

ORIGINAL ARTICLE

Growth inhibitory response and ultrastructural modification of oral-associated candidal reference strains (ATCC) by *Piper betle* L. extract

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Candida species have been associated with the emergence of strains resistant to selected antifungal agents. Plant products have been used traditionally as alternative medicine to ease mucosal fungal infections. This study aimed to investigate the effects of *Piper betle* extract on the growth profile and the ultrastructure of commonly isolated oral candidal cells. The major component of *P. betle* was identified using liquid chromatography-mass spectrophotometry (LC-MS/MS). Seven ATCC control strains of *Candida* species were cultured in yeast peptone dextrose broth under four different growth environments: (i) in the absence of *P. betle* extract; and in the presence of *P. betle* extract at respective concentrations of (ii) 1 mg·mL⁻¹; (iii) 3 mg·mL⁻¹; and (iv) 6 mg·mL⁻¹. The growth inhibitory responses of the candidal cells were determined based on changes in the specific growth rates (μ). Scanning electron microscopy (SEM) was used to observe any ultrastructural alterations in the candida colonies. LC-MS/MS was performed to validate the presence of bioactive compounds in the extract. Following treatment, it was observed that the μ -values of the treated cells were significantly different than those of the untreated cells ($P < 0.05$), indicating the fungistatic properties of the *P. betle* extract. The candidal population was also reduced from an average of 13.44×10^6 to 1.78×10^6 viable cell counts (CFU)·mL⁻¹. SEM examination exhibited physical damage and considerable morphological alterations of the treated cells. The compound profile from LC-MS/MS indicated the presence of hydroxybenzoic acid, chavibetol and hydroxychavicol in *P. betle* extract. The effects of *P. betle* on candida cells could potentiate its antifungal activity.

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INTRODUCTION

Candida species represent a component of the normal flora in the oral cavity. However, under certain favorable conditions, these species can become opportunistic and cause infections in the oral cavity of immunocompromised hosts. This process occurs when there is a change in the ecological balance within the oral cavity that favors *Candida* over other microorganisms. *Candida albicans* has often been reported as the predominant species associated with superficial and systemic fungal infections.¹ Of late, however, the prevalence of *C. albicans* has surpassed by the emergence of non-*Candida albicans* *Candida* species,^{2–4} and increased prescription of antifungal agents⁵ has been suggested to be a contributing factor. The increased number of compromised patients with common endocrine disorders such as diabetes mellitus,⁶ with malnutrition and with smoking habits,⁷ has been identified to be primarily responsible for the development of candidal infections. The wearing of dentures has also resulted in profound alterations in the normal oral flora, providing an opportunity for candida to colonize the underlying mucosa.⁸

The normal carriage rate of *Candida* in the oral cavity varies from 2% to 71%,⁹ but can reach 100% in medically compromised patients

and those on broad-spectrum antibacterial agents.¹⁰ Seven species of *Candida* have been identified in the oral cavity, and among these, *C. albicans* has been reported as the most prevalent pathogen in both mucosal and systemic fungal infections,¹¹ while *C. glabrata* is the second or third most isolated pathogen in patients with oral candidosis.¹² *Candida* possesses a multitude of virulence factors, and a key attribute to its virulence is its adaptability for growth. Thus, an understanding of the physiological growth process of the cells could better explain and support the sustainability of cells growing under unfavorable growth conditions.

Natural products as traditional remedies are in great demand, as they are perceived to have minimal side effect on humans.¹³ Malaysia is well known for its diverse possession of flora and fauna. *Piper betle* L. is a tropical creeper plant belonging to the pepper family. Decoctions prepared from the leaves are used to relieve coughing and asthma and to help in the treatment of halitosis, joint pain and itchiness.¹⁴ It is also popular as an antiseptic that is commonly applied on wounds and lesions for its healing effects.¹⁵ The extract of *P. betle* leaves has been reported to possess anti-oxidative,¹⁶ anti-inflammatory, antibacterial and antifungal activities.^{17–19} The minimal inhibitory concentration

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and minimal fungicidal concentration of *P. betle* extract against candidal species were reported to be within the range of 12.5–25.0 mg·mL⁻¹.¹⁹ This study aimed to investigate the effects of *P. betle* extract on the growth profiles and ultrastructures of seven ATCC control strains of commonly isolated *Candida* species that have been associated with the oral cavity. The data obtained were analyzed to validate the antifungal effects of *P. betle* extract on these strains. Major components of *P. betle* were identified using liquid chromatography-mass spectrophotometry (LC-MS/MS). Findings from the study could provide better understanding of the antifungal effects of the extract on the physiological growth processes of oral candidal strains and the morphology of colonies.

