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Single-nucleotide polymorphisms associated with outcome in metastatic renal cell carcinoma treated with sunitinib

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Background: There are no validated markers that predict response in metastatic renal cell cancer (RCC) patients treated with sunitinib. We aim to study the impact of single-nucleotide polymorphisms (SNPs) that have recently been proposed as predictors of outcome to anti-VEGF-targeted therapy in metastatic RCC in an independent cohort of patients.

Methods: We genotyped 16 key SNPs in 10 genes involved in sunitinib pharmacokinetics, pharmacodynamics and VEGF-independent angiogenesis in patients with metastatic clear-cell RCC treated with sunitinib as the first-line targeted therapy. Association between SNPs, progression-free survival (PFS) and overall survival (OS) were studied by multivariate Cox regression using relevant clinical factors associated with PFS and OS as covariates.

Results: In a series of 88 patients, both PFS and OS were associated significantly with SNP rs1128503 in *ABCB1* ($P=0.027$ and $P=0.025$), rs4073054 in *NR1/3* ($P=0.025$ and $P=0.035$) and rs307821 in *VEGFR3* ($P=0.032$ and $P=0.011$). Progression-free survival alone was associated with rs2981582 in *FGFR2* ($P=0.031$) and rs2276707 in *NR1/2* ($P=0.047$), whereas OS alone was associated with rs2307424 in *NR1/3* ($P=0.048$) and rs307826 in *VEGFR3* ($P=0.013$).

Conclusion: Our results confirm former communications regarding the association between SNPs in *ABCB1*, *NR1/2*, *NR1/3* and *VEGFR3* and sunitinib outcome in clear-cell RCC. Prospective validation of these SNPs is now required.

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Inactivation of the von Hippel–Lindau (*VHL*) tumour-suppressor gene is the most frequent molecular alteration in clear-cell renal cell cancer (RCC). Inactivated *VHL* leads to elevated protein levels of hypoxia-induced factor- α that upregulates vascular endothelial growth factor (VEGF) and platelet-derived growth factor (PDGF) expression. Targeted therapies directed against some of these proteins have significantly improved the perspectives of patients with metastatic RCC. Sunitinib malate is an orally administered tyrosine kinase receptor inhibitor (TKI) that targets VEGF and PDGF receptors, KIT, FLT-3, colony stimulating factor-1 receptor and RET. In a randomised controlled trial, sunitinib significantly prolonged progression-free survival (PFS; 11 vs 5 months, $P < 0.001$) as compared with interferon- α (Motzer *et al*, 2007, 2009). Median overall survival (OS) was 26.4 and 21.8 months, respectively ($P = 0.051$). Sunitinib is a standard treatment option in clear-cell RCC, but other anti-VEGFR and anti-PDGFR-targeted TKIs such as sorafenib, pazopanib and axitinib are also used in different stages of the disease.

Although 50% of RCC patients receiving sunitinib experience an objective response and 43% achieve disease stabilisation, 7% will experience progressive disease (PD) at first evaluation, probably because of intrinsic resistance or other factors (Motzer *et al*, 2009). Moreover, even patients with an initial clinical benefit will finally progress because of acquired resistance or for other reasons. The identification of biomarkers able to predict intrinsic resistance could avoid unnecessary costs and side effects, guiding alternative treatment decisions. On the other hand, the identification of biomarkers for acquired resistance could provide novel directions to develop therapies that block these resistance pathways. Although different mechanisms of resistance have been proposed (Rini and Atkins, 2009), reliable biomarkers predictive of sunitinib sensitivity or primary/secondary resistance are still lacking.

Several clinical and biochemical markers for PFS and OS are available for sunitinib-treated patients (Heng *et al*, 2009; Patil *et al*, 2011). For PFS, these are baseline serum lactate dehydrogenase (LDH) level, the presence of two or more metastatic sites, no prior nephrectomy, Eastern Cooperative Oncology Group Performance Status (ECOG PS) and baseline platelet count. For OS, factors include presence of bone metastases, time between nephrectomy and start of systemic therapy, baseline serum LDH level, baseline haemoglobin, baseline calcium and baseline ECOG. The last five criteria are part of the Memorial Sloan Kettering Cancer Centre (MSKCC) score that categorises patients into a favourable, intermediate- and poor-prognosis group (Motzer *et al*, 2004). These established clinical and biochemical markers are indicators of the general condition of the patient and the extension or stage of the disease. They do not take into account sunitinib pharmacokinetics (absorption, metabolism) or pharmacodynamics (interaction of sunitinib with its molecular targets). Recently, a meta-analysis of pharmacokinetic data from 443 patients treated with sunitinib showed that higher plasma levels of sunitinib and its active metabolite SU12662 were associated with prolonged TTP and OS (Houk *et al*, 2010). Factors influencing the concentration of sunitinib in plasma are dose and schedule of the drug and patient compliance, but importantly, also the concentration of efflux pumps and metabolising enzymes. Moreover, sunitinib efficacy can be influenced by the expression level and variants of the molecular targets of the drug.

Recently, a number of studies have proposed that genetic variability in genes involved in sunitinib pharmacokinetics and pharmacodynamics alter the efficacy of sunitinib (van der Veldt *et al*, 2010; Garcia-Donas *et al*, 2011) or pazopanib (Xu *et al*, 2011a,b) in metastatic RCC. As each of these studies investigated a different set of single-nucleotide polymorphisms (SNPs), these findings need to be validated independently. The aim of the present study is therefore to replicate association of these SNPs to sunitinib

outcome by assessing an independent cohort of patients with metastatic clear-cell RCC treated with first-line sunitinib.

MATERIALS AND METHODS

For this retrospective study, germline DNA samples were collected in the CIT-rein kidney tumour bank and in patients treated at the University Hospitals Leuven. The French-Belgian multicentric CIT-rein kidney tumour bank contains more than 250 frozen kidney tumour samples collected at 20 academic hospitals. We selected the samples of patients with pathologically confirmed clear-cell RCC treated in first line with sunitinib and for whom frozen normal kidney tissue was available. Eligible patients could have received cytokines as systemic treatment for kidney tumours before starting sunitinib as a monotherapy, but they could not have received any other TKI or mTOR (mammalian target of rapamycin) inhibitor before starting sunitinib. To make sure that the effect of sunitinib was accurately measured, patients had to take sunitinib during at least one complete cycle of 28 days and had to reach at least the first evaluation by CT scan. In the whole CIT-rein kidney tumour bank, 79 frozen normal kidney samples corresponded to these selection criteria. In order to extend the series, we added nine patients visiting the University Hospitals Leuven and complying to the same inclusion criteria. As no frozen normal kidney tissue was available for these patients, peripheral blood was sampled during out day clinic from July 2011 till December 2011.

The protocol was approved by the medical ethics review boards of all participating institutions, and signed consent was obtained from all patients. In some cases, we used frozen biologic material from patients who had already died and for whom a general positive advice for the utilisation of remaining tissue was foreseen by the institutional board.

All the patients were treated in routine clinical practice. Drug schedule, dose-reduction policy and timing of radiological assessments were left to the discretion of the attending doctors in accordance with current local practice guidelines. All the patients started their sunitinib therapy at the standard sunitinib dose of 50 mg day⁻¹, 4 weeks on and 2 weeks off. The patient characteristics considered relevant for PFS and OS analysis were the five risk factors according to the MSKCC prognostic criteria and additional factors such as baseline neutrophil count, baseline platelet count, the presence or absence of liver metastases, the presence or absence of a component of sarcomatoid dedifferentiation and the presence or absence of bone metastases. The latter two parameters were associated to outcome on sunitinib in recent publications (Golshayan *et al*, 2009; Beuselinck *et al*, 2011; Patil *et al*, 2011).

