

SHORT COMMUNICATION

## Gene expression profile favoring phenotypic reversion: a clue for mechanism of tumor suppression by NF-IL6 3'UTR

DING GAN LIU<sup>1,\*</sup>, QIU HONG JIANG<sup>1</sup>, YUN YI WEI<sup>1</sup>, LI SUN<sup>1</sup>, BEI BEI FU<sup>1</sup>, FU KUN ZHAO<sup>2</sup>, QIONG ZHOU<sup>1</sup>  
<sup>1</sup>State Key Laboratory of Molecular Biology and <sup>2</sup>Key Laboratory of Proteomics, Institute of Biochemistry and Cell Biology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences, Shanghai 200031, China. E-mail: dgliu@sibs.ac.cn

### ABSTRACT

Transfection of cDNA in 3'untranslated region of human nuclear factor for interleukin-6 (NF-IL6 3'UTR) induced tumor suppression in a human hepatoma cell line. cDNA array analysis was used to reveal changes in gene expression profile leading to tumor suppression. The results indicate that this suppression was not due to activation of dsRNA-dependent protein kinase, nor to inactivation of oncogenes; rather, all the changes in expression of known genes, induced by NF-IL6 3'UTR cDNA may be ascribed to the suppression of cellular malignancy. Therefore, our results imply that this 3'untranslated region may have played role of a regulator of gene expression profile.

**Key words:** cDNA array, NF-IL6, hepatoma, 3'UTR, tumor suppression.

### INTRODUCTION

Studies on the suppression of malignancy of tumor cells are important for the understanding of the mechanism of tumor suppressors and for the establishment of novel therapies of tumors. Since the last decade of the 20<sup>th</sup> century, several 3'untranslated regions of eukaryotic mRNAs were found to have *in vitro* tumor suppressor activity. They caused phenotypic reversion (lowering of tumorigenicity) in cultured tumor cells or malignant cells[1- 6]. However, the molecular mechanisms of these tumor suppressor effects remain elusive.

Several years ago, a regulatory element with tumor suppressor activity, NF-IL6 3'UTR, was found, which caused tumor suppression in a part of mouse malignant DT cells[1, 2]. Recently, in order to test its tumor-suppressive effect in different human tumor cell lines, we transfected it into SMMC7721 cells, a human hepatocarcinoma cell line, and resulted in markedly reduced malignancy (reversion) in significant part of the transfected cell clones. Then, we determined the changes in gene expression profile of a typical revertant clone by using the cDNA array analysis, to pave the way for

elucidating possible molecular mechanisms of this tumor suppression. In this report, we will present results of this analysis.

### MATERIALS AND METHODS

#### Cell lines and plasmids

SMMC7721, purchased from Cell Bank of Chinese Academy of Sciences, Shanghai, is a typical hepatoma cell line[7]. p14-6 is a recombinant pcD2 plasmid harboring a middle part of the 3'untranslated region of the human NF-IL6 cDNA[4], stored in our lab (GenBank accession number M98056). pcDmr Cl 2 is a plasmid containing the same genes as p14-6, but harbors no insert.

#### Transfection of cell lines and malignancy tests

Stable transfection was performed by using the Calcium phosphate transfection kit (Invitrogen) according to the instruction manual. The concentration of cytotoxic antibiotic G418 (Sigma) was 2 mg / ml (valence 1 mg/ml) for all cells, including controls. G418<sup>R</sup> clones appeared in 1-2 week after screening. Cell growth curves determination and soft agar tests were performed according to our previous methods[1]. Nude mice tumorigenicity tests were performed with the Balb/c nu/nu nude mice, 4-6 week age (from the Shanghai Center for Experimental Animals, Chinese Academy of Sciences), which were kept in the sterile rooms of Animal house of our Institute. 5× 10<sup>6</sup> cells per point were injected subcutaneously into the back of animals. Positive tumorigenicity was defined as the appearance of palpable tumor nodules in the injection points one month postinjection.

#### cDNA array analysis

The cDNA array analysis were done in CASarray Co., Shanghai.

\*Corresponding author: Dr. Ding Gan LIU  
Tel: 021-54921135 E-mail: dgliu@sibs.ac.cn

**Abbreviations:** NF-IL6, nuclear factor for interleukin-6 expression. PKR, double-stranded RNA-dependent protein kinase. 3'UTR, 3'untranslated region.

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The cells were directly lysed in culture bottles with Trizol<sup>R</sup> (Invitrogen), then subjected to further mRNA purification, cDNA synthesis and hybridizations. The cDNA array membranes used contained 11,557 known genes and ESTs, and every gene had two spots. The analysis was performed twice to guarantee reproducible results, and passed strict quality control.

