

ORIGINAL ARTICLE

Exome sequencing in dementia with Lewy bodies

MJ Keogh¹, M Kurzawa-Akanbi¹, H Griffin¹, K Douroudis¹, KL Ayers¹, RI Hussein¹, G Hudson¹, A Pyle¹, HJ Cordell¹, J Attems^{2,3}, IG McKeith^{2,3}, JT O'Brien^{2,3}, DJ Burn^{2,3}, CM Morris^{2,3}, AJ Thomas^{2,3} and PF Chinnery^{1,2}

Dementia with Lewy bodies (DLB) is the second most common form of degenerative dementia. Siblings of affected individuals are at greater risk of developing DLB, but little is known about the underlying genetic basis of the disease. We set out to determine whether mutations in known highly penetrant neurodegenerative disease genes are found in patients with DLB. Whole-exome sequencing was performed on 91 neuropathologically confirmed cases of DLB, supplemented by independent *APOE* genotyping. Genetic variants were classified using established criteria, and additional neuropathological examination was performed for putative mutation carriers. Likely pathogenic variants previously described as causing monogenic forms of neurodegenerative disease were found in 4.4% of patients with DLB. The *APOE* $\epsilon 4$ allele increased the risk of disease ($P=0.0001$), conferred a shorter disease duration ($P=0.043$) and earlier age of death ($P=0.0015$). In conclusion, although known pathogenic mutations in neurodegenerative disease genes are uncommon in DLB, known genetic risk factors are present in >60% of cases. *APOE* $\epsilon 4$ not only modifies disease risk, but also modulates the rate of disease progression. The reduced penetrance of reported pathogenic alleles explains the lack of a family history in most patients, and the presence of variants previously described as causing frontotemporal dementia suggests a mechanistic overlap between DLB and other neurodegenerative diseases.

Translational Psychiatry (2016) 6, e728; doi:10.1038/tp.2015.220; published online 2 February 2016

INTRODUCTION

Dementia with Lewy bodies (DLB) is the second most common form of dementia. It affects 5% of the population over 75 years of age,¹ and has a greater impact on healthcare provision than Alzheimer's disease (AD).² The neuropathological hallmark of DLB is widespread α -synuclein-positive neuronal inclusions (Lewy bodies and Lewy neurites) and in addition this is often associated with amyloid deposition.³ Siblings of affected individuals have a 2.3-fold increased risk of developing the disorder,⁴ but little is known about the genetic aetiology of the disease. Although genetic variants in *APOE*,⁵ *GBA*,⁶ *SNCA* and *SCARB2* (ref. 7) have been associated with an increased risk of DLB, only a few families have been described with more than two first-degree relatives,⁸ and no single highly penetrant gene defects have been shown to cause familial forms of the disorder. Using exome sequencing in 91 autopsy-confirmed cases, here we determined whether confirmed or putative pathogenic mutations in genes in known neurodegenerative disease genes are found in patients with DLB.

MATERIALS AND METHODS

Subjects and sample preparation

We studied 91 post-mortem cases conforming to both the clinical and post-mortem diagnostic criteria for DLB.³ Two patients were first-degree relatives (mother and daughter) and two patients were siblings (brothers). The remaining 87 patients had no recorded family history of neurodegenerative disease. Age of onset, disease duration, age of death, neuropathological subtype of Lewy body disease according to McKeith/Newcastle criteria³ and Braak neurofibrillary tangle stage were recorded⁹ (Figure 1). In addition, we assessed Lewy body Braak stages,¹⁰ A β phases¹¹ and stages of cerebral amyloid angiopathy.¹² Of note, none of the cases showed intracytoplasmic TAR DNA-binding protein 43 (TDP-43) inclusions

indicative for frontotemporal lobar degeneration associated with TDP-43 pathology, nor were there neuropathological features consistent with other types of frontotemporal lobar degeneration (see additional Supplementary Methods).

DNA extraction and exome sequencing

DNA was extracted from cerebellum in all the cases. Illumina TruSeq 62 Mb exome capture and sequencing (Illumina HiSeq2000, 100 bp paired-end reads) was performed as described (see additional Supplementary Methods).