The SNPs previously associated with TKI efficacy in RCC were selected from the literature (Table 1). These SNPs are located in genes affecting sunitinib pharmacokinetics (i.e., genes involved in sunitinib absorption, such as *ABCB1*, or metabolism, such as *CYP3A5*, *NR1/2* and *NR1/3*), sunitinib pharmacodynamics (i.e., genes involved in PDGF- and VEGF-dependent angiogenesis such as *HIF1A*, *PDGFRA*, *VEGFR2* and *VEGFR3*) or VEGF-independent alternative pro-angiogenic pathways (*FGFR2*, and *IL8*). DNA was isolated at INSERM U674 in Paris, France, from fresh frozen normal kidney tissue sampled in the nephrectomy specimen using the Qiaquick extraction kit (Qiagen, Valencia, CA, USA) and quantified by fluorometry (Fluoroskan Thermo Labsystems, Cergy-Pontoise, France). DNA was isolated from peripheral blood at the Vesalius Research Center in Leuven with the Qiagen DNA kit (Qiagen) and final DNA concentration quantified with Nanodrop (Nanodrop, Wilmington, DE, USA). High-throughput SNP genotyping was performed at the Vesalius Research Center in Leuven, Belgium, using the Sequenom MassArray platform

Table 1. SNPs linked to sunitinib outcome based on literature evidence

Gene	Polymorphism	SNP ID	Impact on outcome	Reference
Genes involved in pharmacokinetics				
ABCB1	3435C>T	rs1045642	PFS: 15.2 vs 8.4 months if a TCG copy was present in the ABCB1 haplotype composed of rs1045642, rs1128503 and rs2032582 (<i>P</i> = 0.033)	(Van der Veldt <i>et al</i> , 2010)
	1236C>T 2677G>T or G>A	rs1128503 rs2032582		
CYP3A5	6986G>A	rs776746	PFS: not reached for AA and AG genotypes vs 9.3 months for GG genotypes (<i>P</i> = 0.032)	(Van der Veldt <i>et al</i> , 2010)
NR1/2	25385C>T	rs3814055	PFS: 6.7 months for TT genotypes vs 10.8 months for CT and CC genotypes (<i>P</i> = 0.025) OS: 10.2 months for TT genotypes vs 17.1 months for CT and CC genotypes (<i>P</i> = 0.017) OS: 29 vs 22 vs 23 months for the CC, CT and TT variants, respectively (<i>P</i> = 0.03)	(Van der Veldt <i>et al</i> , 2010) (Xu <i>et al</i> , 2011b)
	8055C>T	rs2276707	PFS: 10.8 months for CC and CT genotypes vs 6.7 months for TT genotypes (<i>P</i> = 0.025)	(Van der Veldt <i>et al</i> , 2010)
NR1/3	5719C>T	rs2307424	PFS: 13.3 vs 8.0 months if a CAT copy was absent in the NR1/3 haplotype composed of rs2307424, rs2307418 and r s4073054 (<i>P</i> = 0.017)	(Van der Veldt <i>et al</i> , 2010)
	7738A>C 7837T>G	rs2307418 rs4073054		
Genes involved in pharmacodynamics				
HIF1A	1790G>A	rs11549467	PFS: 44 months for GG genotypes vs 20 weeks for GA genotypes (<i>P</i> = 0.03)	(Xu <i>et al</i> , 2011a)
PDGFRA	1580T>C	rs35597368	OS: 24.2 vs 14.8 months if a GCGT haplotype is present in both alleles of a PDGFRA haplotype composed of rs1800810, rs1800812, rs1800813 and rs35597368 vs patients with GCG–other or other–other haplotypes (<i>P</i> = 0.002)	(Van der Veldt <i>et al</i> , 2010)
VEGFR2	1718T>A	rs1870377	OS: 16.3 months for AA and AT genotypes vs 9.4 months for TT genotypes (<i>P</i> = 0.016)	(Van der Veldt <i>et al</i> , 2010)
VEGFR3	3971G>T	rs307821	PFS: 13.7 months for GG genotypes vs 6.7 months for GT genotypes (<i>P</i> = 0.014)	(Garcia-Donas <i>et al</i> , 2011)
	1480A>G	rs307826	PFS: 13.7 months for AA genotypes vs 3.6 months for AG genotypes (<i>P</i> = 0.0079) OS: 26, 23 and 3.2 months for the AA, AG and GG genotypes, respectively (<i>P</i> = 0.04)	(Garcia-Donas <i>et al</i> , 2011) (Xu <i>et al</i> , 2011b)
Genes involved in alternative proangiogenic pathways				
FGFR2	906C>T	rs2981582	OS: 28.0 months for CC genotypes vs 21.4 months for TT genotypes (<i>P</i> = 0.009)	(Xu <i>et al</i> , 2011b)
IL8	251T>A	rs4073	PFS: 49, 42 and 32 weeks for TT, AT and AA genotypes, respectively (<i>P</i> = 0.01)	(Xu <i>et al</i> , 2011a)
	2767A>T	Rs1126647	PFS: 48, 42 and 27 weeks for AA, AT and TT genotypes, respectively (<i>P</i> = 0.009)	(Xu <i>et al</i> , 2011a)
Abbreviations: SNP = single-nucleotide polymorphism; PFS = progression-free survival; OS = overall survival.				

(Sequenom, San Diego, CA, USA) (Reumers *et al*, 2011). Genotyping analysis was performed by investigators blinded for the clinical data. Overall, 16 SNPs were successfully genotyped, with success rates $\geq 85\%$ for each SNP and an overall average success rate of 96%. We failed to genotype SNP rs1126647 in IL-8 because of technical reasons. For most of the SNPs, genotypes were analysed in the same way as they were communicated in the original reports (i.e., according to dominant, recessive or co-dominant genetic models or in the context of a specific haplotype).

Clinical data were collected at 15 different sites in France and Belgium. The primary objective was PFS and OS, and the secondary objective was RR. We defined PFS as the time between the first day on sunitinib and the date of radiological PD or death. Patients who had not progressed at database closure were censored at last follow-up. Overall survival was defined as the time between the first day on sunitinib and the date of death or last date of

follow-up. Objective response was assessed by the treating doctors and classified as complete response (CR), partial response (PR), stable disease (SD), or PD. Timing for assessments was dictated by individual institution policy.

All patient characteristics were tested in an univariate analysis for association with PFS and OS using Kaplan–Meier statistics and in a multivariate model using Cox proportional hazards. Fisher's exact tests and logistic regression were used to compare the incidence of poor-prognostic variants in patients with PD vs a group with SD, PR or CR as best response. The MSKCC score was used as a covariate in the multivariate analysis, as well as all other variables with a $P \leq 0.2$ on univariate analysis that are not part of the MSKCC score. Results with a P -value of <0.05 were considered as significant in the multivariate analysis. Because this is a confirmatory rather than an exploratory study, SNPs were selected based on literature evidence and, hence, no correction for multiple testing was made.

Table 2. Patient characteristics at diagnosis and at the start of sunitinib treatment and baseline clinical and biochemical parameters associated with PFS and OS

	Total	Median PFS (months)	P-value, HR (95% CI)	95% CI of median PFS
At initial diagnosis				
Male	68% (60/88)	—	—	—
Ethnic origin				
Caucasian	94% (83/88)	—	—	—
Unknown	6% (5/88)	—	—	—
M1 (synchronous metastases)	55% (46/84)	—	—	—
Fuhrman				
Grade 1–3	68% (58/85)	—	—	—
Grade 4	32% (27/85)	—	—	—
Sarcomatoid dedifferentiation				
Present	9% (8/88)	4	0.09	1–Not reached
Absent	91% (80/88)	18	0.37 (0.12–1.18)	12–24
At the start of sunitinib				
ECOG PS				
> 0	49% (43/88)	15	0.08	7–20
0	51% (45/88)	21	0.63 (0.37–1.06)	11–38
Neutrophils				
> 4500 per mm ³	40% (34/85)	9.5	0.13	5–15
< 4500 per mm ³	60% (51/85)	19	0.65 (0.38–1.14)	14–24
Platelets				
> 400,000 per mm ³	15% (13/88)	11	0.25	—
< 400,000 per mm ³	85% (75/88)	18	—	—
Haemoglobin				
Low (< 11.5 g dl ⁻¹ (women) or < 13 g dl ⁻¹ (men))	42% (37/88)	14	0.98	—
Normal	58% (51/88)	18	—	—
LDH				
> 1.5 ULN	10% (8/84)	10.5	0.09	4–19
≤ 1.5 ULN	90% (76/84)	18.0	0.40 (0.14–1.16)	12–25
Corrected calcium				
> 10 mg dl ⁻¹	7% (6/84)	22	0.9	—
≤ 10 mg dl ⁻¹	93% (78/84)	15	—	—
Time from nephrectomy to systemic treatment				
< 12 months	66% (58/88)	18	0.30	—
> 12 months	34% (30/88)	15	—	—
Immunotherapy before sunitinib	28% (24/87)	—	—	—
Site of metastasis				
Lung	84% (74/88)	—	—	—
Liver metastases	18% (16/88)	15	0.59	—
No liver metastases	82% (72/88)	18	—	—
Bone metastases	35% (31/88)	15	0.5	—
No bone metastases	65% (57/88)	18	—	—
Brain	6% (5/88)	—	—	—
MSKCC prognosis				
Favourable	15% (13/85)	Not reached	0.21	8–Not reached
Intermediate	56% (48/85)	15	—	11–21
Poor	28% (24/85)	15	—	4–25

