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REVIEW

Clinical development directions in oncolytic viral therapy

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Oncolytic virotherapy is an emerging experimental treatment platform for cancer therapy. Oncolytic viruses are replicativecompetent viruses that are engineered to replicate selectively in cancer cells with specified oncogenic phenotypes. Multiple DNA and RNA viruses have been clinically tested in a variety of tumors. This review will provide a brief description of these novel anticancer biologics and will summarize the results of clinical investigation. To date oncolytic virotherapy has shown to be safe, and has generated clinical responses in tumors that are resistant to chemotherapy or radiotherapy. The major challenge for researchers is to maximize the efficacy of these viral therapeutics, and to establish stable systemic delivery mechanisms. *Cancer Gene Therapy* (2011) **18**, 305–317; doi:10.1038/cgt.2011.7; published online 25 March 2011

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Introduction

Oncolytic viruses involved in initial cancer therapeutics are non-pathogenic, naturally occurring viruses that are either wild-type or naturally occurring mutants. Specificity to cancer is determined by tumor-specific genetic mutations that result in aberrant protein expression. Adenoviruses are the most widely studied engineered oncolytic viruses clinically. Adenoviral constructs include Onyx 015,^{1,2} CG7060,³ CG7870,⁴ dl922-947,⁵ Ad5-CD/tk-rep,⁶ Ad-delta24,⁷ Ad DF3-E1,⁸ Onyx 411,⁹ OAV001,¹⁰ KD3,¹¹ 01/PEME¹² and Telomelysin.^{13,14}

Historically, evidence of viral oncolytic activity was published in case reports as early as 1912. These reports described rare but dramatic responses in cancer patients recovering from viral syndromes.^{15–29} On the basis of these observations, viruses with low pathogenicity to normal tissue and high oncolytic capacity have been selected for clinical investigation.^{29–35}

This review will focus on the anticancer activity of oncolytic viruses demonstrated in clinical investigation.

Onyx 015

Onyx 015 is a replication-conditional adenovirus genetically modified by deletion of two DNA elements. It was theorized that deletion of the first element, the E1B 55 kDa fragment, would facilitate replication of ONYX 015 in cells with a defective p53 pathway, which commonly occurs in cancer cells, although it has become clear that this virus is not specific for p53-null cells.^{1,36,37} Clinical trials of several hundred patients have shown no evidence of nonspecific viral replication or damage to normal cells at the border of intratumoral injection sites.^{38–51} When administered intravenously (i.v.), dose escalation was limited by transient liver enzyme elevation at a dose of 2×10^{13} particles.^{43,46,52}

Initial phase one investigation of ONYX 015 involved intra-tumor injection in refractory cancer patients. The virus was well tolerated and evidence of activity was suggested.⁵³ A phase II study involving 40 squamous cell carcinoma of the head and neck patients injected intratumorally with 2×10^{11} viral particles for 5 consecutive days revealed cancer-specific viral replication in 7 of 11 patients who underwent biopsy.⁴⁰ No viral replication or toxic effects were identified in normal tissue. No particular toxic effects were observed following 533 viral injections, and nearly 20% of patients demonstrated significant, partial, or complete response (CR) of the injected lesion. In a subsequent phase II study, patients received ONYX 015 (2×10^{11}) particles for 5 consecutive days/21 day cycle) in combination with cisplatin (80 mg m^{-2} once every 21 days) and 5 fluorour-acil ($800-1000 \text{ mg m}^{-2}$ continuos infusion 5 days/21 days).⁴¹ No added toxicity attributable to ONYX 015 was demonstrated in addition to the expected toxicity of cisplatin and 5 fluorouracil. Furthermore, a 63% response rate was observed, which was greater than the expected response rate of 35% from previous publications in the same patient population using similar chemotherapy regimes.41

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Other clinical investigations of ONYX 015 have involved advanced ovarian cancer, ⁴² hepatocellular carcinoma, ⁴⁴ pancreatic cancer^{47,49} and colorectal cancer. ^{48,54} In a meta-analysis⁵⁴ summarizing two intrahepatic arterial infusion trials involving colorectal cancer, survival was compared between patients receiving $< 6 \times 10^{11}$ virus particles per infusion (7 patients) to those receiving $> 6 \times 10^{11}$ virus particles per infusion (28 patients). A significant survival advantage was demonstrated in the high-dose group (359 days) compared with the low-dose group (155 days).⁴⁵

ONYX 015 has been administered safely to >40 patients intravenously. Results show a slightly higher frequency of febrile response with systemic administration than intratumoral administration of ONYX 015, although the frequency was similar to what was observed with intra-arterial infusion. At doses of $<1 \times 10^{13}$ particles per infusion, no significant safety concerns were identified, including situations in which virus was administered in combination with low-dose IL-2 or chemotherapy (paclitaxel, CPT11 and 5 fluorouracil). The presence of ONYX 015 within metastatic malignant disease sites following i.v. infusion was demonstrated; however, evidence of significant tumor regression was not identified.^{43,46,53}

Shanghai Sunway Biotech presently owns the rights to ONYX 015. They also have the rights to a nearly identical virus called H101 (trade name Oncorine), the first oncolvtic virus to be commercialized, which is currently on the market in China following demonstration of improved response and time to disease progression of nasopharyngeal carcinoma in combination with cisplatinbased chemotherapy compared with chemotherapy alone.⁵⁵ Two follow-up products have been introduced, H102, currently in pre-clinical testing, and H103, an oncolytic type 2 adenovirus overexpressing the heat shock protein HSP₇₀, recently tested as a intratumoral vaccination in a completed phase I clinical trial in patients with advanced solid tumors.⁵⁶ Transient and partial regression of distant, un-injected tumors was observed in three patients during this study, and because of promising clinical antitumor activity and positive safety outcome further studies are being pursued.

Telomelysin

Telomelysin is a novel, replication-competent Ad5-based adenoviral construct that incorporates a *human telomerase reverse transcriptase* gene (*hTERT*) promoter. *hTERT* encodes for the catalytic protein subunit of telomerase, a polymerase that acts to stabilize telomere lengths and is highly expressed in tumors but not in normal, differentiated adult cells.⁵⁷ Earlier studies have shown that hTERT promoter can control the expression of exogenous genes in telomerase-positive cancer cells, and can serve as an excellent candidate for cancer-specific control of oncolytic adenoviral replication.⁵⁸

Additional modifications of Telomelysin include the replacement of the normal transcriptional element of viral

E1B gene by an IRES (Internal Ribosomal Entry Site) sequence. Furthermore, Telomelysin is the first replication-competent adenovirus that retains a fully functional viral E3 region. E3 proteins prevent Ad-infected cells from being cleared by cytotoxoic T lymphocyte, tumor necrosis factor, Fas ligand and tumor necrosis factor-related apoptosis-inducing ligand and act to decrease systemic viral clearance.⁵⁹ Other adenoviral therapeutics have dysfunctional or deleted E3 regions for safety considerations. However, removal of the E3 region and rapid clearance of viral therapeutics may also cripple the antitumor effect. Thus, retention of a functional E3 region has the theoretical advantage of optimizing antitumor activity within the constraints of clinical safety by enhanced viral pharmacokinetics and biodistribution.

In vitro studies have validated the selective infectivity and direct cytolysis of Telomelysin in cancer cells but not nonmalignant cells.⁵⁸ In animal experiments, intratumoral injection of Telomelysin demonstrated antitumor activity without significant toxicity to normal organs. Further, distant viral uptake was observed following intratumoral injection when the presence of adenoviral protein was identified in the noninjected tumor following intratumoral treatment of the contralateral tumor.⁵⁸

A phase I study evaluating the tolerability of a single intratumoral injection of Telomelysin was recently completed.⁶⁰ Sixteen patients were entered into three dose escalation cohorts without defining a maximum tolerated dose. There were no clinically significant grade 3 or 4 treatment-related adverse events (AEs). However, multiple grade 1 and 2 AEs were reported, with the most common being fever, chills, fatigue and injection site pain. Nine evaluable patients (one neuroendocrine, three squamous cell carcinomas, three melanomas, one leiomyosarcoma and one salivary cancer) satisfied RECIST criteria for stable disease (SD) at day 28, and seven of these patients had SD at day 56. Six patients had progressive disease at day 28 assessment. Post injection biopsies performed at day 28 on four of the patients with SD revealed intratumoral necrosis. Three of these patients had melanoma.

Viral pharmacokinetic analysis demonstrated the transient presence of systemic Telomelysin dissemination following intratumoral injection early after injection. Evidence of viral replication was demonstrated with detection of late (\geq day 7) viral DNA in three patients; one with elevated malignant tissue hTERT expression demonstrated significant clinical response. Immunohistochemical analysis of viral E1A and hexon was negative 28 days after injection suggesting rapid viral clearance. It is of interest that two patients demonstrated response distant from the injected lesion, which was consistent with animal experience and not previously demonstrated in other oncolytic adenoviral studies.

As a result of the unique modifications built into this adenoviral construct both activity and safety of a single injection approach has been demonstrated. However, despite significant activity in a subset of patients limited clinical relevant responses were observed. As only single-dose intratumoral injection was attempted in this trial it remains unclear what the therapeutic potential is for this agent.

Newcastle virus

Newcastle virus is a paramyxovirus with infectivity normally restricted to fowl. It is an enveloped negativestranded RNA virus, which selectively replicates in human cancer cells that have developed defects in the interferon signaling pathway.⁶¹ Most early studies used Newcastle virus as an oncolysate tumor vaccine. These vaccines were injected into patients to generate an immune response.⁶² In addition, the virus has been given intravenously, intraperitoneally and intratumorally in athymic mice implanted with human cancers, including lung cancer. These preclinical trials have shown few systemic side effects, and have demonstrated evidence of oncolytic activity.^{63–66}

The first report of antitumor activity of the Newcastle virus involved one patient with cervical cancer. Cassel and Garrett injected virus $(2.4 \times 10^{12}$ virus particles) directly into the tumor and demonstrated intratumoral regression of the cancer both at the injection site and at a distant malignant lymph node.⁶⁷ In the mid-1970s, viral-induced oncolysates were studied as vaccines in melanoma, breast, ovarian and colon cancer. Safety and modest evidence of activity were observed.^{68–75}

I.v. infusion of Newcastle virus has been well tolerated. In one placebo-controlled phase II study, 33 patients with advanced cancer received virus and 26 control patients were given placebo treatment. Of the patients treated with virus, seven patients achieved a complete or partial response and one patient had a minor response: these eight patients survived >1 year after treatment. In comparison, none of the control patients had responses. In all, 22 patients receiving virus survived longer than 1 year, whereas only 4 patients in the control group survived 1 year. Eight viral-treated patients survived for >2 years versus none of the control patients.⁷⁶

In another trial of an attenuated Newcastle virus strain, PV701 virus was administered intravenously $(5.9 \times 10^9 \text{ p.f.u. m}^{-2} \text{ to } 24 \times 10^9 \text{ p.f.u. m}^{-2} \text{ every } 28 \text{ days})$ to 79 patients with solid tumors. Side effects were mild and were limited to fever, flu-like symptoms and hypotension. Seven grade 3 AEs were observed, but toxicity decreased with subsequent doses. A maximum tolerated dose following a single infusion was established at 12×10^9 p.f.u. m⁻², and subsequent infusions were tolerated up to 120×10^9 p.f.u. m⁻². Further dose escalation was limited by hypotension. In all, 14 of 62 patients eligible for response assessment maintained SD from 4 months to > 30 months. One patient with squamous cell cancer of the tonsil achieved a CR. Another patient with metastatic colon cancer achieved a partial response. Seven patients achieved minor responses of <50% reduction in tumor size. The presence of viral particles in malignant tissue was confirmed following treatment.

