www.nature.com/tp

ORIGINAL ARTICLE Psychiatric disturbances regulate the innate immune system in CSF of conscious mice

ES Onaivi¹, N Schanz¹ and ZC Lin^{2,3,4}

Environment may affect brain activity through cerebrospinal fluid (CSF) only if there are regulatory molecules or cascades in CSF that are sensitive to external stimuli. This study was designed to identify regulatory activity present in CSF, better elucidating environmental regulation of brain function. By using cannulation-based sequential CSF sampling coupled with mass spectrometry-based identification and quantification of proteins, we show that the naive mouse CSF harbors, among 22 other pathways, the innate immune system as a main pathway, which was downregulated and upregulated, respectively, by acute stressor (AS) and acute cocaine (AC) administrations. Among novel processes and molecular functions, AS also regulated schizophrenia-associated proteins. Furthermore, AC upregulated exosome-related proteins with a false discovery rate of 1.0×10^{-16} . These results suggest that psychiatric disturbances regulate the neuroimmune system and brain disorder-related proteins, presenting a sensitive approach to investigating extracellular mechanisms in conscious and various mouse models of psychiatric disorders.

Translational Psychiatry (2014) 4, e367; doi:10.1038/tp.2014.5; published online 4 March 2014

Keywords: addiction; cocaine; signaling pathways; stress

INTRODUCTION

Neuroimmune systems, especially the innate immune system, are being increasingly implicated in mental disorders.^{1–4} More specifically, the immune systems are postulated to prune dendrites and help maintain neuronal plasticity in the brain.^{5–7} The fact that neuronal plasticity are subject to environmental regulations suggests that these neuroimmune systems be also sensitive to environment, which awaits experimental evidence for validation.

Cerebrospinal fluid (CSF) has been used as a source of biomarkers in humans with neuropsychiatric disorders and as an avenue to deliver therapeutic agents directly into CNS in rodent models.^{8–12} As extracellular molecules in CSF bath brain cells and modulate cellular plasticity constantly,¹³ monitoring the dynamics of these molecules in rodent models may unravel the brain mechanisms underlying behavior and pathogenesis. However, such a monitoring method is missing at present.

Mice are primary biomedical models and research tools and the ability to profile CSF molecules in mice will help better understand the molecular mechanisms for brain function and disorder. Mouse models for human neuropsychiatric disorders allow easy genetic modifications through different approaches and technologies.^{14–18} In most cases, CSF is used as a drug delivery system by implanting intracerebroventricular (ICV) cannulae through skull into the ventricles in brain.^{19,20}

Here we report a stereotaxic approach coupled with protein profiling analysis of sequential multiple CSF samples from the same group of live C57BL/6J mice, dissecting environmental regulation of multiple pathways in CSF of conscious mouse brain.

MATERIALS AND METHODS

Animals and stereotaxic surgery

Adult male mice (25–30 g, C57BL/6 from Charles River Laboratories, Wilmington, MA, USA) were subjected to standard stereotaxic surgery for the implantation of unilateral or bilateral guide cannulae. In brief, the animals were anesthetized and placed in a Kopf stereotaxic frame, and an incision was made to expose the skull. Small holes were drilled and the guide cannula (Plastics One, Roanoke, VA, USA) was lowered 0.5 mm above the lateral ventricles (ICV) and cemented with dental acrylic cement mixed with cyanoacrylate glue to secure the cannula to the skull. Cannula dummies (Plastics One) are inserted into the guide cannulas to keep them patent. The coordinates used for ICV in mm was Ant. \pm 3.5, Vert. \pm 1.3 and Lat. \pm 1.0 according to the atlas of Slotnick and Leonard.

Histological verification of cannula placement

Seven days after surgery, mice were anesthetized with ketamine/xylazine solution and transcardially perfused with 0.9% NaCl followed by 4% paraformaldehyde in 0.1 M phosphate buffer, pH 7.1. The brains were carefully removed and blocked to obtain the forebrain region. The forebrain was postfixed for 48 h, cryoprotected in 20% sucrose overnight and then rapidly frozen in dry ice. A complete set of coronal serial sections was cut through the forebrain at 30 mm, mounted on microscope slides and thionin counterstained to identify the site where the cannula was placed.

Acute stressor (AS) and acute cocaine (AC) treatment

Approximately 30 min after the first withdrawal of CSF, each of nine mice was placed into warm water in a 4-l beaker (three-fourth of the volume filled), followed by forced swimming for 15 min. Wet mice were all dried out under a warm lamp (60 W) and 30 min later withdrawn for a second CSF sample. Next day, each mouse was intraperitoneally injected with cocaine (single dose at 20 mg kg⁻¹), followed by a third CSF withdrawal 30 min later.

