www.nature.com/tp

REVIEW Neuropathology of mood disorders: do we see the stigmata of inflammation?

N Mechawar¹ and J Savitz^{2,3}

A proportion of cases with mood disorders have elevated inflammatory markers in the blood that conceivably may result from stress, infection and/or autoimmunity. However, it is not yet clear whether depression is a neuroinflammatory disease. Multiple histopathological and molecular abnormalities have been found postmortem but the etiology of these abnormalities is unknown. Here, we take an immunological perspective of this literature. Increases in activated microglia or perivascular macrophages in suicide victims have been reported in the parenchyma. In contrast, astrocytic markers generally are downregulated in mood disorders. Impairment of astrocytic function likely compromises the reuptake of glutamate potentially leading to excitotoxicity. Inflammatory cytokines and microglia/macrophage-derived quinolinic acid (QA) downregulate the excitatory amino acid transporters responsible for this reuptake, while QA has the additional effect of inhibiting astroglial glutamine synthetase, which converts glutamate to glutamine. Given that oligodendroglia are particularly vulnerable to inflammation, it is noteworthy that reductions in numbers or density of oligodendrocyte cells are one of the most prominent findings in depression. Structural and/or functional changes to GABAergic interneurons also are salient in postmortem brain samples, and may conceivably be related to early inflammatory insults. Although the postmortem data are consistent with a neuroimmune etiology in a subgroup of depressed individuals, we do not argue that all depression-associated abnormalities are reflective of a neuroinflammatory process or even that all immunological activity in the brain is deleterious. Rather, we highlight the pervasive role of immune signaling pathways in brain function and provide an alternative perspective on the current postmortem literature.

Translational Psychiatry (2016) 6, e946; doi:10.1038/tp.2016.212; published online 8 November 2016

INTRODUCTION

The question of whether mood disorders are neuropathological conditions has been the subject of considerable debate.^{1–3} In the case of major depressive disorder (MDD) and bipolar disorder (BD), there is evidence for subtle histopathological changes observed postmortem that plausibly may affect excitatory and inhibitory circuits involving the prefrontal cortex (PFC) and limbic regions. The most salient of these abnormalities are reductions in the size and/or density of GABAergic neurons and reductions in glial cell densities that are concurrent with altered gene expression. These data have been reviewed in detail elsewhere¹ and are not comprehensively covered here. Instead we focus on one potential cause of the histopathological changes associated with mood disorders—inflammation. That is, we highlight those postmortem cellular and molecular abnormalities that may be directly or indirectly related to immune dysregulation. This is not intended to be a systematic review of the literature. Nor do we argue that all changes observed postmortem in mood disorders are related to inflammation. Rather we provide an alternative perspective on a substantial and diverse body of work.

EVIDENCE FOR IMMUNE DYSREGULATION IN MOOD DISORDERS

A large number of studies have reported increased levels of inflammatory proteins such as tumor necrosis factor (TNF),

interleukin 6 (IL-6) and C-reactive protein (CRP) in the serum or plasma of depressed individuals with MDD and BD, and these results have been confirmed in meta-analyses.⁴⁻⁸ Concentrations of pro-inflammatory cytokines are also reportedly elevated during mania or hypomania.^{9,10} Although most studies have focused on cytokine proteins, additional support for the role of inflammation in mood disorders is derived from gene expression studies of peripheral blood mononuclear cells, which have demonstrated the existence of increased messenger RNA (mRNA) expression of proinflammatory mediators in patients with mood disorders. Notably, Padmos et al.¹¹ performed whole-genome expression profiling on microarrays using purified cluster of differentiation 14 (CD14+) monocytes and reported elevated mRNAs of inflammatory (for example, TNF, IL-1, IL-6, TNF-alpha-induced protein 3), trafficking, survival and mitogen-activated protein kinase pathway genes in BD subjects in various illness phases, and in the affected offspring of other BD subjects. Similarly, the expression of several genes previously implicated in neurological and inflammatory disorders, including TNF, were found to be upregulated in the peripheral blood mononuclear cells of a combined sample of depressed MDD and BD participants.¹⁴ Further, Pandey *et al.*¹³ found that the mRNA expression of IL-1, IL-6 and TNF, as well as their receptors, IL-1R1, IL-1RA and TNFR1 was significantly greater in the lymphocytes from BD patients than those of healthy controls. These data demonstrate the potentially important pathological role of membrane-bound cytokine receptors, which

¹Douglas Mental Health University Institute and Department of Psychiatry, McGill University, Montreal, QC, Canada; ²Laureate Institute for Brain Research, Tulsa, OK, USA and ³Faculty of Community Medicine, The University of Tulsa, Tulsa, OK, USA. Correspondence: Dr J Savitz, Laureate Institute for Brain Research, 6655 S. Yale Avenue, Tulsa, OK 74136, USA.

E-mail: jsavitz@laureateinstitute.org

Received 5 April 2016; revised 8 July 2016; accepted 30 August 2016

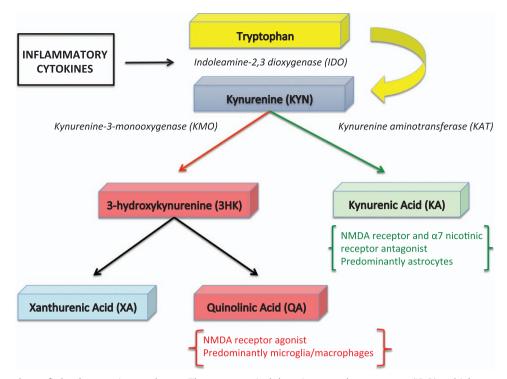


Figure 1. Main branches of the kynurenine pathway. The enzyme indoleamine 2,3 deoxygenase (IDO), which converts tryptophan to kynurenine is upregulated by pro-inflammatory cytokines. Each box represents a metabolite resulting from the oxidation of tryptophan. The black italicized text shows the enzymes that catalyze select steps in the metabolic pathway. NMDA, *N*-methyl-D-aspartate.

mediate the functional and biological effects of cytokines. In a follow-up study by the same group, similar increases in the mRNA expression of cytokines and their membrane-bound receptors were found in hospitalized MDD patients.¹⁵ Soluble cytokine receptors may also be relevant to the pathophysiology of depression. Elevations in the protein concentrations of soluble IL-1R (sIL-1R), sTNFR1, sTNFR2 and sIL-6R also have been reported in patients with mood disorders.^{16–18}

It is not yet clear whether the association between mood disorders and inflammation is causal. Regarding prospective studies, elevated CRP levels have been shown to predict the subsequent development of BD¹⁹ as has been demonstrated several times for CRP or IL-6 in the case of MDD and psychosis.²⁰⁻²² Second, it has been well established that treatment of hepatitis C or melanoma patients with interferon alpha (IFN-a) or interleukin 2 (IL-2) induces a depressive episode in about 40% of patients.²³⁻²⁵ Importantly, there is a temporal disjunction between the 'psychological' and 'physical' manifestations of this immune-stimulating treatment, with the neurovegetative symptoms appearing within 1 week, whereas the mood and cognitive symptoms peak 8-12 weeks post initiation of treatment.^{26,27} Moreover, it is the mood and cognitive symptoms rather than the neurovegetative symptoms that are responsive to antidepressant treatment.^{26,27} Similarly, the typhoid vaccine and low-dose endotoxin, which has been shown to activate microglia in vivo,²⁸ induce transient, mild depressive symptoms in healthy controls.^{29–31} These data prompted investigations into the efficacy of anti-inflammatory medications for the treatment of bipolar depression. Results have been mixed³² although low-dose aspirin, minocycline, infliximab and n-acetyl cysteine show some therapeutic promise.^{33–35} Finally, positron emission tomography studies have found evidence for microglial activation in the PFC and anterior cingulate cortex (ACC) of patients with MDD as well as the hippocampus in patients with BD as indexed by an increase in the distribution volume of the ligand for the translocator protein ligand, TPSO, which is expressed by microglia. 36,37

Inflammatory mediators likely affect neuronal function and neurotransmission by an array of different mechanisms. For instance, pro-inflammatory cytokines such as interferon gamma (IFN-y), IL-1 and TNF can reduce the availability of monoamines (serotonin, dopamine and norepinephrine) by upregulating synaptic reuptake transporters, as well as reducing monoamine synthesis by decreasing the availability of tetrahydrobiopterin (BH4), a co-factor for the enzyme tyrosine hydroxylase (see ref. 38 for a comprehensive review). At the circuit level, these alterations in neurotransmission lead to functional changes in components of the visceromotor network including the ventromedial PFC, insula and hippocampus,^{30,31,39} as well as hypoactivity of a 'reward network' centered on the ventral striatum, thus providing a clear link with the behavioral trait of anhedonia.^{29,40,41} Nevertheless, postmortem studies have generally pointed to abnormalities in glutamatergic and/or GABAergic circuits in BD, and thus immunemediated modulation of the balance between excitatory and inhibitory neurotransmission will be the primary focus here.

As will be discussed below, activated microglia release glutamate^{42–47} and pro-inflammatory cytokines further elevate the risk for excitotoxicity by impairing astrocyte-mediated glutamate recycling.^{48,49} However, inflammatory mediators also may affect glutamatergic signaling indirectly by, for instance, altering the production of neuroactive metabolites of the kynurenine pathway (Figure 1). Specifically, pro-inflammatory cytokines upregulate the enzyme indoleamine 2,3 deoxygenase, increasing the production of kynurenine from tryptophan. Kynurenine in turn is metabolized along two principal branches to form kynurenic acid (KynA) or alternatively, the potentially neurotoxic metabolites, 3HK (3-hydroxykynurenine) and quinolinic acid (QA).⁵⁰ KynA is a pleiotrophic metabolite that is produced by astrocytes and among other roles, acts as a preferential antagonist at the glycine co-agonist site of the *N*-methyl-D-aspartate (NMDA)

receptor.⁵¹ In contrast, QA is produced by microglia and is neurotoxic at elevated concentrations partly via its agonistic effect on the NMDA receptor.⁵² Although QA is usually found in low nanomolar concentrations in the human brain and cerebrospinal fluid, a significant increase in QA levels to micromolar concentrations is observed in patients with inflammatory neurological diseases.⁵³ In addition to its effect on the NMDA receptor, QA has been shown to increase the production of reactive oxygen and nitrogen species,⁵⁴ inhibit the reuptake of glutamate into synaptic vesicles,⁵⁵ promote the formation of hyperphosphorylated tau proteins⁵⁶ and upregulate transcription of pro-inflammatory chemokines and cytokines in astrocytes.⁵⁷

Decreases in KynA and increases in QA in the cerebrospinal fluid of predominantly depressed subjects have been found up to 2 years after a suicide attempt,⁵⁸ and we previously reported a reduction in the ratio of KynA to 3HK and KynA to QA in the serum of patients with both MDD and BD.^{59,60} In addition, the ratio of 3HK to KynA and/or QA to KynA in the serum was inversely correlated with hippocampal volume in both MDD and BD participants, as well as depressed football players with concussion, raising the possibility that neuroactive kynurenine metabolites may affect brain structure in the context of depression.^{59–61}

ETIOLOGY OF THE IMMUNE DYSREGULATION IN MOOD DISORDERS

Although the evidence for immune dysregulation in a subgroup of individuals with mood disorders is robust, the upstream cause of this immune dysregulation is less clear. Below, we discuss three potential, non-mutually exclusive causal factors, stress, pathogens and autoimmunity.

Stress

Stress hormones (for example, epinephrine, norepinephrine and cortisol) and 'hard-wired' sympathetic nervous system innervation of lymphoid organs exert pleiotrophic effects on the immune system via their receptors on immune cells. This stress response causes (potentially chronic) 'low-grade' inflammation characterized by increases in pro-inflammatory mediators such as TNF, IL-6 and IFN-y, as well as impairment of the adaptive immune system leading to increased vulnerability to viral disease, deficient response to vaccination and poorer prognosis for diseases such as cancer.⁶² In complementary studies, Cole and colleagues have demonstrated that social isolation and loneliness is associated with a consistent pattern of gene expression in peripheral blood leukocytes, that is, an increase in inflammatory markers involved in wound healing and bacterial immunity together with a downregulation of transcripts involved in Type I interferon antiviral responses and IgG antibody production.^{63–65} In the brain, this immunological dysregulation may lead to increased activity of the amygdala, deficient hippocampal neuroplasticity, and more generally, morphological and/or atrophic changes to pyramidal neurons.⁶⁶

Pathogens

The impaired adaptive immunity associated with chronic stress may lead to increased susceptibility to the negative sequelae of pathogens, especially viruses. Herpesvirus infections are a salient example. The majority of the population of the United States (US) is seropositive for Epstein–Barr virus and herpes simplex 1 virus,⁶⁷ whereas more than half of the US population is seropositive for cytomegalovirus.⁶⁸ Generally, these viruses remain latent after initial infection but they may undergo reactivation during periods of both physical and psychological stress without producing significant clinical disease. Reactivation is usually accompanied by a noticeable increase in specific antibody titer to the virus, even in the absence of detectable virus. For instance, IgG antibody titers

to cytomegalovirus, Epstein–Barr virus and herpes simplex 1 virus were found to be significantly elevated in medical students during exams compared with when they returned after summer vacation.⁶⁹ Several studies have reported higher IgG cytomegalovirus antibody titers in depressed subjects relative to controls^{70–72} and more recently, the Detroit Neighborhood Health Study showed that individuals with cytomegalovirus antibodies in the top quartile were four times more likely to be depressed than those individuals in the bottom three quartiles of the population.⁷³

Another pathogen that may have a role in the pathophysiology of mood disorders is the protozoan, *Toxoplasma gondii*.⁷⁴ A metaanalysis of 23 studies found that serological evidence of infection with *T. gondii* was associated with a 2.73-fold increased risk of schizophrenia⁷⁵ and several recent studies have reported associations between *T. gondii* seropositivity and depression, mania and suicidal behavior more generally.^{76–79} Like the herpesviruses, *T. gondii* is neurotrophic and additionally encodes proteins with homology to tyrosine hydroxylase and the D2 receptor, raising the possibility that it may modulate dopaminergic neurotransmission.⁸⁰ Further, *T. gondii*-infected mice display increased production of neuroactive kynurenine pathway metabolites in the brain, suggesting another possible mechanistic connection between *T. gondii* and the pathophysiology of mood disorders.⁸¹

Autoimmunity

Certain microbial infections may increase the risk of developing autoimmune disease via molecular mimicry and/or bystander activation and stimulation of pattern recognition receptors.82 There are two predominant sources of evidence for autoimmune illness in depression and psychosis. First, there is a greater prevalence of various autoimmune disorders in patients with mood disorders than that of the general population.^{83–87} Further, there is persuasive epidemiological evidence to suggest that autoimmune disease is a risk factor for the development of de novo mood disorders. For instance, using the Danish Psychiatric Central Register, which included data on 90 000 inpatient admissions for depression, Benros *et al.*⁸⁸ found that a history of any prior autoimmune disease increased the risk of a subsequent diagnosis with a mood disorder by 45%, a history of hospitalization for infection increased the risk of later mood disorders by 62% and the two factors interacted in synergy to increase the risk of subsequent mood disorders by 235%.