MATERIALS AND METHODS

Plant collection and extract preparation

Fresh leaves of *P. betle* were collected from a local farm in Selangor, Malaysia. The specimens were scientifically identified by a botanist from the Institute of Biological Science, Faculty of Science, University of Malaya. The voucher specimen was deposited at the Herbarium of Rimba Ilmu, University of Malaya, under reference number KLU 046620. Crude aqueous extract of the specimen was prepared according to Himratul-Aznita *et al.*¹⁹ The specimen was washed and oven-dried at 60–65 °C for 2 days. The dried specimen was homogenized in distilled water at a ratio of specimen to water of 1 : 10. The homogenate was heated at a high temperature and concentrated to one-third of the original volume. The concentrate was filtered through filter paper (Whatman No. 1) before it was further heated to a final volume of 100 mL. The decoction was then concentrated overnight by freeze-drying (FDU-1200; EYELA, Tokyo, Japan). The powder obtained was kept in sterile Falcon polypropylene conical bottom tubes and was stored at 4 °C. Prior to use, a stock solution of the extract was prepared in sterile distilled water at a concentration of 200 mg·mL⁻¹. Following centrifugation (Jouan A14; Jouan, Saint Herblain, France) for 10 min at 8 000g, the stock was then diluted to the concentrations required for the experiment. The extract was sterilized by filtration using a 0.2 µm nylon syringe filter (Millipore, Billerica, MA, USA).

Microbial strains

Seven strains of oral *Candida*, which included *Candida albicans* ATCC 14053, *Candida dubliniensis* ATCC MYA-2975, *Candida glabrata* ATCC 90030, *Candida krusei* ATCC 14243, *Candida lusitanae* ATCC 64125, *Candida parapsilosis* ATCC 22019 and *Candida tropicalis* ATCC 13803, were purchased from the American Type Culture Collection (ATCC, Manassas, VA, USA) for use in the study. Yeast peptone dextrose (YPD) broth (BD Difco, Sparks, MD, USA) was used to revive the cultures.

Preparation of standard candidal suspension

Stocks of the *Candida* strains were revived in 5 mL of YPD broth and were allowed to propagate overnight at 37 °C in a horizontal incubator shaker. The cells were then harvested by centrifugation at 8 000g (10 °C) for 5 min. The supernatant was discarded, while the pellet was washed twice with sterile saline (8.5 g·L⁻¹). The pellet was resuspended in 40 mL of YPD broth, and the concentration of the suspension was adjusted to an optical density of 0.144 at 550 nm. At this optical density, the cell population was equivalent 1×10⁶ cells per mL or to #0.5 McFarland standard.

Growth profiles of *Candida* species

Five milliliters of candidal suspension (×10⁶ cells per mL) were dispensed into three sterile conical flasks, each containing 40 mL of YPD

broth. Sterile distilled water (5 mL) was added to yield a total volume of 50 mL in each flask. The flasks were incubated at 37 °C (*C. parapsilosis* at 35 °C) for 18 h in a shaking water bath to agitate the suspension continuously. Spectrophotometric assay,²⁰ which is based on continuous monitoring of changes in the optical density of cell growth, was employed. Cell growth was measured periodically at one-hour intervals over a period of 18 h at an optical absorbance of 550 nm. The growth of different candidal species could be distinguished by measuring the changes of specific growth rates (μ), using the equation previously described:^{21–22}

$$\mu = \frac{\ln(N_t/N_0)}{t_2 - t_1}$$

where N_t represented the number of cells at log phase, N_0 represented the number of cells at zero time, t_2 was the time taken to reach plateau and t_1 zero was the time when the cells entered the log phase. The μ -values were distinguished from the exponential phase, during which the cells appearing per unit time were proportional to the present population. The growth of seven candidal species was also determined based on the viable cell counts (CFUs), estimated at 2, 6, 10, 14 and 16 h intervals. The cell suspension was first diluted by serial dilution in a nontoxic diluent (e.g., phosphate-buffered saline, pH 7.2–7.4) before plating. To serve as a positive control, chlorhexidine (1.2 g·L⁻¹)-containing mouth rinse was used in place of the extract.

