

## SHORT COMMUNICATION

## Temperate phages enhance pathogen fitness in chronic lung infection

Emily V Davies<sup>1</sup>, Chloe E James<sup>1,2</sup>, Irena Kukavica-Ibrulj<sup>3</sup>, Roger C Levesque<sup>3</sup>, Michael A Brockhurst<sup>4,5</sup> and Craig Winstanley<sup>1,5</sup><sup>1</sup>Department of Clinical Infection, Microbiology and Immunology, Institute of Infection and Global Health, University of Liverpool, Liverpool, UK; <sup>2</sup>School of Environment and Life Sciences, University of Salford, Manchester, UK; <sup>3</sup>Institut de Biologie Intégrative et des Systèmes (IBIS), Université Laval, Québec, Québec, Canada and <sup>4</sup>Department of Biology, University of York, York, UK

**The Liverpool Epidemic Strain (LES) is a polylysogenic, transmissible strain of *Pseudomonas aeruginosa*, capable of superinfecting existing *P. aeruginosa* respiratory infections in individuals with cystic fibrosis (CF). The LES phages are highly active in the CF lung and may have a role in the competitiveness of the LES *in vivo*. In this study, we tested this by competing isogenic PAO1 strains that differed only by the presence or absence of LES prophages in a rat model of chronic lung infection. Lysogens invaded phage-susceptible populations, both in head-to-head competition and when invading from rare, in the spatially structured, heterogeneous lung environment. Appreciable densities of free phages in lung tissue confirmed active phage lysis *in vivo*. Moreover, we observed lysogenic conversion of the phage-susceptible competitor. These results suggest that temperate phages may have an important role in the competitiveness of the LES in chronic lung infection by acting as anti-competitor weapons.**

*The ISME Journal* (2016) 10, 2553–2555; doi:10.1038/ismej.2016.51; published online 12 April 2016

The LES is a transmissible strain of *Pseudomonas aeruginosa* that causes life-limiting chronic respiratory infections in individuals with CF (Fothergill *et al.*, 2012). Unusually, the LES is capable of superinfection to displace other strains of *P. aeruginosa*, even after years of chronic colonisation (McCallum *et al.*, 2001; Winstanley *et al.*, 2009), but there is no evidence that it can be displaced by other strains (Fothergill *et al.*, 2010; Mowat *et al.*, 2011; Williams *et al.*, 2015). The LES harbours five prophages, highly active in the CF lung (James *et al.*, 2015), four of which have been shown by signature-tagged mutagenesis to be necessary for bacterial competitiveness in a rat model of chronic lung infection (Winstanley *et al.*, 2009; Lemieux *et al.*, 2015). Theoretical and empirical studies suggest that temperate phages could enhance the competitiveness of lysogens by killing phage-susceptible competitors by lysis (Bossi *et al.*, 2003; Brown *et al.*, 2006; Joo *et al.*, 2006; Burns *et al.*, 2014), a form of phage-mediated allelopathy (Stewart and Levin, 1984). However, it is unclear whether this ecological

mechanism operates in the far more complex spatially structured and heterogeneous lung environment.

To test whether temperate phages increase competitive fitness during lung infection, we performed competition experiments between lysogenic and non-lysogenic strains of *P. aeruginosa* in a rat model of chronic lung infection (Winstanley *et al.*, 2009). We constructed antibiotic-resistance-labelled PAO1 LES-Phage Lysogens (PLPLs) (James *et al.*, 2012) using three of the LES phages (LES $\phi$ 2, LES $\phi$ 3 and LES $\phi$ 4), both individually and in combination (for full methods, see Supplementary information). Initial *in vitro* experiments suggested that the triple lysogen (PAO1 $\phi$ triple) was more invasive than any of the constituent single lysogens (Supplementary Figure S1), therefore PAO1 $\phi$ triple was selected for use in the *in vivo* experiments. Competitors were embedded in agar beads in PLPL to PAO1 $\phi$ <sup>-</sup> ratios of 1:5 or 1:1, to model invasion-from-rare and head-to-head competition, respectively. Rats were infected with inoculated agar beads by intubation and monitored for 7 days, after which they were killed. The densities of each competitor in the lungs were quantified (Supplementary Table S2) and the selection rate constant ( $r_{ij}$ ) was calculated as described previously (Lenski *et al.*, 1991).