Known disease genes were defined as those previously shown to cause monogenic forms of Parkinson's disease (PD), AD, frontotemporal lobar dementia and amyotrophic lateral sclerosis (Table 1). Variants were selected with a minor allele frequency of < 0.01 international reference databases. Variants were defined as (1) pathogenic, (2) likely pathogenic, (3) of uncertain significance or (4) benign according to American College of Medical Genetics criteria¹³ (Table 1).

For completeness, exonic variants in genes previously associated with DLB (*GBA*, *APOE*, *SNCA* and *SCARB2*),^{5–7} AD (*APOE*, *TREM2*)¹⁴ or PD (*LRRK2*, *GBA*)¹⁵ were also identified in DLB cases and compared with 93 in-house unrelated disease control exomes.

RESULTS

The mean exome sequencing base coverage depth was 84-fold (s.d.=13) in the 91 DLB cases and 76-fold (s.d.=12) in the 93 controls. There was no difference in the proportion of the exome target covered at >30-fold depth between DLB cases and controls (DLB 84%, s.d.=5; controls 84%, s.d.=3, $P=0.588$).

¹Institute of Genetic Medicine, Newcastle University, Newcastle upon Tyne, UK; ²NIHR Biomedical Research Unit, Newcastle upon Tyne Hospitals NHS Foundation Trust, Campus for Ageing and Vitality, Newcastle upon Tyne, UK and ³Institute for Neuroscience, The Medical School, Newcastle University, Newcastle upon Tyne, UK. Correspondence: Professor PF Chinnery, Institute of Genetic Medicine, Newcastle University, Central Parkway, Newcastle upon Tyne NE2 4HH, UK.

E-mail: pfc25@medschl.cam.ac.uk

Received 5 August 2015; revised 17 October 2015; accepted 13 November 2015

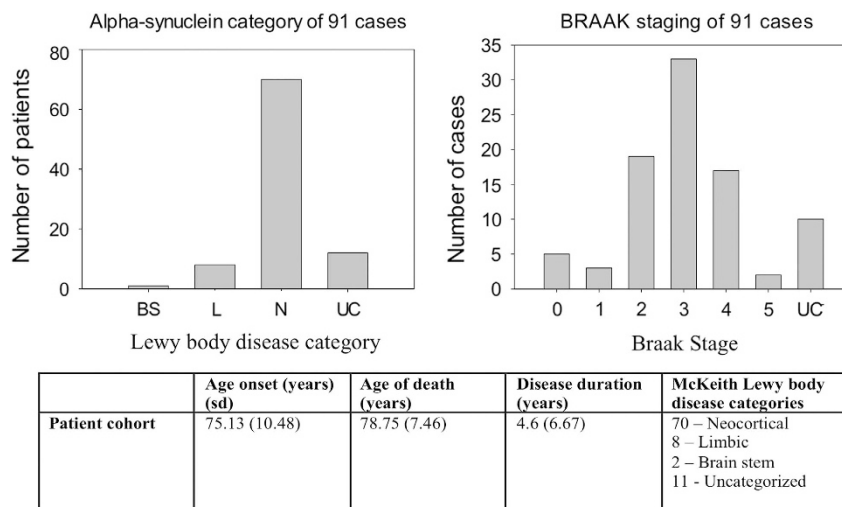


Figure 1. Clinical and pathological characteristics of the 91 dementia with Lewy body (DLB) cases. Top left: frequency of each pathological category (BS, brain stem; L, limbic; N, neocortical; UC, unclassified). Top right: BRAAK neurofibrillary tangle stage of patients (UC, unclassified). Bottom: table of the clinical and pathological data for all the 91 cases of DLB. Data are mean (s.d.). Motor features were defined by documented evidence of a Parkinsonian movement disorder by an assessing clinician.

