



Letter to the Editor

Opposing actions of STAT-1 and STAT-3 on the Bcl-2 and Bcl-x promoters

Dear Editor,

The Bcl-2 family of proteins contains at least 15 members which can have pro- or anti-apoptotic effects. All share at least one of four Bcl-2 homology domains (BH1 to BH4), and inclusion of different BH domains in individual Bcl-2 family members is generally responsible for pro- or anti-apoptotic activity (reviewed in¹). Pro- and anti-apoptotic members can form heterodimers, and their ratio may determine whether a death or survival signal is delivered.² There is strong evidence that the actions of Bcl-2 proteins are exerted at the level of the mitochondrial membrane (reviewed in³). Thus many Bcl-2

members are anchored in the mitochondrial outer membrane, and anti-apoptotic family members such as Bcl-2 and Bcl-x_L form ion channels which increase the rate of proton extrusion from mitochondria.

Some studies have begun to address the transcriptional regulation of anti-apoptotic Bcl-2 genes. In leukaemias, for example, a hybrid transcription factor formed by chromosomal rearrangement increases Bcl-2 expression,⁴ and overexpression of the CCAAT enhancer binding protein, GADD153, reduces Bcl-2 expression with an accompany-

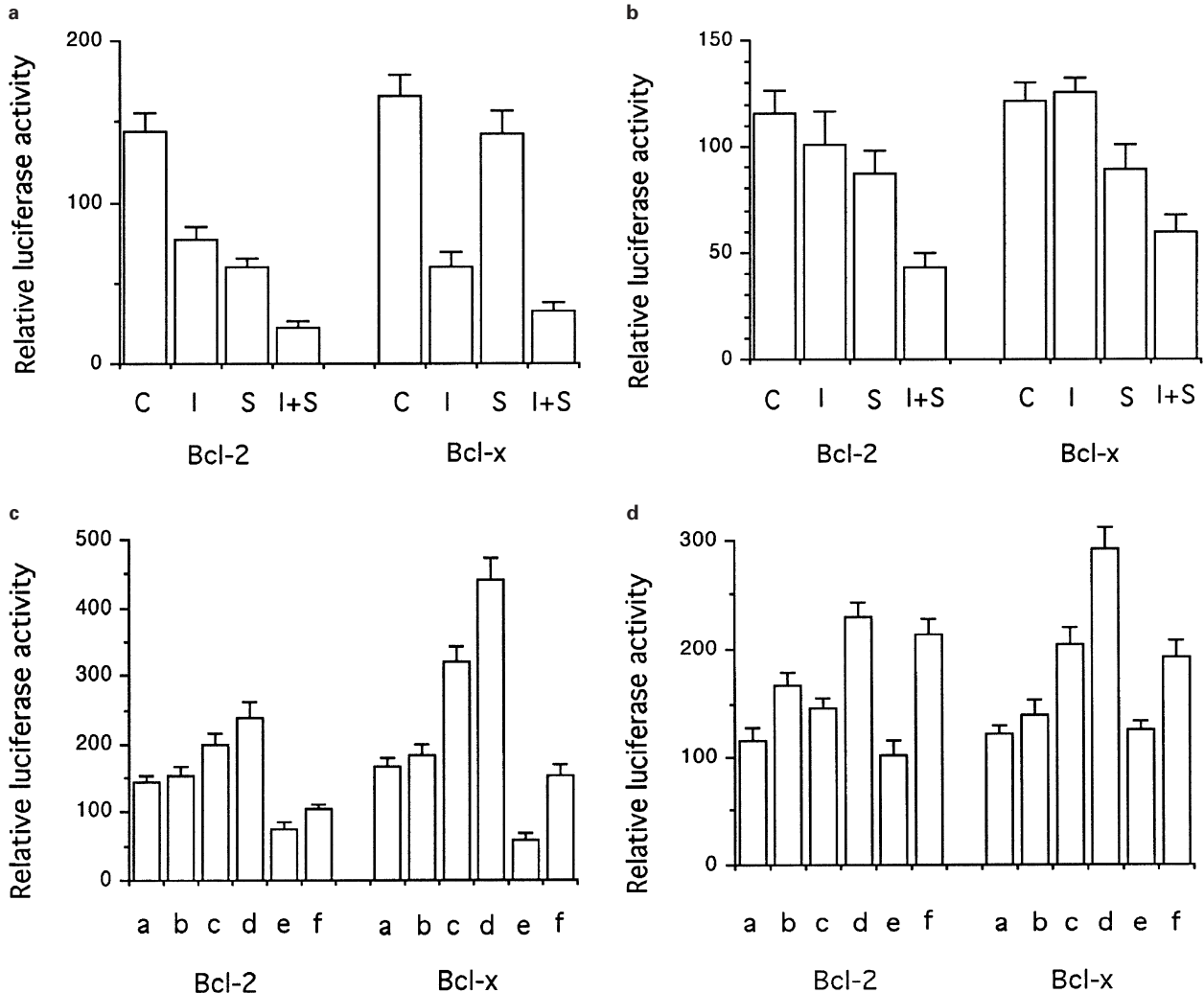


Figure 1 Effects of IFN γ (I), STAT-1 (S) and both IFN γ and STAT-1 (I+S) on control (C) basal activity of the Bcl-2 and Bcl-x promoters in STAT-1 expressing (a) and STAT-1 deficient (b) U3A cells. Effects of CT-1 and STAT-3 on basal activity of the Bcl-2 and Bcl-x promoters in STAT-1 expressing (c) and STAT-1 deficient (d) U3A cells. a: control basal activity; b: STAT-3; c: CT-1, d: CT-1+STAT-3; e: IFN γ ; f: IFN γ +CT-1

ing increase in apoptosis.⁵ In neuronal cells, transactivation of Bcl-2 by the POU factor, Brn-3a, is inhibited by p53, although p53 itself has little effect on basal Bcl-2 expression.⁶ NF- κ B-dependent enhancement of Bcl-x_L expression has been reported to mediate survival signals transduced through CD40 in B cells,⁷ and the transcriptional repressor, Btf, which induces apoptosis, is normally retained in the cytoplasm by Bcl-2 and Bcl-x_L.⁸