¹Department of Biology, William Paterson University, Wayne, NJ, USA; ²Department of Psychiatry, Harvard Medical School, Boston, MA, USA; ³NeuroDiscovery Center, Harvard Medical School, Boston, MA, USA and ⁴Laboratory of Psychiatric Neurogenomics, Division of Alcohol and Drug Abuse, McLean Hospital, Belmont, MA, USA. Correspondence: Dr ZC Lin, Laboratory of Psychiatric Neurogenomics, Division of Alcohol and Drug Abuse, McLean Hospital Mailstop 318, 115 Mill Street, Belmont 02478, MA, USA. E-mail: zhicheng_lin@hms.harvard.edu

Received 3 October 2013; revised 26 November 2013; accepted 7 December 2013

Psychiatric disturbances in CSF of mice ES Onaivi et al

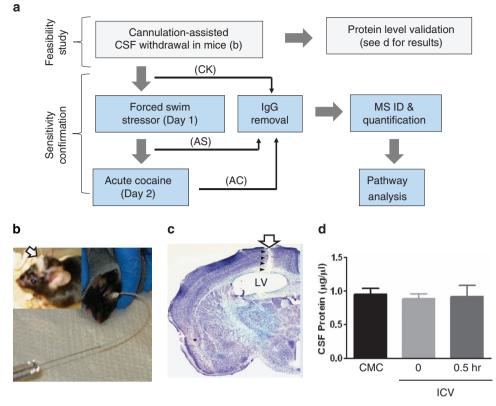


Figure 1. Cannulation-based repeated sampling of cerebrospinal fluid (CSF) from intracerebroventricular (ICV) of mouse brain. (a) Study design. Notice that feasibility study and sensitivity confirmation used two different groups of mice. (b) ICV cannulation and CSF withdrawal. Insert, a close-up of an installed ICV cannula indicated by an open arrow. (c) Postsurgery verification of cannula installed to ICV, based on coronal sectioning of the brain. Open arrow, drilled hole for guide cannula; arrowhead, hole directed into lateral ventricle (LV). (d) Constant protein content in CSF withdrawn from mouse cerebellomedullary cistern (CMC) or ICV with a 0.5-h interval, based on Invitrogen's . NanoOrange quantification technology.

CSF sampling

Sampling from the cerebellomedullary cistern (CMC) followed a published procedure.²¹ To withdraw CSF from the ICV cannula, tubing (~2 cm in length, Plastics One) was connected to the internal (Plastics One) of the cannula at one end and to a 10-µl Hamilton syringe at the other end. Negative pressure was placed on the internal by gently pulling the plunger of the syringe. Each withdrawal was ~2 µl, about 0.5 h before or after treatment. Collected CSF was transferred into an ice-cold tube, followed by -20 °C storage before further processing by mass spectrometry (MS) analysis for identification and quantification of proteins.

MS-based protein profiling

CSF was pooled from nine mice for each condition. We used Pierce's Antibody-Based Albumin/IgG Removal Kit to deplete albumin and IgG before MS analysis at Bioproximity LLC (Chantilly, VA, USA). The samples were run on the Thermo LTQ Velos (San Jose, CA, USA; a dual-pressure linear ion trap mass spectrometer). The Velos utilized the same S-lens as the Q-Exactive to focus the ion cloud as it entered the mass spectrometer and improved overall sensitivity. The linear ion trap enabled very fast sequencing speed of 15 MS/MS scans in 1.8 s. Collision-induced fragmentation was used to fragment peptides for sequence analysis. CID produced peptide ladders from both N- and C-terminal ends of the peptide, allowing for complementary and confirmatory sequence interpretation resulting in high confidence assignments. The limit of identification sensitivity of MS was ~1 fmol.²

MS data analysis

Parsimony type was used to classify protein forms that could have overlapping peptide sequences.²³ PepHits were used to normalize for load variation and normalized PepHits were used to calculate a sensitivity value: base 2 log of ratio (acute stressor (AS)/control (CK), acute cocaine (AC)/AS or AC/CK) for condition-related difference. sensitivity value larger than 1

Translational Psychiatry (2014), 1-7

(twofold upregulation) or less than -1 (twofold downregulation or 0.5-fold) are arbitrarily cutoffs for biological significances.

Pathway analysis

We used the frequently updated MetaCore software/database to map pathways (Thomson Reuters, New York, NY, USA).²⁴ Proteins with biologic significance per sensitivity values were analyzed for pathway regulations or subcellular localizations with significance or enrichment score (-log10Pvalue). In Enrichment by Protein Function interactome analysis, P-values reflect the probability to have that many or more objects of that protein class in an experiment than would be expected by chance (with a positive Z-score). If a Z-score is negative, the P-value represents the probability to have that many or fewer objects of that protein class than would be expected by chance. Protein function categorization was performed by listing of Network Objects or based on Public Ontologies (GO Molecular Functions), both implemented in MetaCore.

