

SCIENTIFIC REPORTS



OPEN

Novel long-chain compounds with both immunomodulatory and MenA inhibitory activities against *Staphylococcus aureus* and its biofilm

Received: 03 October 2016
Accepted: 30 November 2016
Published: 10 January 2017

Seoung-ryoung Choi, Joel Frandsen & Prabakaran Narayanasamy

Menaquinone (MK) biosynthesis pathway is a potential target for evaluating antimicrobials in gram-positive bacteria. Here, 1,4-dihydroxy-2-naphthoate prenyltransferase (MenA) was targeted to reduce methicillin-resistant *Staphylococcus aureus* (MRSA) growth. MenA inhibiting, long chain-based compounds were designed, synthesized and evaluated against MRSA and menaquinone utilizing bacteria in aerobic conditions. The results showed that these bacteria were susceptible to most of the compounds. Menaquinone (MK-4) supplementation rescued MRSA growth, suggesting these compounds inhibit MK biosynthesis. 3a and 7c exhibited promising inhibitory activities with MICs ranging 1–8 µg/mL against MRSA strains. The compounds did not facilitate small colony variant formation. These compounds also inhibited the biofilm growth by MRSA at high concentration. Compounds 3a, 6b and 7c displayed a promising extracellular bactericidal activity against MRSA at concentrations equal to and four-fold less than their respective MICs. We also observed cytokines released from THP-1 macrophages treated with compounds 3a, 6b and 7c and found decreases in TNF-α and IL-6 release and increase in IL-1β. These data provide evidence that MenA inhibitors act as TNF-α and IL-6 inhibitors, raising the potential for development and application of these compounds as potential immunomodulatory agents.

Lipid-soluble vitamin K was discovered in 1929¹ as an essential nutrient for anti-hemorrhage. Its structure and chemical nature were previously reported². It is biosynthesized by many bacteria including *Escherichia coli*, *Mycobacterium tuberculosis* and *Staphylococcus aureus*. Vitamin K can be divided into natural and synthetic forms; Vitamin K₁ (phylloquinone) and K₂ (menaquinone, MK-4) are natural forms stored in the fat tissue and liver. Several homologues of vitamin K₂ (MK-7 to MK-15) are produced by bacteria by changing the length of the isoprenoid chain. Synthetic forms of K₃ (menaphthone or menadione), K₄, and K₅ are used as a nutritional supplement or fungal growth inhibitor. In humans, phylloquinone is supplemented from food while menaquinones (MKs) are produced by conversion of phylloquinone by intestinal bacteria. Humans do not possess a MK biosynthesis pathway, and MK is not utilized in the electron transport chain. Because of this, enzymes involved in either function have recently received attention as potential drug targets to treat Gram-positive pathogens.

Ubiquinone and MK play important roles in the prokaryote electron transport chain by transferring electrons between membrane bound protein complexes³. In Gram-positive bacteria, MK biosynthesis is essential for survival because MK is the only electron carrier in both aerobic and anaerobic conditions. MKs are biosynthesized via two independent classical and futasin pathways^{4,5}. Both pathways utilize chorismate as substrate - derived from phosphoenolpyruvate and D-erythrose-4-phosphate by the shikimate pathway - but do not share other similarities. As shown in Fig. 1, 1,4-dihydroxy-2-naphthoate (DHNA) is synthesized from chorismate by MenFDHCEB enzymes involved in the classical MK pathway. Prenylation by MenA followed by methylation by methyltransferase

Department of Pathology and Microbiology, College of Medicine, University of Nebraska Medical Center, Omaha, Nebraska 68198, USA. Correspondence and requests for materials should be addressed to P.N. (email: p.narayanasamy@unmc.edu)