Table 2. Continued

At initial diagnosis	Total	Median OS (months)	P-value, HR (95% CI)	95% CI of median OS
Male	68% (60/88)	—	—	—
Ethnic origin				
Caucasian	94% (83/88)	—	—	—
Unknown	6% (5/88)	—	—	—
M1 (synchronous metastases)	55% (46/84)	—	—	—
Fuhrman				
Grade 1–3	68% (58/85)	—	—	—
Grade 4	32% (27/85)	—	—	—
Sarcomatoid dedifferentiation				
Present	9% (8/88)	16.5	0.19	5–Not reached
Absent	91% (80/88)	30	0.45 (0.13–1.50)	23–42
At the start of sunitinib				
ECOG PS				
> 0	49% (43/88)	23	0.08	17–34
0	51% (45/88)	35	0.60 (0.34–1.06)	23–Not reached
Neutrophils				
> 4.500 per mm ³	40% (34/85)	22	0.39	—
< 4.500 per mm ³	60% (51/85)	34	—	—
Platelets				
> 400.000 per mm ³	15% (13/88)	27	0.45	—
< 400.000 per mm ³	85% (75/88)	29	—	—
Haemoglobin				
Low (< 11.5 g dl ⁻¹ (women) or < 13 g dl ⁻¹ (men))	42% (37/88)	27	0.42	—
Normal	58% (51/88)	34	—	—
LDH				
> 1.5 ULN	10% (8/84)	24.5	0.19	19–34
≤ 1.5 ULN	90% (76/84)	34	0.51 (0.19–1.38)	23–45
Corrected calcium				
> 10 mg dl ⁻¹	7% (6/84)	42	0.98	—
≤ 10 mg dl ⁻¹	93% (78/84)	29	—	—
Time from nephrectomy to systemic treatment				
< 12 months	66% (58/88)	27	0.13	22–35
> 12 months	34% (30/88)	Not reached	1.58 (0.88–2.85)	19–Not reached
Immunotherapy before sunitinib	28% (24/87)	—	—	—
Site of metastasis				
Lung	84% (74/88)	—	—	—
Liver metastases	18% (16/88)	22	0.60	—
No liver metastases	82% (72/88)	29	—	—
Bone metastases	35% (31/88)	22	0.06	11–34
No bone metastases	65% (57/88)	35	0.54 (0.29–1.03)	24–Not reached
Brain	6% (5/88)	—	—	—
MSKCC prognosis				
Favourable	15% (13/85)	Not reached	0.0097	Not reached–not reached
Intermediate	56% (48/85)	24	—	20–41
Poor	28% (24/85)	27	—	19–42

Abbreviations: PFS = progression-free survival; OS = overall survival; ECOG PS = Eastern Cooperative Oncology Group Performance Status; LDH = lactate dehydrogenase; ULN = upper limit of normal; MSKCC = Memorial Sloan Kettering Cancer Center; HR = hazard ratio; 95% CI = 95% confidence interval. In the univariate analysis, median PFS and median OS were estimated by Kaplan–Meier and P-values were derived from a log-rank test. The impact of the presence of lung metastases and previous immunotherapy was not assessed as these parameters have not been strongly linked to PFS or OS.

Of the 11 clinical parameters assessed in the univariate analysis (for 88 patients), there were 13 missing values (1.3%). For the multivariate analysis, 82 patients with complete data could be included. Statistical analyses were conducted using GraphPad Prism 5 (GraphPad Software, La Jolla, CA, USA) and XLSTAT software (Addinsoft, Paris, France).

RESULTS

We enrolled 88 patients who started sunitinib between November 2005 and July 2011 and closed the follow-up database in April 2012. Table 2 shows the clinical characteristics of enrolled patients. Mean age at diagnosis was 59 years (range 38–84). The majority of patients (>94%) were of Caucasian origin. According to the MSKCC prognostic criteria, 15% of patients were categorised into the favourable risk group, and 56% had intermediate and 28% poor risk.

At the time of analysis, 57 (64.8%) patients had reached progression and 48 (54.5%) had died. The median follow-up was 46.0 months (range 1.0–73.0 months; 95% confidence interval (CI) 42.0–51.0 months) after the start of sunitinib. The median PFS was 15.0 months (95% CI 11.0–23.0 months) and the median OS was 29.0 months (95% CI 23.0–42.0 months). Best response assessment was available in 82 patients (in the 6 remaining patients, there was a clinical benefit, but response assessment was poorly defined in the medical records, and as a consequence, it was unclear whether the best response was either PR or SD in these 6 patients). In all, 6 out of 82 (7.3%) patients had a CR, 30 out of 82 (36.6%) patients a PR, 36 out of 82 (43.9%) SD and 10 out of 88 (11.4%) PD as best response.

For each of these 16 polymorphisms, the respective genotypes, allele frequencies and changes at the amino acid level are given in Table 3. The allele frequencies of the genotyped polymorphisms were similar as previously reported in the dbSNP database (dbSNP

build 136) or 1000 Genomes Project, except for SNPs rs2276707 and rs307821. Their observed minor allele frequencies were slightly higher compared with their frequency reported in dbSNP. In the case of rs11549467, there was only one heterozygous patient. As a consequence, the impact of this SNP could not be analysed.

Next, we assessed the clinical and biochemical parameters associated with PFS and OS (Table 2). The MSKCC score, baseline neutrophil levels and the presence of a sarcomatoid component in the tumour were considered as covariates when assessing the effect of SNPs on PFS. For OS, the MSKCC score, the presence of bone metastases and the presence of a sarcomatoid component in the tumour were considered as covariates. Table 4 and Figures 1–7 show the results of the univariate and multivariable analyses for each of the genotyped SNPs for both PFS and OS after correction for these covariates. In the multivariate analysis, PFS and OS were associated significantly with SNP rs1128503 in *ABCB1* ($P=0.027$ and $P=0.025$), rs4073054 in *NR1/3* ($P=0.025$ and $P=0.035$) and rs307821 in *VEGFR3* ($P=0.032$ and $P=0.011$). Progression-free survival was associated with rs2981582 in *FGFR2* ($P=0.031$) and rs2276707 in *NR1/2* ($P=0.047$). Overall survival was associated with rs2307424 in *NR1/3* ($P=0.048$) and rs307826 in *VEGFR3* ($P=0.013$).

Finally, we also assessed the distribution of various unfavourable SNP genotypes in patients exhibiting a PD vs SD, PR or CR as their best response. On logistic regression, taking into account the MSKCC score, the presence of sarcomatoid dedifferentiation and baseline neutrophil count, the unfavourable genotypes GA/GG in *VEGFR3* rs307826 were significantly more frequent in patients experiencing PD as best response when compared with patients experiencing SD, PR or CR as best response (Table 5).

We could not confirm associations between SNP rs776746 in *CYP3A5*, rs3814055 in *NR1/2*, rs11549467 in *HIFA*, rs1870377 in *VEGFR2* and rs4073 in *IL8* and outcome.

Table 3. Genotype and allele distribution of selected SNPs

Gene	RS ID	Polymorphism	Location or functional consequence	n	Wild-type/wild-type, n (%)	Wild-type/variant, n (%)	Variant/variant, n (%)	Observed minor allele frequency (%)	Minor allele frequency in dbSNP (%)
Sunitinib pharmacokinetics									
<i>ABCB1</i>	rs1045642	3435C>T	I1154I	87	25 (29)	43 (49)	19 (22)	46.6	53.4
	rs1128503	1236C>T	G412G	88	38 (43)	35 (40)	15 (17)	36.9	45.1
	rs2032582	2677G>T or G>A	A893S	80	32 (40)	36 (45)	12 (15)	37.5	41.7
<i>CYP3A5</i>	rs776746	6986G>A	Affecting splicing	75	69 (92)	6 (8)	0 (0)	4.0	3.6
<i>NR1/2</i>	rs3814055	25385C>T	UTR-5'	82	32 (39)	35 (43)	15 (18)	39.6	33.6
	rs2276707	8055C>T	Intron	83	57 (69)	21 (25)	5 (6)	18.7	9.3
<i>NR1/3</i>	rs2307424	5719C>T	P151P	88	45 (51)	32 (36)	11 (12.5)	30.7	33.6
	rs2307418	7738A>C	Intron	86	61 (71)	22 (26)	3 (3)	16.3	15.9
	rs4073054	7837T>G	Intron	87	40 (46)	35 (40)	12 (14)	33.9	40.7
Sunitinib pharmacodynamics									
<i>HIF1A</i>	rs11549467	1790G>A	A588T	84	83 (99)	1 (1)	0 (0)	0.6	1.7
<i>PDGFRA</i>	rs35597368	1580T>C	S478P	88	69 (78)	18 (20)	1 (1)	11.3	13.3
<i>VEGFR2</i>	rs1870377	1718T>A	Q472H	81	46 (57)	28 (35)	7 (9)	25.9	27.5
<i>VEGFR3</i>	rs307821	3971G>T	R1324L	88	64 (73)	23 (26)	1 (1)	14.2	5.8
	rs307826	1480A>G	T494A	88	65 (74)	22 (25)	1 (1)	13.6	10.2
Alternative VEGF-independent proangiogenic pathways									
<i>FGFR2</i>	rs2981582	906C>T	Intron	87	23 (26)	52 (60)	12 (14%)	43.6	45.6
<i>IL8</i>	rs4073	251T>A	5' near gene	79	25 (31)	42 (53)	12 (15%)	41.8	42.5
Abbreviations: SNP = single-nucleotide polymorphism; VEGF = vascular endothelial growth factor; UTR = untranslated region; dbSNP = SNP database.									