Interactions between AS and AC were evaluated by carrying out the interactions between datasets analysis of AS- or AS-regulated proteins (also implemented in MetaCore). This algorithm was to examine whether the objects contained in one dataset had significant numbers of interactions with the other active datasets. A P-value described the probability of the observed ratio of connections with the occurring merely by chance. A Z-score gave a measure of how saturated the potential number of interactions between datasets were. The higher the Z-score the more saturated the object's connections were with objects from the dataset.

RESULTS

The purpose of this study was to map out regulatory pathways in the CSF that may not only mirror but also modulate brain activity. We were focused on pathway analysis because molecular pathways have central roles in both biologic and disease

Psychiatric disturbances in CSF of mice ES Onaivi *et al*

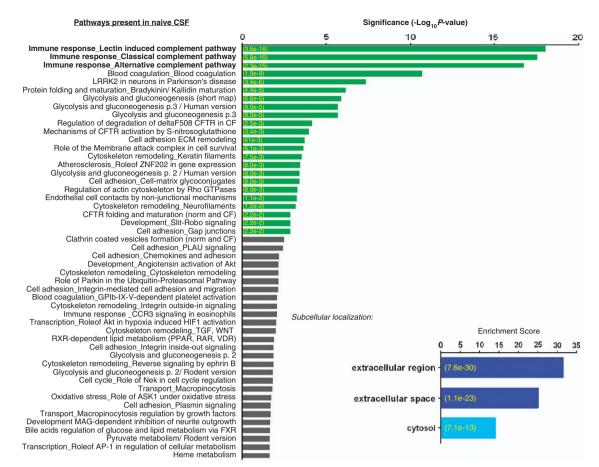


Figure 2. Pathways present in naive cerebrospinal fluid (CSF). Green bar, 23 significant pathways; dark gray bar, not significant. Insert, top two localizations of CSF proteins to extracellular regions. Parenthesis on bar, false discovery rate (FDR).

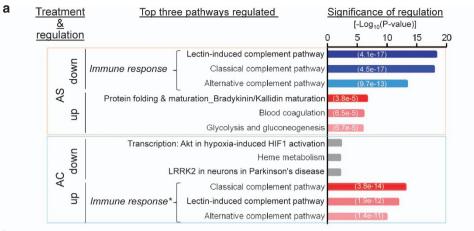
mechanisms.²⁵⁻³² The study design is outlined in Figure 1a. In adult mice, after stereotaxic surgery and unilateral or bilateral cannulation, ICV was used for repeated CSF sampling (Figure 1b) for the determination of dynamics of protein composition in the extracellular space. The correct position of cannula ICV implantation was confirmed by histological examination (Figure 1c, where slightly large appearance of the ventricle could be due to the cannula placement). We found that the average protein level from an ICV sample ($\sim 2 \mu$ I) was neither different from one in the next sample that was collected 0.5 h later, nor different from one in a CMC sample (Figure 1d). These data suggest that 2 µl sampling does not significantly affect protein contents in the subsequent samples from the same mice. Next, we carried out protein profiling analysis of CSF samples from mouse brain before (CK) and after AS, and after AC by MS analysis for identification and quantification of these proteins.

As a result, a total of 268 proteins were detected in the CSF samples at the sensitivity of ~1 fmol; 134 (50.0%) were regulated by AS and 185 (69.0%) were regulated by AC, comparing with CK (see Supplementary Table 1 for a complete list of regulated proteins). Those regulated were mainly peptidase inhibitor activity (see Supplementary Figure 1). Among the AS-regulated proteins, 14 were known ligands (P=0.0000742, Z-score=4.934), 10 were proteases (P=0.0282, Z-score=2.297) and 34 were known enzymes (P=0.0120, Z-score=2.474), based on enrichment by protein function interactome analysis. Among the AC-regulated proteins, 17 were known ligands (P=0.000179, Z-score=4.585), 13 were proteases (P=0.0212, Z-score=2.205) and 45 were known enzymes (P=0.0212, Z-score=2.19). In all, 62.2% of the AS-upregulated proteins were also upregulated proteins were

also downregulated by AC (representing 52.4% of its own; see Supplementary Table 1 for details). On the basis of these enrichments by protein function, both environments regulated more enzymes than ligands or proteases. Only 188 of the proteins were detected in the naive CSF.