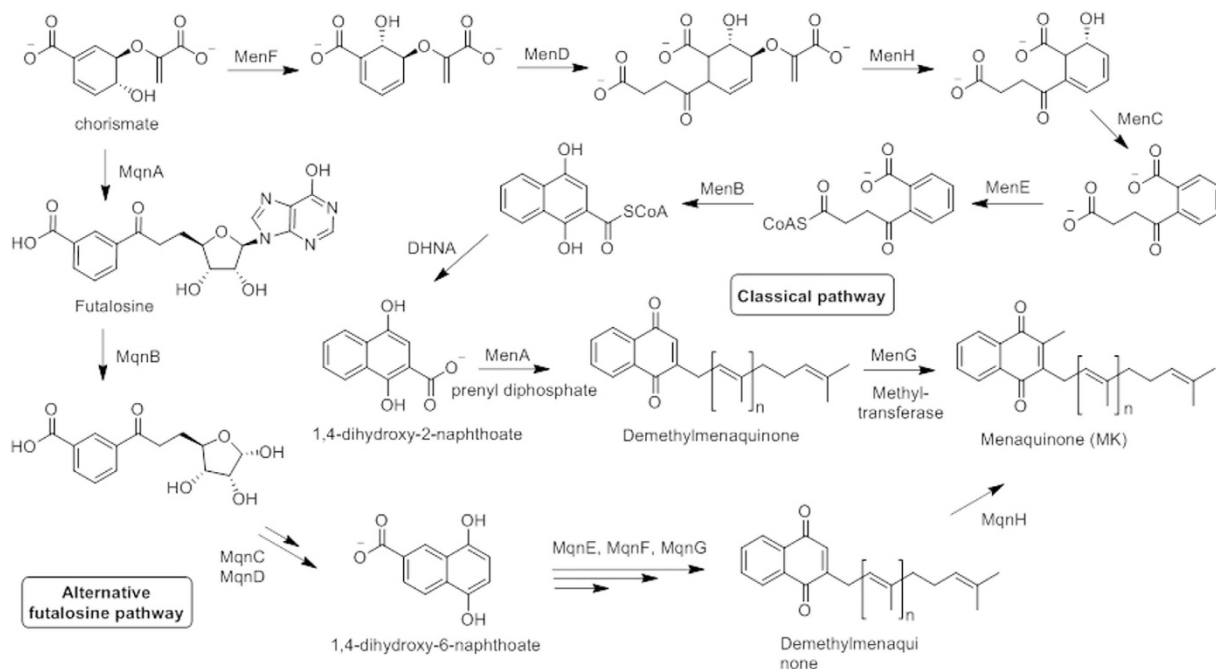


Figure 1. Menaquinone biosynthesis pathways. Classical menaquinone and alternative futasoline pathways from chorismate. MqnA: futasoline synthase, MqnB: futasoline hydrolase, MqnC: dehydropantothine futasoline cyclase, MqnD: 1,4-dihydroxy-6-naphthoate synthase. MenF: isochorismate synthase, MenC: o-succinylbenzoate synthase. MenD: 2-succinyl-5-enolpyruvyl-6-hydroxy-3-cyclohexene-1-carboxylate synthase. MenH: (1R,6R)-6-hydroxy-2-succinylcyclohexa-2,4-diene-1-carboxylate synthase.

(MenG) convert the 2-naphthoate to MK. The futasolin pathway uses MqnABCD enzymes to synthesize 1,4-dihydroxy-6-naphthoate which is converted to MK *via* a series of reactions catalyzed by unknown enzymes.

Inhibitors of enzymes involved in MK biosynthesis demonstrate that targeting these enzymes may lead to therapeutics for treatments of infections by Gram-positive bacteria including *Bacillus subtilis*, *M. tuberculosis*, and *S. aureus*^{6,7}. 4-oxo-4-phenylbutanoate analogues were developed as MenB inhibitors active against MRSA and both replicating and nonreplicating *M. tuberculosis*^{8,9}. Mechanism-based inhibitors of an acyl-CoA synthetase (MenE) have been reported with low micromolar IC₅₀ values^{10–12}. Inhibitors of MenD¹³ and MenC¹⁴ were also discovered, validating the classical menaquinone pathway as a potential target for the development of antibacterial drugs. Here, we report the synthesis and evaluation of potential MenA inhibiting antimicrobials and immunomodulatory compounds.

Results and Discussion

A transmembrane protein 1,4-dihydroxy-2-naphthoate prenyltransferase (MenA) catalyzes formation of demethylmenaquinone (DMMK) *via* consecutive decarboxylation and prenylation of DHNA (Fig. 1). MenA is a promising target for Gram-positive bacteria, especially MRSA and drug-resistant *M.tb*. Benzophenone-based analogues were synthesized and identified as MenA inhibitors against MRSA and nonreplicating and replicating *M.tb*^{15–18}. However, benzoquinone analogues are known to be toxic due to the presence of a benzophenone moiety^{15,19–21}. Previously, we reported that MenA inhibitors possessing a simple, chemical structural backbone of 7-methoxy naphthalene are highly active against Gram-positive bacteria with low micromolar MIC and IC₅₀ values (compound 1 and 2 in Fig. 2)⁶. Here, we synthesized novel compounds to explore structure-activity relationships (compounds 3 to 7 in Fig. 2), determined effect on cytokine expression by macrophages and reduced MRSA growth and its biofilm.

Antimicrobial activity of naphthalene derivatives. MICs were determined against methicillin-resistant *S. aureus* strains and Gram-positive and negative bacteria. We previously reported that compound 1 and 2 exhibited inhibition of both *M.tb* and MenA enzyme activity with IC₅₀ values of 6 and 5 μg/mL, respectively⁶. We synthesized long chain compounds with different functionalities to improve activity. Various active groups were introduced at the terminal part of carbon chain (tail group) to find a lead structure and evaluate its structure-activity relationship against MRSA. As expected, all the synthesized compounds showed no inhibition against Gram-negative *P. aeruginosa*, whereas most of the compounds inhibited growth of Gram-positive bacteria including seven MRSA strains, MSSA, EF and two species of *Mycobacterium*. Compounds 3a, 3b, 3c and 7c showed the most potent inhibitory activities against all Gram-positive bacteria with MIC values ranging from 0.5 to 16 μg/mL (Table 1). These compounds share a phenylethylamine group that plays an important role in growth inhibition of Gram-positive bacteria. Although the compounds are very active, replacing the fluorine in 3a with other halogens did not significantly improve the MIC, and introduction of bicyclic or tricyclic ring

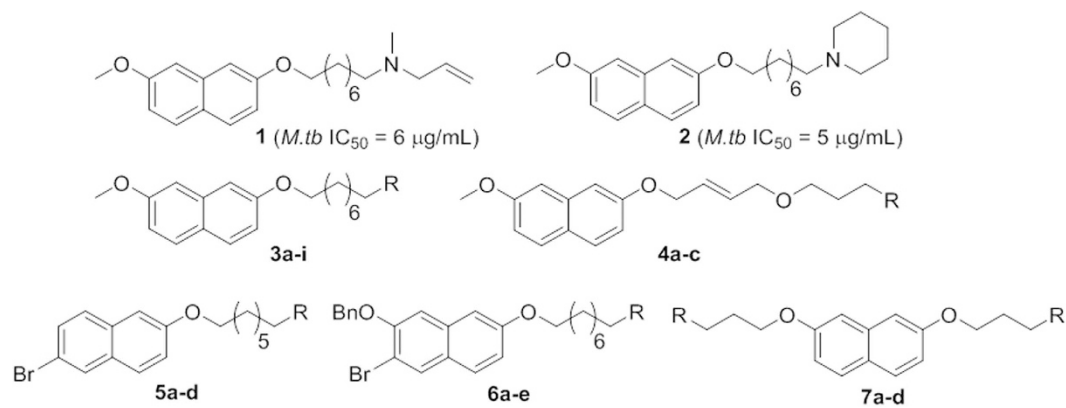


Figure 2. Synthesis of Long chain MenA inhibitors.