Second, there is evidence to suggest that a subset of patients with mood disorders have elevated levels of circulating autoantibodies. For instance, thyroperoxidase antibodies are significantly more common in patients with MDD and BD even after accounting for lithium exposure,^{89,90} a finding that may be related to a shared genetic vulnerability to BD and autoimmune thyroiditis.⁸⁴ In another one of many examples, Ching et al.⁹¹ reported that 2 out of 20 patients with a diagnosis of MDD had raised levels of autoantibodies to glutamic acid decarboxylase 65 (GAD₆₅) and Ro₅₂ in both the serum and cerebrospinal fluid, with one of the patients meeting criteria for stiff-person syndrome. Whether these antibodies have a causal role in the development of the illness or are a consequence of an independent pathological process is still unknown. However, at least some cases of psychosis and/or depression appear to be caused by antibodies that have functional effects on neurotransmission. Dalmau and colleagues first reported the existence of autoantibodies, which cause cross-linking and internalization of the NMDA receptor in limbic encephalitis patients with a variety of psychiatric symptoms including auditory and visual hallucinations, delusions, depression and mania.^{92,93} Further, ~4% of the cases in the Kayser et al.93 series presented with isolated psychiatric symptoms. Subsequently, anti-NMDA receptor antibodies also

have been reported to be present at an increased frequency in patients initially diagnosed with MDD (3%)⁹⁴ and post-partum psychosis (2%).⁹⁵ On the basis of these and other studies, it has become increasingly recognized that a small minority of patients diagnosed with psychosis or depression may in fact have a variant of autoimmune encephalitis. In a recent editorial, Lennox *et al.*⁹⁶ wrote that 'Antibody screening in young people presenting with psychosis, seizures and cognitive disturbance is now part of routine clinical practice in neurological and intensive care settings

[in the United Kingdom]'.

INFLAMMATION-RELATED HISTOPATHOLOGICAL ABNORMALITIES

Postmortem brain studies: methodological considerations

Among the different approaches used to investigate psychiatric illnesses, postmortem brain research offers a unique window through which underlying cellular and molecular alterations can be observed and described with the highest resolution and sensitivity. Moreover, postmortem studies can provide invaluable information on cerebral cells and networks that are only found in humans. This is particularly crucial when it comes to mental illnesses unique to humans such as BD. For instance, our view of human astrocytes has changed considerably in the past few years since Nedergaard and colleagues elegantly highlighted that both the size and complexity of cortical astrocytes are disproportio-nately greater in humans than in rodents.^{97,98} Furthermore, the same team found that the diversity of these cells is more important in the human neocortex, with astrocytic subtypes displaying projections spanning several cortical layers or columns. The great diversity of human astrocytes was subsequently confirmed and extended to other brain regions, with the discovery of novel subtypes in the hippocampus and, more recently, in subcortical regions such as the thalamus and caudate nucleus.^{99,100} The notion that human astrocytes may also display different functional properties than in animal models further highlights the importance of clinical and postmortem studies to better understand the biological roots of mental illnesses.

Despite these advantages, postmortem studies also present some limitations, and the results should always be interpreted with consideration for factors such as: (i) the quality of sample preservation, which is generally assessed by measuring tissue pH and RNA quality (RNA Integrity Number); (ii) the necessity of adequately matched controls (age, death with or without agonal period, postmortem delay, tissue characteristics and so on); and (iii) the amount of information (for example, clinical, psychosocial and so on) available for each donor, which can be considerably increased by psychological autopsies. Even when generated in optimal conditions, postmortem data showing significant differences between cases and controls should be viewed as a 'snapshot' of cellular and molecular events unfolding in brain tissues before death, without precluding the possibility that the observations may reflect cumulative changes having occurred during the course of the illness. Consideration should also be given to possible postmortem changes to brain tissues between death and their preservation by freezing or fixation, although a careful matching of samples should eliminate this concern.

Pro-inflammatory molecules and pathways

Despite the fact that several studies have examined peripheral (plasma and cerebrospinal fluid) inflammatory molecules and mediators in patients with mood disorders, little is currently known about the expression of inflammatory markers in the brain of such individuals. The few postmortem investigations that have addressed this question have all examined cortical areas and are, unfortunately, largely inconsistent. Using real-time polymerase chain reaction to compare the expression of TNF, IL-1, IL-4, IL-5,

IL-6 and IL-13 in the orbitofrontal cortex of suicides vs nonpsychiatric controls, Tonelli *et al.*¹⁰¹ reported a significant increase in the expression of IL-4 in women and of IL-13 in men. As highlighted by the authors, this study had important limitations, such as a lack of diagnosis for the majority of suicides, the absence of toxicological data, and the fact that groups were not matched by age.¹⁰¹ The roles of IL-4 and IL-13 in the brain, similar cytokines released by T-helper 2 cells in allergic inflammation, remain mostly to be determined (but see ref. 102). Interestingly, there is *in vitro* evidence showing that in rat cells, IL-13 can lead to the death of activated microglial cells by enhancing the production of cyclooxygenase-2.¹⁰³ Thus, increased IL-13 expression could represent a mechanism through which microglial activation is kept in check.

In their investigation of teenage suicides (various diagnoses), Pandey *et al.*¹⁰⁴ found that both mRNA and protein levels of proinflammatory cytokines TNF, IL-1 and IL-6 were significantly increased in prefrontal cortex (Brodmann's Area 10; BA10) relative to matched controls. These findings are consistent with the results of a later study by this group on the expression of the Toll-like receptors (TLR) found on macrophages and microglia.¹⁰⁵ They showed that the expression (mRNA and protein) of TLR3 and TLR4 was significantly upregulated in the dorsolateral PFC of depressed suicides vs non-depressed suicides. TLR3 and TLR4 mRNA, but not protein, was similarly increased in depressed non-suicides, suggesting that suicide, independent of diagnosis, is associated with a stronger dysregulation in the expression of these receptors.¹⁰⁵

Focusing on TNF protein expression in prefrontal cortical samples (BA24 and BA46) from 10 adult MDD patients (mostly suicides) and matched controls, Dean et al.¹⁰⁶ also detected a highly significant depression-associated increase in protein levels of transmembrane—but not soluble—TNF that was restricted to BA46. A more extensive follow-up study by these authors showed a greater than three-fold increase in transmembrane TNF expression in ACC (BA24) samples from BD patients, as well as a 51 and 67% decrease in TNFR2 expression in BA46 samples from MDD and BD subjects, respectively.¹⁰⁷ These results suggest illness-specific regional disruptions in TNF signaling. In contrast, Rao et al.¹⁰⁸ did not detect any difference in TNF expression in the frontal cortical samples from BD patients relative to controls. They did however, report significantly increased mRNA and protein levels for IL-1 and IL-1R, as well as of astrocytic (glial fibrillary acidic protein; GFAP) and microglial (CD11b) markers in the same tissues.¹⁰⁸ Using microarrays, a more high-throughput approach, some investigators have reported a widespread upregulation of both pro- and anti-inflammatory cytokines in the dorsolateral PFC of MDD patients,¹⁰⁹ whereas others have been unable to implicate cerebral inflammation in any of brain regions examined in individuals with BD.110

The inconsistencies in the literature presented above could be attributable to differences in experimental approaches or to the diversity of postmortem brain samples. As mentioned above, however, this diversity could be further enhanced by the clinical heterogeneity of patients suffering from mental illness. Molecular differences associated with such heterogeneity were recently highlighted in a postmortem study aimed at examining genes involved in stress and inflammation.¹¹¹ Fillman *et al.*¹¹¹ performed postmortem gene expression analyses with Stanley Array Cohort frontal/prefrontal cortex samples (34 BD, 35 schizophrenia and 35 control). Using eight markers of inflammation (Serpin Family A Member 3, IL-1, IL-1R1, IL-6, IL-8, IL-18, TNF and Prostaglandin-Endoperoxide Synthase 2), the authors were able to cluster the samples into a high inflammation subgroup consisting of 16 samples with schizophrenia (46%), 10 samples with BD (29%) and 9 controls (26%), and a low inflammation subgroup consisting of 19 samples with BD, 20 samples with schizophrenia and 25 controls. The samples also were clustered into high and low stress groups on the basis of the expression of several glucocorticoid

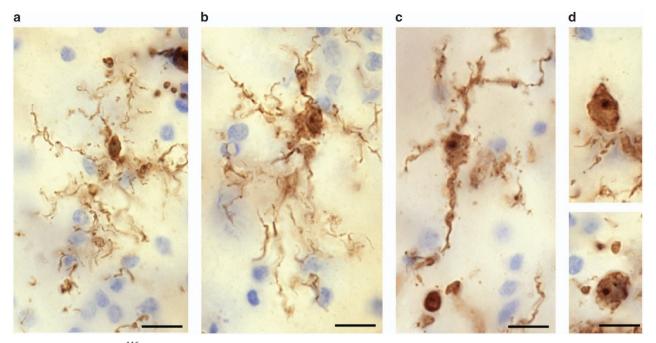


Figure 2. (Torres-Platas *et al.*¹¹⁶): Four main IBA1-IR microglial phenotypes are observed in human dorsal anterior cingulate cortex (ACC). Representative examples are illustrated here for the white matter. (**a**) Ramified microglial cell body and highly ramified processes. (**b**) Primed microglia display a wider cell body compared with the ramified phenotype. (**c**) Reactive microglia present an ameboid-shaped rounder cell body with a few ramified processes, whereas (**d**) ameboid microglia display a characteristic ameboid-shaped cell body extending one or two unramified processes (top panel) or are completely devoid of processes (bottom panel). Scale bars, 10 μm. IR, immunoreactive.

signaling-related markers. The stress and inflammation markers were further combined to yield a high stress/inflammation subgroup and a low stress/inflammation subgroup with the former consisting of 46% of the schizophrenia group, 32% of the BD group and 18% of the controls. In addition, a microarray analysis demonstrated that 57 genes were differentially expressed between the high inflammation/stress subgroup of psychiatric samples and the low inflammation/stress subgroup of controls. A pathway analysis of these 57 genes identified a network of differentially expressed genes involving immune, growth factors, inhibitory signaling and cell death factors.¹¹¹ Thus, a robust immune activation in the brain does seem to be occurring in subgroups of patients with BD and schizophrenia. More recently, Clark et al.¹¹² compared the postmortem mRNA expression of various cytokines in ventrolateral prefrontal cortical samples from patients with depression not otherwise specified to matched controls. Unexpectedly, this study showed that the expression of TNF, IFN-y, IL-13, IL-33, IL-12 and of Chemokine (C-C Motif) Ligand 2 (CCL-2), was significantly decreased in the samples from depressed patients. Furthermore, these findings were correlated with evidence of a compromised kynurenine pathway in the same samples. Taken together, these studies suggest that the important variations in expression levels of inflammation-related molecules and pathways measured in postmortem brain samples potentially reflect the clinical heterogeneity of samples analyzed, both between and within diagnoses.

Microglia

Microglia are the resident immune cells of the brain (Figure 2). In their 'resting' state, these macrophages display a ramified phenotype and are actively implicated in neuronal plasticity.¹¹³ Upon immune activation, microglia release pro-inflammatory cytokines as well as other factors, such as glutamate, chemokines and growth factors, and can undergo graded morphological changes leading up to a highly motile ameboid phenotype.^{114,115}

Other types of macrophages can be found in the brain, such as infiltrated monocytes and perivascular macrophages, both of which are highly responsive to alterations of the blood-brain barrier integrity in pathological conditions. The best way to distinguish these different cell populations in postmortem brain samples remains the observation of their fine morphological properties and spatial distribution in immunostained tissues.¹¹⁶ With this approach, it is particularly easy to distinguish microglia from perivascular macrophages, as the former occupy the parenchyma in non-overlapping domains, whereas the latter are clearly associated with blood vessels.^{114,115} Although many macrophage-specific markers have been identified, it is currently impossible to differentiate macrophage cell populations based on the expression of these markers alone. The scavenger receptor CD163 may represent an exception, as it is more highly expressed by perivascular macrophages than microglia.¹¹⁷

Few studies have examined microglia/macrophages in postmortem samples from individuals having suffered from mood disorders. Steiner et al.¹¹⁸ provided the first evidence suggesting increased microglial activation in psychiatric illnesses. These authors examined the immunohistochemical distribution of human leukocyte antigen-DR, a major histocompatibility complex (MHC class) II cell surface receptor specifically expressed by macrophages, in samples of dorsolateral prefrontal cortex, ACC and mediodorsal thalamus from individuals having died with MDD, BD and schizophrenia as well as matched non-psychiatric controls.¹¹⁸ They observed increased densities of human leukocyte antigen-DR-immunoreactive (-IR) microglial cells in all the three brain regions. Interestingly, this observation concerned samples from individuals having died from suicide, irrespective of diagnosis.¹¹⁸ The same group subsequently published another postmortem study showing increased microglial quinolinic acid-IR within ACC subregions of severely depressed individuals compared with matched controls.¹¹⁹ More recently, support to the notion of depression-associated microglial/immune activation in the brain was provided by independent postmortem

Translational Psychiatry (2016), 1-16

investigations focused on other microglial markers. On the basis of previously established morphometric criteria,¹¹⁶ the first of these studies assessed through stereology the relative abundance of microglial phenotypes immunostained for the macrophagespecific calcium-binding protein IBA1 in the dorsal ACC white matter of depressed suicides and matched sudden-death controls.¹²⁰ Although this comparison suggested a relative increase of primed microglia in depressed suicides, the most striking observation was that samples from depressed suicides displayed significant more blood vessels surrounded by a high density of IBA-IR macrophages than matched controls.¹²⁰ This may reflect increased recruitment of circulating bone marrow-derived monocytes in depressed suicides, a phenomenon that has been associated in mice with anxiety-like behavior in response to repeated social defeat stress.¹²¹ The distinction between resident microglia cells and monocyte-derived macrophages is important because the latter have been more closely linked with neurotoxicity, for instance producing 32 times more QA than resident microglia.122

The postmortem study conducted concomitantly by Schnieder and colleagues suggested that the accumulation of macrophages in the perivascular space is not specific to the ACC. Indeed, their stereological investigation of IBA1 and of CD68-IR cells in the prefrontal white matter also showed an increase in perivascular macrophages in suicides relative to controls.¹²³ In this study, the group of cases was composed of individuals who had suffered from affective disorders or from schizophrenia, and the results were significant only in the subgroup of patients who died by suicide. It remains to be explored whether this phenomenon occurs solely in the white matter or whether the blood vessels in neocortical gray matter are similarly affected. Moreover, it will be important to determine whether the perivascular space is affected in this manner only in regions associated with mood disorders and suicide, such as the ACC and PFC, or whether this is a more global phenomenon in the brain.