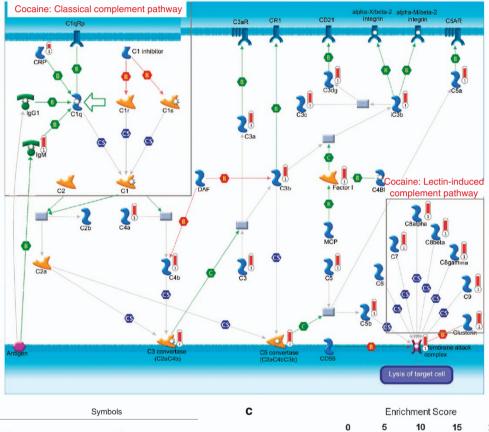
Systematic analysis of the 188 proteins revealed 23 significant pathways in the naive CSF. Among the 23 significant pathways in the naive CSF, the complement pathways of the innate immune system represent the major ones (Figure 2). They included the lectin-induced ($P = 9.59 \times 10^{-19}$, false discovery rate (FDR) = 3.6×10^{-16}), classical ($P = 3.10 \times 10^{-18}$, FDR = 5.8×10^{-16}) and alternative ($P = 1.86 \times 10^{-17}$, FDR = 2.3×10^{-15}) complement pathways. These findings suggest that this immune system may be pruning synapses constantly to facilitate synaptic plasticity in the CNS. Other pathways with less significances were blood coagulation, LRRK2 ($P = 4.83 \times 10^{-8}$), protein folding ($P = 7.39 \times 10^{-7}$) and glycolysis ($P = 1.29 - 2.04 \times 10^{-6}$) pathways.^{33,34} Cell surface matrix and cytoskeleton configurations were also reflected by the CSF pathways. Majority of the 188 proteins were known localized to extracellular region ($P = 1.30 \times 10^{-29}$; Figure 2, insert), consistent with the fact that these were sampled from the extracellular space inside the brain (ICV). In addition to these pathways, responses to wounding $(P = 1.34 \times 10^{-39})$ and stress $(P = 1.71 \times 10^{-35})$ are the top processes, and endopeptidase inhibitions ($P = 3.86 \times 10^{-46}$) or enzyme activity ($P = 4.32 \times 10^{-42}$) regulations are the top molecular functions in the ICV^{35,36} (Supplementary Figure 2).

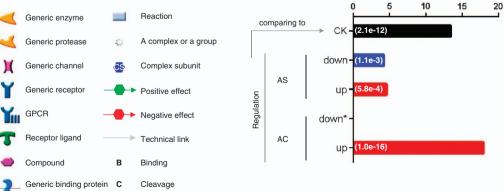
Complement pathways were regulated by both AS and AC but in two opposite directions. Fifteen-minute forced swimming downregulated the complement pathways the most and by contrast, AC upregulated the complement pathways the most by comparing with CK (Figure 3a). As evidenced for the significance





npg





npg

of regulations, the *P*-values were 5.43×10^{-19} for AS downregulation and 1.56×10^{-13} for AC upregulation of the lectininduced pathway; 1.20×10^{-18} for AS downregulation and 1.37×10^{-14} for AC upregulation of the classical pathway; and 3.84×10^{-14} for AS downregulation and 1.38×10^{-16} for AC upregulation of the alternative pathways (Figure 3a). In this study, we did not sample the CSF immediately before the AC, as a control for the AS 'hangover' effects, but when compared with the naive CSF, AC still upregulated these complement pathways the most (Supplementary Figure 3). AS upregulated the protein folding (Bradykinin/Kallidin maturation), blood coagulation and glycolysis pathways all with smaller significances (P = 2.26-- 12.1×10^{-7}). AC downregulated none of the pathways in a significant manner (the latest pathway datasets indicated that AC might downregulate C5, which however is not confirmed yet).

Particular proteins in the regulated complement pathways are indicated in the pathway map (Figure 3b). The AS-downregulated proteins included C3, C4, C5, Factor 1, and the C3 and C5 convertases and the membrane attach complex in the lectin-induced complement pathway. The AC-upregulated proteins included CRP and IgM in the classical pathway, and C7, C8, C9 and clusterine in the lectin-induced complement pathway. This profiling method may help us to better understand how complement pathways are regulated, contributing to the dynamics of circuitry in the conscious CNS.^{6,37}

In addition to the 6 AS-regulated and 3 AC-regulated pathways, AS regulated 36 other pathways (20 downs and 16 ups) and AC upregulated 20 other pathways (Figure 2 and Supplementary Figure 3). Most of them were novel, including cell adhesion remodeling and extracellular cytoskeleton. AS upregulated processes such as the Kallikren-kinin (inflammation) system, cytoskeleton rearrangement and AC upregulated the Kallikren-kinin system as well, along with interleukin-6 signaling and integrin-mediated cell matrix adhesion. AC upregulated endopeptidase regulators significantly ($P = 3.49 \times 10^{-45}$), whereas AS downregulated these molecular functions ($P = 3.49 \times 10^{-22}$, data not shown). These findings suggested that the CSF pathways were sensitive to extensive regulations by environment, which may underlie an extracellular interaction between environment and brain circuitry.