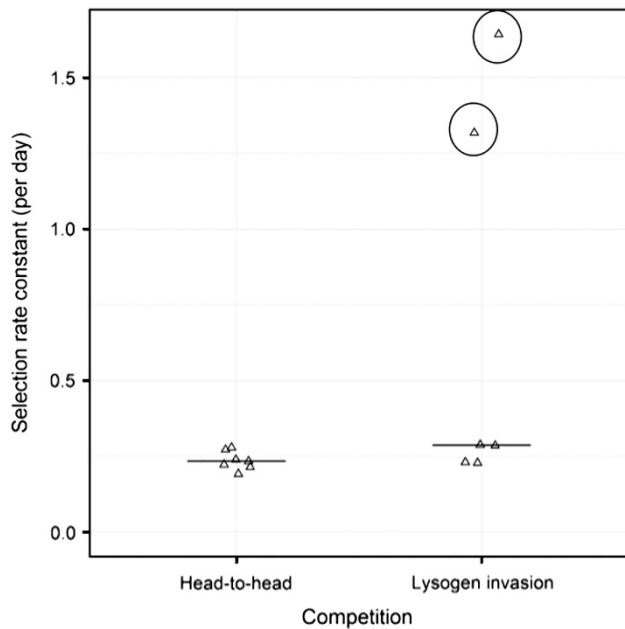
PAO1 $\phi$ triple outcompeted PAO1 $\phi$ <sup>-</sup> *in vivo* (Figure 1), at both initial starting frequencies (competition experiment, 1 sample *t*-test (alt = 0),  $t_6 = 20.3$ ,  $P < 0.001$ ; invasion experiment, 1 sample *t*-test (alt = 0),  $t_3 = 15.8$ ,  $P < 0.01$ ),

Correspondence: C Winstanley, Department of Clinical Infection, Microbiology and Immunology, Institute of Infection and Global Health, University of Liverpool, Ronald Ross Building, 8 West Derby Street, Liverpool L69 7BE, UK.  
E-mail: C.Winstanley@liv.ac.uk

<sup>5</sup>These authors contributed equally to this work.

Received 4 December 2015; revised 19 February 2016; accepted 4 March 2016; published online 12 April 2016

but there was no effect of initial starting frequency on fitness (2-sample *t*-test;  $t_6 = 1.08$ ,  $P = 0.32$ ). Thus, temperate phages improved the competitiveness and invasiveness of lysogens against phage-susceptible populations in chronic lung infection. To confirm the role of phage lysis, we also measured levels of free infective phages in the lung homogenate. We observed appreciable densities of virions in the lungs in both treatments, with no significant difference in mean ( $\pm 1$  s.d.) phage-to-bacterium ratios between the competition ( $0.55 \pm 0.84$ )

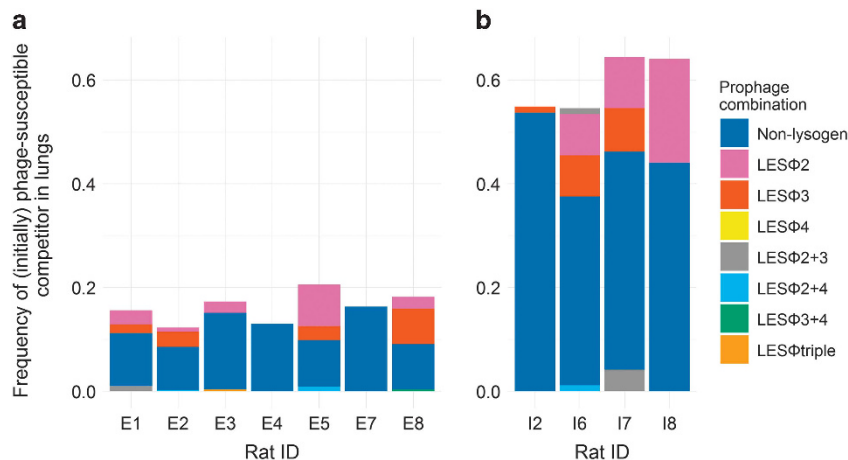


**Figure 1** Selection rate constant for the competition outcome between PAO1 $\phi^-$  and PAO1 $\phi^{\text{triple}}$  at different starting ratios of competitors. Each data point represents the outcome of competition in the lungs of an individual animal after 7 days, with the exception of the two circled data points. These represent two rats that were killed after 2 days, as they were showing symptoms of acute infection, with high bacterial loads (100-fold higher than other lungs after 7 days). These were excluded from statistical analyses.

and invasion ( $0.47 \pm 0.44$ ) treatments (2-sample *t*-test on  $\log_{10}+1$  transformed data;  $t_6 = -0.11$ ,  $P = 0.91$ ).