Glutamate released by activated microglia may induce excitotoxicity and contribute to neuronal damage and/or dysfunction. A recent proton magnetic resonance spectroscopy study reported elevations in glutamate in the basal ganglia and dorsal ACC in patients receiving treatment with IFN-a, and higher levels of glutamate correlated with increases in depressive symptoms during the course of treatment.¹²⁴ Hashimoto *et al.*¹²⁵ reported increased levels of glutamate in the frontal cortex of BD patients and several studies identified increases in the glutamine+glutamate (GLX) signal in multiple brain regions in BD patients.^{126,127} The GLX signal is constituted predominantly by intracellular glutamate and glutamine and is thought to reflect the total glutamatergic pool available for neurotransmission in the form of glutamate or glutamine.¹²⁶ The smaller number of studies that distinguished between glutamate and glutamine, generally suggested elevations in glutamate in patients with BD.¹²⁷ Thus it is unclear how best to interpret the increase in GLX, but the data are at least consistent with glutamate-induced neuronal hyperactivation.¹²⁸

Astrocytes

Astrocytes (Figure 3) have attracted much attention in the search for etiological factors in depression.¹²⁹ Consistent with the changes described above concerning macrophages in ACC white matter, morphometric data generated from the analysis of reconstructed Golgi-stained cells indicated that fibrous astrocytes in this cortical compartment displayed a hypertrophic phenotype in depressed suicides (MDD and BD) relative to controls.¹³⁰ This observation may be indicative of mild astrogliosis in response to local low-grade neuroinflammation. In this regard, it is puzzling that postmortem studies examining the expression (gene and protein) of GFAP, an astrocyte-specific intermediate filament

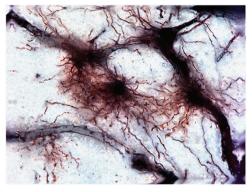


Figure 3. Golgi-stained astrocytes in the human dorsal anterior cingulate cortex (dACC). These cells extend tortuous varicose and thorny processes radiating in all the directions and often are observed to contact adjacent blood vessels, which are also silver-impregnated.

known to be upregulated with inflammation, have consistently found it to be significantly decreased in depressed cases vs matched controls.^{99,131,132} Initially described in prefrontal cortical areas, this downregulation was also recently reported in subcortical regions implicated in depression, but not in other unrelated cortical areas (for example, primary and visual motor cortex), suggesting that specific networks of astrocytes are affected in mood disorders, and that downregulation of GFAP is not a brain-wide phenomenon.⁹⁹

Beyond GFAP, studies have found that other astrocyte-specific genes such as the tropomyosin-related kinase B receptor (TrkB.1) isoform¹³³ and connexins 43 and 30 (refs 134,135) are significantly downregulated in the orbitofrontal and dorsolateral PFC of suicide completers, respectively. The observation of widespread astrocytic gene downregulation could conceivably reflect a loss of astrocytes. This would be consistent with decreases in densities of GFAP-IR cells reported in prefrontal cortical samples from depressed patients.¹³¹ Alternatively, these changes could occur without cell loss and instead be owing to stress-induced structural changes in astrocytes.¹³⁶ Although the underlying mechanisms remain to be elucidated, either a loss or a significant atrophy of astrocytes within fronto-limbic brain regions undoubtedly signifies that communication is altered within these networks in mood disorders and suicide. This likely affects the immune functions of these glial cells, and certainly compromises their essential role in the alutamatergic tripartite synapse.

Glutamate reuptake is critical for regulating glutamate concentrations in the synaptic cleft and maintaining normal synaptic activity, and thus an impairment in glutamate transport may result in excessive or dysregulated glutamate receptor signaling. Under physiological conditions, astrocytes prevent excitotoxicity by maintaining extracellular glutamate in the micromolar range via the high-affinity glutamate transporters, excitatory amino acid transporter (EAAT) 1 and EAAT2.¹³⁷ However, this balance may be disrupted by oxidative stress or inflammation leading to necrosis and/or apoptosis through excessive stimulation of glutamate receptors.⁴⁹ Several researchers^{42–47} have shown that activated microglial cells release glutamate via the system xc – cystine/ glutamate transporter (Xc⁻) that exchanges cysteine for glutamate, and that this process may be accentuated by inflammationinduced downregulation of the astrocytic EAAT1 and/or reversed function (that is, glutamate release) of EAAT2, potentially leading to excitotoxicity.^{48,49}

A microarray analysis of samples of the ACC (BA24) and the dorsolateral PFC (BA9 and BA46) from people with MDD showed reduced expression of the genes coding for the high-affinity

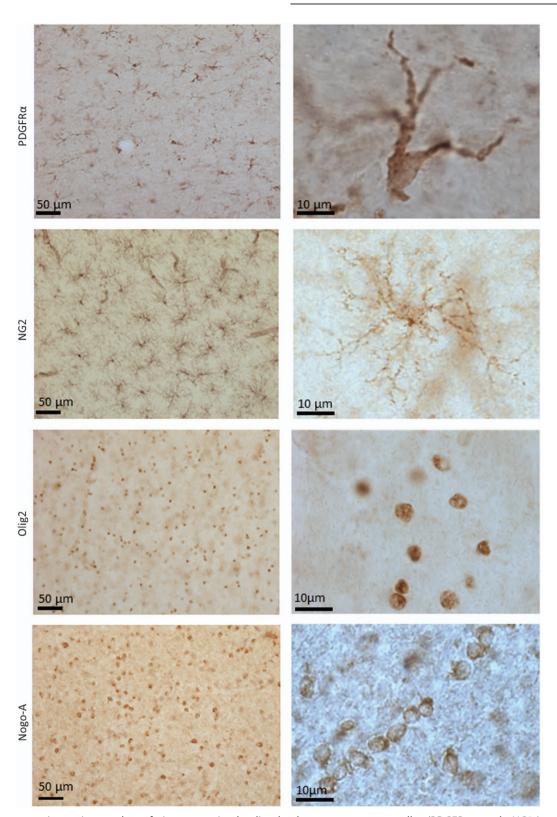


Figure 4. Representative micrographs of immunostained oligodendrocyte precursor cells (PDGFR α - and NG2-immunoreactive), oligodendrocyte-lineage cells (Olig2-immunoreactive) and mature oligodendrocytes (Nogo-A-immunoreactive) in ventromedial prefrontal cortex white matter at low (left column) and high (right column) magnifications. Note the characteristic distribution of Nogo-A+ mature oligodendrocytes along axonal fibers.

glutamate transporters glial excitatory amino acid EAAT1 and EAAT2 along with decreased expression of glutamine synthetase, the enzyme that converts glutamate to glutamine.¹³⁸ These results were subsequently extended to the locus coeruleus¹³⁹ and hippocampus.¹⁴⁰ It is not immediately clear from these studies whether the decrease in glutamate transporter expression was driven by a reduction in glial cell number. However, at least in the case of the locus coeruleus, the depression-associated decrease in EAAT1 and EAAT2 expression was found to be present when expression was normalized to the number of astrocyte cells (but not oligodendrocytes), suggesting an astrocyte-specific dysfunction.¹⁴¹

Regarding BD, the picture appears to be more complicated. Both mRNA and protein levels of EAAT1 were reported to be increased, whereas EAAT2 mRNA and protein levels were decreased relative to controls in samples from the frontal cortex (BA10).¹⁴² As ~ 90% of glutamate is removed from the synapse by EAAT2, Rao et al. hypothesize that their results remain largely consistent with the premise that BD is characterized by hyperglutamatergic signaling.¹⁴² Of note, riluzole, which increases the expression of EAAT2, has shown some efficacy in treatmentresistant depression and as adjunctive therapy for bipolar depression in mostly open-label studies.^{126,143}

Another study found decreased expression of the neuronal transporters, EAAT3 and/or EAAT4 but not EAAT1 or EAAT2 in the striatum of MDD and BD (as well as schizophrenia) samples relative to controls.¹⁴⁴ The decrease in EAAT3 and EAAT4 expression may diminish the capacity of the synapse to clear glutamate, resulting in increased levels of synaptic glutamate in mood disorders.¹⁴⁴ Supporting the potential relevance of EAAT3 function to depression and psychosis, a deletion of the solute carrier family 1 member 1 (SLC1A1) gene that codes for EAAT3 was found to co-segregate with psychosis (BD or schizophrenia) in a large five-generation pedigree, increasing disease risk by 18-fold.¹⁴⁵ Similarly, both rare and more common variants of the genes coding for EAAT1 and EAAT2 have been reported to be associated with BD.¹⁴⁶

Interestingly, autoantibodies to the water channel, aquaporin-4 (AQP4) which are usually associated with neuromyelitis optica, have also been reported in one case of schizophrenia¹⁴⁷ and a case of treatment-resistant depression.¹⁴⁸ EAAT2 and AQP4 exist in astrocytic membranes as a macromolecular complex and the binding of AQP4 IgG has been shown to cause the rapid internalization of both AQP4 and EAAT2.¹⁴⁹

After synaptic uptake into astrocytes by EAATs, glutamate is converted into glutamine by glutamine synthetase and glutamine is, in turn, delivered to neurons where it is re-converted into glutamate. Thus, the expression of glutamine synthetase may affect glutamate cycling and availability to neurons. QA not only inhibits the uptake of glutamate by astrocytes but inhibits astroglial glutamine synthetase¹⁵⁰ perhaps explaining previous reports of decreased glutamine synthetase expression in the amygdala and PFC of suicide completers.^{138,151}

Oligodendroglia

Relative to other cell types, oligodendroglia (Figure 4) are particularly vulnerable to inflammation and/or oxidative stressinduced excitotoxicity during development in both rodents and humans.^{152,153} Lipopolysaccharide is toxic to oligodendrocytes when they are co-cultured with microglia, and in rodents, administration of endotoxin preferentially damages oligodendrocytes; an effect that may be mediated by TLR4 as transgenic mice lacking TLR4, through which endotoxin primarily acts, are less vulnerable to oligodendrocyte damage.^{154,155} Oligodendroglia in turn are capable of modulating immune function by, for instance, producing CCL-2 (previously known as MCP-1) and IL-1 post injury to open up the blood–brain barrier and recruit peripheral immune

Translational Psychiatry (2016), 1–16

cells into the brain parenchyma as well to redirect themselves to areas of demyelination.^{156,157} Interestingly, inflammationassociated white matter damage is associated with pre-term births, and the risk for BD putatively increases monotonically with (shorter) length of gestation.¹⁵⁸ Nevertheless, pre-term birth is more generally associated with a variety psychiatric and neurodevelopmental disorders, and thus the correlation between pre-term birth and risk for mood disorders is not necessary indicative of an underlying inflammatory etiology.

Reductions in numbers or density of glia cells-most likely oligodendroglia cells are one of the most prominent findings in postmortem mood disorder samples.^{1,159} A reduction in the number of glial cells together with an increase in neuronal density initially was found in the subgenual ACC in BD and MDD samples, although the identity of the abnormal glia subtype could not be determined with the nonspecific staining methods applied.¹⁶⁰ These nonspecific glia cell abnormalities were extended to the supragenual ACC, BA9 of the PFC, the amygdala and the entorhinal cortex. Cotter et al. reported a 22% decrease in the glial cell density of layer VI of the supragenual ACC and 30% decrease in the glial cell density in layer V of BA9 in MDD, but not BD, samples.^{161,162} On the other hand, after a stereological analysis of Nissl-stained tissues, Hercher *et al.*¹⁶³ found no difference in glial densities nor in neuronal densities and average neuronal soma size between supracallosal ACC (BA24a) samples from MDD individuals vs matched controls. However, those samples from MDD subjects who had also been alcohol dependent had significantly higher densities of glial cells in this region compared with the samples from controls or from MDD subjects who were not alcohol dependent.¹⁶³ Further, in the BA9 samples derived from BD subjects, Rajkowska et al.¹⁶⁴ did find evidence for a decrease in the density (16-22%) of glial cells in layer III in conjunction with glial cell enlargement. These discrepancies in the literature may be related to various confounders, including treatment with lithium and valproic acid. Indeed, Price and colleagues reported reductions in glial cell density in the amygdala (and to a lesser extent the entorhinal cortex) of MDD samples but not in the BD samples.¹⁶⁵ However, further analysis showed that the reduction in glial cell density was in fact present in the untreated BD cases.¹⁶⁵

Follow-up studies using methods that allowed for the differentiation of oligodendrocytes from other glial cell types, specifically reported reduced numbers of oligodendroglia in layers III and VI of BA9 in BD. 166,167 Similarly, a 19% reduction in oligodendrocyte density in the BD samples that reached trendlevel significance was found in the amygdala,¹⁶⁸ while signs of necrosis or apoptosis-related damage to oligodendrocyte cells have been observed in the BD samples from the caudate.¹⁶⁹ Further, immunoreactivity of myelin basic protein, a surrogate marker of myelination that is expressed by oligodendroglia, was decreased in layer I of the hippocampus in female, but not male subjects with BD.¹⁷⁰ In the case of MDD, the staining of deep white matter in the dorsolateral PFC was reported to be significantly less intense in MDD subjects compared with controls.¹⁷¹ Subsequent studies using specific staining methods or flow activated cell sorting methods, reported reduced numbers of oligodendroglia cells in the amygdala¹⁶⁸ and frontopolar cortex.1

These morphological studies receive support from studies that have measured oligodendrocyte cell-related gene and protein expression changes in mood disorders. For instance, Bahn and colleagues performed a quantitative PCR analysis of BA 9 tissue, finding a significant reduction in mRNA expression of key protein markers of myelination and oligodendrocyte function.¹⁷³ Specifically, the expression of proteolipid protein 1, myelin associated glycoprotein, oligodendrocyte specific protein, myelin oligodendrocyte glycoprotein, and transferrin (TF) was reduced by 2- to 4-fold in the BD patients relative to psychiatrically healthy controls,

while the transcription factors, OLIG2 and SOX10, which are involved in oligodendrocyte differentiation and maturation, were downregulated by 2- to 3-fold in BD.¹⁷³ Although this decrease in oligodendrocyte-related gene expression conceivably could result from cell loss, not all of the oligodendrocyte-related genes were downregulated, suggesting that these abnormalities are more likely to reflect cellular dysfunction than cell death.¹⁷³