### Semi-quantitative RT-PCR

6 genes, including 3 up-regulated and 3 down-regulated as revealed by cDNA array analysis, were chosen to compare their cDNA amounts by using semi-quantitative RT-PCR. The primers were designed using OLIGO version 6.44 software. For control, cDNA of ribosomal protein gene S29, a well-known housekeeping gene, was used. Reverse transcription of total RNAs from CI 1, SMMC7721 and pcDmr was done, respectively, using Superscript II reverse transcriptase (Life Technologies and equal amounts (1  $\mu$ l) of RT mixture were used in a 50  $\mu$ l PCR reaction containing various primer pairs and FD Taq DNA polymerase (Fuhua Co., Shanghai) under the same conditions. The intensities of bands were measured using Tanon gel electrophoresis analysis software, and the values of their strengths relative to S29 were calculated by dividing their measured band intensities by the average S29 band intensities of RT PCR products of corresponding cell lines. The results were shown in a vertical square graph.

## RESULTS AND DISCUSSION

### Establishment of the NF-IL6 transfectants of SMMC7721 cell line

The NF-IL6 3'UTR cDNA (plasmid p14-6) was stably transfected into SMMC7721 cells by using standard calcium phosphate procedure. It was found that this hepatoma cell line has a certain endogenous G418 resistance, and thus a higher concentration (2 mg/ml) of G418 was used for screening. This drug concentration led to the reduction in the number of G418<sup>R</sup> clones; however, all the transfectants contained plasmids, as proved by Southern blots.

### Phenotypic characteristics of the NF-IL6 3'UTR transfectants of SMMC7721

By high concentration G418 screening, 14 independent clones were obtained, 6 of which were found to be contaminated by mycoplasma and excluded. The remaining 8 clones were used in experiments.

These p14-6 transfectants had morphology similar to their parent cells. The majority of clones grew slower than SMMC7721 (see Fig 1B-1) and had reduced ability to form colonies in soft agar medium (Tab 1 and Fig1A; in the figure, only CI 1 and two controls were shown). In the *in vivo* tumorigenicity tests, those transfectant clones that formed less colonies in soft agar also appeared

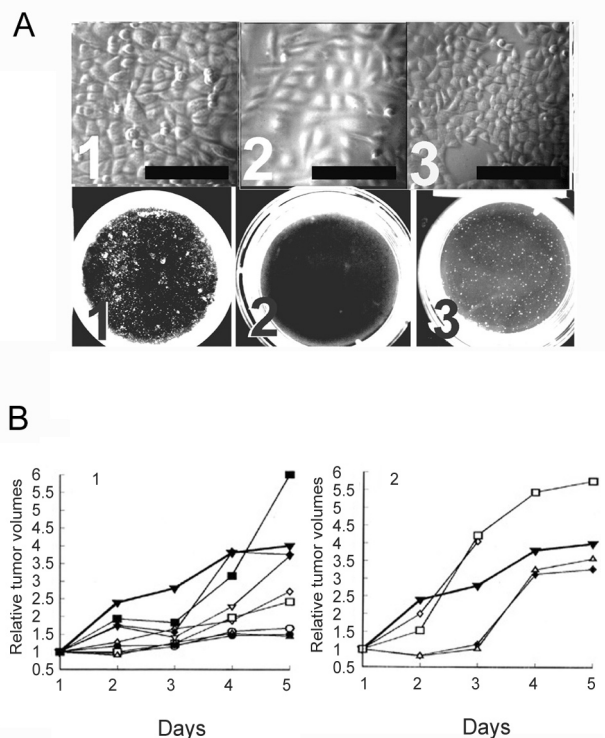


Fig.1. Growth characteristics of cell clones. A, morphology (upper) and soft agar colonies (lower) of SMMC7721 (1), revertant CI 1 (2) and control pcDmr CI 1 (3). Bars: 10  $\mu$ m. B, growth curves of all cell clones. 1, growth curves of 8 NF-IL6 3'UTR transfectants. The signs in the curves are:  $\Delta$ , CI 1;  $\square$ , CI 2;  $\bullet$ , CI 3;  $\square$ , CI 4;  $\square$ , CI 5;  $\nabla$ , CI 6;  $\blacklozenge$ , CI 7;  $\square$ , CI 8. 2, growth curves of control pcDmr clones. The signs in the curves are:  $\square$ , pcDmr CI 1;  $\square$ , pcDmr CI 2;  $\Delta$ , pcDmr CI 3;  $\blacktriangleright$ , pcDmr CI 4. In both figures the thicker curves with  $\blacktriangledown$  signs are growth curves of SMMC7721.

Tab 1. Soft agar colony test of cell clones

Name	Colony formation ability*	Name	Colony formation ability
CI 1	+-	CI 8	+++
CI 2	+	SMMC7721	++++
CI 3	+	pcDmr CI 1	+++
CI 4	++	pcDmr CI 2	+++
CI 5	+-	pcDmr CI 3	+++
CI 6	+	pcDmr CI 4	+++
CI 7	++		

\*+-: 0 - 500 very small-sized colonies per dish. +: 500 - 1000 small-sized colonies per dish; ++: 1000-2000 small- and a few middle-sized (diameter  $\leq$ 0.5 mm) colonies per dish; +++: > 2000, with >10 large (diameter  $\geq$  1mm) colonies, per dish; ++++: > 2000, with > 10 very large (diameter > 2 mm) colonies, per dish.