Table 1. Genes causing monogenic forms of PD, AD, FTLD-ALS, which were analysed for rare protein altering mutations in patients

Inheritance	Disease		
	PD	AD	FTLD-ALS
Autosomal dominant	<i>SNCA</i>	<i>APP</i>	<i>C9orf72</i>
	<i>LRRK2</i>	<i>PSEN-1</i>	<i>SOD1</i>
	<i>UCHL1</i>	<i>PSEN2</i>	<i>MAPT</i>
	<i>GIGYF2</i>		<i>PGRN</i>
	<i>Omi/HTRA2</i>		<i>TARDBP</i>
	<i>EIF4G1</i>		<i>OPTN</i>
			<i>ANG</i>
			<i>CHMP2B</i>
			<i>SQSTM1</i>
			<i>FUS</i>
Autosomal recessive	<i>PARK2</i>		<i>VCP</i>
	<i>PINK1</i>		<i>OPTN</i>
	<i>ATP13A2</i>		
	<i>PLA2G6</i>		
	<i>FBX07</i>		
	<i>DJ-1</i>		

Abbreviation: AD, Alzheimer's disease; ALS, amyotrophic lateral sclerosis; FTLD, frontotemporal lobar degeneration; PD, Parkinson's disease.

Known mendelian disease genes

A total 18 rare heterozygous mutations in 25 patients were observed in genes previously shown to cause autosomal dominant forms of neurodegeneration (Tables 1, 2 and Supplementary Table S1). Three of these variants have been described in patients with AD, PD or frontotemporal lobar degeneration and amyotrophic lateral sclerosis (Patient A:*PSEN2* p.D439A;^{16,17} B:*CHMP2B* p.I29V;¹⁸ and C:*SQSTM1* p.A33V,^{19,20}). In two additional cases (Patient E:*EIF4G1* p.M1134V and F:*SQSTM1* p.P27L), variants in known disease genes affecting highly conserved residues and predicted to be pathogenic by *in silico* software algorithms, were deemed of uncertain significance. Two patients also had variants of uncertain significance in *GIGYF2*, which is also implicated in PD (H:*GIGYF2* p.S66T; G:*GIGYF2* p.S1029C, Table 2). In genes causing autosomal recessive PD, AD or frontotemporal

dementia and amyotrophic lateral sclerosis, only one rare compound heterozygous mutation in *PARK2* was seen (Patient D, p.R275W/p.G430D).

Only patient A had a relevant family history (father affected—deceased and no tissue/DNA available). A clinical description of these cases is shown in the Supplementary Information. All showed typical DLB pathology with cortical LB being present and moderate AD pathology (Table 2).

The mean age at the presentation for the four cases with previously described pathogenic mutations (Patients A–D) was 78.25 years (s.d. = 8.05). Motor symptoms developed in three cases (Patient A, B and D) at a mean of 1.33 years (s.d. = 0.58) after the onset of cognitive symptoms. When patients E and F were included, the mean age of onset was 78.6 (s.d. = 6.68), with motor symptoms developing in four patients (A, B, D and E), and a mean disease duration of 2.3 (s.d. = 1.16) years.

Major risk alleles

GBA, *TREM2* and *LRRK2* had >80% coverage at 30-fold depth in both DLB cases and controls. *APOE* coverage was poor (DLB, 46.2%; controls 48.7% at 30-fold depth) and was therefore genotyped independently (see additional Supplementary Methods). After removing the previously described pathogenic alleles, *APOE* $\epsilon 4$ was significantly associated with DLB compared with controls ($n = 87$, $P = 0.0001$, Table 3). Ten DLB cases had one of five heterozygous *GBA* variants, compared with only three controls ($P = 0.043$). Two *GBA* variants known to be risk factors for PD (p.L370P and p.N296S) were seen only in four patients and no controls. Two patients had variants in *SCARB2* compared with six controls, and no *SNCA* variants were seen. There was no association between DLB and variants in *SCARB2*, *LRRK2* or *TREM2* (Supplementary Table S2).