Growth inhibitory activity of *P. betle* extract

P. betle extract was prepared into stocks of 10, 30 and 60 mg·mL⁻¹. Five milliliters of each stock concentration was dispensed into sterile conical flasks containing 40 mL of YPD broth, followed by 5 mL of the respective candidal suspension (1×10⁶ cells per mL) to yield final concentrations of 1, 3 and 6 mg·mL⁻¹ of the extract. In a similar manner, the culture flasks were placed in a shaking water bath at 37 °C (*C. parapsilosis* was incubated at 35 °C), and the growth of cells in the presence of the extract was measured periodically at 1-h intervals over a period of 18 h. Changes in specific growth rate (μ) were calculated, and the findings were compared with those of the standard. The inhibitory effect of the extract was also determined, based on viable cell counts.

Scanning electron microscope examination

Fresh candidal suspension was cultured on YPD agar, and the colonies were allowed to grow for 24 h. Using a sterile blade, the agar was cut to approximately 1 cm×1 cm and was transferred into a sterile vial to be treated with the 6 mg·mL⁻¹ extract for 4 h. Prior to scanning electron microscope (SEM) processing, the samples were prefixed with a 4% glutaraldehyde solution overnight at 4 °C. The samples were then washed with 0.1 mol·L⁻¹ sodium cacodylate buffer (pH 7.4), followed by fixation in 2% osmium tetroxide in the buffer solution overnight. The next day, the samples were gently washed in distilled water twice for 15 min and were dehydrated in an ascending series of ethanol concentrations (10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, 90% and 95%) for 15 min each. The samples were then dehydrated twice in 100% ethanol for 15 min and subsequently were dehydrated in an ethanol–acetone mixture at ratios of 3 : 1, 1 : 1 and 1 : 3 for 15 min each. Then, the samples were treated three times with pure acetone for 15 min. The samples were processed for critical point drying (Balzers CPD 030; Bal-Tec AG, Balzers, Liechtenstein) for 2 h in liquid CO₂ under 95 bar pressure. Finally, the samples were gold-coated under low pressure with an ion sputter coater (JOEL JFC1100; JEOL, Tokyo, Japan). Any physical changes in the morphology of the candidal cells were observed with a scanning electron microscope.

Liquid chromatography-mass spectrophotometry analysis

Liquid chromatography-mass spectrophotometry (LC-MS/MS) analysis was performed with a Perkin-Elmer FX 15 UHPLC (Flexar autosampler, Flexar binary pump, Flexar column oven, Flexar degasser; PerkinElmer, Waltham, MA, USA) coupled to an AB SCIEX 3200QTrap MS/MS system, in the electrospray ionization negative mode. Ten microliters of the sample were injected onto a Phenomenex Aqua column C₁₈ (50 mm×2.0 mm×5 µm particle size). The mobile phases, consisting of solvent A (water with 0.1% formic acid and 5 mmol·L⁻¹ ammonium formate) and solvent B (acetonitrile with 0.1% formic acid and 5 mmol·L⁻¹ ammonium formate), were used in gradient mode with the following conditions (time/concentration) for B: 0.0 min/5%; 8.0 min/90%; 10.0 min/90%; 10.1 min/5%; 15.0 min/5%; and with a flow rate between 0.25 and 0.4 mL·min⁻¹. Rapid screening was performed at 15 min of run time, and the scan range of MS–MS was *m/z* 50–260. The extract was diluted five times with water and was filtered with a 0.2 µm nylon filter prior to analysis. The spectrum of the unknown component was compared with the spectra of the known components stored in the MS/MS library. The name and molecular weight of the components of the extract were ascertained.

Statistical analysis

All of the data obtained were computed and are expressed as the mean±standard deviation (s.d.) from three independent experiments performed in triplicate (*n*=9). Statistical analysis was performed using SPSS (Statistical Package for the Social Sciences) software (version 17.0; SPSS, Chicago, IL, USA). An independent *t*-test was used to compare the significant differences between controls (untreated) and *P. betle*-treated samples for each individual candidal species. One-way analysis of variance was applied to compare the specific growth rates (μ) of the seven *Candida* species upon exposure to *P. betle* extract. A *P* value <0.05 was considered statistically significant.