Table 4. Univariate and multivariate analyses: association between SNPs and outcome

Gene (a) SNP ID	Polymorphism	No. of pts	Median PFS (months)	P-value (UV)	P-value (MV)	HR	95% CI of HR	95% CI of median PFS (months)
ABCB1 rs1045642 3435C>T	CC	25	14	0.67	NA	NA	NA	NA
	CT	43	15					NA
	TT	19	18					NA
ABCB1 rs1128503 1236C>T	CT+CC	73	19	0.031	0.027	0.464	0.234–0.918	11–25
	TT	15	8					3–21
ABCB1 rs2032582 2677G>T or G>A	GG	32	14	0.45	NA	NA	NA	NA
	GT/GA	36	19					NA
	TT/TA	12	15					NA
ABCB1 TCG copy	Present	16	15	0.68	NA	NA	NA	NA
	Absent	64	19					NA
CYP3A5 rs776746 6986G>A	GG	69	18	0.36	NA	NA	NA	NA
	AG	6	21					NA
NR1/2 s3814055 25385C>T	CC+CT	67	18	0.26	NA	NA	NA	NA
	TT	15	19					NA
NR1/2 rs2276707 8055C>T	CC+CT	78	18	0.0078	0.047	2.978	1.012–8.761	12–25
	TT	5	7					3–19
NR1/3 rs2307424 5719C>T	CC	45	20	0.18	0.155	1.513	0.856–2.675	11–38
	CT+TT	43	15					9–21
NR1/3 rs2307418 7738A>C	AA	61	14	0.45	NA	NA	NA	NA
	AC+CC	27	28					NA
NR1/3 rs4073054 7837T>G	TT	40	12	0.04	0.025	1.864	1.082–3.210	8–19
	TG+GG	47	21					12–38
NR1/3 CAT copy	Present	51	15	0.67	NA	NA	NA	NA
	Absent	36	15					NA
FGFR2 rs2981582 906C>T	TT	12	7.5	0.012	0.031	2.669	1.094–6.511	5–11
	CC	23	14					11–Not reached
IL8 rs4073 251T>A	TT	25	8	0.22	NA	NA	NA	NA
	AA	12	21					NA
PDGFRA rs35597368 1580T>C	TT	69	19	0.088	0.188	1.528	0.813–2.870	11–25
	TC+CC	19	14					8–19
VEGFR2 rs1870377 1718T>A	TT	48	15	0.76	NA	NA	NA	NA
	TA+AA	40	19					NA
VEGFR3 (b) rs3078213971G>T	GT+TT	24	10	0.077	0.032	1.981	1.060–3.702	7–21
	GG	64	18					12–26
VEGFR3 rs307826 1480A>G	AG+GG	23	10	0.022	0.051	1.800	0.996–3.250	6–19
	AA	65	19					14–26
Gene (a) SNP ID	Polymorphism	No. of pts	Median OS (months)	P-value (UV)	P-value (MV)	HR	95% CI of HR	95% CI of median OS (months)
ABCB1 rs1045642 3435C>T	CC	25	45	0.37	NA	NA	NA	NA
	CT	43	27					NA
	TT	19	24					NA
ABCB1 rs1128503 1236C>T	CT+CC	73	34	0.055	0.025	0.415	0.193–0.894	23–45
	TT	15	21					9–30

Table 4. Continued

Gene (a) SNP ID	Polymorphism	No. of pts	Median PFS (months)	P-value (UV)	P-value (MV)	HR	95% CI of HR	95% CI of median PFS (months)
<i>ABCB1</i> rs2032582 2677 G>T or G>A	GG	32	35	0.49	NA	NA	NA	NA
	GT/GA	36	34					NA
	TT/TA	12	24					NA
<i>ABCB1</i> TCG copy	Present	16	26	0.74	NA	NA	NA	NA
	Absent	64	34					NA
<i>CYP3A5</i> rs776746 6986G>A	GG	69	30	0.92	NA	NA	NA	NA
	AG	6	NR					NA
<i>NR1/2</i> s3814055 25385C>T	CC + CT	67	30	0.46	NA	NA	NA	NA
	TT	15	31					NA
<i>NR1/2</i> rs2276707 8055C>T	CC + CT	78	31	0.092	0.080	2.828	0.884–9.044	24–45
	TT	5	12					5–Not reached
<i>NR1/3</i> rs2307424 5719C>T	CC	45	42	0.057	0.048	1.913	1.006–3.636	25–Not reached
	CT + TT	43	23					16–34
<i>NR1/3</i> rs2307418 7738A>C	AA	61	30	0.86	NA	NA	NA	NA
	AC + CC	27	27					NA
<i>NR1/3</i> rs4073054 7837T>G	TT	40	22	0.03	0.035	1.927	1.046–3.549	14–34
	TG + GG	47	35					28–Not reached
<i>NR1/3</i> CAT copy	Present	51	28	0.58	NA	NA	NA	NA
	Absent	36	30					NA
<i>FGFR2</i> rs2981582 906C>T	TT	12	23	0.97	NA	NA	NA	NA
	CC	23	25					NA
<i>IL8</i> rs4073 251T>A	TT	25	23	0.68	NA	NA	NA	NA
	AA	12	31					NA
<i>PDGFRA</i> rs35597368 1580T>C	TT	69	35	0.025	0.302	1.440	0.721–2.875	24–Not reached
	TC + CC	19	23					14–31
<i>VEGFR2</i> rs1870377 1718T>A	TT	48	24	0.63	NA	NA	NA	NA
	TA + AA	40	30					NA
<i>VEGFR3</i> (b) rs307821 3971G>T	GT + TT	24	34	0.056	0.011	2.265	1.202–4.268	11–42
	GG	64	29					23–Not reached
<i>VEGFR3</i> rs307826 1480A>G	AG + GG	23	22	0.0058	0.013	2.223	1.187–4.163	11–34
	AA	65	31					24–Not reached

Abbreviations: SNP = single-nucleotide polymorphism; pts = patients; PFS = progression free survival; OS = overall survival; UV = univariate analysis; MV = multivariate analysis; NA = not applicable; HR = hazard ratio; 95% CI = 95% confidence interval. In the univariate analysis, *P*-values were calculated by a log-rank test. In the multivariate analysis, *P*-values were calculated by Cox proportional hazards. Whenever possible, variants were combined as it was done in the original publications: this was the case for *FGFR2*, *IL8*, *NR1/2*, *VEGFR2* and *VEGFR3*. For *ABCB1* and *NR1/3*, the original publication only reported haplotypes. The haplotypes were tested against PFS and OS in our series and no association with PFS and OS could be shown. Therefore, we checked for each SNP the three subgroups and analysed the PFS and OS curves. For *ABCB1*, when analysing TT vs TC vs CC in rs1045642 or GG vs GT/GA vs TT/TA in rs2032582, the three curves were overlapping for PFS and OS. Only in *ABCB1* rs1128503, when analysing TT vs TC vs CC variants, the CC and CT results were overlapping for PFS and OS and clearly different from the TT results, allowing us to group the results of the CT and CC variants. Concerning *NR1/3*, for SNP rs2307424, the PFS and OS curves of the CT and TT variants were overlapping, but the curves of the CC variant were clearly distinct. For SNP rs2307418, there were only two CC variant patients: they were grouped with the AC variant patients and tested against the AA variant patients. For SNP rs4073054, the PFS and OS curves of the TG and GG variants were overlapping, but the curves of the TT variant were clearly distinct. This distribution allowed us to test the impact of the CC variant in rs2307424, the AA variant in rs2307418 and the TT variant in rs4073054 vs the combination of the other variants. In case of *CYP3A5*, there were no AA variants in our series. For *PDGFRA*, there was only one CC variant: this patient was grouped with the TC variant. The HR for survival for patients with the GT or TT variants in rs307821 in *VEGFR3* vs patients with the GG variant was 2.265, favouring longer survival in patients with the GG genotype. Nevertheless, because of a crossing of the curves, the median OS was longer in the GT and TT variants (see curves).