Phage-mediated invasion by lysogens can be limited by lysogenic conversion of the originally phage-susceptible competitor, creating phage-resistant lysogens (Gama *et al.*, 2013). Having found that lysogenic conversion occurs at a very high frequency *in vitro* (Supplementary Figure S2), we investigated whether the phages also established lysogeny *in vivo*. We calculated for each animal the total lysogen frequency and proportion of each lysogen type for end point populations by screening 46 bacterial lung isolates of the initially phage-free competitor (PAO1 $\phi^-$ ) using a multiplex PCR assay (Supplementary Methods and Supplementary Table S1). We observed appreciable rates of lysogenic conversion at both initial starting frequencies, but with substantial variation between animals (Figure 2). Although the mean ( $\pm 1$  s.d.) total frequency of lysogens was higher in head-to-head competition ( $0.89 \pm 0.03$ ) compared with invasion-from-rare ( $0.56 \pm 0.07$ ) treatment (2-sample *t*-test;  $t_6 = 8.57$ ,  $P < 0.01$ ) (due to a proportion of the PLPL $\phi^{\text{triple}}$  competitor), the rate of lysogenic conversion of PAO1 $\phi^-$  (Figure 2) was similar in both treatments (Mann–Whitney *U* test;  $W = 42.5$ ,  $n_1 = 7$ ,  $n_2 = 4$ ,  $P = 1.00$ ). Lysogenic conversion of PAO1 $\phi^-$  was dominated by the formation of LES $\phi 2$  and LES $\phi 3$  lysogens, suggesting that these phages were most active in the lung, which is consistent with the high free-phage densities of these phages in human CF infections (James *et al.*, 2015).

These data provide important experimental evidence supporting the role for phage-mediated allelopathy as a determinant of pathogen fitness in chronic lung infection. This extends previous studies using observational (James *et al.*, 2015) and insect model approaches (Burns *et al.*, 2014) to confirm, in a clinically relevant environment, that the LES-temperate phages are likely to have had a key role in the global spread of the LES. Crucially, we



**Figure 2** Lysogenic conversion of PAO1 $\phi^-$  after 7 days *in vivo*. Prophage complement of streptomycin-labelled bacteria (initially PAO1 $\phi^-$ ) isolated from rat lungs in (a) head-to-head competition and (b) invasion-from-rare treatments. Height of bars denote the frequency of the competitor out of total bacteria. Data are reported separately for each animal.

demonstrate that phage-mediated allelopathy allows lysogens to invade from rare, even in the complex, spatially structured, heterogeneous host lung environment, which has previously been theoretically predicted, but has never been demonstrated (Gama *et al.*, 2013). In agreement with a recent observational clinical study of the ecological dynamics of the LES and its phages in CF patient sputa, we show the production of appreciable populations of free-phage virions by lysis in the lung (James *et al.*, 2015). We observed lysogenic conversion in the lung, but at rates lower than those observed in liquid *in vitro* environments (Supplementary Figure S2), suggesting that lysogenic conversion may have been impeded in the lung environment. Consistent with this, recent evidence suggests that bacterial populations show strong regional structure within the CF lung, with low rates of mixing between regions of the lung (Jorth *et al.*, 2015). Nevertheless, the transfer of genetic material among strains of *P. aeruginosa* within infections does raise concerns about the potential for the horizontal gene transfer of antibiotic resistance or virulence determinants (Penadés *et al.*, 2015).

## Conflict of Interest

The authors declare no conflict of interest.

## Acknowledgements

This work was funded by a project grant from The Wellcome Trust (089215/Z/09/Z to CW & MAB). EVD was funded by a studentship co-funded by the UK Medical Research Council and the University of Liverpool, and a research visit grant from the Microbiology Society. RCL was funded by Cystic Fibrosis Canada (grant number 2610) and by the Canadian Institute for Health Research (grant reference number 86644).