Majority of the regulated proteins have not been characterized intensively yet. Supplementary Figure 4 shows six of them that were differentially affected by AS and AC. For example, Serpina3C and malate dehydrogenase 1 protein levels were downregulated and upregulated (to 0.3-fold and 5.91-fold) by AS only. Interestingly, the human ortholog Serpina and the human malate dehydrogenase 1 were both associated with Schizophrenia,³⁸⁻⁴ perhaps underlying stressors as a risk factor for this brain disorder. The inflammatory response protein Mug1 and the Felty syndromerelated Eef1a1 protein levels were largely upregulated (3.35-fold and 13.24-fold) by AC only. The copper-carrying protein ceruloplasmin and serum amyloid A4 levels were first downregulated (0.41-fold and 0.1-fold) by AS and then upregulated (2.80-fold and 2.70-fold) by AC. In addition, the levels of cathelicidin antimicrobial peptide were downregulated to 0.17-fold by AS but upregulated to 2.82-fold by AC. The cytoskeletal protein Tubb5 was (up 11.1-fold) regulated by AC only. The GABAA receptor modulator⁴² diazepam-binding inhibitor's levels were upregulated to 5.54-fold by AS but downregulated to 0.48-fold by AC. The levels of candidate signaling protein contactin associated proteinlike 5A were upregulated to 5.17-fold by AS but downregulated to 0.1-fold by AC. Although CSF proteins might have unusual composition, these findings imply new pathways of brain response to changing environment, warranting future investigation of their roles in cellular function of the brain.

More interestingly, AC upregulated proteins associated with extracellular vesicular exosomes, with a high enrichment score of 17.9 and a FDR of 1.0×10^{-16} , with little downregulation of proteins in this category (Figure 3c). In contrast, AS exerted both up- and downregulations with much smaller enrichment scores (4.6 and 4.2). These findings suggest that AC could alter another type of intercellular communication, in addition to neurotransmission.

Finally, interaction analysis showed that all four data sets (up- or downregulations of AS or AC) were overconnected; none were underconnected (Supplementary Table 2). Specifically, AC-upregulated proteins were most significantly overconnected with AS-sensitive proteins (*Z*-score>11.4), as well as AC-downregulated ones (*Z*-score = 9.8). By contract, AC-downregulated proteins were least significantly overconnected with AS-sensitive proteins (*Z*-score < 5.2). Such an overconnected pattern suggests that cocaine-stimulated brain activity be closely related to stress pathways, which merits further study.

DISCUSSION

By using a simple and sensitive method to monitor molecular composition in CSF of conscious mouse brain, we observed, for the first time, that the innate immune system represented the main pathways inside the CSF and that AS and cocaine could both regulate the innate immune system in the brain but in two opposite directions. In addition, we show that environment may regulate exosome-related protein levels in CSF.

To our knowledge, this is the first *in vivo* study on CSF proteins in intact conscious brains of adult mice. Altered concentrations of proteins or small molecules in CSF have been demonstrated for psychiatric disorders such as schizophrenia, bipolar disorder and depression.^{43–46} In fact, many novel proteins are present in the CSF but little is known about their modes of action and function yet. In this study, we report that in both AS- and AC-treated animals more proteins were upregulated and fewer proteins were downregulated (Supplementary Table 1). These results suggest that novel CSF activity remains to be discovered for their role and impact on CNS function and dysfunction. Some of these proteins with known intracellular activity (such as the AS- and ACupregulated Eukaryotic Translation Initiation Factor 4A/Eif4a1 or RAP1, GTP-GDP Dissociation Stimulator 1/Rap1gds1) may have been released into extracellular space by subcellular mechanisms such as formation of extracellular vesicles^{47–49} including exosomes (Figure 3c). Therefore, it will be interesting to examine which of these molecules, including proteins, genetic materials and small molecules, in the CSF are confined to the extracellular vesicles because these extracellular vesicles may affect intracellular activity of brain cells. In addition, the innate immune system may have had an important role in environmental regulations of neuronal activity including morphologic neuroplasticity. One limitation of this study design lies in lack of control for hangover effects of AS on the AC results. However, the AC upregulation of the innate

Figure 3. Cerebrospinal fluid (CSF) pathways regulated by acute stressor (AS) and acute cocaine (AC). (a) Top three pathways downregulated (down) or upregulated (up) by AS (upper portion/brown rectangle) or by AC (lower portion/aquamarine rectangle). Color bars, statistically significant; gray bar, not significant. (b) Pathway map for AS-downregulated lectin-induced complement pathway. Upleft insert, unique proteins of AC-upregulated classical complement pathway; downright insert, unique proteins of AC-upregulated lectin-induced complement pathway. Red thermometer, significantly regulated proteins. (c) Upregulation of exosome-related proteins by AC. Asterisk, not listed among top 50 categories; parenthesis on bar, false discovery rate (FDR). In all, 11.6% of the AC-upregulated; see Supplementary Table 1 for details. Enrichment score ($-\log_{10}p$ -value) of regulation is based on comparison with control (CK; black bar).

[◀]

6

immune system holds true based on comparison with either CK or AS (Supplementary Figure 3).