Herpes simplex virus (HSV)

HSV is a double-stranded DNA virus. Genetic modification enabled the construction of oncolytic virus selectively

activity within malignant tissue. One modification involved inactivation of viral gene ICP6, which encodes the large subunit of ribonucleotide reductase, an enzyme required for viral DNA replication.⁷⁷⁻⁸⁰ This enzyme is expressed abundantly in rapidly dividing tumor cells but is sparse in normal cells. As a consequence, the *ICP6* gene modified HSV-1 replicates selectively in tumor cells. The second gene modification approach consists of deleting another viral gene, the γ -34.5 gene, which functions as the virulence factor during HSV infection.⁸¹ Mutations in this gene also limit replication in non-dividing cells.^{82,83} The oncolytic HSV-1 virus, G207, has been extensively tested in animal models and is currently in clinical trials.⁸⁴⁻⁸⁷ Replication-sensitive HSV1 γ-34.5 viral mutants have been shown to be effective in the treatment of both central nervous system⁸⁸⁻⁹⁰ and non-central nervous system⁹¹⁻⁹⁹ tumors in animal models. Clinical trials involving patients with high-grade glioma, colorectal cancer, non-small cell lung carcinoma and melanoma^{100–103} have demonstrated safety. Four different herpes simplex oncolvtic viruses have been tested in clinical trial. Toxicity includes fever, chills and transient liver enzyme elevation and is greater in patients who have low HSV-1 antibody titers at baseline. However, all patients developed an immune antibody response against HSV antigens within weeks following treatment; thus, significantly less toxicity occurs with continued treatment to patients with high initial HSV antibody titers. PCR analysis of tissue demonstrated the presence of HSV DNA at injection sights.¹⁰⁴ Preclinical results in immune competent models have also suggested immune-mediated distant responses.^{93,94,105–107} Given significant lack of systemic activity of viral-induced oncolysis following local-regional treatment in clinical study, several new vectors carrying immune-stimulating transgenes have been developed (granulocyte-macrophage colony-stimulating factor (GM-CSF), interleukin-2, interleukin-12, B7.1).¹⁰⁷⁻¹¹² Additionally, combination of HSV mutants with chemotherapy or radiotherapy has demonstrated enhanced antitumor activity.^{80,101,113–118} Radiation increased the anticancer activity of HSV when used in pancreatic, glioblastoma and cervical cancer models¹¹⁹ but did not alter the antitumor effect of HSV in prostate cancer. However, high-dose radiation combined with oncolytic HSV virus did improve efficacy in other prostate cancer models.^{102,115,116} Low-dose irradiation also improved efficacy of HSV viral therapy in a cervical cancer model.¹¹⁸ In chemotherapy combination studies, interestingly, both chemotherapy-resistant and -sensitive tumors were equally responsive.¹²⁰ A variety of chemotherapy agents (mitomycin C, cisplatin, methotrexate, taxanes) have demonstrated enhanced antitumor effect when combined with HSV.101

Other studies have evaluated the use of HSV to deliver other genes, such as those that convert benign pro-drugs into cytotoxic agents. In one study, the cytochrome p450 gene and HSV-1 thymidine kinase (TK) gene were delivered using a HSV-1 replication-competent virus via intratumoral injection in a hepatocellular carcinoma model.¹²¹ Cancer regression significantly improved with **Oncolytic viral therapy** RM Eager and J Nemunaitis

the cytochrome p450 conversion of cyclophosphamide to the active metabolite phosphoramide mustard.⁷⁸ Similar results have been produced with the cytosine deaminase transgene.¹²² Interestingly, HSV TK activation of acyclovir or ganciclovir in HSV-infected cells inhibits viral replication without affecting tumor growth.^{95,123–125} This suggests that the *TK* gene and ganciclovir may be useful as a safety valve if persistent virus and related toxicity developed.¹²¹

OncoVEX^{GM-CSF} (BioVax, Worcester, MA) is a replication-competent HSV (HSV-1) that has been modified with several novel genetic enhancements that make it a potent oncolytic and immunogenic vector.^{126–130} The vector contains the coding sequence for human GM-CSF under the control of the human cytomegalovirus immediate early promoter, which has been shown to enhance the immune response.¹³¹ As a safety factor, the gene for TK remains intact preserving sensitivity to clinically effective antiviral agents. This genetic arsenal aims to promote an *in situ* tumor-specific vaccine that is potentiated by viral replication.

A phase I study demonstrated OncoVEX^{GM-CSF} to be well tolerated with local inflammation, erythema and fever being the main AEs. Biological activity was evident by viral replication, local reactions, GM-CSF expression and HSV antigen-associated tumor necrosis was observed.¹³² Tumor flattening, shrinkage and necrosis were noted in tumor types including melanoma, breast, and head and neck in both injected and on-injected tumors. A phase II study of OncoVEX^{GM-CSF} in metastatic mela-

A phase II study of OncoVEX^{GM-CSF} in metastatic melanoma demonstrated a 26% objective response rate including un-injected regional and distant metastatic sites. One-year overall survival rates of 61, 58 and 48% for all patients, all stage IV patients and stage IV M1c patients, respectively.¹³³ A recently published meta-analysis of 2100 patients with stage IV metastatic melanoma reported a 1-year overall survival rate was 25.5%. The median survival time is 16+ months for all patients treated with OncoVEX^{GM-CSF} as well as for the stage IV subset; the median survival time in the meta-analysis was 6.2 months (95% confidence interval, 5.9 months to 6.5 months). Although these data are not directly comparable the results with OncoVEX^{GM-CSF}, they are provocative.

There were 13 objective systemic responses (8 CR, 5 partial response; 26% overall: 6 CR, 3 partial response; 22.5% stage IV), 10 of which continue for >6 months. Response onset was from 2 to 10 months following the first dose. Although local responses often occurred rapidly (after as few as two injections), maximum objective response has been observed as long as 12 months post first dose when biopsy confirmed one patient as disease free. In six patients, distant responses at un-injected sites were documented in the lung, liver, pancreas, regional and distant lymph nodes, and at other soft tissue sites. In most cases, considerably <50% of the overall disease burden was injected. Two patients achieved surgical CRs, one following excision of a newly identified brain metastasis, and one following additional treatment with interleukin-2.

Eighty-five percent of patients had AEs related to OncoVEX^{GM-CSF} all of which were grade 1–2. The most

common AEs were consistent with a mild influenza-like syndrome (fever (54%), chills (40%), nausea (42.5%), fatigue (32%), vomiting (20%) and headache (24%)). There were 21 serious AEs all of which were considered unrelated to OncoVEX^{GM-CSF}. Autoimmune vitiligo was noted in three patients; two of whom achieved CR and one in whom lesions were responding before leaving the study because of non-compliance issues. Considering the relatively benign safety profile in both the phase I and II studies, further evaluation is underway. There is currently a multi-national phase III study in metastatic melanoma in progress and also a phase III clinical study in squamous cell carcinoma of the head and neck is scheduled for the second half of 2010.

Reovirus

Research into the therapeutic potential of reovirus holds particular interest as this double-stranded RNA-containing virus is able to replicate and produce lysis in specifically transformed cells possessing an activated Ras pathway while sparing normal cells.¹³⁴ Although reovirus belongs to the Reoviridae family, which includes rotovirus, infection in humans is usually subclinical and limited to the upper respiratory and gastrointestinal tract.¹³⁵ Three viral serotypes have been isolated and all are commonly found in the environment as this virus possesses a highly stable unenveloped icosahedral capsid, thus it is estimated that nearly half of the population has been exposed and carries antibodies to the virus.¹³⁶⁻¹³⁸ Importantly, reovirus type 3 Dearing strain exhibits replication in cells with an activated Ras signaling pathway, a significant finding because of the link between oncogenesis and mutations in the ras gene and pathway, and it has been demonstrated that this is due to inhibition of double-stranded RNA-activated protein kinase resulting in cell lysis.^{139,140} Although normal mouse fibroblast cells (NIH3T3) do not normally support reovirus replication. NIH3T3 cells transformed with activated ras. epidermal growth factor receptor or V-erb B oncogene (for example, activated ras pathway elements) are lysed by uninhibited reovirus replication. It is now understood that an activated ras pathway, which is present in many ovarian, breast, colon and lung cancers, prevents viralinduced PKR activation and subsequent EIF-2 α -phosphorylation, potentiating cellular protein production and viral replication. In normal cells without ras activation, early viral replication induces EIF-2 α -phosphorylation, which inhibits cell protein synthesis. Thus, reoviruses exhibit preferential oncolytic effects in ras-activated cancer cells.

Reoviruses have demonstrated pre-clinical activity in mouse flank tumor models with cell lines that overexpress certain ras pathway elements. Examples include V-erb-transformed NIH3T3 cells, human V87 glioblastoma cells overexpressing platelet-derived growth factor receptor and ras-transformed C3H-10T1/2 cells.¹⁴⁰ Reoviruses have also demonstrated activity against Lewis lung cancer

metastasis in mice following i.v. administration.¹⁴¹ In this study, 65-80% of the mice tested showed tumor regression.^{141,142} Investigators have recently demonstrated oncolytic activity of reovirus against human cancer cell lines carrying a high percentage of k-ras mutations implanted in mice.⁶¹ The k-ras mutation is observed in 30% of non-small cell lung carcinoma tumors. In addition to demonstrating the susceptibility of human k-raspositive cancer cells to reovirus infection in vitro, this study assessed the ability of reovirus to cause tumor regression and promote survival in immunocompromised mice implanted with human k-ras-positive cancers. Intratumoral injection of virus consistently resulted in major reductions of tumor volume. Of particular significance, i.v. administration of virus to immunocompromised mice consistently resulted in the regression of tumors at remote sites.