RESULTS

Normal growth curves of *Candida* strains

The normal growth curves of all seven candidal strains were cultured under normal, untreated growth conditions. The curves were all sigmoidal, with clear exhibition of the lag, log and stationary phases. Varying durations of the lag and log phases were observed among the different

species. In general, approximately 5–7 h was required by the cells to adapt to the normal growth environment before they were ready to proliferate and enter the log phase. *C. tropicalis* showed the highest growth rates (0.319±0.002) h⁻¹ indicating high proliferation. The others were in the range of (0.141±0.001)–(0.265±0.005) h⁻¹. Based on the enumeration of CFUs, it was shown that the population of candidal species increased gradually from 1.00×10⁵ to 1.61×10¹⁰ CFU·mL⁻¹ over 18 h of incubation (Figure 1).

Growth curves of *Candida* strains following treatment with *P. betle* extract

The patterns of the growth curves of all seven candidal strains were altered and showed deviations from the normal sigmoidal pattern following treatment with *P. betle* extract. Extension of the lag phases and suppression of cell growth were indicated by the reduction in μ -values (Table 1). The growth suppression effect of the extract was found to be concentration-dependent.

At 1 mg·mL⁻¹, the μ -values of *Candida* species were mostly reduced by a range of 15%–42%. The reduction of *C. parapsilosis*, however, was not significant (*P*=0.537). Exposing the candidal cells to 3 mg·mL⁻¹ of the extract drastically reduced the μ -values of all of the *Candida* species to almost half of the untreated cells. *C. dubliniensis* was considered the most susceptible to the extract (97.61%), followed by *C. lusitaniae* (88.68%) and *C. albicans* (88.21%). The μ -value reductions of the four others were comparatively lower, in the range of 48%–71%. The μ -values of all of the species were more than 90% reduced at 6 mg·mL⁻¹ of *P. betle* (*P*<0.05). Except for *C. krusei* (*P*=0.513), significant reductions in the specific growth rates of all of the strains were observed at 6 mg·mL⁻¹ (*P*<0.05). Deviations in the μ -values resulted in extension of the lag and log phases. Based on CFU enumeration, the populations of all of the candidal species also showed reductions of an average of (13.44×10⁶)–(1.78×10⁶) CFU·mL⁻¹ (Figure 1).

Morphology of *Candida* strains following treatment with *P. betle* extract

Treated samples of *Candida* were observed by SEM to investigate any physical changes in the appearance of the cells. Figure 2 shows the SEM images of the untreated and *P. betle*-treated candidal species. The

Table 1 Changes in the specific growth rates (μ) of the seven candidal species that was grown in the absence (untreated) and presence of *Piper betle* extract

<i>Candida</i> species	Specific growth rates (μ)	Untreated	<i>Piper betle</i> extract treated		
			1 mg·mL ⁻¹	3 mg·mL ⁻¹	6 mg·mL ⁻¹
<i>C. albicans</i> ATCC 14053	μ /h ⁻¹	0.263±0.011	0.152±0.008	0.031±0.005	0.005±0.005
	Reduction in μ %	—	42.21	88.21	98.10
<i>C. dubliniensis</i> ATCC MYA-2975	μ /h ⁻¹	0.251±0.010	0.183±0.014	0.006±0.004	0.004±0.003
	Reduction in μ %	—	27.09	97.61	98.41
<i>C. glabrata</i> ATCC 90030	μ /h ⁻¹	0.263±0.004	0.174±0.008	0.099±0.012	0.011±0.006
	Reduction in μ %	—	33.84	62.36	95.82
<i>C. krusei</i> ATCC 14243	μ /h ⁻¹	0.251±0.006	0.151±0.006	0.077±0.007	0.027±0.005
	Reduction in μ %	—	39.84	69.32	89.24
<i>C. lusitaniae</i> ATCC 64125	μ /h ⁻¹	0.265±0.005	0.180±0.009	0.030±0.004	0.012±0.003
	Reduction in μ %	—	32.08	88.68	95.47
<i>C. parapsilosis</i> ATCC 22019	μ /h ⁻¹	0.141±0.001	0.139±0.002	0.074±0.004	0.010±0.003
	Reduction in μ %	—	1.42	47.52	92.91
<i>C. tropicalis</i> ATCC 13803	μ /h ⁻¹	0.319±0.002	0.271±0.004	0.109±0.004	0.008±0.007
	Reduction in μ %	—	15.05	65.83	97.49

ATCC, American Type Culture Collection.

Values were obtained from spectrophotometric assay and expressed as mean±standard deviation of three independent experiments performed in triplicate (*n*=9).