in the terminal end reduced the activity (3d–3f). Replacing the amine group with oxazolidinone also did not improve the activity (3g–3h) but the linezolid control was active. Similarly, the presence of fluorine in the piperidine group did not increase inhibitory activity (3i). Introduction of oxygen in the carbon chain had no effect on inhibitory activity (4a and 4b), but substitution of 4-fluorophenylethylamine at the tail end (4c) maintained activity. Modification of the 7-methoxynaphthalene group by replacing the methoxy group with bromine did not improve inhibitory activity of 4-fluorophenylethylamine compounds (5a–c). However, bromine substituted naphthalene increased activity for piperidine tail end substitution (5d). Likewise, replacement of methoxy group with benzyloxy group did not affect growth inhibition for piperidine terminal end but significantly reduced the activity of 4-fluorophenylethylamine terminal end (6a–e). These data show that a bulkier substitution is needed at any one of the terminal groups. Interestingly, it was found that 7c was the most potent inhibitor against all tested Gram-positive bacteria showing MIC between 1–6 $\mu\text{g}/\text{mL}$. Dual substitution of 4-fluorophenylethylamine increased the activity, but dual substitution of piperidine or allyl amine decreased the activity.

MRSA growth rescued by menaquinone (MK-4). Rescue experiments were performed via resazurin reduction assay to investigate the effect of menaquinone in the presence of various concentrations (0.5–64 $\mu\text{g}/\text{mL}$) of inhibitor 3a or 3c against two MRSA strains USA 700 or 200, respectively. MIC of 3a was 4 $\mu\text{g}/\text{mL}$ against USA 700 and 3c was 2 $\mu\text{g}/\text{mL}$ against USA 200 (Table 1). As shown in Fig. 3 and Figure S1, MIC values for both inhibitors increased in a concentration-dependent manner by addition of MK-4. MICs of both inhibitors moved to 64 $\mu\text{g}/\text{mL}$ in the presence of 0.5 mg/mL MK-4. Similarly, when the media is supplemented with 0.05 mg/mL MK-4, MICs increased to 16 $\mu\text{g}/\text{mL}$. Thus, the rescue experiments suggested that menaquinone biosynthesis is inhibited by 3a and 3c.

Colony morphology. A slow growing auxotrophic bacterial subpopulation resulting from mutations in metabolic genes has very distinctive phenotype compared to its parent wild-type strain. Gene mutations, caused by environmental factors including treatment of antibiotics, metabolic defects and genetic defects, lead to slow growing small colony variants (SCV) of bacteria that are related to electron transport defective strains and thymidylate biosynthesis^{22,23}. Among the clinical bacteria forming SCV, *Pseudomonas aeruginosa*, *Enterococcus faecalis*^{24,25}, and *S. aureus* SCV are a more prominent problem due to their association with intracellular persistence and linked chronic recurrent and antibiotic-resistant infections. Patients with cystic fibrosis, a genetic disorder caused by a mutation in CFTR gene, are especially in danger of higher probability of *S. aureus* SCV infection which can persist intracellularly in the host²⁶. Development of inhibitors that do not induce SCV are urgent and important to treat *S. aureus*. We tested 3a, to analyze induction of SCV by growing MRSA on TSA. MRSA (USA200) was grown on TSA containing 1 \times , 2 \times , 4 \times , or 8 \times MIC of 3a up to 96 h at 37 $^{\circ}\text{C}$ (Figs 3 and S2). No colonies were observed on agar plates that contained more than 2 \times MIC of 3a after 24 h. MK-4 (100 $\mu\text{g}/\text{mL}$) supplemented agar plate containing 8 $\mu\text{g}/\text{mL}$ of 3a showed wild-type colonies, indicating that the inhibitory activity was hampered by MK-4. However, no growth of SCV was observed even after incubation of TSA with 4–16 $\mu\text{g}/\text{mL}$ up to 96 h, suggesting 3a did not induce SCV. MRSA (USA200) was also grown on TSA containing 7c and rifampin (positive control). At 72 h, rifampin showed growth of SCV and confirming that 7c did not induce SCV (Fig. 3C).

Antimicrobial activity against MRSA biofilms. We next investigated if inhibitor 3a exhibits inhibitory activity against MRSA biofilms. Biofilms by Gram-positive bacteria are known to be more resistant to many antibiotics and the required concentration to eradicate biofilm is much higher than MIC²⁷. Minocycline, a tetracycline antibiotic used for skin infection, was used as a control for biofilm eradication. It showed significant decrease in MRSA biofilm growth at concentrations of 0.5 and 0.03 mg/mL compared to the positive control (Fig. 4). Concurrently, Compound 3a showed 10⁴, 10⁶ and 10⁷ fold decrease in MRSA growth from the treated biofilm at 0.03, 0.5 and 1 mg/mL, respectively. Thereby 3a was determined to be effective in reducing biofilm growth.

Growth Inhibition of extracellular MRSA (USA700) in human macrophages by inhibitor. We next investigated the bactericidal activities of inhibitors in human macrophages incubated with MRSA.