Clinically, 18 patients with refractory solid tumors have been treated in a phase I investigation, and a dose of up to 1×10^{10} p.f.u. was well tolerated. Preliminary results identified one patient achieving a CR and one a partial response. Eight maintained SD for a prolonged period.^{62,143}

Phase I clinical studies have been initiated in a range of cancer models, including a dose escalation study performed using intratumoral administration of reovirus in patients with recurrent malignant gliomas in which a maximum tolerated dose was not reached and treatment was well tolerated,¹⁴⁴ as was the i.v. administration of wild-type reovirus in patients with bone and soft tissue sarcomas metastatic to the lung in a phase II open label study.¹⁴⁵ A recent phase I open-label dose escalation study using i.v. administration of reovirus type 3 Dearing (Reolysin, Oncolytics Biotech, Calgary, Alberta, Canada) in patients with advanced cancer was well tolerated and exhibited successful intratumoral localization of reovirus after systemic delivery and confirmed the feasibility of i.v. delivery of high doses of reovirus.¹⁴⁶

Several studies have recently been completed involving the combination of reovirus and radiotherapy or chemotherapy, using taxanes in particular, to achieve synergistic tumor kill. In a phase I clinical study a wildtype reovirus serotype 3 Dearing strain (Reolysin, Oncolytics Biotech) was administered intravenously in combination with a chemotherapeutic agent, gemcitabine, exhibiting disease control for the majority of patients at a well-tolerated dose.¹⁴⁷ Wild-type reovirus serotype 3 Dearing strain was administered in combination with docetaxel in a phase I study in patients with a range of advanced malignancies resulting in objective radiological evidence of anticancer activity and toxicity consistent with that expected from the chemotherapeutic agent alone.¹⁴⁸ Reovirus dose escalation has also been recently evaluated in patients with advanced solid tumors in combination with carboplatin-paclitaxel and because of the promising results in patients with head and neck cancer a phase II study has been initiated for this indication.¹⁴⁹ Marked responses or stabilization in the treated legions for the majority of the patients was achieved in a recently completed phase II clinical study that evaluated the 309

biological effects of intratumoral administration of Reolysin in combination with low-dose radiotherapy in patients with advanced cancer. $^{150}\,$

The early clinical results and wide scope of potential application, as well as the relatively low inherent morbidity and mortality risk because of reovirus's limited pathogenecity in humans, are promising for this oncolytic agent; however, a greater understanding of the circumvention of humoral and cellular immune responses is needed in order to improve the efficacy of this treatment.

Seneca valley virus

A small non-pathogenic picornavirus with potential antineoplastic activity, Seneca Valley Virus-001 (also known by the trade name NTX-010, Neotropix) specifically targets and infects tumor cells with neuroendocrine characteristics, including small cell cancers and carcinoid, and replicates intracellularly resulting in cell lysis.¹⁵¹ It is the representative member of a new genus, Senecavirus. The cytolytic potential of this virus was first examined in neuroendocrine and pediatric tumor cell lines.¹⁵² After promising preclinical results, Seneca Valley Virus-001 (SVV-001) was first tested intravenously in a five-log increment dose escalation phase I study in patients with nueroendocrine cancers, 6 small cell and 24 carcinoidtype, and was found to be well tolerated and showed evidence of intratumoral viral replication in delayed kinetics in the serum viral titer, post-infusion serum titers greater than the dose administered, and positive immunohistochemistry and/or reverse transcriptase-PCR signal for viral antigens in the tumor mass although antibody production was detected.¹⁵³ A phase I single infusion multi-center study is also currently active in pediatric patients with relapsed or refractory neuroblastoma, rhabdomyosarcoma and rare tumors with neuroendocrine features (COG-ADVL0911), and finally, there is an active single infusion phase II randomized Seneca Valley Virus-001 after platinum-based chemotherapy study in patients with extensive-stage small cell lung cancer in which the primary objective is progression-free survival of treated patients compared with placebo (NCCTG-N0923). In addition to the ability of SVV-001, the first nonpathogenic picornavirus to be tested as an oncolytic viral therapy, to specifically target cancer cells, no pre-existing antibodies to the virus are found in humans. Via systemic delivery this agent could potentially be used either as a single agent or in combination with standard cytotoxic therapies.

Vaccinia

Vaccinia is a double-stranded DNA virus and a member of the poxvirus family. Vaccinia virus has tropism for human cells and is highly immunogenic. The immunogenic properties were exploited in the production of smallpox vaccine, leading to the eradication of smallpox. 310

Three techniques have been exploited for the development of oncolytic vaccinia viruses. These include the following: (1) Vaccinia virus has a high efficiency of infection, replicates in the cytoplasm without chromosomal integration, and its 200 kb genome allows the insertion of a large amount of recombinant DNA without loss of infectivity. (2) The immunostimulatory properties of the virus are being harnessed to incite an immune response against cancer cells. (3) Replication-conditional viral mutants are being constructed to target specific cancer types.

In one study,¹⁵⁴ recombinant vaccinia virus was constructed in an effort to enhance the immunogenicity of transfected melanoma cells. The virus expressed a minigene encoding a fusion product that combined an endoplasmic reticulum-targeting signal and the HLA-A201 binding 27-35 peptide. Infection of melanoma cells with this recombinant virus resulted in high levels of cytotoxicity from specific cytotoxoic T lymphocyte clones *in vitro*. In another study, ¹⁵⁵ a recombinant vaccinia virus vector was created containing the tumor-suppressor p53 gene. This virus demonstrated a high level of p53 expression in transfected glioma cells, resulting in high levels of apoptosis. A phase 1 study of intravesical vaccinia virus infection¹⁵⁶ demonstrated that vaccinia virus can be safely administrated into the bladder and found that the treatment was associated with an intense immune response with few clinical side effects.¹⁵⁷ Of the four patients studied, three survived and were free of disease at 4-year follow-up.

Many studies use vaccinia virus as an immunotherapeutic agent. Vaccinia oncolysate has been studied as a vaccine in early stage melanoma.^{156–165} Results suggested a good tolerability and survival advantage compared with historical controls. However, an unpublished prospective controlled trial failed to validate the use of vaccinia oncolysate. The control group did not receive standard care, but instead received live vaccinia virus without tumor oncolysate, which potentially could have affected patient response.

Wild-type vaccinia virus does not selectively infect cancer cells. The virus requires modification to be made replication-conditional. One strategy is to delete the viral TK gene. Although the viral TK gene is necessary for infectivity in normal cells that possess small concentrations of intracellular nucleotide pools, it is not necessary in cancer cells,¹⁵⁸ which possess relatively high concentrations of intracellular nucleotides. Another novel vector involved replacing the viral TK gene with the gene for GM-CSF, creating a mutant vaccinia virus capable of selectively infecting melanoma cells and inducing an antitumor immune response.¹⁵⁸ This virus has been administered intralesionally in a phase one clinical trial involving patients with refractory and/or recurrent melanoma. Injected lesions contained an active inflammatory response and demonstrable viral replication. Two out of seven patients studied had a CR, and three patients had a partial response.¹⁵⁸ Other studies have investigated a vaccinia virus carrying a prostate-specific antigen transgene in the treatment of prostate cancer patients with both minimal disease and metastatic disease. Evidence of cancer-specific immune activation was demonstrated, and tolerability was reasonable. In minimal disease patients with rising prostate-specific antigen following surgery or radiation therapy, 14 of 33 maintained SD for at least 6 months and 6 remained disease free for >2 years. Another mutant vaccinia virus, which deleted the viral *SPI-1* and *SPI-2* genes, resulted in conditional viral replication in cancer cells but not in normal cells.¹⁶⁶ The efficacy of this virus has not yet been tested. Other gene combinations, such as *B7-1*, *ICAM-1* and *LFA3*, have also been added to the vaccinia core construct. Results from animal studies are encouraging.^{167–169}

Conclusion

Encouraging safety profiles and local-regional activity have been demonstrated with a variety of oncolytic viral therapeutics. Unfortunately, the inability to demonstrate systemic response with currently available viral constructs limits future clinical development opportunities. The next generation of oncolytic viral products incorporates numerous modifications and strategies in an attempt to enhance activity. Specific strategies to improve viral immunogenicity and enhance potency include methods to reduce viral clearance, reduce immune inhibition of viral activity, increase intracellular viral release and replication, improve tumor cell specificity, uptake and expression, improve viral replication capacity, combination with other anticancer drugs, and addition of anticancer genes to second-generation vector design. Vector modification to 'arm' oncolytic viruses enables delivery of cancer-toxic genes. Other design modifications increase tumor delivery by the addition of cancer receptor/antigen components and enhance cancer cell expression through cancer promoter modification; these modifications have demonstrated encouraging early results.

Physical shielding to enhance delivery and reduce viral clearance is being tested^{170–175} (plus Rehman 2001). Currently, liposome encapsulated, polymer coated, and cell carrier modes of oncolytic virus delivery are being developed for preclinical testing. The liposome and polymer coated methods can be coated with tumorspecific antibodies, peptides or small molecules to further enhance tumor-specific uptake and delivery.^{176–184} Plas-mapheresis rotation of viral serotype^{170–175} and B-cell suppression have had limited testing as methods to reduce normal immune reactivity against administered viral particles. Restoration of the E3 region of the viral genome or E3 protein activity, in an effort to limit effects of tumor necrosis factor- α through combination with soluble tumor necrosis factor- α receptors, demonstrates positive effect.^{13,14,46,52,185} However, enhancement in viral access and uptake by malignant cells is not the only obstacle in the creation of oncolytic viruses. Clusters of viral particles have been demonstrated to accumulate in malignant tissue following i.v. administration without further

replication or spread. Consequently, new approaches which enhance viral replication and cell to cell spread are under investigation,^{174,186,187} while at the same time attempts are underway to identify more highly replicative viruses. Improving development of tumor-specific promoters to limit viral replication in malignant tissue may enable greater confidence in the utilization of replication aggressive viruses.^{14,188–197} Ultimately these modifications will likely need to be built into viral constructs that deliver molecular targeted therapeutics^{198–202} and can be utilized in combination with traditional therapeutics,^{203–208} thereby creating a 'super' virus.

Overall, it is likely that a combination of these approaches will be required to optimally maximize oncolytic viral activity, thereby expanding potential systemic therapeutic opportunities.

Conflict of interest

The authors declare no conflict of interest.