In the heart, there is also evidence that apoptosis following ischaemia/reperfusion (IR) injury is associated with modulation of Bcl-2 and Bcl-x_L expression. For example, in the isolated perfused rat heart, IR reduces Bcl-2 expression, in parallel with increased AP-1 and reduced NF- κ B.⁹ Moreover, the reduction in apoptosis in IR rabbit hearts produced by phenylephrine preconditioning is associated with an increased Bcl-x/Bax ratio.¹⁰ Since we have observed that activated Signal Transducer and Activator of Transcription (STAT)-1 is an important transactivator in the apoptosis induced by IR injury,¹¹ and others have shown that STAT-3 is involved in the cardioprotective actions of cardiotrophin-1 (CT-1),¹² we have studied whether these pro- and anti-apoptotic effects of STATs-1 and 3 are mediated by regulation of the expression of the Bcl-2 and Bcl-x_L genes.

In U3A-ST1 cells constitutively expressing STAT-1 (Figure 1a), IFN γ , a known activator of STAT-1, reduces basal expression of the Bcl-2 promoter. Transfection with STAT-1 by itself also reduces Bcl-2 promoter activity and overexpression of STAT-1 together with IFN γ treatment produces a reduction significantly greater than by either STAT-1 or IFN γ alone. This presumably reflects phosphorylation of transfected STAT-1 by Janus kinases activated by ligation of the IFN γ receptor. Figure 1a also shows that IFN γ reduces basal activity of the Bcl-x promoter; although the reduction in Bcl-x promoter activity produced by STAT-1 overexpression alone is less pronounced than with the Bcl-2 promoter, it is still significant, and IFN γ treatment of STAT-1 overexpressing cells again results in more profound reduction in Bcl-x promoter activity than IFN γ or STAT-1 alone. The reduction by IFN γ in Bcl-2 and Bcl-x promoter activity is not seen in the STAT-1 deficient cell line U3A, indicating that it is dependent on activation of STAT-1. Moreover, expressing STAT-1 alone or together with IFN γ treatment does reduce activity of both promoters, indicating that the effect of IFN γ can be restored by expressing STAT-1 in the STAT-1 deficient cells (Figure 1b).

Figure 1c shows that CT-1, a STAT-3 activator, can enhance Bcl-2 and Bcl-x activity. Moreover, CT-1 reverses the suppressive effect of IFN γ on the Bcl-2 and Bcl-x promoters (Figure 1c, bars e and f), most markedly in the STAT-1 deficient cell line (Figure 1d). In the STAT-1 deficient cells, both STAT-3 and CT-1 individually enhance Bcl-2 and Bcl-x promoter activity, and CT-1 also has this effect in STAT-1 expressing cells. The combined effects of STAT-3 and CT-1 are again greater than either factor alone.

The data suggest that one mechanism by which STAT-1 mediates IR-induced apoptosis is by reducing expression of anti-apoptotic Bcl-2 and Bcl-x genes. In addition, the cardioprotective effects of CT-1 may be mediated by STAT-3 induced increases in expression of these genes. STAT-1 and STAT-3 homodimers activate distinct sets of genes. In addition, STAT-1 and STAT-3 can also form heterodimers.¹³ The results reported here suggest that, in the heart, the relative proportions of STAT-1 and STAT-3 may affect the transcriptional activity of two anti-apoptotic genes. It is unclear whether STAT-1 and STAT-3 act at a single site to regulate the Bcl-2 and Bcl-x promoters, or whether interaction between separate binding regions is involved. Promoter mapping studies are in progress to resolve this question.

A Stephanou^{*1}, BK Brar¹, RA Knight² and DS Latchman¹

¹Molecular Medicine Unit, Institute of Child Health, 30 Guilford Street, London WC1N 1EH; ²Department of Cystic Fibrosis, Imperial College at the NHLI, Manresa Road, London SW3

*Corresponding author: Molecular Medicine Unit, Institute of Child Health, 30 Guilford Street, London WC1N 1EH. Tel: +44 171 504 9207; Fax: +44 171 373 3310; E-mail: anastasis_stephanou@yahoo.com

1. Adams JM and Cory S (1998) *Science* 281: 1322–1326
2. Oltvai ZN *et al.* (1993) *Cell* 74: 609–619
3. Green DR and Reed JC (1998) *Science* 281: 1309–1312
4. Klampfer L *et al.* (1996) *Proc. Natl. Acad. Sci. USA* 93: 14059–14064
5. Matsumoto M *et al.* (1996) *FEBS Lett.* 395: 143–147
6. Budhram-Mahadeo V *et al.* (1999) *J. Biol. Chem.* 274: 15237–15244
7. Lee HH *et al.* (1999) *Proc. Natl. Acad. Sci. USA* 96: 9136–9141
8. Kasof GM *et al.* (1999) *Mol. Cell. Biol.* 19: 4390–4404
9. Maulik N *et al.* (1999) *FEBS Lett.* 443: 331–336
10. Baghelai K *et al.* (1999) *J. Thor. Cardiovasc. Surg.* 117: 980–986
11. Stephanou A *et al.* (2000) *J. Biol. Chem.*, in press
12. Sheng Z *et al.* (1997) *J. Biol. Chem.* 272: 5783–5791
13. Ihle JN (1996) *Cell* 84: 331–334