DISCUSSION

In this retrospective study, we aim to observe the impact of SNPs that have recently been proposed as predictors of outcome to antiangiogenic therapy in metastatic RCC in an independent

cohort of patients. We observed significant associations between SNPs in genes involved in sunitinib pharmacokinetics (*ABCB1*, *NR1/2* and *NR1/3*), sunitinib pharmacodynamics (*VEGFR3*) and VEGF-independent pro-angiogenic pathways (*FGFR2*) and the therapeutic outcome of sunitinib in metastatic clear-cell RCC patients. For each of these associated SNPs, we observed similar

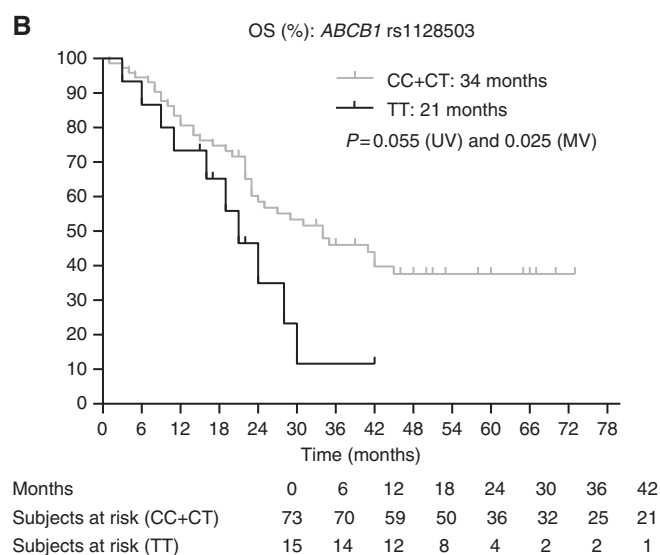
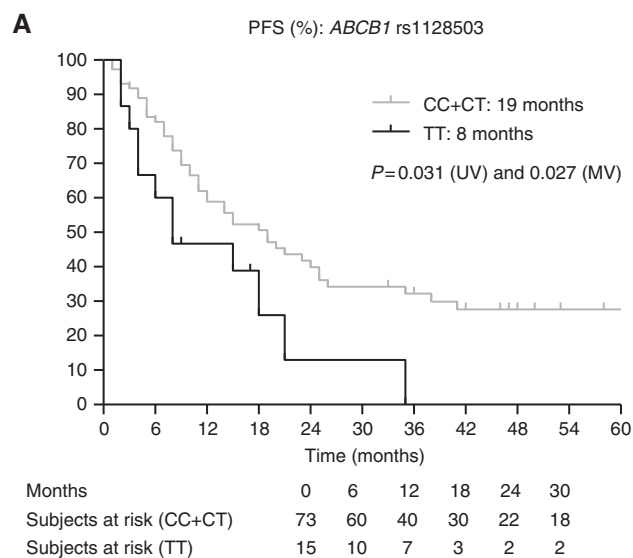


Figure 1. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs1128503 in *ABCB1*. The P -values are indicated for the univariate (UV) and multivariate (MV) analyses.

hazard ratios as reported previously, thereby adding more evidence that these SNPs could be markers associated with outcome on sunitinib. Moreover, for most of these observations, a rationale is available.

As sunitinib was used as a monotherapy and as a first-line treatment, our results were not confounded by concomitant or previous therapies and we could detect significant associations in a series involving only a limited number of patients.

The efflux transporter *ABCB1* (ATP binding cassette member B1, formerly known as P-glycoprotein or MDR1) is expressed in the intestine and liver and involved in the oral absorption and biliary secretion of several anticancer drugs (Dietrich *et al*, 2003). The ABC transporters may also contribute to multidrug resistance in tumours by actively extruding drugs from cancer cells, particularly in RCC cells (Soto-Vega *et al*, 2009; Walsh *et al*, 2009). As a consequence, expression levels and functionality of these drug transporters, for instance due to polymorphisms, may have important consequences for the efficacy of sunitinib. The most common functional SNPs in *ABCB1* are the synonymous 3435C>T (rs1045642) and 1236C>T (rs1128503) changes and the nonsynonymous 2677G>T change (missense A893S/T

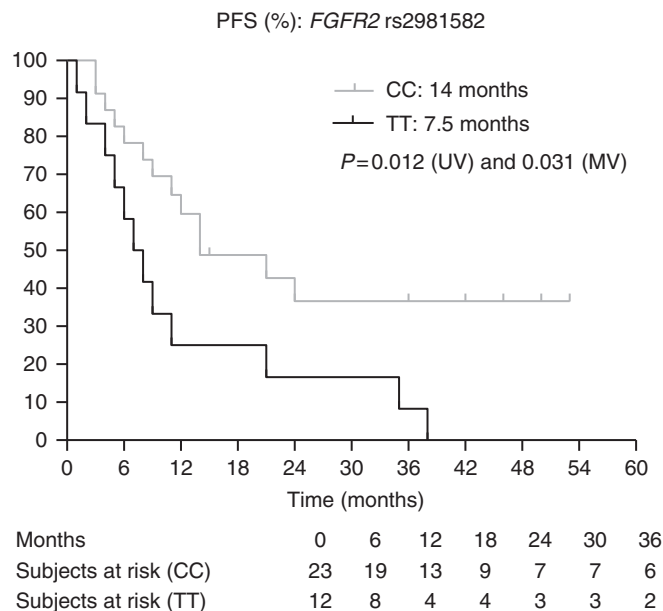


Figure 2. Kaplan–Meier curves for PFS for SNP rs2981582 in *FGFR2*. The P -values are indicated for the univariate (UV) and multivariate (MV) analyses.

rs2032582). Functional studies have shown that the haplotype of these three SNPs (rs1046542, rs1128503 and rs2032582) is a silent mutation and alters the function of the efflux transporter including its substrate specificity. We observed a significant association between the TT variant in rs1128503 1236C>T and shorter PFS and OS. In 89 RCC patients treated with sunitinib, Garcia-Donas *et al* (2011) observed an association, although not significant, between rs1128503 and PFS (HR 1.42, $P=0.089$) and OS (HR 1.75, $P=0.055$), favoring the patients with a C-allele. In 129 RCC patients treated with sunitinib, van der Veldt *et al* (2010) observed that the presence of a TCG haplotype (rs1045642, rs1128503 and rs2032582) in *ABCB1* (and thus the presence of the C-variant in rs1128503) was associated with prolonged PFS ($P=0.033$) and a tendency for prolonged OS ($P=0.078$). In 241 patients treated with pazopanib, the wild-type CC variant of rs1128503 was associated with improved OS compared with the wild-type TT genotypes (28 vs 20 months, $P=0.009$) (Xu *et al*, 2011b).

Fibroblast growth factor receptor 2 (*FGFR2*) is a VEGF-independent pro-angiogenic factor. The TT polymorphism in rs2981582 906C>T leads to increased transcription and expression of *FGFR2* (Meyer *et al*, 2008) and thus possibly to increased VEGF-independent angiogenesis. We observed a significant association between the TT variant in rs2981582 and shorter PFS. Data on the impact of rs2981582 in *FGFR2* on outcome on TKIs are only available in patients treated with pazopanib. In a series of 380 RCC patients, the TT variant was associated with inferior PFS compared with the CC genotype ($P=0.053$) (Xu *et al*, 2011a) and in a group of 241 patients, the TT genotype was associated with inferior OS compared with the CC genotype (median OS 21.4 vs 28.0 months, $P=0.02$) (Xu *et al*, 2011b).

The expression of cytochrome P450 CYP3A4, thought to be the key enzyme for the hepatic biotransformation of sunitinib, is regulated by the ligand-activated nuclear receptors *NR1I2* (pregnane X receptor) and *NR1I3* (constitutive androstane receptor). We observed a significant association between the TT genotype in rs2276707 8055C>T in *NR1I2* and a shorter PFS and OS. van der Veldt *et al* (2010) also found a significant difference in PFS between patients with the CC/CT genotype and patients with the TT genotype, 10.8 vs 6.7 months ($P=0.025$), but they could not confirm these results on multivariate analysis. Concerning

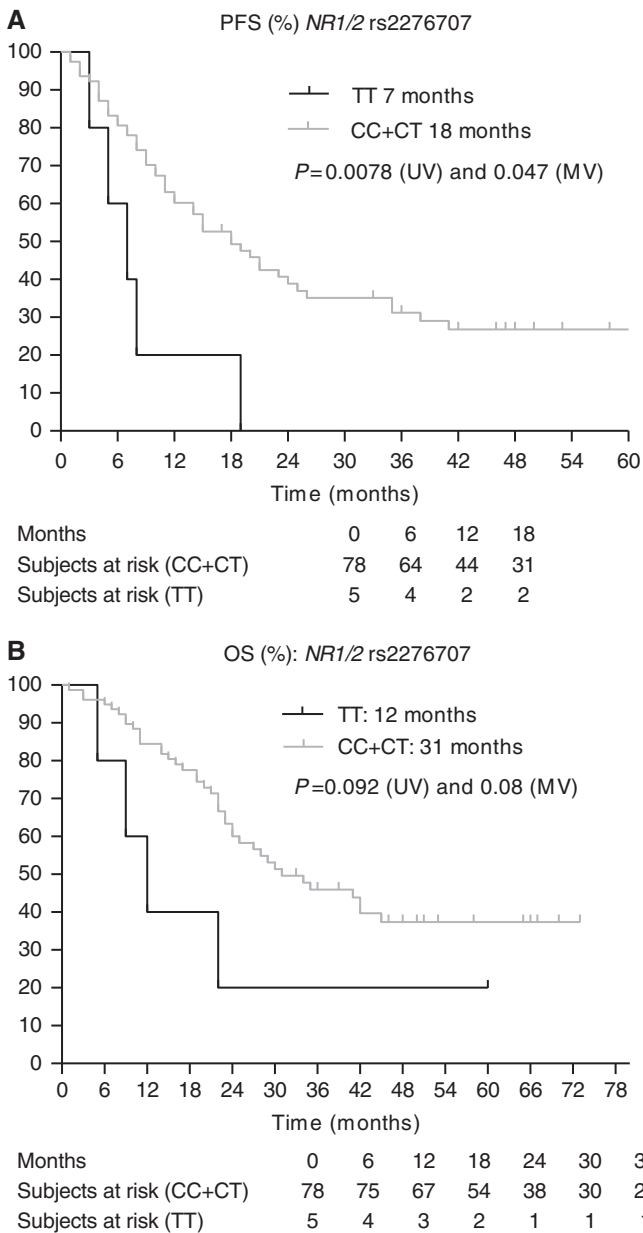


Figure 3. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs2276707 in *NR1/2*. The *P*-values are indicated for the univariate (UV) and multivariate (MV) analyses.