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References

- 1 Bischoff JR, Kirn DH, Williams A, Heise C, Horn S, Muna M *et al.* An adenovirus mutant that replicates selectively in p53-deficient human tumor cells. *Science* 1996; **274**: 373–376.
- 2 Heise C, Sampson-Johannes A, Williams A, McCormick F, Von Hoff DD, Kirn DH. ONYX-015, an E1B geneattenuated adenovirus, causes tumor-specific cytolysis and antitumoral efficacy that can be augmented by standard chemotherapeutic agents. *Nat Med* 1997; **3**: 639–645.
- 3 Yu DC, Sakamoto GT, Henderson DR. Identification of the transcriptional regulatory sequences of human kallikrein 2 and their use in the construction of calydon virus 764, an attenuated replication competent adenovirus for prostate cancer therapy. *Cancer Res* 1999; **59**: 1498–1504.
- 4 Yu DC, Chen Y, Seng M, Dilley J, Henderson DR. The addition of adenovirus type 5 region E3 enables calydon virus 787 to eliminate distant prostate tumor xenografts. *Cancer Res* 1999; **59**: 4200–4203.
- 5 Heise C, Hermiston T, Johnson L, Brooks G, Sampson-Johannes A, Williams A *et al*. An adenovirus E1A mutant that demonstrates potent and selective systemic antitumoral efficacy. *Nat Med* 2000; **6**: 1134–1139.
- 6 Freytag SO, Rogulski KR, Paielli DL, Gilbert JD, Kim JH. A novel three-pronged approach to kill cancer cells selectively: concomitant viral, double suicide gene, and radiotherapy. *Human Gene Ther* 1998; **9**: 1323–1333.
- 7 Fueyo J, Gomez-Manzano C, Alemany R, Lee PS, McDonnell TJ, Mitlianga P *et al.* A mutant oncolytic adenovirus targeting the Rb pathway produces anti-glioma effect *in vivo. Oncogene* 2000; **19**: 2–12.
- 8 Kurihara T, Brough DE, Kovesdi I, Kufe DW. Selectivity of a replication-competent adenovirus for human breast

carcinoma cells expressing the MUC1 antigen. *J Clin Invest* 2000; **106**: 763–771.

- 9 Johnson L. Cytosine deaminase-armed selectively-replicating adenovirus for the treatment of cancer. *Proc Am Assoc Cancer Res* 2002; 43: 3257.
- 10 Hallenbeck PL, Jacubczak J, Ryan P, Hawkins L, Ling X, Forry-Schaudies S. Oncolytic adenoviruses dependent on two prevalent alterations in human cancer: Disregulation of the Rb-pathway and telomerase. *Mol Ther* 2002; **5**: 165.
- 11 Doronin K, Toth K, Kuppuswamy M, Ward P, Tollefson AE, Wold WS. Tumor-specific, replication-competent adenovirus vectors overexpressing the adenovirus death protein. J Virol 2000; 74: 6147–6155.
- 12 Ramachandra M, Rahman A, Zou A, Vaillancourt M, Howe JA, Antelman D *et al.* Re-engineering adenovirus regulatory pathways to enhance oncolytic specificity and efficacy. *Nat Biotechnol* 2001; **19**: 1035–1041.
- 13 Fujiwara T, Urata Y, Tanaka N. Telomerase-specific oncolytic virotherapy for human cancer with the hTERT promoter. *Curr Cancer Drug Targets* 2007; **7**: 191–201.
- 14 Fujiwara T, Urata Y, Tanaka N. Diagnostic and therapeutic application of telomerase-specific oncolytic adenoviral agents. *Front Biosci* 2008; 13: 1881–1886.
- 15 Hansen RM, Libnoch JA. Remission of chronic lymphocytic leukemia after smallpox vaccination. *Arch Intern Med* 1978; **138**: 1137–1138.
- 16 Bousser J, Zittoun R. Prolonged spontaneous remission of chronic lymphoid leukemia. *Nouv Rev Fr Hematol* 1965; 5: 498–501.
- 17 Vladimirskaia EB. A case of prolonged spontaneous remission in a patient with chronic lymphatic leukemia. *Probl Gematol Pereliv Krovi* 1962; 7: 51–54.
- 18 Weintraub LR. Lymphosarcoma. JAMA 1969; 210: 1590–1591.
- 19 Sinkovics JG. Oncolytic viruses and viral oncolysates. Ann Immunol Hung 1986; 26: 271–290.
- 20 Bluming AZ, Ziegler JL. Regression of Burkitt's lymphoma in association with measles infection. *Lancet* 1971; **2**: 105–106.
- 21 Taqi AM, Abdurrahman MB, Yakubu AM, Fleming AF. Regression of Hodgkin's disease after measles. *Lancet* 1981; 1: 1112.
- 22 Bierman HR, Crile DM, Dod KS, Kelly KH, Petrakis NL, White LP *et al.* Remissions in leukemia of childhood following acute infectious disease: staphylococcus and streptococcus, varicella, and feline panleukopenia. *Cancer* 1953; 6: 591–605.
- 23 Pelner L, Fowler GA, Nauts HC. Effects of concurrent infections and their toxins on the course of leukemia. *Acta Med Scand* 1958; **162**(Suppl 338): 1–47.
- 24 London RE. Multiple myeloma: report of a case showing unusual remission lasting two years following severe hepatitis. *Ann Intern Med* 1955; **43**: 191–201.
- 25 Dock G. Influence of complicating diseases upon leukemia. *Am J Med Sci* 1904; **127**: 563–592.
- 26 Bierman HR, Hammon W, Eddie BU, Meyer KF, Shimkin MB. The effect of viruses and bacterial infections on neoplastic diseases. *Cancer Res* 1950; 10: 203–204.
- 27 Hernandez A. Observacion de un case de enfermedad de Hodgkin, con regresion de los sintomas e infartos ganglionares, post-sarampion. *Rev Med Cubana* 1949; 60: 120–125.
- 28 De Pace NG. Sulla scomparsa di un enorme cancro vegetante del callo dell'utero senza cura chirurgica. *Ginecologia* 1912; **9**: 82–88.

- 29 Pack GT. Note of the experimental use of rabies vaccine for melanomatosis. *Arch Dermtol and Syphilol* 1950; **62**: 694–695.
- 30 Southam CM. Present status of oncolytic virus studies. Trans NY Acad Sci 1960; 22: 657–673.
- 31 Asada T. Treatment of human cancer with mumps virus. *Cancer* 1974; **34**: 1907–1928.
- 32 Yamanishi K, Takahashi M, Kurimura T, Ueda S, Minekawa Y. Studies on live mumps virus vaccine. 3. Evaluation of newly developed live mumps virus vaccine. *Biken J* 1970; **13**: 157–161.
- 33 Harris RJC (ed). Carcinolytic viruses. *Biological Approaches to Cancer Chemotherapy*: Academic Press, New York, 1961.
- 34 Harris JE, Sinkovics JG. The Immunology of Malignant Disease. Mosby: St Louis, 1976, 180–182, 464–467, 475–478pp.
- 35 Hoster HA, Zanes RP, von Haam E. Studies in Hodgkin's syndrome. *Cancer Res* 1949; **9**: 473–480.
- 36 Lechner MS, Mack DH, Finicle AB, Crook T, Vousden KH, Laimins LA. Human papillomavirus E6 proteins bind p53 *in vivo* and abrogate p53-mediated repression of transcription. *EMBO J* 1992; **11**: 3045–3052.
- 37 Gannon JV, Lane DP. p53 and DNA polymerase alpha compete for binding to SV40 T antigen. *Nature* 1987; **329**: 456–458.
- 38 Nemunaitis J, Ganly I, Khuri F, Arseneau J, Kuhn J, McCarty T *et al.* Selective replication and oncolysis in p53 mutant tumors with ONYX-015, an E1B-55kD gene-deleted adenovirus, in patients with advanced head and neck cancer: a phase II trial. *Cancer Res* 2000; **60**: 6359–6366.
- 39 Ganly I, Kirn D, Eckhardt G, Rodriguez GI, Soutar DS, Otto R et al. A phase I study of Onyx-015, an E1B attenuated adenovirus, administered intratumorally to patients with recurrent head and neck cancer. *Clin Cancer Res* 2000; 6: 798–806.
- 40 Nemunaitis J, Khuri F, Ganly I, Arseneau J, Posner M, Vokes E *et al.* Phase II trial of intratumoral administration of ONYX-015, a replication-selective adenovirus, in patients with refractory head and neck cancer. *J Clin Oncol* 2001; **19**: 289–298.
- 41 Khuri FR, Nemunaitis J, Ganly I, Arseneau J, Tannock IF, Romel L *et al.* A controlled trial of intratumoral ONYX-015, a selectively-replicating adenovirus, in combination with cisplatin and 5-fluorouracil in patients with recurrent head and neck cancer. *Nat Med* 2000; **6**: 879–885.
- 42 Vasey PA, Shulman LN, Campos S, Davis J, Gore M, Johnston S *et al.* Phase I trial of intraperitoneal injection of the E1B-55-kd-gene-deleted adenovirus ONYX-015 (dl1520) given on days 1 through 5 every 3 weeks in patients with recurrent/refractory epithelial ovarian cancer. *J Clin Oncol* 2002; **20**: 1562–1569.
- 43 Nemunaitis J, Cunningham C, Tong AW, Post L, Netto G, Paulson AS *et al.* Pilot trial of intravenous infusion of a replication-selective adenovirus (ONYX-015) in combination with chemotherapy or IL-2 treatment in refractory cancer patients. *Cancer Gene Ther* 2003; **10**: 341–352.
- 44 Habib N, Salama H, Abd El Latif Abu Median A, Isac Anis I, Abd Al Aziz RA, Sarraf C *et al.* Clinical trial of E1B-deleted adenovirus (dl1520) gene therapy for hepatocellular carcinoma. *Cancer Gene Ther* 2002; **9**: 254–259.
- 45 Reid T, Sze D, Galanis E, Abbruzzese J, Andrews J, Rubin J et al. Intra-arterial administration of a replication-

selective adenovirus ONYX-015 in patients with colorectal carcinoma metastatic to the liver: safety, feasibility and biological activity. (abstr 793). *Proc Am Soc Clin Oncol* 2003; **22**: 198.