Because oligodendrocytes are responsible for the myelination of axons, the abnormalities of oligodendroglia observed postmortem appear congruent with the results of diffusion tensor imaging studies, which have produced evidence of structural abnormalities of white matter tracts connecting the prefrontal cortex with limbic nuclei in MDD¹⁷⁴ and BD.^{175–177} Interestingly, in patients with BD, a recent study reported a significant positive association between several pro-inflammatory cytokines and radial diffusivity, and an inverse association between these cytokines and fractional anisotropy, leading Benedetti et al.¹⁷⁸ to hypothesize that inflammation is associated with demyelination or dysmyelination in BD. Oligodendrocytes are also involved in the turnover of N-acetylaspartate, a marker of neuronal integrity. Males with BD were reported to display a reduced concentration of (N-acetylaspartate), in the dorsolateral PFC (BA8, BA9, BA10 and BA46; ref. 179) and similar findings have been published in the pediatric BD literature.^{180,181} Consistent with these data, *N*acetylaspartate levels increased significantly in MDD patients after treatment with ECT.¹⁸²

Reductions in numbers or density of oligodendroglia in mood disorder samples, postmortem, also receives support from the preclinical literature. In mice, ablation of nerve/glial antigen 2 (NG2)-expressing oligodendrocytes or a chronic social stressinduced decrease in their density, leads to an impairment of astrocytic glutamate reuptake and depressive-like behaviors which can be rescued by repopulation with endogenous oligodendrocyte progenitor cells.¹⁸³

Given the evidence discussed above for mood disorders, it is tempting to draw parallels with the psychopathological features observed in multiple sclerosis (MS). Indeed, MS is a chronic neuroinflammatory disorder that is characterized by a compromised blood-brain barrier, infiltration of immune cells, and loss of myelin and oligodendrocytes.¹⁸⁴ Interestingly, MDD co-morbidity is high in MS, with a lifetime prevalence rate of about 50%, that is, 3- to 5-fold that of the general population.¹⁸⁵ Similarly, BD in MS patients is twice as frequent as in the general population.185 Furthermore, MS patients can also present pseudobulbar affect (emotional incontinence), with prevalence estimates varying widely in the literature (6.5–95%).¹⁸⁶ Pseudobulbar affect, which is often misdiagnosed as MDD or BD, is characterized by inappropriate and uncontrolled crying and/or laughing, and is thought to result from a dishinibition within the corticopontinecerebellar circuits.¹⁸⁷ Given these epidemiological and clinical observations, it can be speculated that the altered connectivity arising from neuroinflammation-related oligodendroglial loss and myelination deficits may represent a common path leading to disorders of mood and affect.

Neurons

The NMDA receptor complex. The NMDA receptor is membranebound ligand-gated Ca²⁺ channel that is assembled from four or five subunits with an obligatory NR1 subunit associated with different combinations of NR2A–D subunits. The NR1 gene is alternatively spliced, producing eight different isoforms of the NR1 subunit, which combine with different NR2 and NR3 subunits to produce NMDA receptors with distinct pharmacological properties. Activation of the receptor requires multiple signals, that is, glutamate binding, AMPA receptor-mediated depolarization of the postsynaptic membrane, and binding of glycine or D-serine to the NR1 subunit.¹⁸⁸ The NMDA receptors are found both in the synapse and in extrasynaptic locations on neurons but are predominantly expressed in the postsynaptic membrane where they interact with an intracellular protein complex termed the postsynaptic density (PSD).

Pro-inflammatory cytokines such as IL-1, IL-6 and TNF can enhance the release of glutamate from presynaptic neurons and increase NMDA receptor currents in postsynaptic neurons potentially leading to excitotoxicity.¹⁸⁹ For instance, IL-1 can enhance NMDA signaling by inducing NR1 (ref. 190) and NR2B (ref. 191) subunit phosphorylation. In neuronal cell cultures, the administration of IFN-a caused dendritic atrophy, an effect that was partially mediated by the NR2A receptor.¹⁹² Similarly, using whole-cell patch-clamp electrophysiological recordings, Di Filippo et al.¹⁹³ reported that IFN- β , a medication that blocks type I interferon receptors and is used to treat infectious and autoimmune diseases such as MS, reduces striatal excitatory postsynaptic potentials through the NR2A NMDA receptor subunit. Further, prenatal inhibition of the kynurenine pathway enzyme, kynurenine monooxygenase, was shown to cause a significant increase in levels of NR2A, NR2B and the postsynaptic NMDA receptor complex protein postsynaptic density protein 95 (PSD-95) at postnatal day 21 together with neuronal excitability and long-term potentiation in the hippocampus.¹⁹⁴

Regarding postmortem studies, a western blot analysis showed approximately 50% reductions in NR2A, NR2B (but not NR1) as well as the anchoring protein, PSD-95, in the anterior region of the PFC of MDD samples.¹⁹⁵ In contrast, elevations in NR2A and NR2C and PSD-95 were reported by the same group to be elevated in the locus coeruleus and lateral amygdala, respectively.^{196,197} A more recent study using quantitative PCR in a large sample of 53 MDD subjects and 32 controls reported increased expression of all the NMDA receptor subunit genes in BA9/BA46 of depressed females, an effect that was accentuated in the individuals who had committed suicide.¹⁹⁸

In BD, in situ hybridization studies generally have reported decreased expression of NR1. Specifically, decreased expression of NR1 was observed in the Cornu Ammonis 3 (CA3) and hippocampal subiculum of BD samples with a history of psychosis,¹⁹⁹ while decreased expression of synapse- associated protein 102 (SAP102), NR1 and NR2A (but not NR2B, NR2C and NR2D) was independently reported in hippocampal samples from individuals with BD (but not schizophrenia) relative to controls.²⁰⁰ Partially consistent with these data, NR1 (but no other NMDA subunit) expression was decreased in the oriens layer of CA1 but not CA2/3.²⁰¹ Regarding the frontal cortex, NR1 expression was reported to be decreased in the dorsolateral PFC of MDD, BD and schizophrenia samples vs controls.²⁰² Similarly, decreased mRNA and protein levels of NR1 and NR3A (but not NR2A or NR2B) together with increased concentrations of inflammatory and excitotoxicity markers such as IL-1, nuclear factor-kB and inducible nitric oxide synthase were reported in BD samples relative to controls.¹⁰⁸

In aggregate, this literature is suggestive of abnormal glutamatergic signaling in depression. Although the changes in expression of the NMDA receptor subunits could conceivably result from the direct actions of pro-inflammatory cytokines, there may be many other factors driving these changes such as compensatory up- or downregulation of receptors as a result of astrocyte dysfunctionmediated decreases or increases in glutamate levels.

Vesicular glutamate transporters. The kynurenine metabolite, xanthurenic acid, which is produced by the transamination of 3HK (Figure 1) and is upregulated in the brain by lipopolysaccharide,²⁰³ is an endogenous inhibitor of vesicular glutamate transporters (VGLUT).²⁰⁴ The vesicular glutamate transporters are responsible for loading glutamate into synaptic vesicles at excitatory neurons thereby regulating presynaptic glutamate release.²⁰⁵ VGLUT1 primarily is expressed in the

cerebral cortex, hippocampus and cerebellar cortex, whereas VGLUT2 transcripts are expressed subcortically (including in dopamine neurons) and in layer V of the cerebral cortex.^{206,207} VGLUT3 is expressed in the subsets of serotonergic, cholinergic and GABAergic interneurons.²⁰⁶

Eastwood and Harrison reported increased VGLUT1 expression in the supragenual ACC (BA24) in BD, a result they hypothesize is indicative of abnormally increased glutamate neurotransmission, rather than, or in addition to, alterations in glutamate metabolism or cycling.²⁰⁸ Consistent with these data, Gray *et al.*¹⁹⁸ reported increased VGLUT1 expression in the dorsolateral PFC of female MDD samples using quantitative PCR. This abnormality may be region-specific since decreased expression of VGLUT1 also has been reported in mood disorders. Using in situ hybridization, Uezato et al.²⁰⁶ reported decreased VGLUT1 mRNA expression in both MDD and BD in the entorhinal cortex as well as decreased VGLUT2 expression in MDD, (trending for BD) in the middle temporal gyrus. Consistent with these data, decreased expression of VGLUT1 was reported in layer V of both MDD and BD samples from the dorsolateral PFC (BA9).²⁰⁹ A reduction in VGLUT1/2 mRNA may be due to structural loss of presynaptic terminals or alternatively a functional decrease in glutamate release while conversely, increased VGLUT expression may be indicative of greater numbers of glutamate neurons and/or greater presynaptic innervation and glutamate neurotransmission.

GABAergic interneurons. Structural and/or functional changes to GABAergic interneurons are one of the most robust postmortem findings in BD.¹ For instance, in the rostral ACC, Woo et al.²¹⁰ found a reduction in the density of NR2A-expressing GABAergic interneurons in subjects with BD (and schizophrenia) relative to controls and in a separate study, the density of GABAergic neurons expressing the GluR5 subunit of the kainate receptor was found to be decreased by 40% in layer II of this region.²¹¹ Similarly, the density of calbindin (CB)-expressing GABAergic neurons in layer II of the supracallosal gyrus was reduced by 33% in BD²¹² and in the dorsolateral cortex, the density of glutamic acid decarboxylase 67 (GAD₆₇) mRNA-containing (GABAergic) neurons in layers II through V of BA9 was decreased by approximately 25-33%.²¹³ These data are supported by additional studies reporting decreased expression of GABAergic neuron-associated proteins such as GAD_{67} in both cortical and subcortical regions, $^{214-217}$ as well as a meta-analysis showing a reduction in the density of CBexpressing neurons in layer VII of BA9 in BD.²¹⁸ Similarly, MDD appears to be characterized by reduced density of GABAergic interneurons labeled with calretinin and/or CB in the dorsolateral PFC,²¹⁹ occipital cortex²²⁰ and auditory cortex.²²¹

These data suggest that GABAergic neurotransmission in at least a subset of local neuronal circuits is attenuated; however, the functional implications are not well understood. In addition to disinhibition of pyramidal neurons, other studies demonstrate that parvalbumin-containing interneurons are critical for the generation of gamma oscillations,^{222,223} which are implicated in normal cortical function (for example, working memory²²⁴). Expression of mRNA for neuronal activity-regulated pentraxin, a protein that is secreted at presynaptic glutamate synapses that terminate on parvalbumin-containing interneurons, is reduced in the dorsolateral PFC of subjects with BD,²²⁵ suggesting that excitatory drive onto this interneuron subclass is disrupted which, in turn, could lead to a disruption of gamma oscillations and associated cortical function.

The mechanisms through which inflammation specifically affects the structure and/or function of GABAergic interneurons is not yet clear. However, there are preclinical data showing that inflammation impacts GABAergic circuits. Multiple rodent studies have reported that prenatal exposure to lipopolysaccharide or polyinosinic: polycytidylic acid (poly I:C) is associated with a reduction in the density of parvalbumin and/or GAD₆₇.staining

neurons in the hippocampus (reviewed in ref. 226). Potentially consistent with these data, Clements *et al.*²²⁷ found a significant reduction in parvalbumin-staining interneurons in layer II of the motor cortex in MS patients vs controls. Stress may have a modulating role in the relationship between inflammation and GABAergic function. For instance, maternal separation stress in rats causes a reduction of GABAergic parvalbumin-expressing interneurons in the PFC; an effect that was reported to be blocked by intracerebroventricular administration of the anti-inflammatory cytokine, IL-10.²²⁸ Similarly, adult offspring born to viral mimetic poly(I:C)-exposed mothers who were subjected to unpredictable subchronic stress during peripubertal development displayed significant reductions of parvalbumin-expressing interneurons in the dentate gyrus.²²⁹

In a different vein, *T. gondii* infection was shown to shift the distribution of GAD₆₇ expression such that GAD₆₇ becomes diffusely located throughout the neuropil rather than clustering on presynaptic termini where it catalyzes GABA synthesis in the brain.²³⁰

CAVEATS

Cause or effect?

Although we have emphasized the potentially detrimental effects of neuroinflammation on the brain, microglial cells also exert neuroprotective effects. Toll-like receptor signaling caused by cellular damage or infectious agents triggers a rapid activation of microglial cells, clearing away debris, promoting angiogenesis, neurogenesis, increasing the recruitment of oligodendrocyte progenitor cells, favoring remyelination and conferring neuroprotection.²³¹ However, without the rapid clearance of myelin debris or toxins from the brain, microglial cells may instead mount a detrimental pro-inflammatory response, resulting in demyelination, synaptic dysfunction and ultimately neurodegeneration.^{231–233} Thus a delicate equilibrium between under- and overactivation of microglial cells may determine whether the conditions conducive to neuronal repair or neuronal damage, predominate. There are likely to be many mechanisms beyond removal of debris and oligodendroglia-genesis underlying these neuroprotective effects. For instance, activated macrophages and microglial cells express high-affinity glutamate transporters and glutamine synthetase during both acute and chronic central nervous system inflammation, potentially partially compensating for astrocytic dysfunction.²³⁴ Mice given intraperitoneal injections of 1.0 mg kg⁻¹ of endotoxin for four consecutive days to globally activate central nervous system microglia, are protected from a subsequent traumatic brain injury challenge²³⁵ by microglia-mediated stripping of inhibitory GABAergic synapses, which facilitates NMDA receptor signaling allowing for greater expression of neurotrophic and anti-apoptotic molecules such as brain-derived neurotrophic factor and B-cell lymphoma 2 (Bcl-2).²³⁶ Thus conceivably, the increased expression of glutamaterelated genes together with the decreased expression of GABA observed in postmortem tissue in BD may be an adaptive response to injury rather than a cause of neuronal pathology.

Misattribution of function

The assumption that molecules that are involved in the peripheral immune response automatically play the same functional role in the brain is potentially faulty. A case in point is the recent discovery that a variant in the gene coding for a component of the complement (CA4) pathway constitutes a risk factor for schizophrenia.²³⁷ Rather than (or at least in addition to) defending against microbial infection, CA4 in the brain appears to have a key role in synaptic pruning during neurodevelopment.²³⁷ Thus postmortem alterations in inflammation-related proteins do not

necessarily reflect an immune-related etiology—at least in the classical sense of the word.

CONCLUSION AND FUTURE DIRECTIONS

The most prominent morphological abnormalities observed in postmortem samples from individuals with BD include decreases in the density and/or number of GABAergic neurons and glial cells. The literature describing microglial activation in mood disorders is smaller but appears robust. These data are complemented by gene expression studies, which are consistent with functional deficits in myelination, astrocytic glutamate recycling and in the regulation of inhibitory/excitatory neurotransmission. Immune processes could conceivably account for these abnormalities, but with the possible exception of the microglia data it is not possible to draw a direct link between neuroinflammation and histopathology. It is likely that there is a great deal of heterogeneity within the MDD and BD syndromes with respect to pathophysiology and etiology such that postmortem samples encompass patients with a range of conditions that appear clinically related but are neurobiologically distinct. This lack of a precise and biologically verifiable definition of illness, together with contrasting experimental approaches presumably contribute to the inconsistencies observed within the literature. Future studies should give greater consideration to these factors to better clarify the possible contribution of neuroinflammation to the etiology of mood disorders.