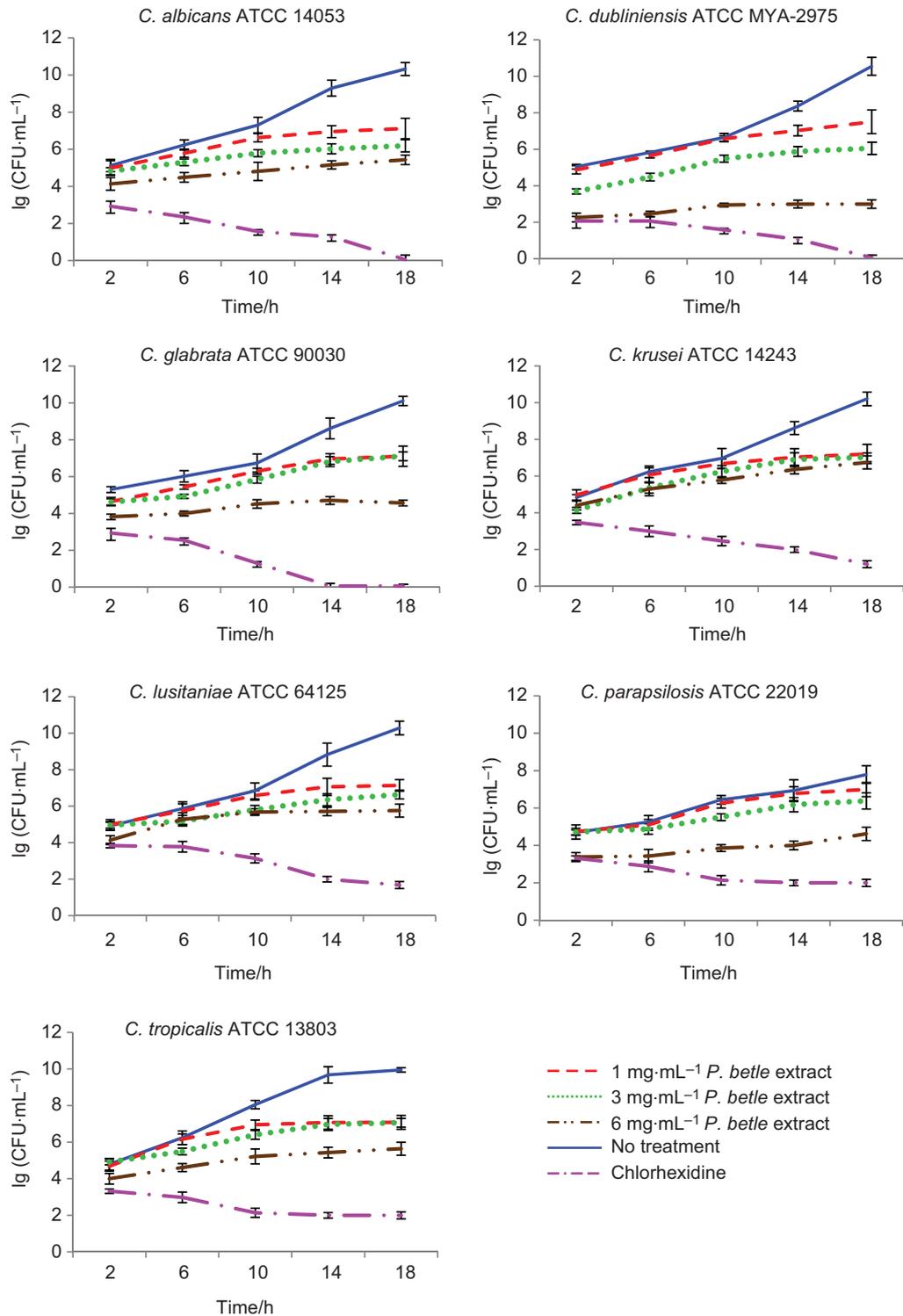


Figure 1 The population of candidal species under treatment with *P. betle* extract at 1 , 3 and 6 mg·mL⁻¹. Chlorhexidine was used as a reference. The data are expressed as the mean±standard deviation of three independent experiments performed in triplicate ($n=59$).

non-treated cells were entirely intact, and they had attained optimum cell sizes within the range of (3.13 $\mu\text{m} \times 2.33 \mu\text{m}$)–(6.65 $\mu\text{m} \times 1.95 \mu\text{m}$) (Table 2). These non-treated cells were smooth-surfaced and rounded, and some were elongated in their well-developed structures. It was observed that the cells were in the active dividing state, as the interconnecting processes and buds were present.