- 46 Nemunaitis J, Cunningham C, Buchanan A, Blackburn A, Edelman G, Maples P *et al.* Intravenous infusion of a replication-selective adenovirus (ONYX-015) in cancer patients: safety, feasibility and biological activity. *Gene Ther* 2001; 8: 746–759.
- 47 Mulvihill S, Warren R, Venook A, Adler A, Randlev B, Heise C *et al.* Safety and feasibility of injection with an E1B-55 kDa gene-deleted, replication-selective adenovirus (ONYX-015) into primary carcinomas of the pancreas: a phase I trial. *Gene Ther* 2001; **8**: 308–315.
- 48 Reid T, Galanis E, Abbruzzese J, Sze D, Wein LM, Andrews J *et al.* Hepatic arterial infusion of a replicationselective oncolytic adenovirus (dl1520): phase II viral, immunologic, and clinical endpoints. *Cancer Res* 2002; **62**: 6070–6079.
- 49 Hecht JR, Bedford R, Abbruzzese JL, Lahoti S, Reid TR, Soetikno RM *et al.* A phase I/II trial of intratumoral endoscopic ultrasound injection of ONYX-015 with intravenous genetitabine in unresectable pancreatic carcinoma. *Clin Cancer Res* 2003; **9**: 555–561.
- 50 Hamid O, Varterasian ML, Wadler S, Hecht JR, Benson III A, Galanis E *et al.* Phase II trial of intravenous CI-1042 in patients with metastatic colorectal cancer. *J Clin Oncol* 2003; **21**: 1498–1504.
- 51 Makower D, Rozenblit A, Kaufman H, Edelman M, Lane ME, Zwiebel J *et al.* Phase II clinical trial of intralesional administration of the oncolytic adenovirus ONYX-015 in patients with hepatobiliary tumors with correlative p53 studies. *Clin Cancer Res* 2003; **9**: 693–702.
- 52 Nemunaitis J, Senzer N, Sarmiento S, Zhang YA, Arzaga R, Sands B *et al.* A phase I trial of intravenous infusion of ONYX-015 and enbrel in solid tumor patients. *Cancer Gene Ther* 2007; **14**: 885–893.
- 53 Nemunaitis J, Edelman J. Selectively replicating viral vectors. *Cancer Gene Ther* 2002; **9**: 987–1000.
- 54 Reid T, Galanis E, Abbruzzese J, Sze D, Andrews J, Romel L et al. Intra-arterial administration of a replication-selective adenovirus (dl1520) in patients with colorectal carcinoma metastatic to the liver: a phase I trial. *Gene Ther* 2001; 8: 1618–1626.
- 55 Kirn D. The end of the beginning: oncolytic virotherapy achieves clinical proof-of-concept. *Mol Ther* 2006; **13**: 237–238.
- 56 Li JL, Liu HL, Zhang XR, Xu JP, Hu WK, Liang M et al. A phase I trial of intratumoral administration of recombinant oncolytic adenovirus overexpressing HSP70 in advanced solid tumor patients. *Gene Ther* 2009; 16: 376–382.
- 57 Shay JW, Zou Y, Hiyama E, Wright WE. Telomerase and cancer. *Hum Mol Genet* 2001; **10**: 677–685.
- 58 Kawashima T, Kagawa S, Kobayashi N, Shirakiya Y, Umeoka T, Teraishi F *et al.* Telomerase-specific replication-selective virotherapy for human cancer. *Clin Cancer Res* 2004; **10**(1 Pt 1): 285–292.
- 59 Lichtenstein DL, Toth K, Doronin K, Tollefson AE, Wold WS. Functions and mechanisms of action of the adenovirus E3 proteins. *Int Rev Immunol* 2004; 23: 75–111.
- 60 Nemunaitis J, Tong AW, Nemunaitis M, Senzer N, Phadke AP, Bedell C *et al.* A phase I study of telomerase-specific replication competent oncolytic adenovirus (telomelysin) for various solid tumors. *Mol Ther* 2010; **18**: 429–434.

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- 61 Robert MS, Lorence RM, Groene WS, Rabin H, Von Borstel RW (inventors). Treatment of neoplasms with viruses. 1999.
- 62 Stojdl DF, Lichty B, Knowles S, Marius R, Atkins H, Sonenberg N. *et al.* Exploiting tumor-specific defects in the interferon pathway with a previously unknown oncolytic virus. *Nat Med* 2000; **6**: 821–825.
- 63 Lorence RM, Roberts MS, Groene WS, Rabin H. Replication-competent, oncolytic Newcastle disease virus for cancer therapy. *Monogr Virol Basel, Karger* 2001; 22: 160–182.
- 64 Lorence RM, Reichard KW, Katubig BB, Reyes HM, Phuangsab A, Mitchell BR *et al.* Complete regression of human neuroblastoma xenografts in athymic mice after local Newcastle disease virus therapy. *J Natl Cancer Inst* 1994; **86**: 1228–1233.
- 65 Lorence RM, Katubig BB, Reichard KW, Reyes HM, Phuangsab A, Sassetti MD *et al.* Complete regression of human fibrosarcoma xenografts after local Newcastle disease virus therapy. *Cancer Res* 1994; **54**: 6017–6021.
- 66 Lorence RM, Pecora AL, Major PP, Hotte SJ, Laurie SA, Roberts MS, *et al.* Overview of phase I studies of intravenous adminstration of PV701, an oncolytic virus. *Curr Opin Mol Ther* 2003; **5**: 618–624.
- 67 Cassel WA, Garrett RE. Newcastle disease virus as an antineoplastic agent. *Cancer* 1965; 18: 863–868.
- 68 Batliwalla FM, Bateman BA, Serrano D, Murray D, Macphail S, Maino VC *et al.* A 15-year follow-up of AJCC stage III malignant melanoma patients treated postsurgically with Newcastle disease virus (NDV) oncolysate and determination of alterations in the CD8 T cell repertoire. *Mol Med (Cambridge, MA)* 1998; 4: 783–794.
- 69 Schirrmacher V, Ahlert T, Probstle T, Steiner HH, Herold-Mende C, Gerhards R *et al.* Immunization with virus-modified tumor cells. *Semin Oncol* 1998; **25**: 677–696.
- 70 Cassel WA, Murray DR, Phillips HS. A phase II study on the postsurgical management of Stage II malignant melanoma with a Newcastle disease virus oncolysate. *Cancer* 1983; **52**: 856–860.
- 71 Cassel WA, Murray DR. A ten-year follow-up on stage II malignant melanoma patients treated postsurgically with Newcastle disease virus oncolysate. *Med Oncol Tumor Pharmacother* 1992; 9: 169–171.
- 72 Sinkovics J, Horvath J. New developments in the virus therapy of cancer: a historical review. *Intervirology* 1993; **36**: 193–214.
- 73 Schlag P, Manasterski M, Gerneth T, Hohenberger P, Dueck M, Herfarth C *et al.* Active specific immunotherapy with Newcastle-disease-virus-modified autologous tumor cells following resection of liver metastases in colorectal cancer. First evaluation of clinical response of a phase II-trial. *Cancer Immunol Immunother* 1992; **35**: 325–330.
- 74 Kirchner HH, Anton P, Atzpodien J. Adjuvant treatment of locally advanced renal cancer with autologous virusmodified tumor vaccines. World J Urol 1995; 13: 171–173.
- 75 Haas C, Strauss G, Moldenhauer G, Iorio RM, Schirrmacher V. Bispecific antibodies increase T-cell stimulatory capacity *in vitro* of human autologous virus-modified tumor vaccine. *Clin Cancer Res* 1998; **4**: 721–730.
- 76 Csatary LK, Eckhardt S, Bukosza I, Czegledi F, Fenyvesi C, Gergely P *et al.* Attenuated veterinary virus vaccine for the treatment of cancer. *Cancer Detect Prev* 1993; 17: 619–627.

- 77 Mineta T, Rabkin SD, Martuza RL. Treatment of malignant gliomas using ganciclovir-hypersensitive, ribonucleotide reductase-deficient herpes simplex viral mutant. *Cancer Res* 1994; 54: 3963–3966.
- 78 Chase M, Chung RY, Chiocca EA. An oncolytic viral mutant that delivers the CYP2B1 transgene and augments cyclophosphamide chemotherapy. *Nat Biotechnol* 1998; 16: 444–448.
- 79 Boviatsis EJ, Scharf JM, Chase M, Harrington K, Kowall NW, Breakefield XO *et al.* Antitumor activity and reporter gene transfer into rat brain neoplasms inoculated with herpes simplex virus vectors defective in thymidine kinase or ribonucleotide reductase. *Gene Ther* 1994; 1: 323–331.
- 80 Varghese S, Rabkin SD. Oncolytic herpes simplex virus vectors for cancer virotherapy. *Cancer Gene Ther* 2002; **9**: 967–978.
- 81 Chou J, Kern ER, Whitley RJ, Roizman B. Mapping of herpes simplex virus-1 neurovirulence to gamma 134.5, a gene nonessential for growth in culture. *Science* 1990; 250: 1262–1266.
- 82 Chou J, Roizman B. The gamma 1(34.5) gene of herpes simplex virus 1 precludes neuroblastoma cells from triggering total shutoff of protein synthesis characteristic of programed cell death in neuronal cells. *Proc Natl Acad Sci USA* 1992; **89**: 3266–3270.
- 83 McKie EA, MacLean AR, Lewis AD, Cruickshank G, Rampling R, Barnett SC *et al.* Selective *in vitro* replication of herpes simplex virus type 1 (HSV-1) ICP34.5 null mutants in primary human CNS tumours–evaluation of a potentially effective clinical therapy. *Br J Cancer* 1996; 74: 745–752.
- 84 Martuza RL. Act locally, think globally. *Nat Med* 1997; **3**: 1323.
- 85 Alemany R, Gomez-Manzano C, Balague C, Yung WK, Curiel DT, Kyritsis AP *et al.* Gene therapy for gliomas: molecular targets, adenoviral vectors, and oncolytic adenoviruses. *Exp Cell Res* 1999; **252**: 1–12.
- 86 Pennisi E. Will a twist of viral fate lead to a new cancer treatment? *Science* 1996; **274**: 342–343.
- 87 Mineta T, Rabkin SD, Yazaki T, Hunter WD, Martuza RL. Attenuated multi-mutated herpes simplex virus-1 for the treatment of malignant gliomas. *Nat Med* 1995; 1: 938–943.
- 88 Andreansky SS, He B, Gillespie GY, Soroceanu L, Markert J, Chou J *et al.* The application of genetically engineered herpes simplex viruses to the treatment of experimental brain tumors. *Proc Natl Acad Sci USA* 1996; **93**: 11313–11318.
- 89 Chambers R, Gillespie GY, Soroceanu L, Andreansky S, Chatterjee S, Chou J *et al.* Comparison of genetically engineered herpes simplex viruses for the treatment of brain tumors in a scid mouse model of human malignant glioma. *Proc Natl Acad Sci USA* 1995; **92**: 1411–1415.
- 90 Kesari S, Randazzo BP, Valyi-Nagy T, Huang QS, Brown SM, MacLean AR *et al.* Therapy of experimental human brain tumors using a neuroattenuated herpes simplex virus mutant. *Lab Invest J Tech Methods Pathol* 1995; **73**: 636–648.
- 91 Lambright ES, Kang EH, Force S, Lanuti M, Caparrelli D, Kaiser LR *et al.* Effect of preexisting anti-herpes immunity on the efficacy of herpes simplex viral therapy in a murine intraperitoneal tumor model. *Mol Ther* 2000; 2: 387–393.
- 92 Randazzo BP, Bhat MG, Kesari S, Fraser NW, Brown SM. Treatment of experimental subcutaneous

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human melanoma with a replication-restricted herpes simplex virus mutant. *J Invest Dermatol* 1997; **108**: 933–937.