It will be particularly importance for future postmortem studies to be carried out using thoroughly characterized samples, that is, case and control tissues for which substantial clinical information is available and complemented retrospectively with psychosocial data. Moreover, in addition to common tissue-based immunohistochemical and molecular approaches, the field should increasingly take advantage of techniques allowing for single cell analyses, such as laser capture microdissection and fluorescenceactivated cell sorting. These approaches offer the opportunity to explore and compare, within a given brain region or circuit, the epigenetic and molecular profiles of different cellular subtypes (neurons vs glia, subtypes of neurons and glia, macrophages and so on). Such characterizations should lead to a more detailed understanding of the causes and consequences of cerebral neuroinflammation in mood disorders.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

NM, a CIHR New Investigator and FRQ-S scholar, is supported by CIHR grant MOP-111022 and by an ERA-NET NEURON (FRQ-S) team grant. JS receives support from the NIMH (K01MH096077) and the Laureate Institute for Brain Research. We thank Dr Susana Gabriela Torres-Platas for supplying the image of the golgi-stained astrocytes (Figure 3) and Dr Arnaud Tanti for the oligodendrocyte cell micrographs (Figure 4).

REFERENCES

- 1 Savitz JB, Price JL, Drevets WC. Neuropathological and neuromorphometric abnormalities in bipolar disorder: view from the medial prefrontal cortical network. *Neurosci Biobehav Rev* 2014; **42**: 132–147.
- 2 Weinberger DR, Radulescu E. Finding the elusive psychiatric "Lesion" with 21stcentury neuroanatomy: a note of caution. *Am J Psychiatry* 2016; **173**: 27–33.
- 3 Zipursky RB, Reilly TJ, Murray RM. The myth of schizophrenia as a progressive brain disease. *Schizophr Bull* 2013; **39**: 1363–1372.
- 4 Dargel AA, Godin O, Kapczinski F, Kupfer DJ, Leboyer M. C-reactive protein alterations in bipolar disorder: a meta-analysis. *J Clin Psychiatry* 2015; **76**: 142–150.

- 5 Munkholm K, Brauner JV, Kessing LV, Vinberg M. Cytokines in bipolar disorder vs. healthy control subjects: a systematic review and meta-analysis. J Psychiatr Res 2013; 47: 1119–1133.
- 6 Modabbernia A, Taslimi S, Brietzke E, Ashrafi M. Cytokine alterations in bipolar disorder: a meta-analysis of 30 studies. *Biol Psychiatry* 2013; 74: 15–25.
- 7 Haapakoski R, Mathieu J, Ebmeier KP, Alenius H, Kivimaki M. Cumulative metaanalysis of interleukins 6 and 1beta, tumour necrosis factor alpha and C-reactive protein in patients with major depressive disorder. *Brain Behav Immun* 2015; **49**: 206–215.
- 8 Dowlati Y, Herrmann N, Swardfager W, Liu H, Sham L, Reim EK et al. A metaanalysis of cytokines in major depression. Biol Psychiatry 2010; 67: 446–457.
- 9 Fiedorowicz JG, Prossin AR, Johnson CP, Christensen GE, Magnotta VA, Wemmie JA. Peripheral inflammation during abnormal mood states in bipolar I disorder. J Affect Disord 2015; 187: 172–178.
- 10 Munkholm K, Weikop P, Kessing LV, Vinberg M. Elevated levels of IL-6 and IL-18 in manic and hypomanic states in rapid cycling bipolar disorder patients. *Brain Behav Immun* 2015; **43**: 205–213.
- 11 Padmos RC, Hillegers MH, Knijff EM, Vonk R, Bouvy A, Staal FJ et al. A discriminating messenger RNA signature for bipolar disorder formed by an aberrant expression of inflammatory genes in monocytes. Arch Gen Psychiatry 2008; 65: 395–407.
- 12 Drexhage RC, Hoogenboezem TH, Versnel MA, Berghout A, Nolen WA, Drexhage HA. The activation of monocyte and T cell networks in patients with bipolar disorder. *Brain Behav Immun* 2011; 25: 1206–1213.
- 13 Pandey GN, Ren X, Rizavi HS, Zhang H. Abnormal gene expression of proinflammatory cytokines and their receptors in the lymphocytes of patients with bipolar disorder. *Bipolar Disord* 2015; **17**: 636–644.
- 14 Savitz J, Frank MB, Victor T, Bebak M, Marino JH, Bellgowan PS *et al.* Inflammation and neurological disease-related genes are differentially expressed in depressed patients with mood disorders and correlate with morphometric and functional imaging abnormalities. *Brain Behav Immun* 2013; **31**: 161–171.
- 15 Rizavi HS, Ren X, Zhang H, Bhaumik R, Pandey GN. Abnormal gene expression of proinflammatory cytokines and their membrane-bound receptors in the lymphocytes of depressed patients. *Psychiatry Res* 2016; 240: 314–320.
- 16 Teixeira AL, de Sousa RT, Zanetti MV, Brunoni AR, Busatto GF, Zarate CA, Jr et al. Increased plasma levels of soluble TNF receptors 1 and 2 in bipolar depression and impact of lithium treatment. Hum Psychopharmacol 2015; 30: 52–56.
- 17 Bai YM, Su TP, Li CT, Tsai SJ, Chen MH, Tu PC *et al.* Comparison of proinflammatory cytokines among patients with bipolar disorder and unipolar depression and normal controls. *Bipolar Disord* 2015; **17**: 269–277.
- 18 Morch RH, Dieset I, Faerden A, Hope S, Aas M, Nerhus M et al. Inflammatory evidence for the psychosis continuum model. *Psychoneuroendocrinology* 2016; 67: 189–197.
- 19 Wium-Andersen MK, Orsted DD, Nordestgaard BG. Elevated C-reactive protein and late-onset bipolar disorder in 78 809 individuals from the general population. Br J Psychiatry 2015.
- 20 Pasco JA, Nicholson GC, Williams LJ, Jacka FN, Henry MJ, Kotowicz MA et al. Association of high-sensitivity C-reactive protein with *de novo* major depression. Br J Psychiatry 2010; **197**: 372–377.
- 21 Khandaker GM, Pearson RM, Zammit S, Lewis G, Jones PB. Association of serum interleukin 6 and C-reactive protein in childhood with depression and psychosis in young adult life: a population-based longitudinal study. *JAMA Psychiatry* 2014; **71**: 1121–1128.
- 22 Tully PJ, Baumeister H, Bengel J, Jenkins A, Januszewski A, Martin S *et al.* The longitudinal association between inflammation and incident depressive symptoms in men: the effects of hs-CRP are independent of abdominal obesity and metabolic disturbances. *Physiol Behav* 2015; **139**: 328–335.
- 23 Capuron L, Raison CL, Musselman DL, Lawson DH, Nemeroff CB, Miller AH. Association of exaggerated HPA axis response to the initial injection of interferon-alpha with development of depression during interferon-alpha therapy. Am J Psychiatry 2003; 160: 1342–1345.
- 24 Capuron L, Hauser P, Hinze-Selch D, Miller AH, Neveu PJ. Treatment of cytokineinduced depression. *Brain Behav Immun* 2002; 16: 575–580.
- 25 Wichers M, Maes M. The psychoneuroimmuno-pathophysiology of cytokineinduced depression in humans. Int J Neuropsychopharmacol 2002; 5: 375–388.
- 26 Capuron L, Miller AH. Cytokines and psychopathology: lessons from interferon-alpha. *Biol Psychiatry* 2004; 56: 819–824.
- 27 Capuron L, Miller AH. Immune system to brain signaling: neuropsychopharmacological implications. *Pharmacol Ther* 2011; **130**: 226–238.
- 28 Sandiego CM, Gallezot JD, Pittman B, Nabulsi N, Lim K, Lin SF et al. Imaging robust microglial activation after lipopolysaccharide administration in humans with PET. Proc Natl Acad Sci USA 2015; 112: 12468–12473.
- 29 Eisenberger NI, Berkman ET, Inagaki TK, Rameson LT, Mashal NM, Irwin MR. Inflammation-induced anhedonia: endotoxin reduces ventral striatum responses to reward. *Biol Psychiatry* 2010; **68**: 748–754.

- 30 Hannestad J, Subramanyam K, Dellagioia N, Planeta-Wilson B, Weinzimmer D, Pittman B et al. Glucose metabolism in the insula and cingulate is affected by systemic inflammation in humans. J Nucl Med 2012; 53: 601–607.
- 31 Harrison NA, Brydon L, Walker C, Gray MA, Steptoe A, Critchley HD. Inflammation causes mood changes through alterations in subgenual cingulate activity and mesolimbic connectivity. *Biol Psychiatry* 2009; 66: 407–414.
- 32 Ayorech Z, Tracy DK, Baumeister D, Giaroli G. Taking the fuel out of the fire: evidence for the use of anti-inflammatory agents in the treatment of bipolar disorders. J Affect Disord 2015; 174: 467–478.
- 33 Berk M, Copolov DL, Dean O, Lu K, Jeavons S, Schapkaitz I et al. N-acetyl cysteine for depressive symptoms in bipolar disorder--a double-blind randomized placebo-controlled trial. Biol Psychiatry 2008; 64: 468–475.
- 34 Savitz J, Preskorn S, Teague TK, Drevets D, Yates W, Drevets W. Minocycline and aspirin in the treatment of bipolar depression: a protocol for a proof-of-concept, randomised, double-blind, placebo-controlled, 2x2 clinical trial. *BMJ Open* 2012; 2: e000643.
- 35 Raison CL, Rutherford RE, Woolwine BJ, Shuo C, Schettler P, Drake DF et al. A randomized controlled trial of the tumor necrosis factor antagonist infliximab for treatment-resistant depression: the role of baseline inflammatory biomarkers. JAMA Psychiatry 2013; 70: 31–41.
- 36 Setiawan E, Wilson AA, Mizrahi R, Rusjan PM, Miler L, Rajkowska G et al. Role of translocator protein density, a marker of neuroinflammation, in the brain during major depressive episodes. JAMA Psychiatry 2015; 72: 268–275.
- 37 Haarman BC, Riemersma-Van der Lek RF, de Groot JC, Ruhe HG, Klein HC, Zandstra TE et al. Neuroinflammation in bipolar disorder - A [(11)C]-(R)-PK11195 positron emission tomography study. Brain Behav Immun 2014; 40: 219–225.
- 38 Miller AH, Raison CL. The role of inflammation in depression: from evolutionary imperative to modern treatment target. *Nat Rev Immunol* 2015; **16**: 22–34.
- 39 Harrison NA, Doeller CF, Voon V, Burgess N, Critchley HD. Peripheral inflammation acutely impairs human spatial memory via actions on medial temporal lobe glucose metabolism. *Biol Psychiatry* 2014; 76: 585–593.
- 40 Capuron L, Pagnoni G, Drake DF, Woolwine BJ, Spivey JR, Crowe RJ et al. Dopaminergic mechanisms of reduced basal ganglia responses to hedonic reward during interferon alfa administration. Arch Gen Psychiatry 2012; 69: 1044–1053.
- 41 Felger JC, Li Z, Haroon E, Woolwine BJ, Jung MY, Hu X et al. Inflammation is associated with decreased functional connectivity within corticostriatal reward circuitry in depression. *Mol Psychiatry* 2016; 21: 1358.
- 42 Piani D, Spranger M, Frei K, Schaffner A, Fontana A. Macrophage-induced cytotoxicity of N-methyl-D-aspartate receptor positive neurons involves excitatory amino acids rather than reactive oxygen intermediates and cytokines. *Eur J Immunol* 1992; 22: 2429–2436.
- 43 Takaki J, Fujimori K, Miura M, Suzuki T, Sekino Y, Sato K. L-glutamate released from activated microglia downregulates astrocytic L-glutamate transporter expression in neuroinflammation: the 'collusion' hypothesis for increased extracellular L-glutamate concentration in neuroinflammation. J Neuroinflammation 2012; 9: 275.
- 44 Evonuk KS, Baker BJ, Doyle RE, Moseley CE, Sestero CM, Johnston BP et al. Inhibition of system Xc(-) transporter attenuates autoimmune inflammatory demyelination. J Immunol 2015; 195: 450–463.
- 45 Zhao W, Xie W, Le W, Beers DR, He Y, Henkel JS et al. Activated microglia initiate motor neuron injury by a nitric oxide and glutamate-mediated mechanism. J Neuropathol Exp Neurol 2004; 63: 964–977.
- 46 Tilleux S, Hermans E. Down-regulation of astrocytic GLAST by microglia-related inflammation is abrogated in dibutyryl cAMP-differentiated cultures. *J Neurochem* 2008; **105**: 2224–2236.
- 47 Kigerl KA, Ankeny DP, Garg SK, Wei P, Guan Z, Lai W *et al.* System x(c)(-) regulates microglia and macrophage glutamate excitotoxicity *in vivo. Exp Neurol* 2012; 233: 333–341.
- 48 Dantzer R, Walker AK. Is there a role for glutamate-mediated excitotoxicity in inflammation-induced depression? *J Neural Transm (Vienna)* 2014; **121**: 925–932.
- 49 Fontana AC. Current approaches to enhance glutamate transporter function and expression. J Neurochem 2015; 134: 982–1007.
- 50 Dantzer R, O'Connor JC, Lawson MA, Kelley KW. Inflammation-associated depression: from serotonin to kynurenine. *Psychoneuroendocrinology* 2011; 36: 426–436.
- 51 Kessler M, Terramani T, Lynch G, Baudry M. A glycine site associated with N-methyl-D-aspartic acid receptors: characterization and identification of a new class of antagonists. *J Neurochem* 1989; **52**: 1319–1328.
- 52 Schwarcz R, Whetsell WO Jr, Mangano RM. Quinolinic acid: an endogenous metabolite that produces axon-sparing lesions in rat brain. *Science* 1983; **219**: 316–318.
- 53 Stone TW. Kynurenines in the CNS: from endogenous obscurity to therapeutic importance. *Prog Neurobiol* 2001; **64**: 185–218.