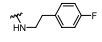
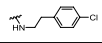
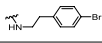
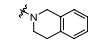
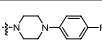
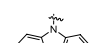
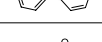
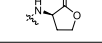
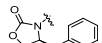
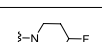
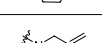
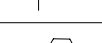
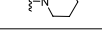
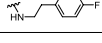
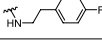
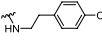
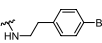
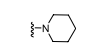
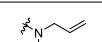
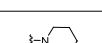
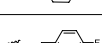
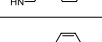
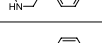

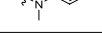
#	R	<i>Staphylococcus aureus</i> isolates, MIC (ug/mL)							MSSA	EF ¹	PA ²	MA ³	Mab ⁴
		USA100	USA200	USA300	USA400	USA500	USA700	USA800					
1		—	—	16	16	—	16	—	32	8	128	—	—
2		—	—	16	16	—	16	—	16	16	>64	—	—
3a		4	2	4	4	4	4	6	4	1	>64	2	6
3b		6	2	6	4	8	8	6	4	0.5	>64	4	12
3c		6	2	4	4	6	4	6	4	1	>64	4	12
3d		—	>128	—	64	128	32	—	32	—	—	—	—
3e		—	—	—	—	—	>128	—	—	>128	—	—	—
3f		>128	—	—	—	—	>128	—	—	—	—	—	—
3g		—	—	—	>128	—	>128	—	>128	—	>128	—	—
3h		—	—	—	>128	—	>128	—	>128	—	>128	—	—
3i		—	—	—	—	—	>128	—	—	—	—	—	—
4a		—	128	—	—	—	128	—	—	—	—	—	—
4b		—	—	—	—	—	128	—	—	—	>128	—	—
4c		8	—	8	8	16	16	16	16	—	>64	8	—
5a		16	4	4	8	—	4	—	—	2	—	—	—
5b		>128	16	32	64	—	64	—	—	16	—	—	—
5c		32	8	32	16	—	32	—	—	8	—	—	—
5d		8	—	—	4	—	16	—	—	—	—	—	—
6a		—	8	—	8	—	16	64	16	4	—	—	—
6b		6	4	6	6	12	16	12	4	2	>128	2	—
6c		>128	—	—	>128	>128	>128	>128	>128	—	—	—	—
6d		—	—	>128	>128	—	64	—	64	16	—	—	—
6e		—	128	>128	—	—	128	—	—	—	—	—	—
7a		—	>128	—	—	—	128	—	—	—	—	—	—
7b		—	—	—	—	—	>128	—	—	—	—	—	—
7c		3	2	3	1	8	2	4	2	1	>128	3	6
7d		—	—	—	—	—	>128	—	—	—	—	—	—
Linezolid	—	0.5	1	1	1	0.5	1	0.5	1	>32	1	—	—
Minomycin	—	—	—	—	0.5	<0.06	—	—	—	16	—	—	—

Table 1. Minimum inhibitory concentration against MRSA strains and other bacteria. ¹EF: *Enterococcus faecalis*. ²PA: *Pseudomonas aeruginosa*. ³MA: *Mycobacterium avium*. ⁴Mab: *Mycobacterium abscessus*, MSSA: Methicillin susceptible *Staphylococcus aureus*, (—): not determined.