In this study we have shown that the lateral ventricles in adult mouse brain, after stereotaxic surgery and cannulation, unilaterally or bilaterally, can be utilized for repeated CSF sampling for the determination of and alterations in protein dynamics in the brain. Previous continuous CSF sampling in mice from the brain and cisternum magnum had been controversial because of low volume and production rate in mice compared with rats and humans. However, with new and sensitive protein detection technology, we report constant CSF protein content of $1 \mu q \mu l^{-1}$ sensitivity from the multiple CSF sampling from the lateral ventricles. Further studies will be necessary to localize brainarea-specific alterations associated with the global molecular protein dynamics predicted by this model. That approach will be much straightforward dissection of brain regions to determine associated protein dynamics with specific function for further analysis. This will provide a powerful tool to determine the roles of such identified proteins and transcription factors and specific brain regions in neuropsychiatric disorders.

As mentioned above, this study at its current stage has major limitations. The controls used were not necessarily the best controls, particularly considering the serial nature of the design and the preliminary nature of the development of the approach: stressed versus non-stressed and cocaine versus saline-treated would have been much better comparisons. As of the limited design, we are unable to evaluate yet the time courses of the regulations, in particular.

In summary, we have used stereotaxic surgical approaches coupled with protein profiling analysis of sequential multiple CSF sampling in conscious mouse models for neuropsychiatric disturbances and demonstrated the potentials of monitoring molecular dynamics in nervous system extracellular space in mouse brain. This method, when combined with site-specific investigation of neurobiological mechanisms in the brain, provides a powerful tool to screen and identify the screen-associated realtime changes in nonsynaptic signal transmission in such animal models of brain disorders.

CONFLICT OF INTEREST

The authors declare no conflict of interest.

ACKNOWLEDGMENTS

We are indebted to Dr Patricia Tagliaferro for the helpful guide and assistance during the stereotaxic surgery and histological verification of guide-cannula placement, to Melinda Baker for the analytical assistance and to the anonymous reviewer for insightful suggestions. This study was supported by WPUNJ-Center for research, the Dean Dr Ken Wolf support for student workers with ESO, NIH grant DA032890 to ESO and NIH grants DA031573 and DA02409 to ZCL.

AUTHOR CONTRIBUTIONS

ESO and ZCL designed and performed the animal experiments including CSF withdrawal; ESO proof-read the manuscript; NS provided laboratory technical assistance during stereotaxic cannulation and withdrawal of CSF. ZCL conceived the idea, developed the project, analyzed the data and wrote the manuscript.

REFERENCES

- Gold PW, Licinio J, Pavlatou MG. Pathological parainflammation and endoplasmic reticulum stress in depression: potential translational targets through the CNS insulin, klotho and PPAR-γ systems. *Mol Psychiatry* 2013; **18**: 154–165.
- 2 Wei L, Simen A, Mane S, Kaffman A. Early life stress inhibits expression of a novel innate immune pathway in the developing hippocampus. *Neuropsychopharmacology* 2012; **37**: 567–580.

