19,20} a promising and a validated therapeutic target to effectively treat type 2 diabetes mellitus and obesity, showing an IC₅₀ value of 27.6 μ M, while **2** was inactive against protein tyrosine phosphatase 1B at levels up to 100 μ M.

ACKNOWLEDGEMENTS

We acknowledge the financial support by grants from the Global R&D Center (GRDC, NRF-2010-00719) programs of the National Research Foundation of Korea (NRF) funded by the Ministry of Science, ICT and Future Planning of Korea (MSIFP).

- Fenical, W. & Jensen, P. R. Developing a new resource for drug discovery: marine actinomycete bacteria. *Nat. Chem. Biol.* **2**, 666–673 (2006).
- Bugni, T. S. & Ireland, C. M. Marine-derived fungi: a chemically and biologically diverse group of microorganisms. *Nat. Prod. Rep.* **21**, 143–163 (2004).
- Saleem, M. *et al.* Marine natural products of fungal origin. *Nat. Prod. Rep.* **24**, 1142–1152 (2007).
- Rateb, M. E. & Ebel, R. Secondary metabolites of fungi from marine habitats. *Nat. Prod. Rep.* **28**, 290–334 (2011).
- Lee, D.-S. *et al.* Penicillinolide A: a new anti-inflammatory metabolite from the marine fungus *Penicillium* sp. SF-5292. *Mar. Drugs* **11**, 4510–4526 (2013).
- Onodera, H. *et al.* (Kyowa Hakko Kirin Co., Ltd.). Proliferation promoting agent for neural stem cells. US2011/0020931 A1 (2011).
- Greshock, T. J. *et al.* Isolation, structure elucidation, and biomimetic total synthesis of versicolamide B, and the isolation of antipodal (-)-stephacidin A and (+)-notoamide B from *Aspergillus versicolor* NRRL 35600. *Angew. Chem. Int. Ed. Engl.* **47**, 3573–3577 (2008).
- Takayama, H. *et al.* Stereochemical studies on the *Uncaria* alkaloid, 3-oxo-7-hydroxy-3,7-secorhynchophylline: the absolute configuration of 3-hydroxyoxindole derivatives. *Tetrahedron* **55**, 6841–6846 (1999).
- Mugishima, T. *et al.* Absolute stereochemistry of citrinadins A and B from marine-derived fungus. *J. Org. Chem.* **70**, 9430–9435 (2005).
- Tsukamoto, S. *et al.* Notoamides F-K, prenylated indole alkaloids isolated from a marine-derived *Aspergillus* sp. *J. Nat. Prod.* **71**, 2064–2067 (2008).
- Bond, R. F., Boeyens, J. C. A., Holzapfel, C. W. & Steyn, P. S. Cyclopiamines A and B novel oxindole metabolites of *Penicillium cyclopium* westling. *J. Chem. Soc. Perkin Trans. 1*, 1751–1761 (1979).
- Steyn, P. S., Vlegaar, R. & Rabie, C. J. Alkaloids from *Aspergillus caespitosus*. *Phytochemistry* **20**, 538–539 (1981).
- Pimenta, E. F. *et al.* Use of experimental design for the optimization of the production of new secondary metabolites by two *Penicillium* species. *J. Nat. Prod.* **73**, 1821–1832 (2010).
- Kato, H. *et al.* Notoamides A–D: Prenylated indole alkaloids isolated from a marine-derived fungus, *Aspergillus* sp. *Angew. Chem. Int. Ed. Engl.* **46**, 2254–2256 (2007).
- Grubbs, A. W., Artman, G. D. III, Tsukamoto, S. & Williams, R. M. A. Concise total synthesis of the notoamides C and D. *Angew. Chem. Int. Ed. Engl.* **46**, 2257–2261 (2007).
- Tsukamoto, S. *et al.* Isolation of antipodal (-)-versicolamide B and notoamides L-N from a marine-derived *Aspergillus* sp. *Org. Lett.* **11**, 1297–1300 (2009).
- Cai, S. *et al.* Isolation and photoinduced conversion of 6-epi-stephacidins from *Aspergillus taichungensis*. *Org. Lett.* **15**, 2168–2171 (2013).
- Mercado-Marin, E. V. *et al.* Total synthesis and isolation of citrinalin and cyclopiamine congeners. *Nature* **509**, 318–324 (2014).
- Vintonyak, V. V., Antonchick, A. P., Rauh, D. & Waldmann, H. The therapeutic potential of phosphatase inhibitors. *Curr. Opin. Chem. Biol.* **13**, 272–283 (2009).
- Johnson, T. O., Ermolieff, J. & Jirousek, M. R. Protein tyrosine phosphatase 1B inhibitors for diabetes. *Nat. Rev. Drug. Discov.* **1**, 696–709 (2002).

Supplementary Information accompanies the paper on The Journal of Antibiotics website (<http://www.nature.com/ja>)