Some physical changes and morphological alterations in the candidal cells were observed following treatment with *P. betle* extract. It was found that the cells were slightly smaller. Based on the independent *t*-test, the mean lengths of *C. albicans*, *C. glabrata* and *C. lusitanae* were significantly different ($P<0.05$) between the untreated and *P. betle*-treated cells. The widths of all of the candidal

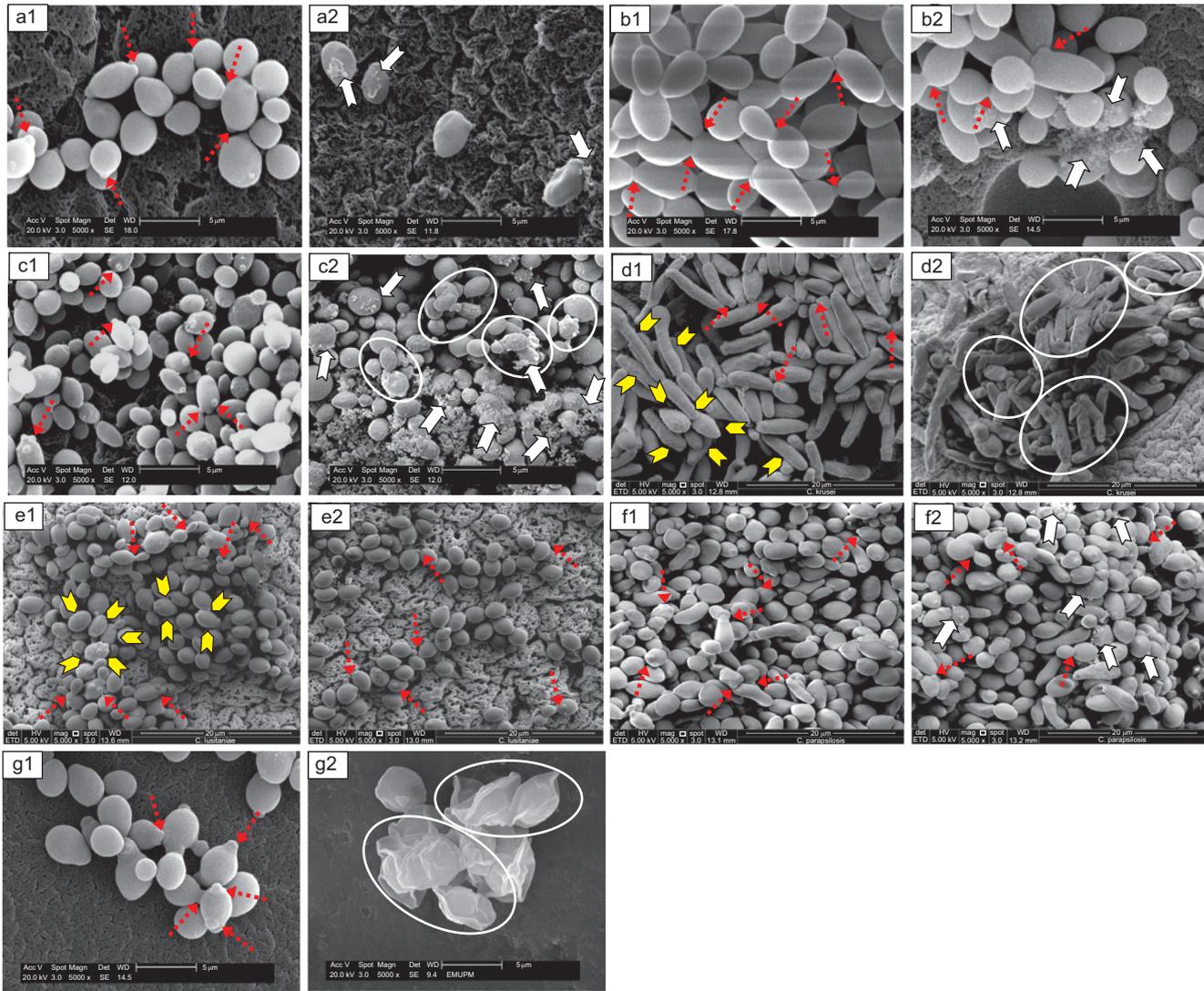


Figure 2 Composite micrographs illustrating the morphological changes of seven *Candida* species treated with *P. betle* extract, compared to untreated candidal cells. (a) *C. albicans*; (b) *C. dubliniensis*; (c) *C. glabrata*; (d) *C. krusei*; (e) *C. lusitanae*; (f) *C. parapsilosis*; (g) *C. tropicalis*. 1, Control, untreated; 2, treated with *P. betle* extract. Magnification: $\times 5000$. Σ , Dense deposits; \cdots , buds and dividing state; \bigcirc , decomposition and shrunk; \blacktriangleright , punctates.