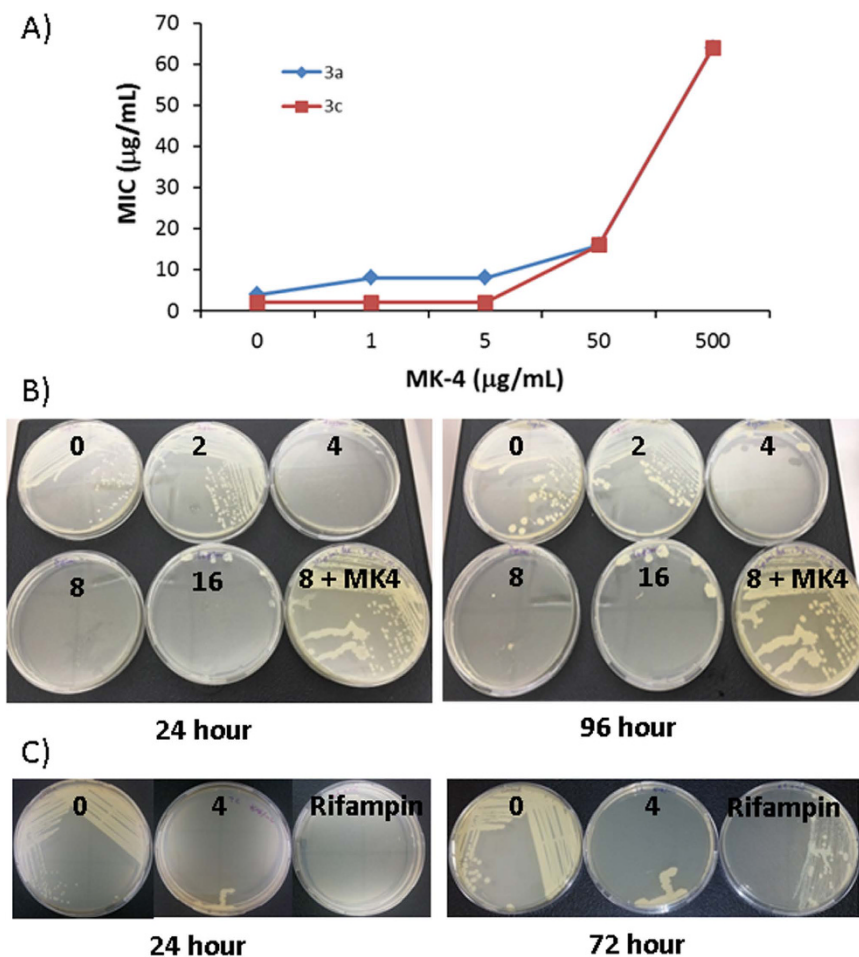


Figure 3. Menaquinone rescue (A) and colony morphology (B). (A) MRSA (USA 700 for 3a, USA 200 for 3c) was treated with various concentrations of menaquinone (MK-4) in the presence of inhibitor. (B) MRSA (USA 200) was incubated on TSA containing various concentrations of 3a (0, 2, 4, 8, 16 $\mu\text{g}/\text{mL}$ and 8 $\mu\text{g}/\text{mL}$ + 100 $\mu\text{g}/\text{mL}$ MK4) up to 96 h. (C) MRSA (USA 200) was incubated on TSA containing 7c and rifampin (positive control) up to 72 h.

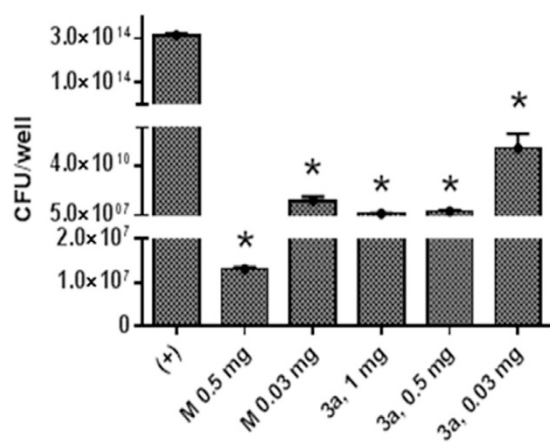


Figure 4. Antibacterial activity against MRSA biofilm. MRSA (USA700) biofilms were treated with 3a or minocycline for 24 h and serially diluted and plated on TSA plates to determine CFU. Data represents the mean \pm SEM of triplicate ($n = 3$). M: minocycline, Statistical differences were determined using Student's t test: * $p < 0.001$ compared to (+) positive control.

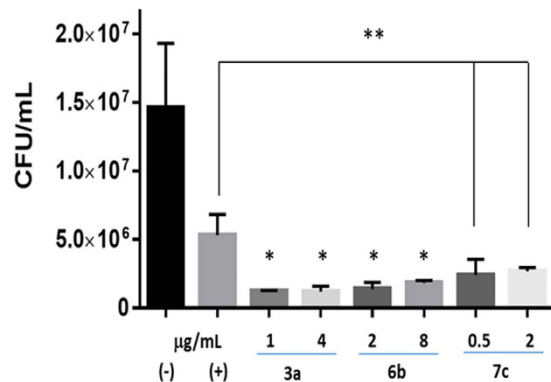


Figure 5. Macrophage-mediated inhibition of MRSA growth with inhibitors. Human monocytes were differentiated into macrophages (MDM) in the presence of MCSF. MDMs were incubated with MRSA (USA 700) in the presence of inhibitor for 24 h at 37 °C. Media were serially diluted and plated on TSA to determine CFU. (-): No MDM and No inhibitor, (+): No inhibitor. Data represents the mean ± SEM of triplicate (n = 3). Statistical differences were determined using Student's *t* test: **p* < 0.002, ***p* < 0.01 compared to (+) positive control.

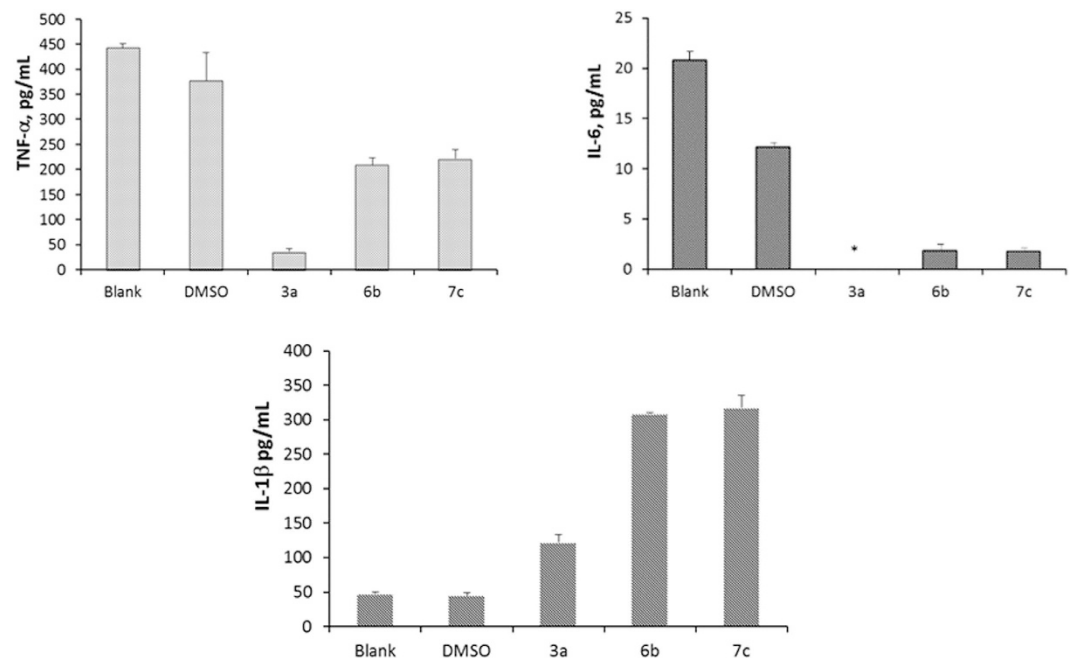


Figure 6. TNF-α, IL-6 and IL-1β released from macrophages treated with long-chain inhibitors. THP-1 macrophages were treated with **3a**, **6b** or **7c** at concentrations equal to their MICs for 24 h at 37 °C. Culture supernatants were analyzed for cytokines. Data represents the mean ± SEM of triplicate samples (n = 3). Blank: No inhibitor was added. DMSO (<0.05%), *out of range below the detection limit.