NR1/3, we observed a significant association between the TT variant in SNP rs4073054 and shorter PFS and OS. Prolonged PFS (13.3 vs 8.0 months, $P=0.017$) was found in 136 patients with absence of a CAT copy in the *NR1/3* haplotype (rs2307424, rs2307418 and rs4073054; $P=0.021$) (van der Veldt *et al*, 2010). This corresponds with our results, as rs4073054 concerns the T in the CAT haplotype. We also observed a significant association between the CC genotype in rs 2307424 in *NR1/3* and better OS, but there is no external validation at this moment for these results and we could not link this finding to the observations of van der Veldt *et al* (2010).

Platelet-derived growth factor receptor- α is one of the molecular targets of sunitinib. On univariate analysis, we observed a significant association between the TT variant in rs35597368 1580T/C in *PDGFRA* and longer PFS and OS. The T in rs35597368 corresponds to the T in the GCGT haplotype composed of four SNPs in the gene (rs1800810, rs1800812, rs1800813 and

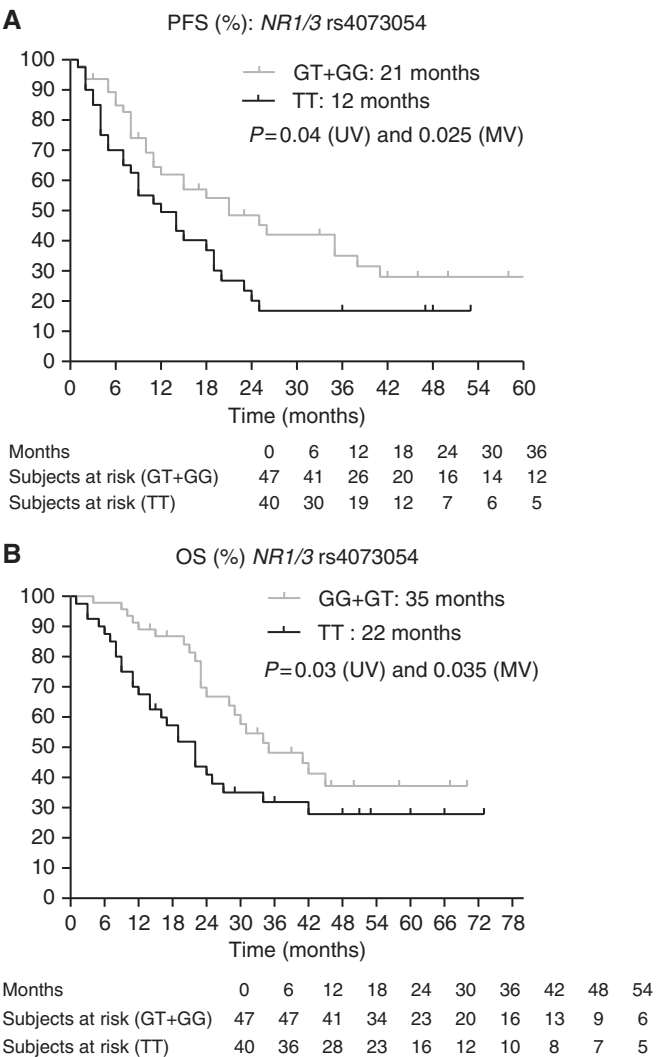


Figure 4. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs4073054 in *NR1/3*. The *P*-values are indicated for the univariate (UV) and multivariate (MV) analyses.

rs35597368). van der Veldt *et al* (2010) observed on univariate analysis a better OS in patients with a GCGT haplotype in both alleles (GCGT–GCGT), and thus in patients with a TT variant of the SNP, whereas patients with a GCG–other or other–other haplotype had a poorer median OS: 24.2 vs 14.8 months ($P=0.002$ on univariate analysis but 0.108 on multivariate analysis). We could not confirm the association with PFS and OS on multivariate analysis. The functional impact of this SNP is presently unknown.

The *VEGFR3* signalling is involved in embryonic angiogenesis, adult lymphangiogenesis and tumoural angiogenesis (Partanen *et al*, 1999; Valtola *et al*, 1999) and is one of the main targets of sunitinib. We observed a significant association between the GT or TT variant in rs307821 3971G>T in *VEGFR3* and shorter PFS and OS. Note that because of a crossing of the curves, the median OS was longer in the GT and TT variants than in the GG variants of rs307821. Nevertheless, the HR for survival for patients with the GT or TT variants in rs307821 in *VEGFR3* vs patients with the GG variant was 2.265 (95% CI 1.202–4.238). The crossing of the curves is probably because of the limited number of patients in our series. We also observed a significant association between the AG or GG variant in rs307826 1480A>G and shorter OS. In a series of 89 RCC patients treated with sunitinib, TTP for the GT variant of rs307821 was 6.7 months vs 13.7 months for patients with the GG genotype ($P=0.00085$) and TTP for the GA variant of

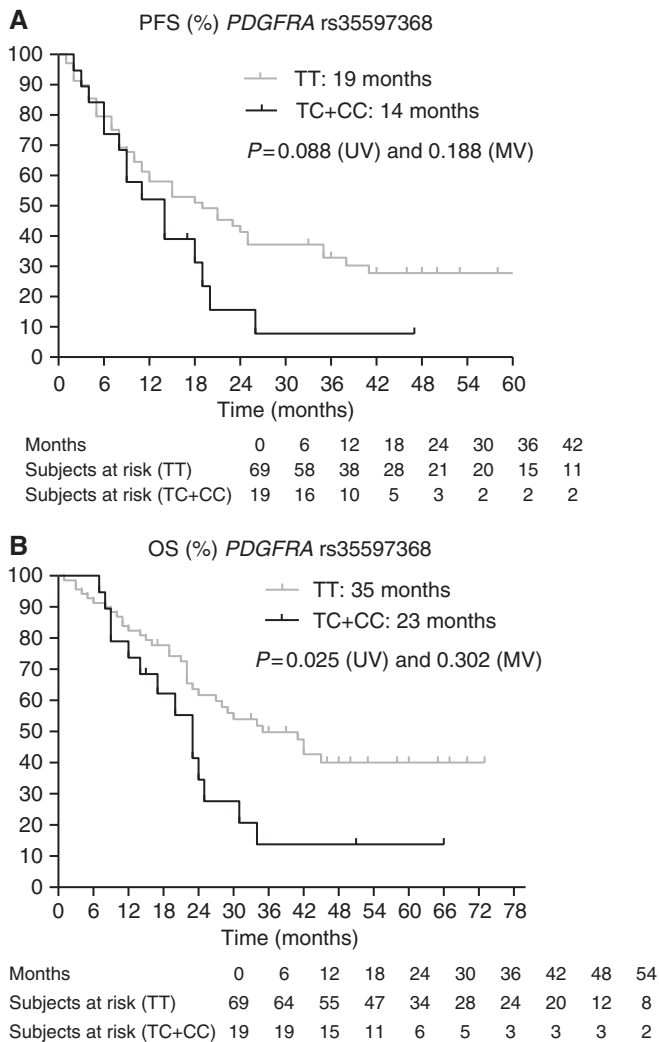


Figure 5. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs35597368 in *PDGFRA*. The *P*-values are indicated for the univariate (UV) and multivariate (MV) analyses.

rs307826 was 3.6 months vs 13.7 months for patients with the AA genotype ($P=0.00049$). There was no significant association with OS (Garcia-Donas *et al*, 2011). In 228 patients treated with pazopanib, OS was 26 months in the AA variant vs 23 months in the AG variant ($P=0.04$) of rs307826 but, surprisingly, these authors did not find any association between the SNP and PFS (Xu *et al*, 2011a,b). This matches the observation of van der Veldt *et al* (2010), who reported no significant effect of rs307826 on PFS after sunitinib treatment.