- 3 Terrando N, Eriksson LI, Ryu JK, Yang T, Monaco C, Feldmann M *et al.* Resolving postoperative neuroinflammation and cognitive decline. *Ann Neurol* 2011; **70**: 986–995.
- 4 Raison CL, Rye DB, Woolwine BJ, Vogt GJ, Bautista BM, Spivey JR *et al.* Chronic interferon-alpha administration disrupts sleep continuity and depth in patients with hepatitis C: association with fatigue, motor slowing, and increased evening cortisol. *Biol Psychiatry* 2010; **68**: 942–949.
- 5 Boulanger LM. Immune proteins in brain development and synaptic plasticity. *Neuron* 2009; **64**: 93–109.
- 6 Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, Yamasaki R *et al.* Microglia sculpt postnatal neural circuits in an activity and complementdependent manner. *Neuron* 2012; **74**: 691–705.
- 7 Di Filippo M, Chiasserini D, Gardoni F, Viviani B, Tozzi A, Giampà C et al. Effects of central and peripheral inflammation on hippocampal synaptic plasticity. *Neurobiol Dis* 2013; **52**: 229–236.
- 8 Bateman RJ, Xiong C, Benzinger TL, Fagan AM, Goate A, Fox NC *et al.* Clinical and biomarker changes in dominantly inherited Alzheimer's disease. *N Engl J Med* 2012; 367: 795–804.
- 9 Papadopoulos MC, Verkman AS. Aquaporin water channels in the nervous system. *Nat Rev Neurosci* 2013; **14**: 265–277.
- 10 Höftberger R, Sabater L, Ortega A, Dalmau J, Graus F. Patient with homer-3 antibodies and cerebellitis. *JAMA Neurol* 2013; **70**: 506–509.
- 11 Parnetti L, Castrioto A, Chiasserini D, Persichetti E, Tambasco N, El-Agnaf O et al. Cerebrospinal fluid biomarkers in Parkinson disease. Nat Rev Neurol 2013; 9: 131–140.
- 12 Luykx JJ, Bakker SC, Lentjes E, Neeleman M, Strengman E, Mentink Let al. Genome-wide association study of monoamine metabolite levels in human cerebrospinal fluid. *Mol Psychiatry* 2014; **19**: 228–234.
- 13 Stephan AH, Barres BA, Stevens B. The complement system: an unexpected role in synaptic pruning during development and disease. *Annu Rev Neurosci* 2012; 35: 369–389.
- 14 Cramer PE, Cirrito JR, Wesson DW, Lee CY, Karlo JC, Zinn AE et al. ApoE-directed therapeutics rapidly clear β-amyloid and reverse deficits in AD mouse models. *Science* 2012; **335**: 1503–1506.
- 15 Han S, Tai C, Westenbroek RE, Yu FH, Cheah CS, Potter GB et al. Autistic-like behaviour in Scn1a+/- mice and rescue by enhanced GABA-mediated neurotransmission. *Nature* 2012; 489: 385–390.
- 16 Giovanoli S, Engler H, Engler A, Richetto J, Voget M, Willi R et al. Stress in puberty unmasks latent neuropathological consequences of prenatal immune activation in mice. Science 2013; 339: 1095–1099.
- 17 Niwa M, Jaaro-Peled H, Tankou S, Seshadri S, Hikida T, Matsumoto Y et al. Adolescent stress-induced epigenetic control of dopaminergic neurons via glucocorticoids. *Science* 2013; **339**: 335–339.
- 18 Chaudhury D, Walsh JJ, Friedman AK, Juarez B, Ku SM, Koo JW et al. Rapid regulation of depression-related behaviours by control of midbrain dopamine neurons. *Nature* 2013; **493**: 532–536.
- 19 Vom Berg J, Prokop S, Miller KR, Obst J, Kälin RE, Lopategui-Cabezas I et al. Inhibition of IL-12/IL-23 signaling reduces Alzheimer's disease-like pathology and cognitive decline. Nat Med 2012; 18: 1812–1819.
- 20 Kallupi M, Cannella N, Economidou D, Ubaldi M, Ruggeri B, Weiss F *et al.* Neuropeptide S facilitates cue-induced relapse to cocaine seeking through activation of the hypothalamic hypocretin system. *Proc Natl Acad Sci USA* 2010; **107**: 19567–19572.
- 21 DeMattos RB, Bales KR, Parsadanian M, O'Dell MA, Foss EM, Paul SM et al. Plaqueassociated disruption of CSF and plasma amyloid-beta (Abeta) equilibrium in a mouse model of Alzheimer's disease. J Neurochem 2002; 81: 229–236.
- 22 Second TP, Blethrow JD, Schwartz JC, Merrihew GE, MacCoss MJ, Swaney DL et al. Dual-pressure linear ion trap mass spectrometer improving the analysis of complex protein mixtures. Anal Chem 2009; 81: 7757–7765.
- 23 Yang X, Dondeti V, Dezube R, Maynard DM, Geer LY, Epstein J. *et al.* DBParser: web-based software for shotgun proteomic data analyses. *J Proteome Res* 2004; 3: 1002–1008.
- 24 Ekins S, Nikolsky Y, Bugrim A, Kirillov E, Nikolskaya T. Pathway mapping tools for analysis of high content data. *Methods Mol Biol* 2007; **356**: 319–350.
- 25 Moylan S, Maes M, Wray NR, Berk M. The neuroprogressive nature of major depressive disorder: pathways to disease evolution and resistance, and therapeutic implications. *Mol Psychiatry* 2013; **18**: 595–606.
- 26 Balu DT, Li Y, Puhl MD, Benneyworth MA, Basu AC, Takagi S et al. Multiple risk pathways for schizophrenia converge in serine racemase knockout mice, a mouse model of NMDA receptor hypofunction. Proc Natl Acad Sci USA 2013; 110: E2400–E2409.
- 27 Rahrmann EP, Watson AL, Keng VW, Choi K, Moriarity BS, Beckmann DA et al. Forward genetic screen for malignant peripheral nerve sheath tumor formation identifies new genes and pathways driving tumorigenesis. *Nat Genet* 2013; 45: 756–766.



