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ORIGINAL ARTICLE

In vitro activity of two old antibiotics against clinical isolates of methicillin-resistant Staphylococcus aureus

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The objective of this paper was to investigate the *in vitro* effects of fusidic acid combined with fosfomycin against methicillin-resistant *Staphylococcus aureus* (MRSA). In all, 196 MRSA strains isolated from three clinical specimens of human infections from hospitals in China were used in this study. The checkerboard method was used to determine whether combinations act synergistically against these strains. The susceptibility results for fusidic acid and fosfomycin were interpreted according to the guidelines of the Clinical and Laboratory Standards Institute. The combination of fusidic acid and fosfomycin demonstrated the following interactions: 87.76% (172/196) synergism, 12.24% (24/196) indifference and no antagonism was seen (minimum and maximum fractional inhibitory concentration index 0.14 and 0.75, respectively). Thus, combinations of fusidic acid and fosfomycin show synergism for most of the MRSA isolates tested in this study, and may be a future therapeutic alternative for infections caused by MRSA.

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Keywords: fosfomycin; fractional inhibitory concentration (FIC); fusidic acid; methicillin-resistant *Staphylococcus aureus* (MRSA); synergism

INTRODUCTION

It is well known that methicillin-resistant Staphylococcus aureus (MRSA) causes life-threatening infections, such as meningitis, bone marrow destruction, sepsis, endocarditis, septic pulmonary emboli and extensive soft tissue infection in humans; MRSA has become the most prevalent pathogen worldwide.^{1,2} Currently, over 60% of S. aureus isolates from intensive care units in the USA and over 50% of isolates from inpatient non-intensive care unit hospital settings are caused by MRSA.^{3,4} In Asia, MRSA is highly endemic in health care settings with prevalence rates of >50% in many hospitals. In Hong Kong, MRSA is known to be endemic in hospitals since mid 1980s.⁵ Vancomycin is an effective agent in treating serious infections due to MRSA.6 However, other possibilities should be explored for those patients unable to tolerate the use of this drug or in the case of development of resistance to vancomycin among the strains of MRSA.⁷ In recent years, new antistaphylococcal antibiotics, such as linezolid, daptomycin, tygecicline or telavancin, have been developed, but their cost is high. The infections caused by MRSA are serious and are difficult to treat. Therefore, we attempted to find an effective and safe combination of antibiotics to treat the infections caused by MRSA. A number of old antibiotic compounds, such as fosfomycin and fusidic acid, are reemerging as valuable alternatives for the treatment of difficult-to-treat infections.8

Fusidic acid has a high degree of activity against *S. aureus*, including MRSA. ^{9–11} However, the development of resistance during treatment with fusidic acid when this antibiotic is used alone has been reported. ^{5,12,13} Resistance to fusidic acid can be produced readily in

the laboratory by growing *S. aureus* in the presence of increasing concentrations of this antibiotic.¹⁴

Fosfomycin is a structurally unique antibiotic that is chemically unrelated to any other antimicrobial agent. Fosfomycin is a phosphoenolpyruvate analog that irreversibly inhibits enolpyruvate transferase resulting in the prevention of formation of *N*-acetyl-muramic acid, the first stage of peptidoglycan synthesis of the bacterial cell wall. Fosfomycin is bactericidal against MRSA and has also been reported to directly interact with the immune system by affecting T lymphocyte function. Fosfomycin is a structurally unique antibiotic that is chemically unrelated to an appropriate transferase resulting in the prevention of the propriate transferase resulting in the prevention of the propriate transferase resulting in the prevention of the pre

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MATERIALS AND METHODS

Bacterial isolates

We collected and evaluated 196 methicillin-resistant *S. aureus* strains from three Chinese hospitals (PLA General Hospital 98 strains, Beijing Hospital 33 strains and Peking Union Medical College Hospital 65 strains) in this study. This isolates were collected from the blood (29%), nose (7%), sputum (21%), wound sites (15%), skin (23%), urine (3%), unspecified (3%) of the patients. All the strains were identified by VITEK-2 system (Biomerieux, Marcy l'Etoile, France). *S. aureus* American type culture collection 25923 was used for quality control in the latex agglutination test. SLIDEX Staph Plus (Biomerieux, France) is a rapid latex agglutination test for the identification of *S. aureus* strains. Disk agar diffusion method (Kirby-Bauer) was used to test susceptibility. Broth dilution testing was used for determining *in vitro* effects of fusidic acid combined with fosfomycin. *S. aureus* ATCC 29213 was used as quality control strain for these tests.¹⁸