Our study has several potential limitations. (1) It was a retrospective analysis of patients treated in several centres without a central protocol dictating schedule and dose modifications or timing of radiological assessments. (2) Because our patients were mainly white, the relevance of these polymorphisms needs to be assessed in other ethnic groups, in whom the described polymorphisms may be less frequent. (3) We failed to genotype SNP rs1126647 in IL-8 because of technical reasons. (4) In case of rs11549467 there was only one heterozygous patient. As a consequence, the impact of this SNP could not be analysed. (5) Concerning SNPs in *ABCB1* and *NRI3*, in literature, only results of associations with haplotypes were available. (6) Finally, there was a better outcome in our series (PFS 15.0 and OS 29.0 months) compared with the outcome on sunitinib in the pivotal trial (PFS 11.0 and OS 26.0 months; Motzer *et al*, 2007). This

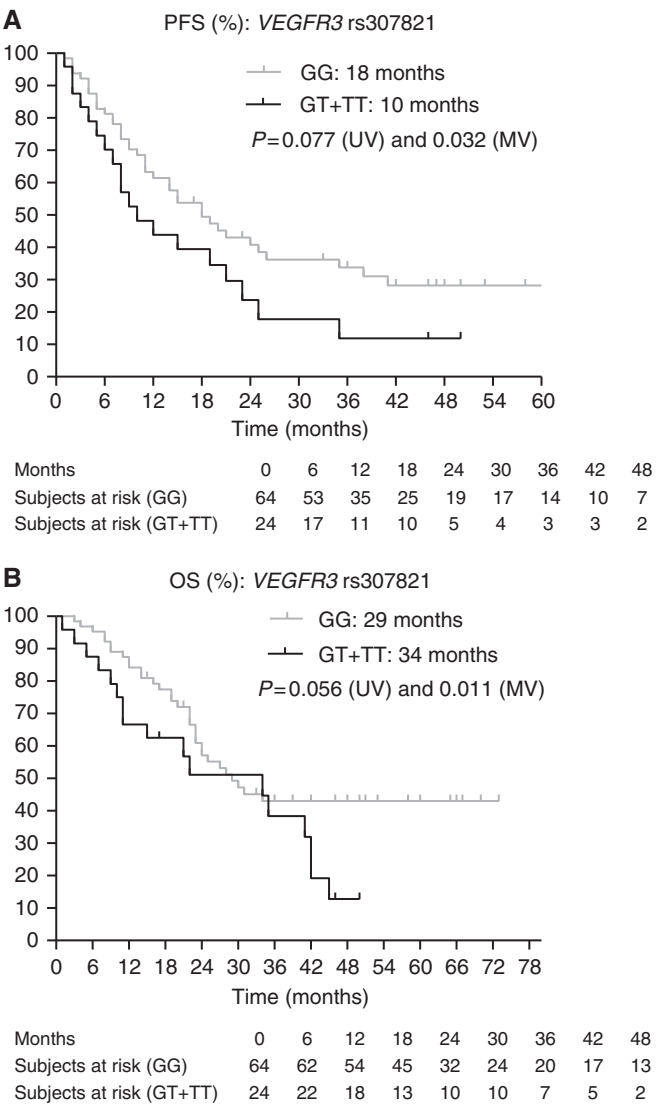


Figure 6. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs307821 in *VEGFR3*. The *P*-values are indicated for the univariate (UV) and multivariate (MV) analyses. Note that because of a crossing of the curves, the median OS was longer in the GT and TT variants than in the GG variants of rs307821. Nevertheless, the HR for survival for patients with the GT or TT variants in rs307821 in *VEGFR3* vs patients with the GG variant was 2.265 (95% CI 1.202–4.238).

difference is likely because of the patient selection in our series: all the patients had to complete at least one cycle of sunitinib and had to reach at least the first evaluation by CT scan.

CONCLUSIONS

We confirmed several associations between polymorphisms in genes linked to pharmacokinetics and pharmacodynamics of sunitinib and therapeutic outcome of patients receiving sunitinib for metastatic RCC. These associations had previously been described in other series of patients treated with sunitinib or pazopanib.

The impact of SNPs in pathways linked to pharmacokinetics and pharmacodynamics of sunitinib shows that besides acquired genetic characteristics of tumour cells, patient’s germline genetic variation may also affect the efficacy of anticancer therapy.

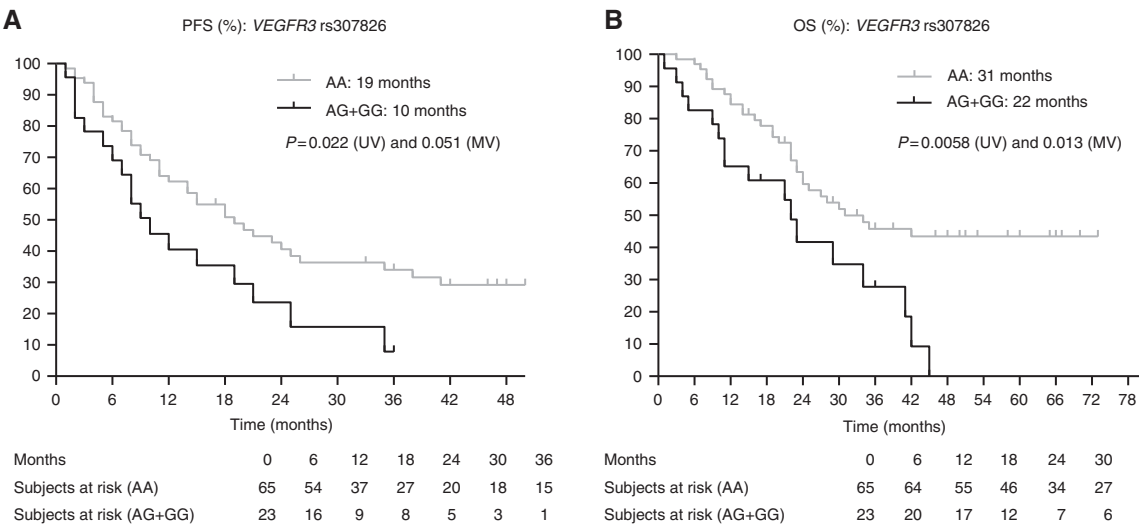


Figure 7. (A and B) Kaplan–Meier curves for PFS and OS for SNP rs307826 in *VEGFR3*. The *P*-values are indicated for the univariate (UV) and multivariate (MV) analyses.

Table 5. Distribution of SNP genotypes in patients exhibiting progressive disease and partial response as the best response						
Gene (a)	SNP ID	In patients with PD as their best response (n = 10)	In patients with SD, PR or CR as their best response (n = 78)	P-Value by Fisher's exact	Adjusted P-value by logistic regression	Odds ratio (95% CI)
Genes involved in pharmacokinetics						
ABCB1	rs1045642	CC 2/10 (20%)	CC 23/77 (30%)	NS	—	—
	rs1128503	TT 3/10 (30%)	TT 12/78 (15%)	NS	—	—
	rs2032582	TT or TA 2/8 (25%)	TT or TA 10/72 (14%)	NS	—	—
	TCG copy	Not present 7/7 (100%)	Not present 54/70 (77%)	NS	—	—
CYP3A5	rs776746	GG 8/8 (100%)	GG 61/67 (91%)	NS	—	—
NR1/2	rs3814055	TT 3/8 (38%)	TT 12/74 (16%)	NS	—	—
	rs2276707	TT 2/8 (25%)	TT 3/75 (4%)	0.02	NS	—
NR1/3	rs2307424	CC 6/10 (60%)	CC 36/78 (46%)	NS	—	—
	rs2307418	AA 8/10 (80%)	AA 49/78 (63%)	NS	—	—
	rs4073054	TT 7/10 (70%)	TT 31/78 (40%)	0.08	NS	—
	CAT copy	Present 8/10 (80%)	Present 40/77 (52%)	0.09	NS	—
Genes involved in pharmacodynamics						
PDGFRA	rs35597368	TT 8/10 (80%)	TT 61/78 (78%)	NS	—	—
VEGFR2	rs1870377	TT 7/10 (70%)	TT 41/78 (53%)	NS	—	—
VEGFR3	rs307821	GT + TT 5/10 (50%)	GT + TT 18/78 (23%)	0.07	0.05	5.763 (0.986–33.693)
	rs307826	GA + GG 6/10 (60%)	GA + GG 17/78 (22%)	0.01	0.02	7.011 (1.372–42.209)
Genes in alternative proangiogenic factors						
FGFR2	rs2981582	TT 2/10 (20%)	TT 10/77 (13%)	NS	—	—
IL8	rs4073	AA 1/9 (11%)	AA 11/70 (16%)	NS	—	—
Abbreviations: PR = partial response; PD = progressive disease; CR = complete response; SNP = single-nucleotide polymorphism; 95% CI = 95% confidence interval; SD = stable disease; NS = nonsignificant. The logistic regression analysis was adjusted for the presence of sarcomatoid dedifferentiation, the MSKCC score and baseline neutrophils. Variants were combined as follows: ABCB1: a TCG copy was linked to better outcome in van der Veldt et al (2010). Therefore, we analysed the impact of CC in rs1045642, TT in rs1128503 and TT (or TA) in rs2032582; CYP3A5: the GG variant was linked to poor outcome in van der Veldt et al (2010); NR1/2 rs3814055 and rs2276707: the TT variant was linked to poor outcome in van der Veldt et al (2010); NR1/3: a CAT copy was linked to poor outcome in van der Veldt et al (2010). Therefore, we analysed the impact of CC in rs2307424, AA in rs2307418 and TT in rs4073054; FGFR2: the TT variant was linked to poor outcome in Xu et al (2011a,b); IL8: the AA variant was linked to poor outcome in Xu et al (2011a,b); PDGFRA: the TT variant was linked to poor outcome in van der Veldt et al (2010); VEGFR2: the TT variant was linked to poor outcome in van der Veldt et al (2010); VEGFR3 rs307821: the GT/TT variant was linked to poor outcome in Garcia-Donas et al (2011); VEGFR3 rs307826: the GA/GG variant was linked to poor outcome in Garcia-Donas et al (2011).						