- 28 Kim SY, Adhikari A, Lee SY, Marshel JH, Kim CK, Mallory CS et al. Diverging neural pathways assemble a behavioural state from separable features in anxiety. Nature 2013; 496: 219–223.
- 29 Cui G, Jun SB, Jin X, Pham MD, Vogel SS, Lovinger DM *et al.* Concurrent activation of striatal direct and indirect pathways during action initiation. *Nature* 2013; **494**: 238–242.
- 30 Baxt LA, Garza-Mayers AC, Goldberg MB. Bacterial subversion of host innate immune pathways. *Science* 2013; **340**: 697–701.
- 31 Lu MS, Johnston CA. Molecular pathways regulating mitotic spindle orientation in animal cells. *Development* 2013; 140: 1843–1856.
- 32 Braunschweig U, Gueroussov S, Plocik AM, Graveley BR, Blencowe BJ. Dynamic integration of splicing within gene regulatory pathways. *Cell* 2013; 152: 1252–1269.
- 33 Williams FM, Carter AM, Hysi PG, Surdulescu G, Hodgkiss D, Soranzo N et al. Ischemic stroke is associated with the ABO locus: the EuroCLOT study. Ann Neurol 2013; 73: 16–31.
- 34 Jickling GC, Ander BP, Stamova B, Zhan X, Liu D, Rothstein L*et al.* RNA in blood is altered prior to hemorrhagic transformation in ischemic stroke. *Ann Neurol* 2013. : doi: 10.1002/ana.23883; e-pub ahead of print.
- 35 Sashindranath M, Sales E, Daglas M, Freeman R, Samson AL, Cops EJ et al. The tissue-type plasminogen activator-plasminogen activator inhibitor 1 complex promotes neurovascular injury in brain trauma: evidence from mice and humans. Brain 2012; 135: 3251–3264.
- 36 Vandenbroucke RE, Dejonckheere E, Van Lint P, Demeestere D, Van Wonterghem E, Vanlaere I *et al.* Matrix metalloprotease 8-dependent extracellular matrix cleavage at the blood-CSF barrier contributes to lethality during systemic inflammatory diseases. *J Neurosci* 2012; **32**: 9805–9816.
- 37 Berg A, Zelano J, Thams S, Cullheim S. The extent of synaptic stripping of motoneurons after axotomy is not correlated to activation of surrounding glia or downregulation of postsynaptic adhesion molecules. *PLoS One* 2013; 8: e59647.
- 38 Arion D, Unger T, Lewis DA, Levitt P, Mirnics K. Molecular evidence for increased expression of genes related to immune and chaperone function in the prefrontal cortex in schizophrenia. *Biol Psychiatry* 2007; 62: 711–721.
- 39 Fillman SG, Cloonan N, Catts VS, Miller LC, Wong J, McCrossin T *et al.* Increased inflammatory markers identified in the dorsolateral prefrontal cortex of individuals with schizophrenia. *Mol Psychiatry* 2013; **18**: 206–214.

- 40 Vawter MP, Ferran E, Galke B, Cooper K, Bunney WE, Byerley W. Microarray screening of lymphocyte gene expression differences in a multiplex schizophrenia pedigree. *Schizophr Res* 2004; 67: 41–52.
- 41 Vawter MP, Shannon Weickert C, Ferran E, Matsumoto M, Overman K, Hyde TM *et al.* Gene expression of metabolic enzymes and a protease inhibitor in the prefrontal cortex are decreased in schizophrenia. *Neurochem Res* 2004; **29**: 1245–1255.
- 42 Christian CA, Herbert AG, Holt RL, Peng K, Sherwood KD, Pangratz-Fuehrer S et al. Endogenous positive allosteric modulation of GABA(A) receptors by diazepam binding inhibitor. *Neuron* 2013; **78**: 1063–1074.
- 43 Regenold WT, Phatak P, Marano CM, Sassan A, Conley RR, Kling MA. Elevated cerebrospinal fluid lactate concentrations in patients with bipolar disorder and schizophrenia: implications for the mitochondrial dysfunction hypothesis. *Biol Psychiatry* 2009; 65: 489–494.
- 44 Rothermundt M, Falkai P, Ponath G, Abel S, Bürkle H, Diedrich M et al. Glial cell dysfunction in schizophrenia indicated by increased S100B in the CSF. *Mol Psy*chiatry 2004; **9**: 897–899.
- 45 Thompson PM, Kelley M, Yao J, Tsai G, van Kammen DP. Elevated cerebrospinal fluid SNAP-25 in schizophrenia. *Biol Psychiatry* 2003; **53**: 1132–1137.
- 46 Raison CL, Dantzer R, Kelley KW, Lawson MA, Woolwine BJ, Vogt G et al. CSF concentrations of brain tryptophan and kynurenines during immune stimulation with IFN-alpha: relationship to CNS immune responses and depression. *Mol Psychiatry* 2010; **15**: 393–403.
- 47 Raposo G, Stoorvogel W. Extracellular vesicles: exosomes, microvesicles, and friends. J Cell Biol 2013; 200: 373–383.
- 48 Lee Y, El Andaloussi S, Wood MJ. Exosomes and microvesicles: extracellular vesicles for genetic information transfer and gene therapy. *Hum Mol Genet* 2012; 21: R125–R134.
- 49 Belting M, Wittrup A. Nanotubes, exosomes, and nucleic acid-binding peptides provide novel mechanisms of intercellular communication in eukaryotic cells: implications in health and disease. *J Cell Biol* 2008; **183**: 1187–1191.

This work is licensed under a Creative Commons Attribution-NonCommercial-NoDerivs 3.0 Unported License. To view a copy of this license, visit http://creativecommons.org/licenses/by-nc-nd/3.0/

Supplementary Information accompanies the paper on the Translational Psychiatry website (http://www.nature.com/tp)