**Tab 3.** Genes whose regulation is induced by NF-IL6 3'UTR, found by cDNA array analysis

GenBank Accession Number	Name	Expression ratio CI 1/SMMC7721(average)	Known functions
<i>1. Up-regulated genes</i>			
AB036063	p53-induced ribonucleotide reductase small subunit 2 homolog	2.05	Tumor suppressor related to p53[9]
S59049 (NM_002922)	regulator of G protein signalling 1	2.51	Negative regulator of G protein signalling[16]
AJ133355	zinc finger protein 237	2.27	Putative regulator (DNA-binding)
U12767	nuclear receptor subfamily 4 group A member 3	2.84	Putative regulator (DNA-binding)[14]
AL079310	high-mobility-group protein 2-like 1	2.40	Putative DNA binding protein
U87460	G-protein-coupled receptor 37 (endothelin receptor type B-like)	3.25	Function unknown
NM_020233	x 006 protein	2.55	Function unknown
BE301841	hypothetical protein (DKFZp434N1923)	11.5	Function unknown
<i>2 Down-regulated genes</i>			
AF012281	PDZ domain containing protein 1	0.45	Overexpressed in some epithelial tumors[12]
M26326	keratin 18	0.42	binds to keratin 8 to provide resistance to Fas-mediated apoptosis[13]
K00558	tubulin $\alpha$ , ubiquitous	0.40	Microtubule component
D31885	ADP-ribosylation factor 6-like Interacting protein	0.47	Activator of signal transduction pathways
J03592	solute carrier family 25 (mitochondrial carrier; adenine nucleotide translocator), member 6	0.28	ADP/ATP translocase[14]
BE222923	highly similar to spectrin $\beta$ chain	0.34	Essential for TGF- $\beta$ signalling[15]
AF000989	thymosin $\beta$ 4, Y chromosome	0.46	Related to cell differentiation; increasing expression enhances metastasis[16]
X06256	integrin $\alpha$ 5 (fibronectin receptor)	0.29	Cell growth regulator[17]
AL035297	human gene from PAC 747 L4	0.27	Function unknown
AK001187	hypothetical protein FLJ14642	0.37	Function unknown
AA885521	KIAA1409 protein	0.40	Function unknown

duced ribonucleotide reductase small subunit 2 homolog, and regulator of G protein signalling 1) were directly or indirectly related to tumor suppression, and their enhanced expression favored this suppression. Among those genes that were down-regulated, the first gene (PDZ domain-containing protein 1) is reported to be overexpressed in some epithelial tumors[12]; therefore, its reduced expression may contribute to the tumor suppression. The functions of other down-regulated genes include resistance to apoptosis (keratin 18), activation of signal transduction (ADP-ribosylation factor 6-like interacting protein), and activation of cell motility (thymosin  $\beta$ 4) etc; clearly, their down-regulated expression are unfavorable

for tumor growth and metastasis. Therefore, the changes in the expression of above genes, induced by the transfected NF-IL6 3'UTR, all favored to the reversion of cellular phenotype. No oncogene, that was detectably up- or down-regulated, was found by the cDNA array analysis.

We have also performed analysis on expressed proteins of these cell lines by using 2D electrophoresis (will be published elsewhere). The majority of results of 2D electrophoresis were different with cDNA array results. For example, maspin[18] and annexin III[19] were found up-regulated in 2D analysis. Besides technical reasons, this difference may suggest that the expression of a gene from transcription to final protein product is more com-

plicated than that anticipated by our present knowledge.

### *Possible mechanism(s) of NF-IL6 3'UTR tumor suppression*

It might be thought that some oncogenes were inactivated by the insertion of transfected plasmids, leading to tumor suppression. However, this does not seem to be the case, because no oncogene was found differentially and significantly expressed.

The activation of dsRNA-dependent protein kinase (PKR) by 3'UTR RNA[22] was used to explain the tumor suppressor effect of  $\alpha$ -tropomyosin; however, this explanation is not satisfactory enough, because, first, some 3'UTRs of mRNA, like the 3'UTR of NF-IL6, do not have stable double stranded structure; second, as the activation of PKR leads to general suppression of protein synthesis, the up-regulation of protein expression (like the maspin and annexin III in our case) is hard to be explained.

To sum up, we are inclined to believe that the transfection of NF-IL6 3'UTR cDNA might possibly play some regulatory role for the expression of a group of genes, either in up- or down-regulated ways. It is well known that the 3'UTRs are sites for interactions with trans-acting factors, and these interactions, as a whole, may be responsible for the maintenance of normal cell life. Therefore, the exogenous 3'UTR might also act as a regulator of gene expression. If such supposition is correct, its molecular mechanism of tumor suppression should be found in the analysis of its regulatory functions.

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