Although there was no difference in the age of onset of DLB in *APOE* $\epsilon 4$ allele carriers when compared with non-*APOE* $\epsilon 4$ allele carriers ($P = 0.227$), the *APOE* $\epsilon 4$ allele carriers had a shorter disease duration following diagnosis ($P = 0.036$), and died at an earlier age ($P = 0.005$) than non-*APOE* $\epsilon 4$ carriers (Figure 2, Table 3). There was no association between the presence of variants in *GBA*, *SNCA*, *SCARB2*, *LRRK2*, *PARK2* or *ATP13A2* and age of onset, disease duration, age of death, neurofibrillary Braak stage or the presence of motor symptoms.

Table 2. The frequency of potentially pathogenic variants in DLB cases and controls

Patient	Pathogenicity	Allele information and protein alteration										Functional prediction of variant					Neuropathology					Pathogenicity
		Gene	Chromosome	Position	R/V	Predicted protein change	Previously reported phenotype	MAF ESP 6500	MAF 1000G	ExAC	MAF	SIFT	PolyPhen2	Mutation-taster	CADD score (scaled)	NFT Braak stage	Braak PD stage	Aβ phase	TDP-43	CAA	ACMG criteria	
A	P	CHMP2B	3	87289899	A/G	p.I29V	FTLD	0.00015	—	0.0001237	T	N	D	14.1	4	5/6	4	+ve	CA1	2	(1) Same amino acid as previously reported (PS1) (2) Well established functional studies show a deleterious effect (PS3)	
B	P	PARK2	6	162206852	G/A	p.R275W	PD	0.001999	0.0005	0.00206	D	D	D	33	4	6	3	NT	—	(1) Same amino acid as previously reported (PS1) (2) Well established functional studies show a deleterious effect (PS3)		
B	P	PARK2	1	161771240	C/T	p.G430D	AD	0.000231	—	0.0001076	D	D	D	34	4	6	4	NT	—	(1) Same amino acid as previously reported (PS1) (2) Well established functional studies show a deleterious effect (PS3)		
C	LP	PSEN2	1	227083249	A/C	p.D439A	AD	0.00015	—	0.00003764	D	D	D	26.9	4	6	4	NT	—	(1) Same amino acid as previously reported (PS1) (2) Multiple lines of computational evidence (PP3) (3) Missense with low rate of benign variability (PP2)		
D	LP	SQSTM1	5	179250906	C/T	p.A33V	FTLD	0.000769	0.0018	0.001523	T	N	N	11.9	3/4	6	3	+ve	CA1	2	(1) Same amino acid as previously reported (PS1) (2) Multiple lines of computational evidence (PP3) (3) Missense with low rate of benign variability (PP2)	
E	US	EIF4G1	3	184046450	A/G	p.M1134V	U	0.00015	—	0.0002224	D	D	D	26.3	3	5/6	2	+ve	CA1	2	(1) Same amino acid as previously reported (PS1) (2) Multiple lines of computational evidence (PP3) (3) Missense with low rate of benign variability (PP2)	
F	US	SQSTM1	5	179250888	C/T	p.P27L	U	0.00008	—	0.00003349	T	D	N	12.8	3	5/6	2	+ve	CA1	2	(1) Same amino acid as previously reported (PS1) (2) Multiple lines of computational evidence (PP3) (3) Missense with low rate of benign variability (PP2)	
G	US	GIGYF2	2	233709083	C/G	p.S1029C	U	0.00123	—	0.0007833	D	D	D	23.2	5	6	4	-ve	—	2	(1) Computational evidence supports a deleterious effect (2) Well established functional studies show a deleterious effect (PS3)	
H	US	GIGYF2	2	233655546	G/C	p.S66T	U	0.00008	—	0.0001813	T	D	D	23.7	4	5	3	NT	—	2	(1) Computational evidence supports a deleterious effect (2) Well established functional studies show a deleterious effect (PS3)	