Monocyte-derived macrophages were obtained by incubating monocytes in the presence of MCSF in DMEM media containing 10% human serum. Adherent MDMs were incubated with MRSA (1×10^6 CFU/mL) and treated with inhibitors at concentrations of MIC and MIC/4 for 24 h. Extracellular MRSA surviving in media was determined by counting colonies. As seen in control experiments in Fig. 5, a 3 fold decrease of extracellular anti-bacterial activity by MDMs was observed in culture media, indicating that MRSA was phagocytized and killed by MDMs. Our inhibitors were then tested in the presence or absence of MDMs. Antibacterial activities of inhibitors (**3a**, **6b**, **7c**) tested were increased by 6–12 fold compared to activities without MDMs and by 1.5 – 3-fold compared to activities with MDMs. This result suggests that these inhibitors prevented extracellular bacterial growth at their MIC and MIC/4, but use of a concentration greater than the MIC is required for complete bactericidal activity. However, we did not observe concentration-dependent antibacterial activities when MDMs were treated with MRSA and inhibitors at different concentrations.

Cytokine secretion by THP-1 macrophages treated with MenA inhibitors. Since MenA inhibitors enhanced the bactericidal activity of macrophages, we next examined whether macrophages treated with inhibitors display an altered cytokine profile. The cytokines released from THP-1 macrophages were assessed from culture supernatants using a Luminex system. Among 10 cytokines analyzed, IL-8 was most abundant in ~25 ng/ml (Figure S2) but there was no significant difference in IL-8 expression in the presence or absence of inhibitors. Interestingly, we found that amounts of TNF- α and IL-6 released from THP-1 macrophages were decreased in the presence of inhibitors **3a**, **6b** and **7c** whereas these inhibitors stimulated the release of IL-1 β at concentrations equal to their MICs (Fig. 6). These intriguing results suggest that this class of small molecules, in addition to antibiotic activity, possesses potential to be developed as TNF- α and IL-6 suppressors for use as immunomodulatory therapy for the treatment of inflammatory diseases. In addition, IL-1 β was also regulated by the same MenA inhibitors.

The MenA inhibitors are composed of three main groups: naphthol, carbon chain linker, and amine tail group. We explored this class of inhibitor by modifying these three main backbone structures to find a more potent lead structure against *S. aureus*. We found that **3a-c** and **5a** have potent activity against MRSA strains, MSSA and other bacteria with MIC values ranging 1–16 $\mu\text{g}/\text{mL}$. As expected, all compounds showed no activity against Gram-negative *P. aeruginosa*. Interestingly, **7c** showed very low MIC values against MK utilizing bacteria (Table 1), suggesting it is a potential lead compound for future optimization. MK rescue experiment confirmed the involvement of the MK biosynthetic pathway in reducing MRSA growth. MRSA strains treated with **3a** or **7c** were rescued by MK4, supporting disruption of electron transfer chain by MenA inhibitor. IC₅₀ analysis also confirmed that MenA is the target for compounds. **3a** was found to inhibit MRSA biofilm growth, which is defined as a sessile microbial community. *S. aureus* biofilms colonize medical implants and host tissue and contribute to the persistence of chronic infections. The cells are protected in an extracellular polymeric matrix in which their physiologies are altered with respect to gene expression and protein production. Thus, biofilm formation reduces susceptibility to antimicrobials and requires use of larger quantity of antimicrobials. As expected, minocycline as a reference did not completely inhibit MRSA (USA 700) biofilm at 8300 \times MIC (broth MIC < 0.06 $\mu\text{g}/\text{mL}$, Table 1), showing 10⁶-fold reduction compared to the positive control. Likewise, treatment of MRSA biofilm with **3a** (250 \times MIC) inhibited the MRSA growth by 10⁵ fold. The results suggest that MenA inhibitor **3a** is highly effective in preventing MRSA growth.

Development of antibiotics that are active against both extracellular and intracellular pathogens is important to eradicate infectious bacteria. MenA inhibitors in the presence of macrophages showed promising activities against extracellular MRSA. We observed macrophage stimulation by MenA inhibitors alters the cytokine-release profile, causing a reduction of TNF- α and IL-6, and an increase of IL-1 β . These three cytokines play a vital role in immune defense by responding to pathogens and inflammatory states. They have also been implicated as novel immunomodulatory therapeutic targets.

The pro- and anti-inflammatory IL-6 participates in the host immune defense. IL-6 binding to either form of its receptors (transmembrane and soluble IL-6 receptors) triggers biological signal transductions, affecting various cell populations²⁸. However, dysregulation of IL-6 production is a major causative factor of autoimmunity and chronic inflammatory diseases. There are various drugs that block IL-6 signaling pathways by targeting IL-6, IL-6 receptors or gp130 in development stage or market. For example, monoclonal antibody (tocilizumab) to IL-6R has been approved and used for treatment of rheumatoid arthritis, indicating that IL-6 blockade is a novel strategy for inflammatory diseases. Since macrophages are one of major sources of IL-6, reducing or blocking IL-6 release from macrophages by an anti-IL-6 reagent is an alternative strategy in development of therapeutic agent. As shown in Fig. 6, it is observed that treatment of THP-1 macrophages with MenA inhibitors (**3a**, **6b**, or **7c**) significantly reduced IL-6 release, providing a rational strategy for the development of immunomodulatory agent and prompting us to investigate mechanism of action of MenA inhibitor that affects IL-6 release of macrophage.