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Antimicrobials

All antibiotics were obtained from the National Institute for the control of Pharmaceutical and Biological Products (Beijing, China). The purity of fusidic acid was 96% and fosfomycin 99%. Antibiotic powders were used to prepare stock solutions at concentrations of $1024\,\mu g\,ml^{-1}$ as recommended by the Clinical and Laboratory Standards Institute. 18

Minimum inhibitory concentration (MIC) determination and synergy testing

Initially, all the isolates were tested against a single compound using the broth microdilution method according to Clinical and Laboratory Standards Institute standards. Synergy tests were performed in 96-well broth microdilution plates, containing two antimicrobial agents in twofold dilutions dispensed in a checkerboard format.¹⁹

Fosfomycin was dispended alone in the first row with concentration ranging from 0.5 to $256\,\mu g\,ml^{-1}$, whereas fusidic acid was dispended in the first column with concentration ranging from 0.03 to $2\,\mu g\,ml^{-1}$. For seven strains, fusidic acid concentration ranging from 0.25 to $16\,\mu g\,ml^{-1}$ was used. The concentrations were set according to the MIC values obtained in a preliminary test. Inocula were prepared by suspending growth from agar plates in Mueller–Hinton broth to a density of 0.5 McFarland standard and were diluted 1:10 to produce a final inoculum of 1.5×10^5 colony-forming units per ml with a multipoint inoculators. The trays were incubated aerobically overnight. Standard quality control strains were incubated with each run. Interpretations of the antimicrobial combinations were based on calculation of the fractional inhibitory concentration index (FICI) for each drug pair. The FICI was calculated as the MIC of drug A and B in combination divided by the MIC of drug A or B alone, according to the following formula: 20,21

 $FICI = (MIC \ of \ DrugA \ in \ combination/MIC \ of \ DrugA \ alone) \\ + (MIC \ of \ the \ DrugB \ in \ combination/MIC \ of \ the \ DrugB \ alone)$

FICI≤0.5	Synergism
0.5 <fici≤4< td=""><td>Indifferent</td></fici≤4<>	Indifferent
FICI>4	Antagonistic

Statistical analysis

Statistical analysis was performed by using the SAS statistical program.

RESULTS

MIC for fusidic acid combined with fosfomycin against MRSA

The MIC $_{50}$ and MIC $_{90}$ for the strains tested in combination, respectively, were fusidic acid 0.03 and 0.125 µg ml $^{-1}$, and for fosfomycin 8 and 16 µg ml $^{-1}$, respectively, whereas alone they were fusidic acid 0.125/0.5 µg ml $^{-1}$ and fosfomycin 64/128 µg ml $^{-1}$, respectively. Fusidic acid MIC values for 9.2% of the strains were > 1 µg ml $^{-1}$, whereas fosfomycin MIC values for 66.8% of the strains were ≥ 64 µg ml $^{-1}$ (Table 1). However, when they were combined fusidic acid MIC values for 99.7% of the strains were ≤ 1 µg ml $^{-1}$. Although fosfomycin MIC values for all of the strains were ≤ 32 µg ml $^{-1}$.

Table 1 Minimum inhibitory concentration (MIC) (in $\mu g \, ml^{-1}$) of fusidic acid combined with fosfomycin in *Staphylococcus aureus* (*N*=196)

	Single			Combination		
Antimicrobial	MIC ₅₀	MIC ₉₀	MIC range	MIC ₅₀	MIC ₉₀	MIC range
Fusidic acid Fosfomycin	0.125 64	0.5 128	0.03–8 8–256	0.0300 8	0.125 16	0.0008–2 0.5–32

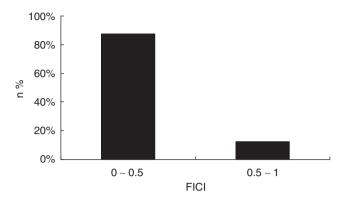


Figure 1 Fractional inhibitory concentration index (FICI) of fusidic acid combined with fosfomycin in methicillin-resistant *Staphylococcus aureus* (N=196).