Abbreviations: ACMG, American College of Medical Genetics; AD, Alzheimer's disease; CA1, CA1 division of the hippocampus; CAA, cerebral amyloid angiopathy; DLB, dementia with Lewy body; FTLD, frontotemporal lobar degeneration; MAF, minor allele frequency; PD, Parkinson's disease; R, reference allele; U, unknown or not described; V, variant allele. The number of patients covered at > 30-fold sequence depth, and the number of case and control patients carrying each mutation is shown. Functional predictions were performed by SIFT, PolyPhen2 and MutationTaster. Variants were classified as: (1) pathogenic, if the same alleles had previously been described in patients with neurodegenerative disease; (2) likely pathogenic, if the alleles were in previously known neurodegenerative disease genes and *in silico* predictions supported a pathogenic role; and (3) possibly pathogenic, if *in silico* predictions supported a pathogenic role, and the gene had previously been associated with a Mendelian neurodegenerative disease. See Supplementary Material for citations. Neuropathology scores according to existing accepted diagnostic criteria as outlined in Supplementary Methods are shown.

DISCUSSION

Exome sequencing of 91 cases of pathologically confirmed DLB identified four patients harbouring previously described pathogenic mutations neurodegenerative disease genes based on current diagnostic criteria (*PSEN2*, *CHMP2B*, *SQSTM1*, *PARK2*); possible pathogenic mutations in two (*EIF4G1* and *SQSTM1*); and two further cases with mutations in *GIGYF2*, which has previously been associated with autosomal dominant PD. The central question is: are these variants causing DLB, or are they co-incident findings? The role of *GIGYF2* in PD remains contentious,²¹ and the p.D439A variant in *PSEN2* may have incomplete penetrance,¹⁷ and is thus found in control databases along with the *CHMP2B* and *SQSTM1* variants. Providing definitive proof of pathogenicity is therefore challenging, and there are arguments in both directions.

On one hand, the variants detected in *PSEN2*, *CHMP2B*, *SQSTM1* and *PARK2* are exceptionally rare in the general population.²² Given the clinical, pathological and mechanistic overlap between DLB and the neurodegenerative disorders where these disease genes were first described, it is plausible that they are contributing to the neuropathology. For example, in families with familial AD due to *PSEN2* mutations, up to 64% of cases have extensive Lewy body deposition at autopsy.²³ The CHMP2B protein has been shown to be found in association with Lewy bodies in post-

mortem cases of DLB,²⁴ and *SQSTM1* deficiency has been shown to enhance α -synuclein accumulation in mice.²⁵ The *SQSTM1* p. A33V variant was previously described in five cases of fronto-temporal dementia.^{19,20} Recently, this allele was also detected in a patient with young-onset AD.²⁶ Although seen in 0.0012% of controls, the p.A33V variant has now been seen in 8/1060 (0.007%) of patients with a neurodegenerative disease (including our study)^{19,20,26} suggesting a broad association with neurodegenerative disorders ($P=0.0037$, chi squared with Yate's correction). These findings support the notion that rare, incompletely penetrant pathogenic alleles cause overlapping syndromes of neurodegeneration, perhaps explaining why previously ascribed variants for frontotemporal dementia were also found in our DLB cases. Pathogenic mutations with a reduced penetrance will also be detected in healthy individuals (as for *PSEN2* p.D439A¹⁷), and their presence in a control cohort does not preclude their potential to cause disease.²² This may explain why none of the four patients harbouring established pathogenic mutations reported a relevant family history.

On the other hand, the clinical and pathological phenotype of these five cases was wholly typical of DLB: how can this be reconciled with known pathogenic compound heterozygous mutations in *PARK2*, which typically presents with dystonia in early adult life? These findings highlight the challenges of using exome or whole-genome sequencing in a clinical context: is rare pathogenic mutation in a known disease gene more likely to be causing a variant phenotype, or is the phenotype so unusual that the variants must be a co-incident finding? This will be difficult to resolve in individual cases, but the ongoing reporting of rare putative disease alleles, linked to rich phenotypic data, is an essential step in generating global data sets, which will ultimately provide definitive evidence of pathogenicity.²²