## References

- Bossi L, Fuentes JA, Mora G, Figueroa-Bossi N. (2003). Prophage contribution to bacterial population dynamics. *J Bacteriol* **185**: 6467–6471.
- Brown SP, Le Chat L, De Paepe M, Taddei F. (2006). Ecology of microbial invasions: amplification allows virus carriers to invade more rapidly when rare. *Curr Biol* **16**: 2048–2052.
- Burns N, James CE, Harrison E. (2014). Polylysogeny magnifies competitiveness of a bacterial pathogen *in vivo*. *Evol Appl* **8**: 346–351.
- Fothergill JL, Mowat E, Ledson MJ, Walshaw MJ, Winstanley C. (2010). Fluctuations in phenotypes and genotypes within populations of *Pseudomonas aeruginosa* in the cystic fibrosis lung during pulmonary exacerbations. *J Med Microbiol* **59**: 472–481.
- Fothergill JL, Walshaw MJ, Winstanley C. (2012). Transmissible strains of *Pseudomonas aeruginosa* in cystic fibrosis lung infections. *Eur Respir J* **40**: 227–238.

- Gama JA, Reis AM, Domingues I, Mendes-Soares H, Matos AM, Dionisio F. (2013). Temperate bacterial viruses as double-edged swords in bacterial warfare. *PLoS One* **8**: e59043.
- James C, Fothergill J, Kalwij H, Hall A, Cottell J, Brockhurst M *et al.* (2012). Differential infection properties of three inducible prophages from an epidemic strain of *Pseudomonas aeruginosa*. *BMC Microbiol* **12**: 216.
- James CE, Davies EV, Fothergill JL, Walshaw MJ, Beale CM, Brockhurst MA *et al.* (2015). Lytic activity by temperate phages of *Pseudomonas aeruginosa* in long-term cystic fibrosis chronic lung infections. *ISME J* **9**: 1391–1398.
- Joo J, Gunny M, Cases M, Hudson P, Albert R, Harvill E. (2006). Bacteriophage-mediated competition in *Bordetella* bacteria. *Proc Biol Sci* **273**: 1843–1848.
- Jorth P, Staudinger BJ, Wu X, Hisert KB, Hayden H, Garudathri J *et al.* (2015). Regional isolation drives bacterial diversification within cystic fibrosis lungs. *Cell Host Microbe* **18**: 307–319.
- Lemieux A-A, Jeukens J, Kukavica-Ibrulj I, Fothergill JL, Boyle B, Laroche J *et al.* (2015). Genes required for free phage production are essential for *Pseudomonas aeruginosa* chronic lung infections. *J Infect Dis* **213**: 395–402.
- Lenski RE, Rose MR, Simpson SC, Tadler SC. (1991). Long-term experimental evolution in *Escherichia coli*. I. adaptation and divergence during 2,000 generations. *Am Nat* **138**: 1315–1341.
- McCallum SJ, Corkill J, Gallagher M, Ledson MJ, Hart CA, Walshaw MJ. (2001). Superinfection with a transmissible strain of *Pseudomonas aeruginosa* in adults with cystic fibrosis chronically colonised by *P. aeruginosa*. *Lancet* **358**: 558–560.
- Mowat E, Paterson S, Fothergill JL, Wright EA, Ledson MJ, Walshaw MJ *et al.* (2011). *Pseudomonas aeruginosa* population diversity and turnover in cystic fibrosis chronic infections. *Am J Respir Crit Care Med* **183**: 1674–1679.
- Penadés JR, Chen J, Quiles-Puchalt N, Carpena N, Novick RP. (2015). Bacteriophage-mediated spread of bacterial virulence genes. *Curr Opin Microbiol* **23**: 171–178.
- Stewart FM, Levin BR. (1984). The population biology of bacterial viruses: why be temperate. *Theor Popul Biol* **26**: 93–117.
- Williams D, Evans B, Haldenby S, Walshaw MJ, Brockhurst MA, Winstanley C *et al.* (2015). Divergent, coexisting, *Pseudomonas aeruginosa* lineages in chronic cystic fibrosis lung infections. *Am J Respir Crit Care Med* **191**: 775–785.
- Winstanley C, Langille MGI, Fothergill JL, Kukavica-Ibrulj I, Paradis-Bleau C, Sanschagrin F *et al.* (2009). Newly introduced genomic prophage islands are critical determinants of *in vivo* competitiveness in the liverpool epidemic strain of *Pseudomonas aeruginosa*. *Genome Res* **19**: 12–23.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

Supplementary Information accompanies this paper on The ISME Journal website (<http://www.nature.com/ismej>)