cells, however, were not significant, except for that of *C. albicans* ($P=0.011$) (Table 2). Despite the minimal changes in the sizes of the cells, alterations of their morphology might be an indication of

the inhibitory effects of the extract, which in one way or another affected the growth profile of *Candida* cells. Some candidal cells shrank and became flaccid due to the decomposition of the cell

Table 2 Deviations in the sizes of candidal cells following treatment of *Piper betle* extract

<i>Candida</i> species	Normal		<i>P. betle</i> extract treated	
	Length/ μm	Width/ μm	Length/ μm	Width/ μm
<i>C. albicans</i> ATCC 14053	3.93 \pm 0.30	3.07 \pm 0.15	3.20 \pm 0.51 ^a	2.73 \pm 0.19 ^a
<i>C. dubliniensis</i> ATCC MYA-2975	4.11 \pm 0.48	2.40 \pm 0.15	3.57 \pm 0.47	2.43 \pm 0.13
<i>C. glabrata</i> ATCC 90030	3.13 \pm 0.39	2.33 \pm 0.39	2.43 \pm 0.23 ^a	1.87 \pm 0.45
<i>C. krusei</i> ATCC 14243	6.65 \pm 0.52	1.95 \pm 0.16	6.00 \pm 0.71	1.85 \pm 0.48
<i>C. lusitanae</i> ATCC 64125	3.80 \pm 0.44	2.56 \pm 0.25	3.16 \pm 0.24 ^a	2.40 \pm 0.21
<i>C. parapsilosis</i> ATCC 22019	4.56 \pm 0.34	2.36 \pm 0.28	4.04 \pm 0.63	2.32 \pm 0.25
<i>C. tropicalis</i> ATCC 13803	3.80 \pm 0.51	2.80 \pm 0.18	3.77 \pm 0.59	2.90 \pm 0.33

ATCC, American Type Culture Collection.

^a $P < 0.05$ comparing to the untreated (normal) candidal cell sizes.

Values are expressed as mean \pm standard deviation of nine determinations ($n=9$).

Table 3 Tentative identification of major compounds in crude extract of betel leaves (*Piper betle*) by LC-MS/MS

Peak	R _t /min	Tentative names of compounds	MW	m/z
1	3.49	Hydroxybenzoic acid (C ₇ H ₆ O ₃)	138	137.0
2	4.27	Chavibetol (C ₁₀ H ₁₂ O ₂)	164	163.0
3	5.47	Hydroxychavicol (C ₉ H ₁₀ O ₂)	150	149.0

LC-MS/MS, liquid chromatography-mass spectrophotometry; MW, molecular weight; R_t, retention time; m/z, mass-to-charge ratio.

wall. Among the treated candidal cells, deposition of heavy, mesh-like extracellular matrix was observed, which resulted in a fluffy appearance around the cells (Figure 2).

Identification of the main constituents of *Piper betle*

The chromatogram of LC-MS/MS and the associated analytical data showed that the main constituents of *P. betle* leaf extract corresponded to hydroxychavicol, chavibetol and hydroxybenzoic acid. It was clearly demonstrated that hydroxychavicol was a predominant component in crude *P. betle* extract (Table 3).

DISCUSSION

The selection of the ATCC reference strains *C. albicans*, *C. dubliniensis*, *C. glabrata*, *C. krusei*, *C. lusitaniae*, *C. parapsilosis* and *C. tropicalis* was based on various reports of the prevalence of *Candida* species in the oral cavity.^{2,10,23–24} Although the reference strains were isolated originally from blood, similar strains have also been reported as present in the oral cavity.^{25–27}

It was found that *P. betle* extract exhibited varying degrees of growth inhibitory effects on species of *Candida* without displaying cytotoxic effects on normal cell lines.²⁸ *C. dubliniensis* appeared to be the most susceptible strain to *P. betle*, compared to the others. Disruption of the normal physiological growth of candidal cells was indicated by the deviation of the growth curves from the normal pattern. Upon the addition of *P. betle* extract to the growth environment, the log phase of *Candida* species was reduced and shifted to the right. The presence of a higher concentration of *P. betle* extract affected the specific growth rates of seven candidal species. Therefore, this extract demonstrated fungistatic activity towards seven candidal species and successfully suppressed the cells, causing them to become dormant and unable to proliferate actively.