- 93 Toda M, Rabkin SD, Martuza RL. Treatment of human breast cancer in a brain metastatic model by G207, a replication-competent multimutated herpes simplex virus 1. *Human Gene Ther* 1998; 9: 2177–2185.
- 94 Toda M, Rabkin SD, Kojima H, Martuza RL. Herpes simplex virus as an *in situ* cancer vaccine for the induction of specific anti-tumor immunity. *Human Gene Ther* 1999; **10**: 385–393.
- 95 Yoon SS, Carroll NM, Chiocca EA, Tanabe KK. Cancer gene therapy using a replication-competent herpes simplex virus type 1 vector. *Ann Surg* 1998; **228**: 366–374.
- 96 Coukos G, Makrigiannakis A, Kang EH, Caparelli D, Benjamin I, Kaiser LR *et al.* Use of carrier cells to deliver a replication-selective herpes simplex virus-1 mutant for the intraperitoneal therapy of epithelial ovarian cancer. *Clin Cancer Res* 1999; **5**: 1523–1537.
- 97 Lambright ES, Caparrelli DJ, Abbas AE, Toyoizumi T, Coukos G, Molnar-Kimber KL *et al.* Oncolytic therapy using a mutant type-1 herpes simplex virus and the role of the immune system. *Ann Thorac Surg* 1999; **68**: 1756–1760; discussion 1761–1752.
- 98 Cozzi PJ, Burke PB, Bhargav A, Heston WD, Huryk B, Scardino PT *et al.* Oncolytic viral gene therapy for prostate cancer using two attenuated, replicationcompetent, genetically engineered herpes simplex viruses. *Prostate* 2002; **53**: 95–100.
- 99 Coukos G, Rubin SC, Molnar-Kimber KL. Application for recombinant herpes simplex virus-1 (HSV-1) for the treatment of malignancies outside the central nervous system. *Gene Ther Mol Biol* 1999; **3**: 79–89.
- 100 Kucharczuk JC, Randazzo B, Chang MY, Amin KM, Elshami AA, Sterman DH *et al.* Use of a 'replicationrestricted' herpes virus to treat experimental human malignant mesothelioma. *Cancer Res* 1997; **57**: 466–471.
- 101 Toyoizumi T, Mick R, Abbas AE, Kang EH, Kaiser LR, Molnar-Kimber KL. Combined therapy with chemotherapeutic agents and herpes simplex virus type 1 ICP34.5 mutant (HSV-1716) in human non-small cell lung cancer. *Human Gene Ther* 1999; **10**: 3013–3029.
- 102 Advani SJ, Chung SM, Yan SY, Gillespie GY, Markert JM, Whitley RJ *et al.* Replication-competent, nonneuroinvasive genetically engineered herpes virus is highly effective in the treatment of therapy-resistant experimental human tumors. *Cancer Res* 1999; **59**: 2055–2058.
- 103 Carroll NM, Chiocca EA, Takahashi K, Tanabe KK. Enhancement of gene therapy specificity for diffuse colon carcinoma liver metastases with recombinant herpes simplex virus. *Ann Surg* 1996; **224**: 323–329; discussion 329–330.
- 104 Papanastassiou V, Rampling R, Fraser M, Petty R, Hadley D, Nicoll J et al. The potential for efficacy of the modified (ICP 34.5(-)) herpes simplex virus HSV1716 following intratumoural injection into human malignant glioma: a proof of principle study. *Gene Ther* 2002; 9: 398–406.
- 105 Todo T, Rabkin SD, Sundaresan P, Wu A, Meehan KR, Herscowitz HB *et al.* Systemic antitumor immunity in experimental brain tumor therapy using a multimutated, replication-competent herpes simplex virus. *Human Gene Ther* 1999; **10**: 2741–2755.

- 106 Endo T, Toda M, Watanabe M, Iizuka Y, Kubota T, Kitajima M *et al. In situ* cancer vaccination with a replication-conditional HSV for the treatment of liver metastasis of colon cancer. *Cancer Gene Ther* 2002; **9**: 142–148.
- 107 Toda M, Martuza RL, Kojima H, Rabkin SD. In situ cancer vaccination: an IL-12 defective vector/replicationcompetent herpes simplex virus combination induces local and systemic antitumor activity. J Immunol 1998; 160: 4457–4464.
- 108 Carew JF, Kooby DA, Halterman MW, Kim SH, Federoff HJ, Fong Y. A novel approach to cancer therapy using an oncolytic herpes virus to package amplicons containing cytokine genes. *Mol Ther* 2001; **4**: 250–256.
- 109 Todo T, Martuza RL, Dallman MJ, Rabkin SD. *In situ* expression of soluble B7-1 in the context of oncolytic herpes simplex virus induces potent antitumor immunity. *Cancer Res* 2001; **61**: 153–161.
- 110 Wong RJ, Patel SG, Kim S, DeMatteo RP, Malhotra S, Bennett JJ *et al.* Cytokine gene transfer enhances herpes oncolytic therapy in murine squamous cell carcinoma. *Human Gene Ther* 2001; **12**: 253–265.
- 111 Bennett JJ, Malhotra S, Wong RJ, Delman K, Zager J, St-Louis M *et al.* Interleukin 12 secretion enhances antitumor efficacy of oncolytic herpes simplex viral therapy for colorectal cancer. *Ann Surg* 2001; **233**: 819–826.
- 112 Parker JN, Gillespie GY, Love CE, Randall S, Whitley RJ, Markert JM. Engineered herpes simplex virus expressing IL-12 in the treatment of experimental murine brain tumors. *Proc Natl Acad Sci USA* 2000; **97**: 2208–2213.
- 113 Jorgensen TJ, Katz S, Wittmack EK, Varghese S, Todo T, Rabkin SD *et al.* Ionizing radiation does not alter the antitumor activity of herpes simplex virus vector G207 in subcutaneous tumor models of human and murine prostate cancer. *Neoplasia* 2001; **3**: 451–456.
- 114 Chahlavi A, Todo T, Martuza RL, Rabkin SD. Replication-competent herpes simplex virus vector G207 and cisplatin combination therapy for head and neck squamous cell carcinoma. *Neoplasia* 1999; **1**: 162–169.
- 115 Advani SJ, Sibley GS, Song PY, Hallahan DE, Kataoka Y, Roizman B *et al.* Enhancement of replication of genetically engineered herpes simplex viruses by ionizing radiation: a new paradigm for destruction of therapeutically intractable tumors. *Gene Ther* 1998; **5**: 160–165.
- 116 Bradley JD, Kataoka Y, Advani S, Chung SM, Arani RB, Gillespie GY *et al.* Ionizing radiation improves survival in mice bearing intracranial high-grade gliomas injected with genetically modified herpes simplex virus. *Clin Cancer Res* 1999; **5**: 1517–1522.
- 117 Chung SM, Advani SJ, Bradley JD, Kataoka Y, Vashistha K, Yan SY *et al.* The use of a genetically engineered herpes simplex virus (R7020) with ionizing radiation for experimental hepatoma. *Gene Ther* 2002; **9**: 75–80.
- 118 Blank SV, Rubin SC, Coukos G, Amin KM, Albelda SM, Molnar-Kimber KL. Replication-selective herpes simplex virus type 1 mutant therapy of cervical cancer is enhanced by low-dose radiation. *Human Gene Ther* 2002; 13: 627–639.
- 119 Spear MA, Sun F, Eling DJ, Gilpin E, Kipps TJ, Chiocca EA *et al.* Cytotoxicity, apoptosis, and viral replication in tumor cells treated with oncolytic ribonucleotide reductase-defective herpes simplex type 1 virus (hrR3) combined with ionizing radiation. *Cancer Gene Ther* 2000; **7**: 1051–1059.
- 120 Coukos G, Makrigiannakis A, Kang EH, Rubin SC, Albelda SM, Molnar-Kimber KL. Oncolytic herpes simplex virus-1 lacking ICP34.5 induces p53-independent death

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and is efficacious against chemotherapy-resistant ovarian cancer. *Clin Cancer Res* 2000; **6**: 3342–3353.

- 121 Pawlik TM, Nakamura H, Yoon SS, Mullen JT, Chandrasekhar S, Chiocca EA *et al.* Oncolysis of diffuse hepatocellular carcinoma by intravascular administration of a replication-competent, genetically engineered herpesvirus. *Cancer Res* 2000; **60**: 2790–2795.
- 122 Nakamura H, Mullen JT, Chandrasekhar S, Pawlik TM, Yoon SS, Tanabe KK. Multimodality therapy with a replication-conditional herpes simplex virus 1 mutant that expresses yeast cytosine deaminase for intratumoral conversion of 5-fluorocytosine to 5-fluorouracil. *Cancer Res* 2001; **61**: 5447–5452.
- 123 Miyatake S, Martuza RL, Rabkin SD. Defective herpes simplex virus vectors expressing thymidine kinase for the treatment of malignant glioma. *Cancer Gene Ther* 1997; **4**: 222–228.
- 124 Aghi M, Chou TC, Suling K, Breakefield XO, Chiocca EA. Multimodal cancer treatment mediated by a replicating oncolytic virus that delivers the oxazaphosphorine/ rat cytochrome P450 2B1 and ganciclovir/herpes simplex virus thymidine kinase gene therapies. *Cancer Res* 1999; **59**: 3861–3865.
- 125 Todo T, Rabkin SD, Martuza RL. Evaluation of ganciclovir-mediated enhancement of the antitumoral effect in oncolytic, multimutated herpes simplex virus type 1 (G207) therapy of brain tumors. *Cancer Gene Ther* 2000; 7: 939–946.
- 126 Fruh K, Ahn K, Djaballah H, Sempe P, van Endert PM, Tampe R et al. A viral inhibitor of peptide transporters for antigen presentation. *Nature* 1995; 375: 415–418.
- 127 York IA, Roop C, Andrews DW, Riddell SR, Graham FL, Johnson DC. A cytosolic herpes simplex virus protein inhibits antigen presentation to CD8+ T lymphocytes. *Cell* 1994; 77: 525–535.
- 128 Todo T, Martuza RL, Rabkin SD, Johnson PA. Oncolytic herpes simplex virus vector with enhanced MHC class I presentation and tumor cell killing. *Proc Natl Acad Sci USA* 2001; 98: 6396–6401.
- 129 Martuza RL. Conditionally replicating herpes vectors for cancer therapy. *J Clin Invest* 2000; **105**: 841–846.
- 130 Berger C, Xuereb S, Johnson DC, Watanabe KS, Kiem HP, Greenberg PD *et al.* Expression of herpes simplex virus ICP47 and human cytomegalovirus US11 prevents recognition of transgene products by CD8(+) cytotoxic T lymphocytes. *J Virol* 2000; **74**: 4465–4473.
- 131 Kaufman HL, Deraffele G, Mitcham J, Moroziewicz D, Cohen SM, Hurst-Wicker KS *et al.* Targeting the local tumor microenvironment with vaccinia virus expressing B7.1 for the treatment of melanoma. *J Clin Invest* 2005; 115: 1903–1912.
- 132 Hu JC, Coffin RS, Davis CJ, Graham NJ, Groves N, Guest PJ *et al.* A phase I study of OncoVEXGM-CSF, a second-generation oncolytic herpes simplex virus expressing granulocyte macrophage colony-stimulating factor. *Clin Cancer Res* 2006; **12**: 6737–6747.
- 133 Senzer NN, Kaufman HL, Amatruda T, Nemunaitis M, Reid T, Daniels G *et al.* Phase II clinical trial of a granulocyte-macrophage colony-stimulating factor-encoding, second-generation oncolytic herpesvirus in patients with unresectable metastatic melanoma. *J Clin Oncol* 2009; 27: 5763–5771.
- 134 Hashiro G, Loh PC, Yau JT. The preferential cytotoxicity of reovirus for certain transformed cell lines. *Arch Virol* 1977; 54: 307–315.