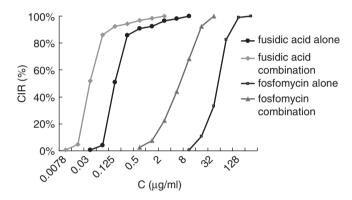


Figure 2 The cumulative inhibition ratio (CIR) of fusidic acid and/or fosfomycin against methicillin-resistant *Staphylococcus aureus* (*N*=196). C, concentration.

FICI of fusidic acid combined with fosfomycin for MRSA

Of the strains tested, 172 isolates (87.75%) showed synergistic interaction (FICI \leq 0.5) and 24 isolates (12.25%) showed indifference (0.5 < FICI < 1) (Figure 1). No antagonism (minimum and maximum fractional inhibitory concentration index 0.14 and 0.75, respectively) was observed.

The cumulative inhibition ratio of fusidic acid and/or fosfomycin against MRSA

Cumulative inhibition ratio of fusidic acid and/or fosfomycin against MRSA are shown in Figure 2. The curve of the cumulative inhibition ratio was moved to the left when the two drugs were used together, which suggested the potential utility of this combination.

DISCUSSION

MRSA has reemerged as a global public health problem.¹² Over the last decade, MRSA has become increasingly common in hospitals and in community settings.^{22–24} Combination antimicrobial is an important treatment approach for infections. Synergy is one of the most common reasons for using combination antimicrobial therapy. Thus, the search for combinations of antibiotics might yield more effective treatment options and fewer side effects.

Some compounds are unable to inhibit or kill bacteria by themselves, but can block bacterial mechanisms of resistance, enhancing the activity of other antimicrobials administered in combination. As the emergence of resistance has been associated with monotherapy, 25 some authors found that fusidic acid in combination with linezolid

avoided the appearance of resistance to the partner drug.²⁶ Some data suggests a synergistic activity of fosfomycin with various antistaphylococcal agents.¹⁶ Moreover, fosfomycin is far less expensive than linezolid.²⁵ Although fosfomycin has limited *in vitro* activity against MRSA, our data suggests its combination with fusidic acid is synergistic against MRSA. Moreover, fosfomycin has the advantage of low toxity and high safety.

In our study using 196 MRSA isolates from hospitals in China, fusidic acid combined with fosfomycin was synergistic with 87.76% of strains: 12.24% showed indifference and no antagonistic were observed. Most of the isolates evaluated in the study were intermediate susceptible or resistant to fusidic acid and fosfomycin. Most of the isolates had MIC values within the susceptible range for fusidic acid, whereas only 66.8% for fosfomycin. However, in combinations, fusidic acid MIC values for 99.7% of the strains were $\leq 1 \text{ ml}^{-1}$, whereas fosfomycin MIC values for all of the strains were $\leq 32 \,\mu \text{g ml}^{-1}$. From our in vitro results, the combination of fusidic acid with fosfomycin is active at lower concentrations than the single agents. This combination could also prevent the emergence of resistance to current antistaphylococcal drugs. Thus fosfomycin and fusidic acid should be used in combination when treating MRSA infections to prevent the emergence of resistance to either of these antibiotics. From the cumulative inhibition ratio curve we can still see the notable difference when fusidic acid and fosfomycin were tested in combination, supporting the combination as a future therapeutic alternative provided that pharmacokinetics could be optimized for both drugs.

In conclusion, combinations of fusidic acid with fosfomycin have significant synergistic activity against most of the strains of MRSA that were tested. The presence of fofomycin and fusidic acid is crucial to prevent further emergence of these resistant strains. For this reason, the combinations may be useful in the treatment of infections caused by MRSA. Further studies, especially animal models will be required to establish if the *in vitro* synergistic activity observed with fosfomycin and fusidic acid is also evident *in vivo*.

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