Those cells grown in the presence of *P. betle* extract could have experienced environmental stress, which might have influenced their ability to use nutrients efficiently, thereby slowing their growth. In addition, their metabolisms might have been deactivated while waiting for the environment to revert to the normal condition. This effect was previously observed for *Candida* species when growth curves based on capacitance were obtained.²⁹ It was observed that the fungistatic effect of the extract was concentration-dependent. The higher the concentration was, the more the critical turbidity was delayed, resulting in a longer lag phase. Fathilah et al.¹⁸ reported that the early settlers of dental plaque also experienced bacteriostatic effects when treated with *P. betle* extract, indicating that *P. betle* extract has a significant antimicrobial activity against a broad spectrum of oral microorganisms.

SEM images demonstrated that there were alterations in the morphology of *Candida* cells following treatment with *P. betle* extract. Dense deposits were observed when a matrix was formed on the surface of the cells, as indicated by the arrows (⇔) in Figure 2 (a2, b2, c2 and f2). Some species were able to retain their structures and their

buds while they were still in the actively dividing state, as indicated by the open arrows (•••••) in Figure 2 (a1, b1–b2, c1, d1, e1–e2, f1–f2 and g1). The destructive effects of the extract on the cells might have been minimal, but there was a possibility of certain cell wall constituents that were less firmly bound to the rigid glucan-chain network being lost during the extract treatment. As a result, the ions' and nutrients' uptake mechanisms, which normally occur on the cell surfaces, could have been restricted. Figure 2 (c2, d2 and g2) clearly demonstrates that the cells endured decomposition of the cell wall, some which shrank (circled; ○) and showed apparent loss of cell density. This finding explains the reduction in sizes of the treated cells relative to the untreated cells. In addition, the punctate appearance (arrow head; ▶), which was observed to be randomly positioned on the surface of untreated *C. krusei*³⁰ and *C. lusitaniae*, disappeared following treatment with the extract, which illustrated the direct effects of *P. betle* extract on the candidal cell walls. Similar findings were also reported by Nakamura et al.³¹ with regard to the effects on the morphology and ultrastructure of yeasts.

The extract of *P. betle* leaf has been reported to possess various chemical constituents,^{32–34} that could be isolated from the solvent extract. In the present study, hydroxychavicol, chavibetol and hydroxybenzoic acid were among the bioactive constituents found present in the aqueous extract of *P. betle*. They were similar to the types that were previously identified as present in the other solvent extracts. Hydroxychavicol, which is a phenol, was the major compound in *P. betle* extract. In the presence of hydroxychavicol, the extract might have the tendency to act as an antioxidant and a chemopreventive agent, and it might possess anticarcinogenic activities.³⁵ There have been several previous studies reporting the antibacterial and antifungal activities of hydroxychavicol.³⁶ Chavibetol, an isomer of eugenol, was regarded as one of the most active components against Gram-positive and Gram-negative bacteria.³⁷ Hydroxybenzoic acid is a phenolic derivative of benzoic acid, and it has also been reported to possess antifungal effects on the mycelia growth of *Eutypa lata*.³⁸ The antimicrobial action has been shown to be determined by more than one compound,³⁹ which are responsible not only for the antimicrobial activity but also for the synergistic effect. The mechanism of action of these components is expected to be similar to those of other terpenes and phenolic compounds, which allow for the adherence of *Candida* to host tissue surfaces before it can penetrate to target sites. The distortion of wall components and the disruption of the cytoplasmic membrane cause coagulation of the cell contents, thus leading to a loss of structural integrity and of the ability of the membrane to act as a permeability barrier.⁴⁰ These changes can be attributed to the fungistatic activity and affect the physiological functions of *Candida*. In addition, the extensive loss of the cell contents and the efflux of critical molecules and ions due to high concentrations of extract might initiate an autolytic process that results in cell death.

In conclusion, this study showed that betel leaf extract possessed potent fungistatic activity on *Candida* species. The suppression of cell growth and the alterations in morphology diminished the population of *Candida* and reduced the likelihood of its invading and colonizing the oral tissues. The presence of bioactive components in the crude aqueous extract of *P. betle* also suggests that betel leaves have the potential to be used as an antifungal agent in oral health care products in the future.

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