Although the size of our study cohort limited the potential to discover new disease genes and risk loci, and did not include exclusion of repeat expansions such as *C9orf72*, we saw enrichment of *GBA* alleles and *APOE* $\epsilon 4$ alleles in DLB. In total, 48 patients (55.2%) possessed an *APOE* $\epsilon 4$ allele, with 5 (5.7%) having a variant in *GBA*, together with four (4.4%) having likely pathogenic alleles (potentially with incomplete penetrance). Therefore, 62.6% of patients harbour a risk factor or potentially pathogenic allele. This could explain why DLB is a relatively common disorder in the population, with an increased risk of

Table 3. APOE genotype of all cases (excluded confirmed pathogenic variants) and controls

Study size	APOE genotype							
	4/4	3/3	2/2	4/3	3/2	4/2	$\epsilon 4$ carrier	
Controls	93	1	54	2	24	12	0	25
DLB patients	87	3	33	0	45	6	0	49
P-value	0.35	0.0076	0.50	0.0004	0.22	1.0	0.0001	

Abbreviation: DLB, dementia with Lewy body. Comparison between groups (patients $n=87$, controls $n=91$) performed by Fisher's exact test. *APOE* $\epsilon 4$ carrier determined by the presence of at least one *APOE* $\epsilon 4$ allele.

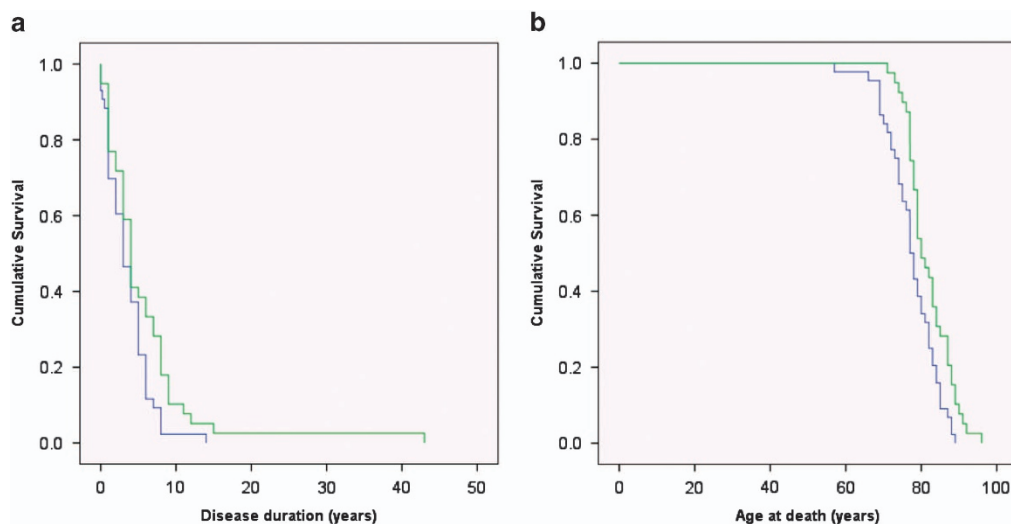


Figure 2. Kaplan–Meier survival curves for DLB patients by APOE allele. Kaplan–Meier survival curves for DLB patients by APOE allele carrying at least one *APOE* $\epsilon 4$ allele ($n=43$, blue line), compared with non-*APOE* $\epsilon 4$ carriers ($n=39$, green line). Despite there being no significant difference in the age of onset of the DLB (see Results), *APOE* $\epsilon 4$ carriers (a) lived for a shorter period of time following diagnosis ($P=0.036$, log rank, Mantel–Cox test), and thus (b) died at a younger age ($P=0.005$, log rank, Mantel–Cox test) than non-*APOE* $\epsilon 4$ carriers. DLB, dementia with Lewy body.

disease within families, but few pedigrees suggestive of highly penetrant alleles. Finally, the association between *APOE* genotype and clinical progression has, to our knowledge, not been previously described, and has implications for cohort stratification in treatment studies.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