- 54 Platenik J, Stopka P, Vejrazka M, Stipek S. Quinolinic acid-iron(ii) complexes: slow autoxidation, but enhanced hydroxyl radical production in the Fenton reaction. *Free Rad Res* 2001; **34**: 445–459.
- 55 Tavares RG, Tasca CI, Santos CE, Wajner M, Souza DO, Dutra-Filho CS. Quinolinic acid inhibits glutamate uptake into synaptic vesicles from rat brain. *Neuroreport* 2000; **11**: 249–253.
- 56 Rahman A, Ting K, Cullen KM, Braidy N, Brew BJ, Guillemin GJ. The excitotoxin quinolinic acid induces tau phosphorylation in human neurons. *PLoS One* 2009; 4: e6344.
- 57 Guillemin GJ, Croitoru-Lamoury J, Dormont D, Armati PJ, Brew BJ. Quinolinic acid upregulates chemokine production and chemokine receptor expression in astrocytes. *Glia* 2003; **41**: 371–381.
- 58 Bay-Richter C, Linderholm KR, Lim CK, Samuelsson M, Traskman-Bendz L, Guillemin GJ et al. A role for inflammatory metabolites as modulators of the glutamate N-methyl-d-aspartate receptor in depression and suicidality. Brain Behav Immun 2015; 43: 110–117.
- 59 Savitz J, Dantzer R, Wurfel BE, Victor TA, Ford BN, Bodurka J et al. Neuroprotective kynurenine metabolite indices are abnormally reduced and positively associated with hippocampal and amygdalar volume in bipolar disorder. *Psychoneuroendocrinology* 2015; **52**: 200–211.
- 60 Savitz J, Drevets WC, Smith CM, Victor TA, Wurfel BE, Bellgowan PS *et al.* Putative neuroprotective and neurotoxic kynurenine pathway metabolites are associated with hippocampal and amygdalar volumes in subjects with major depressive disorder. *Neuropsychopharmacology* 2015; **40**: 463–471.
- 61 Meier TB, Savitz J, Singh R, Teague TK, Bellgowan PS. Smaller dentate gyrus and CA2 and CA3 volumes are associated with kynurenine metabolites in collegiate football athletes. J Neurotrauma 2016; 33: 1349–1357.
- 62 Glaser R, Kiecolt-Glaser JK. Stress-induced immune dysfunction: implications for health. *Nat Rev Immunol* 2005; **5**: 243–251.
- 63 Cole SW, Levine ME, Arevalo JM, Ma J, Weir DR, Crimmins EM. Loneliness, eudaimonia, and the human conserved transcriptional response to adversity. *Psychoneuroendocrinology* 2015; **62**: 11–17.
- 64 Cole SW. Human social genomics. PLoS Genet 2014; 10: e1004601.
- 65 Cole SW, Hawkley LC, Arevalo JM, Cacioppo JT. Transcript origin analysis identifies antigen-presenting cells as primary targets of socially regulated gene expression in leukocytes. *Proc Natl Acad Sci USA* 2011; **108**: 3080–3085.
- 66 Nagaraja AS, Sadaoui NC, Dorniak PL, Lutgendorf SK, Sood AK. SnapShot: stress and disease. *Cell Metab* 2016; **23**: 388–388 e381.
- 67 Schillinger JA, Xu F, Sternberg MR, Armstrong GL, Lee FK, Nahmias AJ *et al.* National seroprevalence and trends in herpes simplex virus type 1 in the United States, 1976-1994. *Sex Transm Dis* 2004; **31**: 753–760.
- 68 Colugnati FA, Staras SA, Dollard SC, Cannon MJ. Incidence of cytomegalovirus infection among the general population and pregnant women in the United States. BMC Infect Dis 2007; 7: 71.
- 69 Glaser R, Kiecolt-Glaser JK, Speicher CE, Holliday JE. Stress, loneliness, and changes in herpesvirus latency. *J Behav Med* 1985; **8**: 249–260.
- 70 Miller GE, Stetler CA, Carney RM, Freedland KE, Banks WA. Clinical depression and inflammatory risk markers for coronary heart disease. *Am J Cardiol* 2002; **90**: 1279–1283.
- 71 Appels A, Bar FW, Bar J, Bruggeman C, de Baets M. Inflammation, depressive symptomtology, and coronary artery disease. *Psychosom Med* 2000; 62: 601–605.
- 72 Rector JL, Dowd JB, Loerbroks A, Burns VE, Moss PA, Jarczok MN *et al.* Consistent associations between measures of psychological stress and CMV antibody levels in a large occupational sample. *Brain Behav Immun* 2014; **38**: 133–141.
- 73 Simanek AM, Cheng C, Yolken R, Uddin M, Galea S, Aiello AE. Herpesviruses, inflammatory markers and incident depression in a longitudinal study of Detroit residents. *Psychoneuroendocrinology* 2014; **50**: 139–148.
- 74 Yolken RH, Torrey EF. Are some cases of psychosis caused by microbial agents? A review of the evidence. *Mol Psychiatry* 2008; **13**: 470–479.
- 75 Torrey EF, Bartko JJ, Lun ZR, Yolken RH. Antibodies to Toxoplasma gondii in patients with schizophrenia: a meta-analysis. Schizophr Bull 2007; 33: 729–736.
- 76 Sugden K, Moffitt TE, Pinto L, Poulton R, Williams BS, Caspi A. Is *Toxoplasma gondii* infection related to brain and behavior impairments in humans? Evidence from a population-representative birth cohort. *PLoS One* 2016; **11**: e0148435.
- 77 Arling TA, Yolken RH, Lapidus M, Langenberg P, Dickerson FB, Zimmerman SA et al. Toxoplasma gondii antibody titers and history of suicide attempts in patients with recurrent mood disorders. J Nerv Ment Dis 2009; **197**: 905–908.
- 78 Duffy AR, Beckie TM, Brenner LA, Beckstead JW, Seyfang A, Postolache TT et al. Relationship between Toxoplasma gondii and mood disturbance in women veterans. Mil Med 2015; 180: 621–625.
- 79 Dickerson F, Stallings C, Origoni A, Vaughan C, Katsafanas E, Khushalani S et al. Antibodies to *Toxoplasma gondii* in individuals with mania. *Bipolar Disord* 2014; 16: 129–136.
- 80 Carruthers VB, Suzuki Y. Effects of *Toxoplasma gondii* infection on the brain. *Schizophr Bull* 2007; **33**: 745–751.

- 81 Notarangelo FM, Wilson EH, Horning KJ, Thomas MA, Harris TH, Fang Q *et al.* Evaluation of kynurenine pathway metabolism in *Toxoplasma gondii*-infected mice: implications for schizophrenia. *Schizophr Res* 2014; **152**: 261–267.
- 82 Floreani A, Leung PS, Gershwin ME. Environmental basis of autoimmunity. *Clin Rev Allergy Immunol* 2016; **50**: 287–300.
- 83 Korczak DJ, Pereira S, Koulajian K, Matejcek A, Giacca A. Type 1 diabetes mellitus and major depressive disorder: evidence for a biological link. *Diabetologia* 2011; 54: 2483–2493.
- 84 Vonk R, van der Schot AC, Kahn RS, Nolen WA, Drexhage HA. Is autoimmune thyroiditis part of the genetic vulnerability (or an endophenotype) for bipolar disorder? *Biol Psychiatry* 2007; 62: 135–140.
- 85 Eaton WW, Pedersen MG, Nielsen PR, Mortensen PB. Autoimmune diseases, bipolar disorder, and non-affective psychosis. *Bipolar Disord* 2010; 12: 638–646.
- 86 Kosmidis MH, Bozikas VP, Giannouli V, Karavatos A, Fokas K. Familial comorbidity of bipolar disorder and multiple sclerosis: genetic susceptibility, coexistence or causal relationship? *Behav Neurol* 2012; 25: 341–349.
- 87 Davison K. Autoimmunity in psychiatry. Br J Psychiatry 2012; 200: 353-355.
- 88 Benros ME, Waltoft BL, Nordentoft M, Ostergaard SD, Eaton WW, Krogh J et al. Autoimmune diseases and severe infections as risk factors for mood disorders: a nationwide study. JAMA Psychiatry 2013; 70: 812–820.
- 89 Pop VJ, Maartens LH, Leusink G, van Son MJ, Knottnerus AA, Ward AM et al. Are autoimmune thyroid dysfunction and depression related? J Clin Endocrinol Metab 1998; 83: 3194–3197.
- 90 Kupka RW, Nolen WA, Post RM, McElroy SL, Altshuler LL, Denicoff KD *et al.* High rate of autoimmune thyroiditis in bipolar disorder: lack of association with lithium exposure. *Biol Psychiatry* 2002; **51**: 305–311.
- 91 Ching KH, Burbelo PD, Carlson PJ, Drevets WC, ladarola MJ. High levels of Anti-GAD65 and Anti-Ro52 autoantibodies in a patient with major depressive disorder showing psychomotor disturbance. *Journal of neuroimmunology* 2010; 222: 87–89.
- 92 Dalmau J, Lancaster E, Martinez-Hernandez E, Rosenfeld MR, Balice-Gordon R. Clinical experience and laboratory investigations in patients with anti-NMDAR encephalitis. *Lancet Neurol* 2011; **10**: 63–74.
- 93 Kayser MS, Titulaer MJ, Gresa-Arribas N, Dalmau J. Frequency and characteristics of isolated psychiatric episodes in anti-N-methyl-d-aspartate receptor encephalitis. *JAMA Neurol* 2013; **70**: 1133–1139.
- 94 Steiner J, Walter M, Glanz W, Sarnyai Z, Bernstein HG, Vielhaber S et al. Increased prevalence of diverse N-methyl-D-aspartate glutamate receptor antibodies in patients with an initial diagnosis of schizophrenia: specific relevance of IgG NR1a antibodies for distinction from N-methyl-D-aspartate glutamate receptor encephalitis. JAMA Psychiatry 2013; 70: 271–278.
- 95 Bergink V, Armangue T, Titulaer MJ, Markx S, Dalmau J, Kushner SA. Autoimmune encephalitis in postpartum psychosis. Am J Psychiatry 2015; 172: 901–908.
- 96 Lennox BR, Coles AJ, Vincent A. Antibody-mediated encephalitis: a treatable cause of schizophrenia. Br J Psychiatry 2012; 200: 92–94.
- 97 Oberheim NA, Wang X, Goldman S, Nedergaard M. Astrocytic complexity distinguishes the human brain. *Trends Neurosci* 2006; 29: 547–553.
- 98 Oberheim NA, Takano T, Han X, He W, Lin JH, Wang F et al. Uniquely hominid features of adult human astrocytes. J Neurosci 2009; 29: 3276–3287.
- 99 Torres-Platas SG, Nagy C, Wakid M, Turecki G, Mechawar N. Glial fibrillary acidic protein is differentially expressed across cortical and subcortical regions in healthy brains and downregulated in the thalamus and caudate nucleus of depressed suicides. *Mol Psychiatry* 2016; 21: 509–515.
- 100 Sosunov AA, Wu X, Tsankova NM, Guilfoyle E, McKhann GM 2nd, Goldman JE. Phenotypic heterogeneity and plasticity of isocortical and hippocampal astrocytes in the human brain. J Neurosci 2014; 34: 2285–2298.
- 101 Tonelli LH, Stiller J, Rujescu D, Giegling I, Schneider B, Maurer K *et al.* Elevated cytokine expression in the orbitofrontal cortex of victims of suicide. *Acta Psychiatr Scand* 2008; **117**: 198–206.
- 102 Gadani SP, Cronk JC, Norris GT, Kipnis J. IL-4 in the brain: a cytokine to remember. J Immunol 2012; **189**: 4213–4219.
- 103 Yang MS, Ji KA, Jeon SB, Jin BK, Kim SU, Jou I et al. Interleukin-13 enhances cyclooxygenase-2 expression in activated rat brain microglia: implications for death of activated microglia. J Immunol 2006; 177: 1323–1329.
- 104 Pandey GN, Rizavi HS, Ren X, Fareed J, Hoppensteadt DA, Roberts RC et al. Proinflammatory cytokines in the prefrontal cortex of teenage suicide victims. J Psychiatr Res 2012; 46: 57–63.
- 105 Pandey GN, Rizavi HS, Ren X, Bhaumik R, Dwivedi Y. Toll-like receptors in the depressed and suicide brain. J Psychiatr Res 2014; **53**: 62–68.
- 106 Dean B, Tawadros N, Scarr E, Gibbons AS. Regionally-specific changes in levels of tumour necrosis factor in the dorsolateral prefrontal cortex obtained postmortem from subjects with major depressive disorder. J Affect Disord 2010; 120: 245–248.
- 107 Dean B, Gibbons AS, Tawadros N, Brooks L, Everall IP, Scarr E. Different changes in cortical tumor necrosis factor-alpha-related pathways in schizophrenia and mood disorders. *Mol Psychiatry* 2013; **18**: 767–773.