Several monoclonal antibody drugs such as infliximab (Remicade), etanercept (Enbrel), adalimumab (Humira), certolizumab pegol (Cimzia) have been developed against tumor necrosis factor (TNF- α) overproduction to treat inflammatory and autoimmune diseases including rheumatoid arthritis, psoriatic arthritis, ankylosing spondylitis and Crohn's disease. It is also suggested that a combination of TNF inhibitors and antibiotics might be beneficial for treatment of *S. aureus* sepsis²⁹. TNF- α is released by monocytes, lymphocytes and macrophages, and facilitates macrophage apoptosis in response to *M.tb*^{30–32}. TNF- α also recruits other immune cells to sites of inflammation. During immune response to pathogens or stimuli, macrophages synthesize and secrete TNF to the extracellular space via a series of secretion pathways. Many trafficking proteins are involved to deliver TNF outside the plasma membrane. Our MenA inhibitor potentially inhibited TNF release from THP-1 macrophages by either inhibiting one of trafficking proteins involved in secretion pathways, or binding to a membrane receptor that blocks activation of macrophages during the immune response. Earlier, it was reported that adenosine as agonist reduces TNF- α release from macrophage by activating the cAMP-elevating A(2A) adenosine receptor (AR) subtype^{33,34}. However, further study will be conducted to identify the anti-inflammatory role of these MenA inhibitors.

Unlike TNF- α and IL-6, we observed an increase in IL-1 β release from THP-1 macrophages by stimulation by MenA inhibitors. Pro-inflammatory cytokine IL-1 β is important in defending infection, but dysregulation of IL-1 β is responsible for chronic inflammatory diseases and acute tissue injuries³⁵. These results indicate that stimuli-mediated secretion of cytokines is more complicated. It is challenging to develop selective stimulating agents that block the release of cytokines of interest.

A series of long-chain compounds were developed to inhibit growth of MRSA and other gram-positive bacteria that utilize menaquinone under aerobic conditions. MenA inhibitors displayed a promising extracellular bactericidal activity against growth of MRSA and its biofilm. More importantly, naphthalene-based MenA inhibitors showed potential as inhibitors of extracellular bacterial growth and as immunomodulatory agents to reduce the release of IL-6 and TNF- α from macrophages. These MenA inhibitors could be used to reduce macrophage cytokine release and it would provide insight in the development of pharmaceuticals for MRSA septic shock and other inflammatory diseases.

Methods

Materials. All chemicals were purchased from Aldrich (St. Louis, MO, USA) and Across (Geel, Belgium). Microbial species are purchased from ATCC (Manassas, VA, USA). NMR spectroscopies were recorded on a Varian Unity/Inova-500 NB (500 MHz, Varian Medical Systems Inc. Palo Alto, CA, USA). Chemical shifts are reported in parts per million (ppm) downfield from TMS. Molecular weight is determined using MALDI TOF/TOF (Applied Biosystems, CA, USA).

Minimum inhibitory concentration (MIC) against *S. aureus*. 96-well plates were seeded with 5×10^5 CFU/mL in a volume of 100 μ L of tryptic soy broth (TSB) per well for *Staphylococcus aureus* assays. Compounds were dissolved in DMSO and serially diluted as 1:2 dilutions to give the final concentration ranging from 0.25 to 128 μ g/mL per well. The final concentration of DMSO in the assay plates did not exceed 2%. Plates were incubated at 37 °C for 18–20 hours and MIC was determined by reading OD₆₂₅. For *M. avium* and *M. abscessus*, 7H9 broth media was used for determination of MICs. The final concentration was 5×10^5 CFU/mL in each well. The plate was incubated at 37 °C for 7–8 days and MICs were determined using a Tecan microplate reader (OD₆₂₅) and resazurin reduction assay.

Menaquinone rescue experiments with MRSA. Resazurin reduction assay was performed in 96-well plate. MRSA was cultured in TSB until OD₆₂₅ reached 0.4–0.5 and diluted and plated in 96-well plate. MRSA (5×10^5 CFU/mL/well) was treated with menaquinone (MK4, 0, 1, 5, 50, 500 μ g/mL) in the presence of 0.5–128 μ g/mL of inhibitor. The plate was incubated overnight at 37 °C followed by 20 μ L of resazurin (0.15 mg/mL) was added to each well. The plate was incubated for 2–4 h and fluorescence was measured at 560 nm excitation/590 nm emission.

Cytotoxicity Assay. To determine cytotoxicity of the inhibitors, THP-1 cells were differentiated into macrophages by PMA (7.5 ng/mL) for 24 h. THP-1 macrophages (5×10^5 /well) were treated with inhibitors in 48-well plates for 24 h at 37 °C in 5% carbon dioxide. Cells were washed with PBS buffer three times and serum-free culture medium was added to each well. Cytotoxicity was assessed using resazurin assay.