- 135 Rosen L, Evans HE, Spickard A. Reovirus infections in human volunteers. *Am J Hyg* 1963; 77: 29–37.
- 136 Tai JH, Williams JV, Edwards KM, Wright PF, Crowe Jr JE, Dermody TS. Prevalence of reovirus-specific antibodies in young children in Nashville, Tennessee. J Infect Dis 2005; 191: 1221–1224.
- 137 Jackson GG, Muldoon RL. Viruses causing common respiratory infection in man. IV. Reoviruses and Adenoviruses. J Infect Dis 1973; **128**: 811–866.
- 138 Adams DJ, Spendlove JC, Spendlove RS, Barnett BB. Aerosol stability of infectious and potentially infectious reovirus particles. *Appl Environ Microbiol* 1982; **44**: 903–908.
- 139 Strong JE, Coffey MC, Tang D, Sabinin P, Lee PW. The molecular basis of viral oncolysis: usurpation of the Ras signaling pathway by reovirus. *EMBO J* 1998; **17**: 3351–3362.
- 140 Coffey MC, Strong JE, Forsyth PA, Lee PW. Reovirus therapy of tumors with activated Ras pathway. *Science* 1998; 282: 1332–1334.
- 141 Hirasawa K, Yoon C, Nishikawa SG, Waisman DM, Lee PW. Reovirus therapy of metastatic cancer models in immune-competent mice. *Proc Am Assoc Cancer Res* 2001; 42: 2427.
- 142 Hirasawa K, Nishikawa SG, Norman KL, Alain T, Kossakowska A, Lee PW. Oncolytic reovirus against ovarian and colon cancer. *Cancer Res* 2002; **62**: 1696–1701.
- 143 Morris DG, Forsyth PA, Paterson AHG, Fonseca K, DiFrancesco LM, Thompson BG *et al.* A phase I clinical trial evaluating intralesional Reolysin (reovirus) in histologically confirmed malignancies. *Proc Am Soc Clin Oncol* 2002; **21**: 24. Abstract 92.
- 144 Forsyth P, Roldan G, George D, Wallace C, Palmer CA, Morris D *et al.* A phase I trial of intratumoral administration of reovirus in patients with histologically confirmed recurrent malignant gliomas. *Mol Ther* 2008; **16**: 627–632.
- 145 Soefje SA, Sarantopoulos J, Sankhala KK, Mita AC, Mahany JJ, Carmona T *et al.* A phase II study of intravenous reolysin (wild-type reovirus) in the treatment of patients with bone and soft tissue sarcomas metastatic to the lung. *J Clin Oncol* 2008; **26**: Abstract 10568.
- 146 Vidal L, Pandha HS, Yap TA, White CL, Twigger K, Vile RG *et al.* A phase I study of intravenous oncolytic reovirus type 3 Dearing in patients with advanced cancer. *Clin Cancer Res* 2008; **14**: 7127–7137.
- 147 Arkenau H, Evans J, Lokelma M, Roxburgh P, Morisson R, Coffey M et al. A phase I study of the combination of intravenous Reolysin (REO) and gemcitabine (GEM) in patients (pts) with advanced cancer. J Clin Oncol, 2009 ASCO Ann Meeting Proc 2009; 27 (15S, May 20 Suppl): Abstract 3584.
- 148 Comins C, Spicer J, Protheroe A, Mukherji D, Coffey M, Thompson B et al. A phase I study to evaluate systemic wild-type reovirus (REOLYSIN) in combination with docetaxel in patients with advanced malignancies. J Immunother 2008; 31: 951.
- 149 Karapanagiotou E, Pandha H, Hall G, Chester J, Melcher A, de Bono J *et al.* Phase I trial of oncolytic reovirus (reolysin) in combination with carboplatin/ paclitaxel in patients with advanced solid cancers. *J Immunother* 2008; **31**: 952.
- 150 Saunders M, Anthoney A, Coffey M, Mettinger K, Thompson B, Melcher A *et al.* Results of a phase II study to evaluate the biological effects of intratumoral (ITu)

- reolysin in combination with low dose radiotherapy (RT) in patients (Pts) with advanced cancers. *J Clin Oncol, 2009* ASCO Ann Meeting Proc 2009; **27**(15S, May 20 Suppl): e14514.
- 151 Reddy PS, Burroughs KD, Hales LM, Ganesh S, Jones BH, Idamakanti N *et al.* Seneca Valley virus, a systemically deliverable oncolytic picornavirus, and the treatment of neuroendocrine cancers. *J Natl Cancer Inst* 2007; **99**: 1623–1633.
- 152 Morton CL, Houghton PJ, Kolb EA, Gorlick R, Reynolds CP, Kang MH *et al.* Initial testing of the replication competent Seneca Valley virus (NTX-010) by the pediatric preclinical testing program. *Pediatr Blood Cancer* 2010; 55: 295–303.
- 153 Rudin CM, Senzer N, Stephenson J, Loesch D, Burroughs K, Police SR *et al.* Phase I study of intravenous Seneca Valley virus (NTX-010), a replication competent oncolytic virus, in patients with neuroendocrine (NE) cancers. *J Clin Oncol* 2009; **27**(15s, Suppl): Abstract 4629.
- 154 Schutz A, Oertli D, Marti WR, Noppen C, Padovan E, Spagnoli GC *et al.* Immunogenicity of nonreplicating recombinant vaccinia expressing HLA-A201 targeted or complete MART-1/Melan-A antigen. *Cancer Gene Ther* 2001; 8: 655–661.
- 155 Timiryasova TM, Chen B, Fodor I. Replication-deficient vaccinia virus gene therpay vector: evaluation of exogenous gene expression mediated by PUV-inactivated virus in glioma cells. *J Gene Med* 2001; **3**: 468–477.
- 156 Gomella LG, Mastrangelo MJ, McCue PA, Maguire HJ, Mulholland SG, Lattime EC. Phase i study of intravesical vaccinia virus as a vector for gene therapy of bladder cancer. J Urol 2001; **166**: 1291–1295.
- 157 Wallack MK, Sivanandham M, Balch CM, Urist MM, Bland KI, Murray D *et al.* Surgical adjuvant active specific immunotherapy for patients with stage III melanoma: the final analysis of data from a phase III, randomized, double-blind, multicenter vaccinia melanoma oncolysate trial. *J Am Coll Surg* 1998; **187**: 69–77; discussion 77–69.
- 158 Mastrangelo MJ, Maguire Jr HC, Eisenlohr LC, Laughlin CE, Monken CE, McCue PA *et al.* Intratumoral recombinant GM-CSF-encoding virus as gene therapy in patients with cutaneous melanoma. *Cancer Gene Ther* 1999; 6: 409–422.
- 159 Wallack MK, Bash JA, Leftheriotis E, Seigler H, Bland K, Wanebo H *et al.* Positive relationship of clinical and serologic responses to vaccinia melanoma oncolysate. *Arch Surg* 1987; **122**: 1460–1463.
- 160 Wallack MK, Scoggin SD, Sivanandham M. Active specific immunotherapy with vaccinia melanoma oncolysate. *Mt Sinai J Med NY* 1992; **59**: 227–233.
- 161 Scoggin SD, Sivanandham M, Sperry RG, Wallack MK. Active specific adjuvant immunotherapy with vaccinia melanoma oncolysate. *Ann Plast Surg* 1992; 28: 108–109.
- 162 Mukherjee S, Haenel T, Himbeck R, Scott B, Ramshaw I, Lake RA *et al.* Replication-restricted vaccinia as a cytokine gene therapy vector in cancer: persistent transgene expression despite antibody generation. *Cancer Gene Ther* 2000; **7**: 663–670.
- 163 van Ophoven A, Gitlitz B, Tso CL, Dolan N, Stiles A, Kulmaticki A *et al.* Phase I dose escalation trials of vaccinia (VV)-MUC-1-IL-2 gene therapy in patient with advanced MUC-1+ prostate cancer (CAP) and non small cell lung cancer (NSCLC) B. *Proc Am Soc Clin Oncol* 2000; 19: abstract 1811.