- 108 Rao JS, Harry GJ, Rapoport SI, Kim HW. Increased excitotoxicity and neuroinflammatory markers in postmortem frontal cortex from bipolar disorder patients. *Mol Psychiatry* 2010; **15**: 384–392.
- 109 Shelton RC, Claiborne J, Sidoryk-Wegrzynowicz M, Reddy R, Aschner M, Lewis DA et al. Altered expression of genes involved in inflammation and apoptosis in frontal cortex in major depression. *Mol Psychiatry* 2011; 16: 751–762.
- 110 Elashoff M, Higgs BW, Yolken RH, Knable MB, Weis S, Webster MJ et al. Metaanalysis of 12 genomic studies in bipolar disorder. J Mol Neurosci 2007; 31: 221–243.
- 111 Fillman SG, Sinclair D, Fung SJ, Webster MJ, Shannon Weickert C. Markers of inflammation and stress distinguish subsets of individuals with schizophrenia and bipolar disorder. *Transl Psychiatry* 2014; **4**: e365.
- 112 Clark SM, Pocivavsek A, Nicholson JD, Notarangelo FM, Langenberg P, McMahon RP *et al.* Reduced kynurenine pathway metabolism and cytokine expression in the prefrontal cortex of depressed individuals. *J Psychiatry Neurosci* 2016; **41**: 150226.
- 113 Tremblay ME, Lowery RL, Majewska AK. Microglial interactions with synapses are modulated by visual experience. *PLoS Biol* 2010; **8**: e1000527.
- 114 Guillemin GJ, Microglia Brew BJ. macrophages, perivascular macrophages, and pericytes: a review of function and identification. J Leukoc Biol 2004; 75: 388–397.
- 115 Kettenmann H, Hanisch UK, Noda M, Verkhratsky A. Physiology of microglia. *Physiol Rev* 2011; **91**: 461–553.
- 116 Torres-Platas SG, Comeau S, Rachalski A, Bo GD, Cruceanu C, Turecki G *et al.* Morphometric characterization of microglial phenotypes in human cerebral cortex. *J Neuroinflammation* 2014; **11**: 12.
- 117 Galea I, Felton LM, Waters S, van Rooijen N, Perry VH, Newman TA. Immune-tobrain signalling: the role of cerebral CD163-positive macrophages. *Neurosci Lett* 2008; **448**: 41–46.
- 118 Steiner J, Bielau H, Brisch R, Danos P, Ullrich O, Mawrin C et al. Immunological aspects in the neurobiology of suicide: elevated microglial density in schizophrenia and depression is associated with suicide. J Psychiatr Res 2008; 42: 151–157.
- 119 Steiner J, Walter M, Gos T, Guillemin GJ, Bernstein HG, Sarnyai Z *et al.* Severe depression is associated with increased microglial quinolinic acid in subregions of the anterior cingulate gyrus: evidence for an immune-modulated glutamatergic neurotransmission? *J Neuroinflammation* 2011; **8**: 94.
- 120 Torres-Platas SG, Cruceanu C, Chen GG, Turecki G, Mechawar N. Evidence for increased microglial priming and macrophage recruitment in the dorsal anterior cingulate white matter of depressed suicides. *Brain Behav Immun* 2014; **42**: 50–59.
- 121 Wohleb ES, Powell ND, Godbout JP, Sheridan JF. Stress-induced recruitment of bone marrow-derived monocytes to the brain promotes anxiety-like behavior. J Neurosci 2013; 33: 13820–13833.
- 122 Espey MG, Chernyshev ON, Reinhard JF Jr, Namboodiri MA, Colton CA. Activated human microglia produce the excitotoxin quinolinic acid. *Neuroreport* 1997; 8: 431–434.
- 123 Schnieder TP, Trencevska I, Rosoklija G, Stankov A, Mann JJ, Smiley J et al. Microglia of prefrontal white matter in suicide. J Neuropathol Exp Neurol 2014; 73: 880–890.
- 124 Haroon E, Woolwine BJ, Chen X, Pace TW, Parekh S, Spivey JR et al. IFN-alphainduced cortical and subcortical glutamate changes assessed by magnetic resonance spectroscopy. Neuropsychopharmacology 2014; 39: 1777–1785.
- 125 Hashimoto K, Sawa A, Iyo M. Increased levels of glutamate in brains from patients with mood disorders. *Biol Psychiatry* 2007; **62**: 1310–1316.
- 126 Yuksel C, Ongur D. Magnetic resonance spectroscopy studies of glutamaterelated abnormalities in mood disorders. *Biol Psychiatry* 2010; **68**: 785–794.
- 127 Gigante AD, Bond DJ, Lafer B, Lam RW, Young LT, Yatham LN. Brain glutamate levels measured by magnetic resonance spectroscopy in patients with bipolar disorder: a meta-analysis. *Bipolar Disord* 2012; 14: 478–487.
- 128 Stork C, Renshaw PF. Mitochondrial dysfunction in bipolar disorder: evidence from magnetic resonance spectroscopy research. *Mol Psychiatry* 2005; **10**: 900–919.
- 129 Rajkowska G, Stockmeier CA. Astrocyte pathology in major depressive disorder: insights from human postmortem brain tissue. *Curr Drug Targets* 2013; 14: 1225–1236.
- 130 Torres-Platas SG, Hercher C, Davoli MA, Maussion G, Labonte B, Turecki G et al. Astrocytic hypertrophy in anterior cingulate white matter of depressed suicides. *Neuropsychopharmacology* 2011; 36: 2650–2658.
- 131 Webster MJ, O'Grady J, Kleinman JE, Weickert CS. Glial fibrillary acidic protein mRNA levels in the cingulate cortex of individuals with depression, bipolar disorder and schizophrenia. *Neuroscience* 2005; **133**: 453–461.
- 132 Cobb JA, O'Neill K, Milner J, Mahajan GJ, Lawrence TJ, May WL et al. Density of GFAP-immunoreactive astrocytes is decreased in left hippocampi in major depressive disorder. *Neuroscience* 2016; **316**: 209–220.

- 133 Ernst C, Deleva V, Deng X, Sequeira A, Pomarenski A, Klempan T et al. Alternative splicing, methylation state, and expression profile of tropomyosin-related kinase B in the frontal cortex of suicide completers. Arch Gen Psychiatry 2009; 66: 22–32.
- 134 Ernst C, Nagy C, Kim S, Yang JP, Deng X, Hellstrom IC *et al.* Dysfunction of astrocyte connexins 30 and 43 in dorsal lateral prefrontal cortex of suicide completers. *Biol Psychiatry* 2011; **70**: 312–319.
- 135 Nagy C, Suderman M, Yang J, Szyf M, Mechawar N, Ernst C et al. Astrocytic abnormalities and global DNA methylation patterns in depression and suicide. *Mol Psychiatry* 2015; 20: 320–328.
- 136 Tynan RJ, Beynon SB, Hinwood M, Johnson SJ, Nilsson M, Woods JJ et al. Chronic stress-induced disruption of the astrocyte network is driven by structural atrophy and not loss of astrocytes. Acta Neuropathol 2013; 126: 75–91.
- 137 Bak LK, Schousboe A, Waagepetersen HS. The glutamate/GABA-glutamine cycle: aspects of transport, neurotransmitter homeostasis and ammonia transfer. J Neurochem 2006; 98: 641–653.
- 138 Choudary PV, Molnar M, Evans SJ, Tomita H, Li JZ, Vawter MP *et al.* Altered cortical glutamatergic and GABAergic signal transmission with glial involvement in depression. *Proc Natl Acad Sci USA* 2005; **102**: 15653–15658.
- 139 Bernard R, Kerman IA, Thompson RC, Jones EG, Bunney WE, Barchas JD et al. Altered expression of glutamate signaling, growth factor, and glia genes in the locus coeruleus of patients with major depression. *Mol Psychiatry* 2011; 16: 634–646.
- 140 Medina A, Burke S, Thompson RC, Bunney W Jr, Myers RM, Schatzberg A et al. Glutamate transporters: a key piece in the glutamate puzzle of major depressive disorder. J Psychiatr Res 2013; 47: 1150–1156.
- 141 Chandley MJ, Szebeni K, Szebeni A, Crawford J, Stockmeier CA, Turecki G et al. Gene expression deficits in pontine locus coeruleus astrocytes in men with major depressive disorder. J Psychiatry Neurosci 2013; 38: 276–284.
- 142 Rao JS, Kellom M, Reese EA, Rapoport SI, Kim HW. Dysregulated glutamate and dopamine transporters in postmortem frontal cortex from bipolar and schizophrenic patients. J Affect Disord 2012; 136: 63–71.
- 143 Zarate CA Jr, Payne JL, Quiroz J, Sporn J, Denicoff KK, Luckenbaugh D *et al*. An open-label trial of riluzole in patients with treatment-resistant major depression. *Am J Psychiatry* 2004; **161**: 171–174.
- 144 McCullumsmith RE, Meador-Woodruff JH. Striatal excitatory amino acid transporter transcript expression in schizophrenia, bipolar disorder, and major depressive disorder. *Neuropsychopharmacology* 2002; 26: 368–375.
- 145 Myles-Worsley M, Tiobech J, Browning SR, Korn J, Goodman S, Gentile K et al. Deletion at the SLC1A1 glutamate transporter gene co-segregates with schizophrenia and bipolar schizoaffective disorder in a 5-generation family. Am J Med Genet B Neuropsychiatr Genet 2013; 162B: 87–95.
- 146 Fiorentino A, Sharp SI, McQuillin A. Association of rare variation in the glutamate receptor gene SLC1A2 with susceptibility to bipolar disorder and schizophrenia. *Eur J Hum Genet* 2015; 23: 1200–1206.
- 147 Dahm L, Ott C, Steiner J, Stepniak B, Teegen B, Saschenbrecker S et al. Seroprevalence of autoantibodies against brain antigens in health and disease. Ann Neurol 2014; 76: 82–94.
- 148 Iorio R. Treatment-resistant depression and aquaporin-4 autoantibodies: is there a link? Biol Psychiatry 2015; 78: e1–e2.
- 149 Hinson SR, Roemer SF, Lucchinetti CF, Fryer JP, Kryzer TJ, Chamberlain JL et al. Aquaporin-4-binding autoantibodies in patients with neuromyelitis optica impair glutamate transport by down-regulating EAAT2. J Exp Med 2008; 205: 2473–2481.
- 150 Guillemin GJ. Quinolinic acid, the inescapable neurotoxin. FEBS J 2012; 279: 1356–1365.
- 151 Sequeira A, Mamdani F, Ernst C, Vawter MP, Bunney WE, Lebel V et al. Global brain gene expression analysis links glutamatergic and GABAergic alterations to suicide and major depression. PLoS One 2009; 4: e6585.
- 152 Deng W. Neurobiology of injury to the developing brain. Nat Rev Neurol 2010; 6: 328–336.
- 153 Juurlink BH. Response of glial cells to ischemia: roles of reactive oxygen species and glutathione. *Neurosci Biobehav Rev* 1997; 21: 151–166.
- 154 Lehnardt S, Lachance C, Patrizi S, Lefebvre S, Follett PL, Jensen FE *et al.* The tolllike receptor TLR4 is necessary for lipopolysaccharide-induced oligodendrocyte injury in the CNS. *J Neurosci* 2002; 22: 2478–2486.
- 155 Lehnardt S, Massillon L, Follett P, Jensen FE, Ratan R, Rosenberg PA et al. Activation of innate immunity in the CNS triggers neurodegeneration through a Tolllike receptor 4-dependent pathway. Proc Natl Acad Sci USA 2003; 100: 8514–8519.
- 156 Moyon S, Dubessy AL, Aigrot MS, Trotter M, Huang JK, Dauphinot L et al. Demyelination causes adult CNS progenitors to revert to an immature state and express immune cues that support their migration. J Neurosci 2015; 35: 4–20.
- 157 Fernandez-Castaneda A, Gaultier A. Adult oligodendrocyte progenitor cells -Multifaceted regulators of the CNS in health and disease. *Brain Behav Immun* 2016; **57**: 1–7.

- 158 Nosarti C, Reichenberg A, Murray RM, Cnattingius S, Lambe MP, Yin L *et al.* Preterm birth and psychiatric disorders in young adult life. *Arch Gen Psychiatry* 2012; **69**: E1–E8.
- 159 Edgar N, Sibille E. A putative functional role for oligodendrocytes in mood regulation. *Transl Psychiatry* 2012; **2**: e109.
- 160 Ongur D, Drevets WC, Price JL. Glial reduction in the subgenual prefrontal cortex in mood disorders. Proc Natl Acad Sci USA 1998; 95: 13290–13295.
- 161 Cotter D, Mackay D, Landau S, Kerwin R, Everall I. Reduced glial cell density and neuronal size in the anterior cingulate cortex in major depressive disorder. Arch Gen Psychiatry 2001; 58: 545–553.
- 162 Cotter D, Mackay D, Chana G, Beasley C, Landau S, Everall IP. Reduced neuronal size and glial cell density in area 9 of the dorsolateral prefrontal cortex in subjects with major depressive disorder. *Cereb Cortex* 2002; **12**: 386–394.
- 163 Hercher C, Parent M, Flores C, Canetti L, Turecki G, Mechawar N. Alcohol dependence-related increase of glial cell density in the anterior cingulate cortex of suicide completers. J Psychiatry Neurosci 2009; 34: 281–288.
- 164 Rajkowska G, Halaris A, Selemon LD. Reductions in neuronal and glial density characterize the dorsolateral prefrontal cortex in bipolar disorder. *Biol Psychiatry* 2001; **49**: 741–752.
- 165 Bowley MP, Drevets WC, Ongur D, Price JL. Low glial numbers in the amygdala in major depressive disorder. *Biol Psychiatry* 2002; **52**: 404–412.
- 166 Vostrikov VM, Uranova NA, Orlovskaya DD. Deficit of perineuronal oligodendrocytes in the prefrontal cortex in schizophrenia and mood disorders. *Schizophr Res* 2007; 94: 273–280.
- 167 Uranova NA, Vostrikov VM, Orlovskaya DD, Rachmanova VI. Oligodendroglial density in the prefrontal cortex in schizophrenia and mood disorders: a study from the Stanley Neuropathology Consortium. *Schizophr Res* 2004; 67: 269–275.
- 168 Hamidi M, Drevets WC, Price JL. Glial reduction in amygdala in major depressive disorder is due to oligodendrocytes. *Biol Psychiatry* 2004; 55: 563–569.
- 169 Uranova N, Orlovskaya D, Vikhreva O, Zimina I, Kolomeets N, Vostrikov V et al. Electron microscopy of oligodendroglia in severe mental illness. Brain Res Bull 2001; 55: 597–610.
- 170 Chambers JS, Perrone-Bizzozero NI. Altered myelination of the hippocampal formation in subjects with schizophrenia and bipolar disorder. *Neurochem Res* 2004; **29**: 2293–2302.
- 171 Regenold WT, Phatak P, Marano CM, Gearhart L, Viens CH, Hisley KC. Myelin staining of deep white matter in the dorsolateral prefrontal cortex in schizophrenia, bipolar disorder, and unipolar major depression. *Psychiatry Res* 2007; **151**: 179–188.
- 172 Hayashi Y, Nihonmatsu-Kikuchi N, Yu X, Ishimoto K, Hisanaga SI, Tatebayashi Y. A novel, rapid, quantitative cell-counting method reveals oligodendroglial reduction in the frontopolar cortex in major depressive disorder. *Mol Psychiatry* 2011; 16: 1155–1158.
- 173 Tkachev D, Mimmack ML, Ryan MM, Wayland M, Freeman T, Jones PB et al. Oligodendrocyte dysfunction in schizophrenia and bipolar disorder. *Lancet* 2003; 362: 798–805.
- 174 Myung W, Han CE, Fava M, Mischoulon D, Papakostas Gl, Heo JY et al. Reduced frontal-subcortical white matter connectivity in association with suicidal ideation in major depressive disorder. *Transl Psychiatry* 2016; 6: e835.
- 175 Houenou J, Wessa M, Douaud G, Leboyer M, Chanraud S, Perrin M *et al.* Increased white matter connectivity in euthymic bipolar patients: diffusion tensor tractography between the subgenual cingulate and the amygdalohippocampal complex. *Mol Psychiatry* 2007; **12**: 1001–1010.
- 176 McIntosh AM, Maniega SM, Lymer GK, McKirdy J, Hall J, Sussmann JE *et al.* White matter tractography in bipolar disorder and schizophrenia. *Biol Psychiatry* 2008; 64: 1088–1092.
- 177 Sussmann JE, Lymer GK, McKirdy J, Moorhead TW, Munoz Maniega S, Job D *et al.* White matter abnormalities in bipolar disorder and schizophrenia detected using diffusion tensor magnetic resonance imaging. *Bipolar Disord* 2009; 11: 11–18.
- 178 Benedetti F, Poletti S, Hoogenboezem TA, Mazza E, Ambree O, de Wit H et al. Inflammatory cytokines influence measures of white matter integrity in bipolar disorder. J Affect Disord 2016; 202: 1–9.
- 179 Molina V, Sanchez J, Sanz J, Reig S, Benito C, Leal I et al. Dorsolateral prefrontal N-acetyl-aspartate concentration in male patients with chronic schizophrenia and with chronic bipolar disorder. Eur Psychiatry 2007; 22: 505–512.
- 180 Chang K, Adleman N, Dienes K, Barnea-Goraly N, Reiss A, Ketter T. Decreased N-acetylaspartate in children with familial bipolar disorder. *Biol Psychiatry* 2003; 53: 1059–1065.
- 181 Sassi RB, Stanley JA, Axelson D, Brambilla P, Nicoletti MA, Keshavan MS et al. Reduced NAA levels in the dorsolateral prefrontal cortex of young bipolar patients. Am J Psychiatry 2005; 162: 2109–2115.
- 182 Zhang J, Narr KL, Woods RP, Phillips OR, Alger JR, Espinoza RT. Glutamate normalization with ECT treatment response in major depression. *Mol Psychiatry* 2013; 18: 268–270.