This study was funded by the NHS National Institute of Health Research Biomedical Research Unit for Lewy body dementia at Newcastle upon Tyne Hospitals NHS Foundation Trust and Newcastle University. Tissue for this study was provided by Newcastle Brain Tissue Resource, which is funded in part by a grant from the UK Medical Research Council and by Brains for Dementia Research, a joint venture between Alzheimer's Society and Alzheimer's Research UK. MJK is a Wellcome Trust Clinical Research Training Fellow. PFC is a Wellcome Trust Senior Fellow in Clinical Science and National Institute for Health Research Senior Investigator. He receives funding from the Medical Research Council and the National Institute for Health Research Biomedical Research Centre for Ageing and Age-Related Disease award to the Newcastle upon Tyne Foundation Hospitals National Health Service Trust. The funding sources had no role in study design, data collection/analysis, the writing of the paper or the decision of when or where to publish it. The views expressed here are the views of the authors and not necessarily those of the NHS, NIHR or the Department of Health.

REFERENCES

- Rahkonen T, Eloniemi-Sulkava U, Rissanen S, Vatanen A, Viramo P, Sulkava R. Dementia with Lewy bodies according to the consensus criteria in a general population aged 75 years or older. *J Neurol Neurosurg Psychiatry* 2003; **74**: 720–724.
- Bostrom F, Jonsson L, Minthon L, Londos E. Patients with Lewy body dementia use more resources than those with Alzheimer's disease. *Int J Geriatr Psychiatry* 2007; **22**: 713–719.
- McKeith IG, Dickson DW, Lowe J, Emre M, O'Brien JT, Feldman H *et al*. Diagnosis and management of dementia with Lewy bodies: third report of the DLB Consortium. *Neurology* 2005; **65**: 1863–1872.
- Nervi A, Reitz C, Tang MX, Santana V, Piriz A, Reyes D *et al*. Familial aggregation of dementia with Lewy bodies. *Arch Neurol* 2011; **68**: 90–93.
- Lane R, He Y, Morris C, Leverenz JB, Emre M, Ballard C. BuChE-K and *APOE* epsilon4 allele frequencies in Lewy body dementias, and influence of genotype and hyperhomocysteinemia on cognitive decline. *Mov Disord* 2009; **24**: 392–400.
- Clark LN, Katsaklis LA, Wolf Gilbert R, Dorado B, Ross BM, Kisselev S *et al*. Association of glucocerebrosidase mutations with dementia with Lewy bodies. *Arch Neurol* 2009; **66**: 578–583.
- Bras J, Guerreiro R, Darwent L, Parkkinen L, Ansorge O, Escott-Price V *et al*. Genetic analysis implicates *APOE*, *SNCA* and suggests lysosomal dysfunction in the etiology of dementia with Lewy bodies. *Hum Mol Genet* 2014; **23**: 6139–6146.
- Harding AJ, Das A, Kril JJ, Brooks WS, Duffy D, Halliday GM. Identification of families with cortical Lewy body disease. *Am J Med Genet B Neuropsychiatr Genet* 2004; **128**: 118–122.
- Braak H, Alafuzoff I, Arzberger T, Kretschmar H, Del Tredici K. Staging of Alzheimer disease-associated neurofibrillary pathology using paraffin sections and immunocytochemistry. *Acta Neuropathol* 2006; **112**: 389–404.
- Alafuzoff I, Ince PG, Arzberger T, Al-Sarraj S, Bell J, Bodi I *et al*. Staging/typing of Lewy body related alpha-synuclein pathology: a study of the BrainNet Europe Consortium. *Acta Neuropathol* 2009; **117**: 635–652.
- Thal DR, Rub U, Orantes M, Braak H. Phases of A beta-deposition in the human brain and its relevance for the development of AD. *Neurology* 2002; **58**: 1791–1800.