- 164 Eder JP, Kantoff PW, Roper K, Xu GX, Bubley GJ, Boyden J et al. A phase I trial of a recombinant vaccinia virus expressing prostate-specific antigen in advanced prostate cancer. *Clin Cancer Res* 2000; **6**: 1632–1638.
- 165 Gulley J, Chen AP, Dahut W, Arlen PM, Bastian A, Steinberg SM *et al.* Phase I study of a vaccine using recombinant vaccinia virus expressing PSA (rV-PSA) in patients with metastatic androgen-independent prostate cancer. *Prostate* 2002; **53**: 109–117.
- 166 Naik AM, Xu H, Alexander HR, Bartlett DL. A mutant vaccinia virus with improved tumor selectivity. *Proc 54th Ann SSO Cancer Symp* 2001; Washington, DC.
- 167 Freund YR, Mirsalis JC, Fairchild DG, Brune J, Hokama LA, Schindler-Horvat J et al. Vaccination with a recombinant vaccinia vaccine containing the B7-1 co-stimulatory molecule causes no significant toxicity and enhances T cell-mediated cytotoxicity. Int J Cancer 2000; 85: 508–517.
- 168 Shankar P, Schlom J, Hodge JW. Enhanced activation of rhesus T cells by vectors encoding a triad of costimulatory molecules (B7-1, ICAM-1, LFA-3). *Vaccine* 2001; 20: 744–755.
- 169 Kalus RM, Kantor JA, Gritz L, Gomez Yafal A, Mazzara GP, Schlom J et al. The use of combination vaccinia vaccines and dual-gene vaccinia vaccines to enhance antigen-specific T-cell immunity via T-cell costimulation. Vaccine 1999; 17: 893–903.
- 170 Wolff G, Worgall S, van Rooijen N, Song WR, Harvey BG, Crystal RG. Enhancement of *in vivo* adenovirus-mediated gene transfer and expression by prior depletion of tissue macrophages in the target organ. *J Virol* 1997; **71**: 624–629.
- 171 Fisher KD. Polymer-coated adenovirus can be programmed to infect cells using selected ligands and shows extended plasma circulation following intravenous injection. *Third International Meeting on Oncolytic Viruses as Cancer Therapeutics*, Banff, Alberta 2003, p 17.
- 172 Green NK, Herbert CW, Hale SJ, Hale AB, Mautner V, Harkins R *et al.* Extended plasma circulation time and decreased toxicity of polymer-coated adenovirus. *Gene Ther* 2004; **11**: 1256–1263.
- 173 Fisher KD, Green NK, Hale A, Subr V, Ulbrich K, Seymour LW. Passive tumour targeting of polymer-coated adenovirus for cancer gene therapy. *J Drug Target* 2007; **15**: 546–551.
- 174 Wohlfahrt ME, Beard BC, Lieber A, Kiem HP. A capsidmodified, conditionally replicating oncolytic adenovirus vector expressing TRAIL Leads to enhanced cancer cell killing in human glioblastoma models. *Cancer Res* 2007; 67: 8783–8790.
- 175 Fisher PB, Gopalkrishnan RV, Chada S, Ramesh R, Grimm EA, Rosenfeld MR *et al.* mda-7/IL-24, a novel cancer selective apoptosis inducing cytokine gene: from the laboratory into the clinic. *Cancer Biol Ther* 2003; 2 (4 Suppl 1): S23–S37.
- 176 Douglas JT, Rogers BE, Rosenfeld ME, Michael SI, Feng M, Curiel DT. Targeted gene delivery by tropism-modified adenoviral vectors. *Nat Biotechnol* 1996; 14: 1574–1578.
- 177 Haisma HJ, Pinedo HM, Rijswijk A, der Meulen-Muileman I, Sosnowski BA, Ying W *et al.* Tumor-specific gene transfer via an adenoviral vector targeted to the pan-carcinoma antigen EpCAM. *Gene Ther* 1999; 6: 1469–1474.
- 178 Goldman CK, Rogers BE, Douglas JT, Sosnowski BA, Ying W, Siegal GP *et al.* Targeted gene delivery to Kaposi's sarcoma cells via the fibroblast growth factor receptor. *Cancer Res* 1997; **57**: 1447–1451.

316

- 180 Grill J, Van Beusechem VW, Van Der Valk P, Dirven CM, Leonhart A, Pherai DS *et al.* Combined targeting of adenoviruses to integrins and epidermal growth factor receptors increases gene transfer into primary glioma cells and spheroids. *Clin Cancer Res* 2001; 7: 641–650.
- 181 Sebestyen Z, de Vrij J, Magnusson M, Debets R, Willemsen R. An oncolytic adenovirus redirected with a tumor-specific T-cell receptor. *Cancer Res* 2007; **67**: 11309–11316.
- 182 Rogers RP, Strominger JL, Speck SH. Epstein-Barr virus in B lymphocytes: viral gene expression and function in latency. *Adv Cancer Res* 1992; **58**: 1–26.
- 183 Ulasov IV, Zhu ZB, Tyler MA, Han Y, Rivera AA, Khramtsov A *et al.* Survivin-driven and fiber-modified oncolytic adenovirus exhibits potent antitumor activity in established intracranial glioma. *Human Gene Ther* 2007; **18**: 589–602.
- 184 Ulasov IV, Rivera AA, Han Y, Curiel DT, Zhu ZB, Lesniak MS. Targeting adenovirus to CD80 and CD86 receptors increases gene transfer efficiency to malignant glioma cells. *J Neurosurg* 2007; **107**: 617–627.
- 185 Fujiwara T, Tanaka N. Theranostic application of telomerase-specific oncolytic adenovirus for human cancer. *Nippon Rinsho* 2007; 65: 1913–1922.
- 186 Doronin K, Toth K, Kuppuswamy M, Krajcsi P, Tollefson AE, Wold WS. Overexpression of the ADP (E3-11.6K) protein increases cell lysis and spread of adenovirus. *Virology* 2003; 305: 378–387.
- 187 Cheng J, Sauthoff H, Huang Y, Kutler DI, Bajwa S, Rom WN *et al.* Human matrix metalloproteinase-8 gene delivery increases the oncolytic activity of a replicating adenovirus. *Mol Ther* 2007; **15**: 1982–1990.
- 188 Delgado-Enciso I, Cervantes-Garcia D, Martinez-Davila IA, Ortiz-Lopez R, Alemany-Bonastre R, Silva-Platas CI *et al.* A potent replicative delta-24 adenoviral vector driven by the promoter of human papillomavirus 16 that is highly selective for associated neoplasms. *J Gene Med* 2007; 9: 852–861.
- 189 Kim E, Kim JH, Shin HY, Lee H, Yang JM, Kim J et al. Ad-mTERT-delta19, a conditional replication-competent adenovirus driven by the human telomerase promoter, selectively replicates in and elicits cytopathic effect in a cancer cell-specific manner. *Human Gene Ther* 2003; 14: 1415–1428.
- 190 Vile RG, Hart IR. *In vitro* and *in vivo* targeting of gene expression to melanoma cells. *Cancer Res* 1993; 53: 962–967.
- 191 Gao Z, Fields JZ, Boman BM. Tumor-specific expression of anti-mdr1 ribozyme selectively restores chemosensitivity in multidrug-resistant colon-adenocarcinoma cells. *Int J Cancer* 1999; 82: 346–352.
- 192 Savontaus MJ, Sauter BV, Huang TG, Woo SL. Transcriptional targeting of conditionally replicating adenovirus to dividing endothelial cells. *Gene Ther* 2002; 9: 972–979.
- 193 Thompson GA, Boston RS, Lyznik LA, Hodges TK, Larkins BA. Analysis of promoter activity from an alpha-zein gene 5' flanking sequence in transient expression assays. *Plant Mol Biol* 1990; 15: 755–764.

- 194 Thompson EM, Nagata S, Tsuji FI. Vargula hilgendorfii luciferase: a secreted reporter enzyme for monitoring gene expression in mammalian cells. *Gene* 1990; **96**:
- 257–262.
 195 Shirakawa T, Hamada K, Zhang Z, Okada H, Tagawa M, Kamidono S *et al.* A cox-2 promoter-based replication-selective adenoviral vector to target the cox-2-expressing human bladder cancer cells. *Clin Cancer Res* 2004; 10: 4342–4348.
- 196 Kawakami K, Kawakami M, Joshi BH, Puri RK. Interleukin-13 receptor-targeted cancer therapy in an immunodeficient animal model of human head and neck cancer. *Cancer Res* 2001; **61**: 6194–6200.
- 197 Kawakami K, Takeshita F, Puri RK. Identification of distinct roles for a dileucine and a tyrosine internalization motif in the interleukin (IL)-13 binding component IL-13 receptor alpha 2 chain. J Biol Chem 2001; 276: 25114–25120.
- 198 Zhang YA, Nemunaitis J, Samuel SK, Chen P, Shen Y, Tong AW. Antitumor activity of an oncolytic adenovirusdelivered oncogene small interfering RNA. *Cancer Res* 2006; **66**: 9736–9743.
- 199 Tong AW. Small RNAs and non-small cell lung cancer. Curr Mol Med 2006; 6: 339–349.
- 200 DeWeese TL, van der Poel H, Li S, Mikhak B, Drew R, Goemann M et al. A phase I trial of CV706, a replicationcompetent, PSA selective oncolytic adenovirus, for the treatment of locally recurrent prostate cancer following radiation therapy. *Cancer Res* 2001; **61**: 7464–7472.
- 201 Ther M (ed). A Phase I/II dose escalation trial of the intra prostatic injection of CG7870, a prostate specific antigendependent oncolytic adenovirus in patients with locally recurrent prostate cancer following definitive radiotherapy. *Sixth Annual Meeting of the American Society of Gene Therapy*; June 4–8; Washington, DC, 2003.
- 202 Ther M (ed). Circulating oncolytic and wild type adenovirus levels in clinical trial patients treated with CG7870. Sixth Annual Meeting of the American Society of Gene Therapy; June 4–8; Washington, DC, 2003.
- 203 Alonso MM, Gomez-Manzano C, Bekele BN, Yung WK, Fueyo J. Adenovirus-based strategies overcome temozolomide resistance by silencing the O6-methylguanine-DNA methyltransferase promoter. *Cancer Res* 2007; 67: 11499–11504.
- 204 Pan Q, Liu B, Liu J, Cai R, Liu X, Qian C. Synergistic antitumor activity of XIAP-shRNA and TRAIL expressed by oncolytic adenoviruses in experimental HCC. *Acta Oncol* 2008; **47**: 135–144.
- 205 Pan Q, Liu B, Liu J, Cai R, Wang Y, Qian C. Synergistic induction of tumor cell death by combining cisplatin with an oncolytic adenovirus carrying TRAIL. *Mol Cell Biochem* 2007; **304**: 315–323.
- 206 Jordan MA, Toso RJ, Thrower D, Wilson L. Mechanism of mitotic block and inhibition of cell proliferation by taxol at low concentrations. *Proc Natl Acad Sci USA* 1993; 90: 9552–9556.
- 207 Kohn KW, Jackman J, O'Connor PM. Cell cycle control and cancer chemotherapy. *J Cell Biochem* 1994; **54**: 440–452.
- 208 Lamfers ML, Idema S, Bosscher L, Heukelom S, Moeniralm S, van der Meulen-Muileman IH *et al.* Differential effects of combined Ad5- delta 24RGD and radiation therapy in *in vitro* versus *in vivo* models of malignant glioma. *Clin Cancer Res* 2007; **13**: 7451–7458.