Moreover, germline DNA is inherited, fixed and relatively insensitive to time and environmental factors, which makes it more reliable than nucleotide and protein biomarkers linked to the tumour.

If the impact of these and other SNPs on outcome on sunitinib could be validated prospectively in independent series, scoring systems based on the combination of several unfavourable or favourable SNPs could be elaborated. When combining these SNPs with clinical and biochemical parameters associated with outcome, we will probably be able to predict more precisely the chance of response to sunitinib and identify primary resistant patients in order to orient them towards other therapies, avoiding unnecessary side effects and costs. Similarly, we will be able to predict more accurately disease progression, which is the time point of secondary resistance to sunitinib. Polymorphisms could also help us to identify those patients whose ideal starting dose of sunitinib could be higher than the usual 50 mg daily, for instance, patients with genotypes and haplotypes leading to lower sunitinib plasma levels.

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CONFLICT OF INTEREST

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REFERENCES

- Beuselinck B, Oudard S, Rixe O, Wolter P, Blesius A, Ayllon J, Elaidi R, Schöffski P, Barrascout E, Morel A, Escudier B, Lang H, Zucman-Rossi J, Medioni J (2011) Negative impact of bone metastasis on outcome in clear cell renal cell carcinoma treated with Sunitinib. *Ann Oncol* **22**: 794–800.
- Dietrich CG, Geier A, Oude Elferink RPJ (2003) ABC of oral bioavailability: transporters as gatekeepers in the gut. *Gut* **52**: 1788–1795.
- Garcia-Donas J, Esteban E, Leandro-García LJ, Castellano DE, Del Alba AG, Climent MA, Arranz JA, Gallardo E, Puente J, Bellmunt J, Mellado B, Martínez E, Moreno F, Font A, Robledo M, Rodríguez-Antona C (2011) Single nucleotide polymorphism associations with response and toxic effects in patients with advanced renal-cell carcinoma treated with first-line sunitinib: a multicentre, observational, prospective study. *Lancet Oncol* **12**: 1143–1150.
- Golshayan AR, George S, Heng DY, Elson P, Wood LS, Mekhail TM, Garcia JA, Aydin H, Zhou M, Bukowski RM, Rini BI (2009) Metastatic sarcomatoid renal cell carcinoma treated with vascular endothelial growth factor-targeted therapy. *J Clin Oncol* **27**: 235–241.
- Heng DY, Xie W, Regan MM, Warren MA, Golshayan AR, Sahi C, Eigl BJ, Ruether JD, Cheng T, North S, Venner P, Knox JJ, Chi KN, Kollmannsberger C, McDermott DF, Oh WK, Atkins MB, Bukowski RM, Rini BI, Choueiri TK (2009) Prognostic factors for overall survival in patients with metastatic renal cell carcinoma treated with vascular endothelial growth factor-targeted agents: results from a large, multicenter study. *J Clin Oncol* **27**: 5794–5799.
- Houk BE, Bello CL, Poland B, Rosen LS, Demetri GD, Motzer RJ (2010) Relationship between exposure to sunitinib and efficacy and tolerability endpoints in patients with cancer: results of a pharmacokinetic/pharmacodynamic meta-analysis. *Cancer Chemother Pharmacol* **66**: 357–371.
- Meyer KB, Maia AT, O'Reilly M, Teschendorff AE, Chin SF, Caldas C, Ponder BA (2008) Allele-specific up-regulation of FGFR2 increases susceptibility to breast cancer. *PLoS Biol* **5**: e108.
- Motzer R, Bacik J, Mazumbar M (2004) Prognostic factors for survival of patients with stage IV renal cell carcinoma: Memorial Sloan Kettering Cancer Center experience. *Clin Cancer Res* **10**: 6202S–6203S.
- Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Oudard S, Negrier S, Szczylik C, Pili R, Bjarnason GA, Garcia-del-Muro X, Sosman JA, Solska E, Wilding G, Thompson JA, Kim ST, Chen I, Huang X, Figlin RA (2009) Overall survival and updated results for sunitinib compared with interferon alfa in patients with metastatic renal cell carcinoma. *J Clin Oncol* **27**: 3584–3590.
- Motzer RJ, Hutson TE, Tomczak P, Michaelson MD, Bukowski RM, Rixe O, Oudard S, Negrier S, Szczylik C, Kim ST, Chen I, Bycott PW, Baum CM, Figlin RA (2007) Sunitinib versus interferon alfa in metastatic renal-cell carcinoma. *N Engl J Med* **356**: 115–124.
- Partanen TA, Alitalo K, Miettinen M (1999) Lack of lymphatic vascular specificity of vascular endothelial growth factor 3 in 185 vascular tumors. *Cancer* **86**: 2406–2412.
- Patil S, Figlin R, Hutson T, Michaelson M, Negrier S, Kim T, Huang X, Motzer R (2011) Prognostic factors for progression free survival and overall survival with sunitinib targeted therapy and with cytokine as first-line therapy in patients with metastatic renal cell carcinoma. *Ann Oncol* **22**: 295–300.
- Reumers J, De Rijk P, Zhao H, Liekens A, Smeets D, Cleary J, Van Loo P, Van Den Bossche M, Cathoor K, Sabbe B, Despiere E, Vergote I, Hilbush B,

- Lambrechts D, Del-Favero J (2011) Optimized filtering reduces the error rate in detecting genomic variants by short-read sequencing. *Nat Biotechnol* **30**: 61–68.
- Rini BI, Atkins MB (2009) Resistance to targeted therapy in renal-cell carcinoma. *Lancet Oncol* **10**: 992–1000.
- Soto-Vega E, Arroyo C, Richaud-Patin Y, Garcia-Carrasco M, Vazquez-Lavista LG, Llorente L (2009) P-glycoprotein activity in renal clear cell carcinoma. *Urol Oncol* **27**: 363–366.
- Valtola R, Salven P, Heikkilä P, Taipale J, Joensuu H, Rehn M, Pihlajaniemi T, Weich H, deWaal R, Alitalo K (1999) VEGFR-3 and its ligand VEGF-C are associated with angiogenesis in breast cancer. *Am J Pathol* **154**: 1381–1390.
- van der Veldt AA, Eechoute K, Gelderblom H, Gietema J, Guchelaar HJ, van Erp NP, van den Eertwegh AJ, Haanen JB, Mathijssen RH, Wessels JA (2010) Genetic polymorphisms associated with a prolonged progression-free survival in patients with metastatic renal cell cancer treated with sunitinib. *Clin Cancer Res* **17**: 620–629.
- Walsh N, Larkin A, Kennedy S, Connolly L, Ballot J, Ooi W, Gullo G, Crown J, Clynes M, O'Driscoll L (2009) Expression of multidrug resistance markers ABCB1 (MDR-1/P-gp) and ABCC1 (MRP-1) in renal cell carcinoma. *BMC Urol* **9**: 6.
- Xu CF, Bing NX, Ball HA, Rajagopalan D, Sternberg CN, Hutson TE, de Souza P, Xue ZG, McCann L, King KS, Ragone LJ, Whittaker JC, Spraggs CF, Cardon LR, Mooser VE, Pandite LN (2011a) Pazopanib efficacy in renal cell carcinoma: evidence for predictive genetic markers in angiogenesis-related and exposure-related genes. *J Clin Oncol* **29**: 2557–2564.
- Xu CF, Ball HA, Bing N, Sternberg C, Xue Z, McCann L, King K, Spraggs C, Mooser V, Pandite LN (2011b) Association of genetic markers in angiogenesis- or exposure-related genes with overall survival in pazopanib-treated patients with advanced renal cell carcinoma. *J Clin Oncol* **29**(Suppl 7): abstr 303.