- 183 Birey F, Kloc M, Chavali M, Hussein I, Wilson M, Christoffel DJ et al. Genetic and Stress-Induced Loss of NG2 Glia Triggers Emergence of Depressive-like Behaviors through Reduced Secretion of FGF2. *Neuron* 2015; 88: 941–956.
- 184 Stadelmann C, Wegner C, Bruck W. Inflammation, demyelination, and degeneration - recent insights from MS pathology. *Biochim Biophys Acta* 2011; **1812**: 275–282.
- 185 Politte LC, Huffman JC, Stern TA. Neuropsychiatric manifestations of multiple sclerosis. Prim Care Companion J Clin Psychiatry 2008; 10: 318–324.
- 186 Johnson B, Nichols S. Crying and suicidal, but not depressed. Pseudobulbar affect in multiple sclerosis successfully treated with valproic acid: Case report and literature review. *Palliat Support Care* 2015; **13**: 1797–1801.
- 187 Ahmed A, Simmons Z. Pseudobulbar affect: prevalence and management. Ther Clin Risk Manag 2013; 9: 483–489.
- 188 Ghasemi M, Phillips C, Trillo L, De Miguel Z, Das D, Salehi A. The role of NMDA receptors in the pathophysiology and treatment of mood disorders. *Neurosci Biobehav Rev* 2014; **47**: 336–358.
- 189 Kawasaki Y, Zhang L, Cheng JK, Ji RR. Cytokine mechanisms of central sensitization: distinct and overlapping role of interleukin-1beta, interleukin-6, and tumor necrosis factor-alpha in regulating synaptic and neuronal activity in the superficial spinal cord. J Neurosci 2008; 28: 5189–5194.
- 190 Zhang RX, Liu B, Li A, Wang L, Ren K, Qiao JT et al. Interleukin 1beta facilitates bone cancer pain in rats by enhancing NMDA receptor NR-1 subunit phosphorylation. *Neuroscience* 2008; **154**: 1533–1538.
- 191 Balosso S, Maroso M, Sanchez-Alavez M, Ravizza T, Frasca A, Bartfai T et al. A novel non-transcriptional pathway mediates the proconvulsive effects of interleukin-1beta. Brain 2008; **131**(Pt 12): 3256–3265.
- 192 Kessing CF, Tyor WR. Interferon-alpha induces neurotoxicity through activation of the type I receptor and the GluN2A subunit of the NMDA receptor. *J Interferon Cytokine Res* 2015; **35**: 317–324.
- 193 Di Filippo M, Tozzi A, Arcangeli S, de lure A, Durante V, Di Gregorio M et al. Interferon-beta1a modulates glutamate neurotransmission in the CNS through CaMKII and GluN2A-containing NMDA receptors. *Neuropharmacology* 2016; **100**: 98–105.
- 194 Forrest CM, Khalil OS, Pisar M, Darlington LG, Stone TW. Prenatal inhibition of the tryptophan-kynurenine pathway alters synaptic plasticity and protein expression in the rat hippocampus. *Brain Res* 2013; **1504**: 1–15.
- 195 Feyissa AM, Chandran A, Stockmeier CA, Karolewicz B. Reduced levels of NR2A and NR2B subunits of NMDA receptor and PSD-95 in the prefrontal cortex in major depression. *Prog Neuropsychopharmacol Biol Psychiatry* 2009; **33**: 70–75.
- 196 Karolewicz B, Stockmeier CA, Ordway GA. Elevated levels of the NR2C subunit of the NMDA receptor in the locus coeruleus in depression. *Neuropsychopharmacology* 2005; **30**: 1557–1567.
- 197 Karolewicz B, Szebeni K, Gilmore T, Maciag D, Stockmeier CA, Ordway GA. Elevated levels of NR2A and PSD-95 in the lateral amygdala in depression. Int J Neuropsychopharmacol 2009; 12: 143–153.
- 198 Gray AL, Hyde TM, Deep-Soboslay A, Kleinman JE, Sodhi MS. Sex differences in glutamate receptor gene expression in major depression and suicide. *Mol Psychiatry* 2015; 20: 1057–1068.
- 199 Scarr E, Pavey G, Sundram S, MacKinnon A, Dean B. Decreased hippocampal NMDA, but not kainate or AMPA receptors in bipolar disorder. *Bipolar Disord* 2003; 5: 257–264.
- 200 McCullumsmith RE, Kristiansen LV, Beneyto M, Scarr E, Dean B, Meador-Woodruff JH. Decreased NR1, NR2A, and SAP102 transcript expression in the hippocampus in bipolar disorder. *Brain Res* 2007; **1127**: 108–118.
- 201 Benes FM, Lim B, Matzilevich D, Subburaju S, Walsh JP. Circuitry-based gene expression profiles in GABA cells of the trisynaptic pathway in schizophrenics versus bipolars. *Proc Natl Acad Sci USA* 2008; **105**: 20935–20940.
- 202 Beneyto M, Meador-Woodruff JH. Lamina-specific abnormalities of NMDA receptor-associated postsynaptic protein transcripts in the prefrontal cortex in schizophrenia and bipolar disorder. *Neuropsychopharmacology* 2008; 33: 2175–2186.
- 203 Walker AK, Budac DP, Bisulco S, Lee AW, Smith RA, Beenders B et al. NMDA receptor blockade by ketamine abrogates lipopolysaccharide-induced depressive-like behavior in C57BL/6J mice. *Neuropsychopharmacology* 2013; 38: 1609–1616.
- 204 Neale SA, Copeland CS, Uebele VN, Thomson FJ, Salt TE. Modulation of hippocampal synaptic transmission by the kynurenine pathway member xanthurenic acid and other VGLUT inhibitors. *Neuropsychopharmacology* 2013; 38: 1060–1067.
- 205 Daniels RW, Collins CA, Chen K, Gelfand MV, Featherstone DE, DiAntonio A. A single vesicular glutamate transporter is sufficient to fill a synaptic vesicle. *Neuron* 2006; **49**: 11–16.
- 206 Uezato A, Meador-Woodruff JH, McCullumsmith RE. Vesicular glutamate transporter mRNA expression in the medial temporal lobe in major depressive disorder, bipolar disorder, and schizophrenia. *Bipolar Disord* 2009; **11**: 711–725.

- 207 Vigneault E, Poirel O, Riad M, Prud'homme J, Dumas S, Turecki G et al. Distribution of vesicular glutamate transporters in the human brain. Front Neuroanatomy 2015; **9**: 23.
- 208 Eastwood SL, Harrison PJ. Markers of glutamate synaptic transmission and plasticity are increased in the anterior cingulate cortex in bipolar disorder. *Biol Psychiatry* 2010; 67: 1010–1016.
- 209 Gilabert-Juan J, Varea E, Guirado R, Blasco-Ibanez JM, Crespo C, Nacher J. Alterations in the expression of PSA-NCAM and synaptic proteins in the dorsolateral prefrontal cortex of psychiatric disorder patients. *Neurosci Lett* 2012; **530**: 97–102.
- 210 Woo TU, Walsh JP, Benes FM. Density of glutamic acid decarboxylase 67 messenger RNA-containing neurons that express the N-methyl-D-aspartate receptor subunit NR2A in the anterior cingulate cortex in schizophrenia and bipolar disorder. Arch Gen Psychiatry 2004; 61: 649–657.
- 211 Woo TU, Shrestha K, Amstrong C, Minns MM, Walsh JP, Benes FM. Differential alterations of kainate receptor subunits in inhibitory interneurons in the anterior cingulate cortex in schizophrenia and bipolar disorder. *Schizophr Res* 2007; **96**: 46–61.
- 212 Cotter D, Landau S, Beasley C, Stevenson R, Chana G, MacMillan L et al. The density and spatial distribution of GABAergic neurons, labelled using calcium binding proteins, in the anterior cingulate cortex in major depressive disorder, bipolar disorder, and schizophrenia. *Biol Psychiatry* 2002; **51**: 377–386.
- 213 Woo TU, Kim AM, Viscidi E. Disease-specific alterations in glutamatergic neurotransmission on inhibitory interneurons in the prefrontal cortex in schizophrenia. *Brain Res* 2008; **1218**: 267–277.
- 214 Guidotti A, Auta J, Davis JM, Di-Giorgi-Gerevini V, Dwivedi Y, Grayson DR et al. Decrease in reelin and glutamic acid decarboxylase67 (GAD67) expression in schizophrenia and bipolar disorder: a postmortem brain study. Arch Gen Psychiatry 2000; 57: 1061–1069.
- 215 Veldic M, Guidotti A, Maloku E, Davis JM, Costa E. In psychosis, cortical interneurons overexpress DNA-methyltransferase 1. Proc Natl Acad Sci USA 2005; 102: 2152–2157.
- 216 Thompson M, Weickert CS, Wyatt E, Webster MJ. Decreased glutamic acid decarboxylase(67) mRNA expression in multiple brain areas of patients with schizophrenia and mood disorders. J Psychiatr Res 2009; 43: 970–977.
- 217 Heckers S, Stone D, Walsh J, Shick J, Koul P, Benes FM. Differential hippocampal expression of glutamic acid decarboxylase 65 and 67 messenger RNA in bipolar disorder and schizophrenia. *Arch Gen Psychiatry* 2002; **59**: 521–529.
- 218 Kim S, Webster MJ. Correlation analysis between genome-wide expression profiles and cytoarchitectural abnormalities in the prefrontal cortex of psychiatric disorders. *Mol Psychiatry* 2010; **15**: 326–336.
- 219 Rajkowska G, O'Dwyer G, Teleki Z, Stockmeier CA, Miguel-Hidalgo JJ. GABAergic neurons immunoreactive for calcium binding proteins are reduced in the prefrontal cortex in major depression. *Neuropsychopharmacology* 2007; 32: 471–482.
- 220 Maciag D, Hughes J, O'Dwyer G, Pride Y, Stockmeier CA, Sanacora G et al. Reduced density of calbindin immunoreactive GABAergic neurons in the occipital cortex in major depression: relevance to neuroimaging studies. *Biol Psy*chiatry 2010; 67: 465–470.
- 221 Smiley JF, Hackett TA, Bleiwas C, Petkova E, Stankov A, Mann JJ et al. Reduced GABA neuron density in auditory cerebral cortex of subjects with major depressive disorder. J Chem Neuroanatomy 2015; 76: 108–121.
- 222 Gonzalez-Burgos G, Cho RY, Lewis DA. Alterations in cortical network oscillations and parvalbumin neurons in schizophrenia. *Biol Psychiatry* 2015; **77**: 1031–1040.
- 223 Gonzalez-Burgos G, Lewis DA. GABA neurons and the mechanisms of network oscillations: implications for understanding cortical dysfunction in schizophrenia. *Schizophr Bull* 2008; **34**: 944–961.
- 224 Yamamoto J, Suh J, Takeuchi D, Tonegawa S. Successful execution of working memory linked to synchronized high-frequency gamma oscillations. *Cell* 2014; 157: 845–857.
- 225 Kimoto S, Zaki MM, Bazmi HH, Lewis DA. Altered markers of cortical gammaaminobutyric acid neuronal activity in schizophrenia: role of the NARP gene. JAMA Psychiatry 2015; 72: 747–756.
- 226 Crowley T, Cryan JF, Downer EJ, O'Leary OF. Inhibiting neuroinflammation: the role and therapeutic potential of GABA in neuro-immune interactions. *Brain Behav Immun* 2016; 54: 260–277.
- 227 Clements RJ, McDonough J, Freeman EJ. Distribution of parvalbumin and calretinin immunoreactive interneurons in motor cortex from multiple sclerosis post-mortem tissue. *Exp Brain Res* 2008; **187**: 459–465.
- 228 Wieck A, Andersen SL, Brenhouse HC. Evidence for a neuroinflammatory mechanism in delayed effects of early life adversity in rats: relationship to cortical NMDA receptor expression. *Brain Behav Immun* 2013; **28**: 218–226.

- 229 Giovanoli S, Weber L, Meyer U. Single and combined effects of prenatal immune activation and peripubertal stress on parvalbumin and reelin expression in the hippocampal formation. *Brain Behav Immun* 2014; 40: 48–54.
- 230 Brooks JM, Carrillo GL, Su J, Lindsay DS, Fox MA, Blader JJ. Toxoplasma gondii infections alter GABAergic synapses and signaling in the central nervous system. *Mbio* 2015; 6: e01428–15.
- 231 Rivest S. Regulation of innate immune responses in the brain. *Nat Rev Immunol* 2009; **9**: 429–439.
- 232 Ransohoff RM, Perry VH. Microglial physiology: unique stimuli, specialized responses. Ann Rev Immunol 2009; 27: 119–145.
- 233 Loane DJ, Kumar A. Microglia in the TBI brain: the good, the bad, and the dysregulated. *Exp Neurol* 2015; 275: 316–327.
- 234 Gras G, Porcheray F, Samah B, Leone C. The glutamate-glutamine cycle as an inducible, protective face of macrophage activation. J Leukoc Biol 2006; 80: 1067–1075.
- 235 Chen Z, Jalabi W, Shpargel KB, Farabaugh KT, Dutta R, Yin X et al. Lipopolysaccharide-induced microglial activation and neuroprotection against experimental brain injury is independent of hematogenous TLR4. J Neurosci 2012; 32: 11706–11715.

- 236 Chen Z, Jalabi W, Hu W, Park HJ, Gale JT, Kidd GJ *et al.* Microglial displacement of inhibitory synapses provides neuroprotection in the adult brain. *Nat Commun* 2014; **5**: 4486.
- 237 Sekar A, Bialas AR, de Rivera H, Davis A, Hammond TR, Kamitaki N et al. Schizophrenia risk from complex variation of complement component 4. Nature 2016; 530: 177–183.

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/ by/4.0/

© The Author(s) 2016