- Thal DR, Ghebremedhin E, Orantes M, Wiestler OD. Vascular pathology in Alzheimer disease: correlation of cerebral amyloid angiopathy and arteriosclerosis/lipohyalinosis with cognitive decline. *J Neuropathol Exp Neurol* 2003; **62**: 1287–1301.
- Richards S, Aziz N, Bale S, Bick D, Das S, Gastier-Foster J *et al*. Standards and guidelines for the interpretation of sequence variants: a joint consensus recommendation of the American College of Medical Genetics and Genomics and the Association for Molecular Pathology. *Genet Med* 2015; **17**: 405–423.
- Guerreiro R, Wojtas A, Bras J, Carrasquillo M, Rogaeva E, Majounie E *et al*. TREM2 variants in Alzheimer's disease. *N Engl J Med* 2013; **368**: 117–127.
- An XK, Peng R, Li T, Burgunder JM, Wu Y, Chen WJ *et al*. LRRK2 Gly2385Arg variant is a risk factor of Parkinson's disease among Han-Chinese from mainland China. *Eur J Neurol* 2008; **15**: 301–305.
- Lleo A, Blesa R, Gendre J, Castellvi M, Pastor P, Queralt R *et al*. A novel presenilin 2 gene mutation (D439A) in a patient with early-onset Alzheimer's disease. *Neurology* 2001; **57**: 1926–1928.
- Sassi C, Guerreiro R, Gibbs R, Ding J, Lupton MK, Troakes C *et al*. Investigating the role of rare coding variability in Mendelian dementia genes (*APP*, *PSEN1*, *PSEN2*, *GRN*, *MAPT*, and *PRNP*) in late-onset Alzheimer's disease. *Neurobiol Aging* 2014; **35**: 2881.e1–2881.e6.
- Parkinson N, Ince PG, Smith MO, Highley R, Skibinski G, Andersen PM *et al*. ALS phenotypes with mutations in *CHMP2B* (charged multivesicular body protein 2B). *Neurology* 2006; **67**: 1074–1077.
- Fecto F, Yan J, Vemula SP, Liu E, Yang Y, Chen W *et al*. SQSTM1 mutations in familial and sporadic amyotrophic lateral sclerosis. *Arch Neurol* 2011; **68**: 1440–1446.
- Le Ber I, Camuzat A, Guerreiro R, Bouya-Ahmed K, Bras J, Nicolas G *et al*. SQSTM1 mutations in French patients with frontotemporal dementia or frontotemporal dementia with amyotrophic lateral sclerosis. *JAMA Neurol* 2013; **70**: 1403–1410.
- Bras J, Simon-Sanchez J, Federoff M, Morgado A, Januario C, Ribeiro M *et al*. Lack of replication of association between *GIGYF2* variants and Parkinson disease. *Hum Mol Genet* 2009; **18**: 341–346.
- MacArthur DG, Manolio TA, Dimmock DP, Rehm HL, Shendure J, Abecasis GR *et al*. Guidelines for investigating causality of sequence variants in human disease. *Nature* 2014; **508**: 469–476.
- Leverenz JB, Fishel MA, Peskind ER, Montine TJ, Nochlin D, Steinbart E *et al*. Lewy body pathology in familial Alzheimer disease: evidence for disease- and mutation-specific pathologic phenotype. *Arch Neurol* 2006; **63**: 370–376.
- Tanikawa S, Mori F, Tanji K, Kakita A, Takahashi H, Wakabayashi K. Endosomal sorting related protein *CHMP2B* is localized in Lewy bodies and glial cytoplasmic inclusions in alpha-synucleinopathy. *Neurosci Lett* 2012; **527**: 16–21.
- Tanji K, Odagiri S, Miki Y, Maruyama A, Nikaido Y, Mimura J *et al*. p62 deficiency enhances alpha-synuclein pathology in mice. *Brain Pathol* 2014; **25**: 552–564.
- Cuyvers E, van der Zee J, Bettens K, Engelborghs S, Vandenbulcke M, Robberecht C *et al*. Genetic variability in *SQSTM1* and risk of early-onset Alzheimer dementia: a European early-onset dementia consortium study. *Neurobiol Aging* 2015; **36**: 2005.e15–2005.e22.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

Supplementary Information accompanies the paper on the Translational Psychiatry website (<http://www.nature.com/tp>)