***S. aureus* biofilm assay.** Biofilm assays were performed using a method previously described with modification³⁶. A 96 well plate was precoated with 20% human plasma (Sigma) in 0.05 M carbonate buffer, pH 9.6 at 4 °C for 24 h. MRSA was cultured overnight in TSB containing 3% NaCl and 0.5% glucose. The overnight culture was diluted with TSB/NaCl/Glucose media until OD₆₀₀ reached 0.05. After carefully removing the liquid from the 96 well plate, 200 μ L of MRSA dilution was added to each well in triplicate. The plate was incubated at 37 °C for 24 h and carefully washed with PBS twice. MH broth (200 μ L) containing a compound was added to each well in triplicate and incubated at 37 °C for 24 h. After washing with PBS twice, the biofilm was scraped in 100 μ L of PBS and serially diluted and plated on TSA to determine CFU.

Colony morphology. MRSA was streaked and then incubated on tryptic soy agar (TSA) containing 0, 2, 4, 8 or 16 μ g/mL of inhibitor **3a** or 4 μ g/mL of **7C** or 0.025 μ g/mL of rifampin at 37 °C for 96 h. MK-4 (100 μ g/mL) was supplemented in TSA containing 8 μ g/mL of **3a** for the rescue experiment.

MenA inhibitor activity against extracellular *S. aureus* in the presence of human MDMs. Human monocyte-derived macrophages (MDM) were cultured at a concentration of 1×10^6 cells/mL/well in a 24 well plate³⁷. MRSA (USA 700) was cultured in TSB (10 mL) until OD₆₂₅ reached 0.4–0.5 and then centrifuged at 14,000 rpm for 10 min. The pellets were resuspended in 10 mL of DMEM supplemented with 10% human serum (no antibiotics). The bacteria medium was diluted with DMEM containing an inhibitor to obtain 1×10^6 CFU/mL and added to human MDMs (MOI = 1) in a 24-well plate. After 24 hour infection at 37 °C, the supernatants were serially diluted for CFU determination (TSA plates).

Quantitation of cytokines. THP-1 cells were differentiated into macrophages by treatment with PMA (7.5 ng/mL). Levels of the human cytokines (GM-CSF, IFN- γ , IL-1 β , IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, TNF- α) released from THP-1 macrophages were determined using a multiplex kit from Life Technologies according to the manufacturer's instructions. In brief, the 96 well filter plate was pre-wetted, and then 25 μ L of the diluted bead suspension was added to each well and washed twice. 100 μ L of samples and standards were added to each well. The plate was incubated at room temperature for 2 h on a shaker. Following washing, biotinylated detection antibody was added and incubated for 1 h. After washing, streptavidin-RPE was added to each well and incubated for 0.5 h. The plate was again washed, resuspended in 100 μ L of the washing buffer, and read on the Luminex xMAP™ system (Bio-Rad, AtheNA multi-Lyte™, USA). All samples were run in triplicate and standards were run in duplicate.

Statistical analysis. Statistical analysis was performed using Student's t-test. Data are represented as mean \pm SEM. Data are significant at $p < 0.05$.

References

1. Dam, H. The antihemorrhagic vitamin of the chick: occurrence and chemical nature. *Nature* **135** 652–653 (1935).
2. MacCorquodale, D. W., Binkley, S. B., Thayer, S. A. & Doisy, E. A. Constitution of vitamin K1. *J. Am. Chem. Soc.* **61**, 1928–1929, doi: 10.1021/ja01876a510 (1939).
3. Meganathan, R. Biosynthesis of menaquinone (vitamin K2) and ubiquinone (coenzyme Q): a perspective on enzymatic mechanisms. *Vitam Horm* **61**, 173–218 (2001).
4. Hiratsuka, T. *et al.* An alternative menaquinone biosynthetic pathway operating in microorganisms. *Science (Washington, DC, US)* **321**, 1670–1673, doi: 10.1126/science.1160446 (2008).
5. Bentley, R. & Meganathan, R. Biosynthesis of vitamin K (menaquinone) in bacteria. *Microbiol. Rev.* **46**, 241–280 (1982).

6. Choi, S. R. *et al.* Discovery of bicyclic inhibitors against menaquinone biosynthesis. *Future Med Chem* **8**, 11–16, doi: 10.4155/fmc.15.168 (2016).
7. Choi, S. R., Larson, M. A., Hinrichs, S. H. & Narayanasamy, P. Development of potential broad spectrum antimicrobials using C-symmetric 9-fluorenone alkyl amine. *Bioorganic & medicinal chemistry letters*, doi: 10.1016/j.bmcl.2016.02.087 (2016).
8. Li, X. *et al.* CoA Adducts of 4-Oxo-4-Phenylbut-2-enoates: Inhibitors of MenB from the M. tuberculosis Menaquinone Biosynthesis Pathway. *ACS medicinal chemistry letters* **2**, 818–823, doi: 10.1021/ml200141e (2011).
9. Matarlo, J. S. *et al.* A Methyl 4-Oxo-4-phenylbut-2-enoate with *in Vivo* Activity against MRSA that Inhibits MenB in the Bacterial Menaquinone Biosynthesis Pathway. *ACS Infect Dis* **2**, 329–340, doi: 10.1021/acsinfecdis.6b00023 (2016).
10. Lu, X., Zhang, H., Tonge, P. J. & Tan, D. S. Mechanism-based inhibitors of MenE, an acyl-CoA synthetase involved in bacterial menaquinone biosynthesis. *Bioorg. Med. Chem. Lett.* **18**, 5963–5966, doi: 10.1016/j.bmcl.2008.07.130 (2008).
11. Lu, X. *et al.* Stable analogues of OSB-AMP: potent inhibitors of MenE, the o-succinylbenzoate-CoA synthetase from bacterial menaquinone biosynthesis. *ChemBiochem* **13**, 129–136 (2012).
12. Matarlo, J. S. *et al.* Mechanism of MenE inhibition by acyl-adenylate analogues and discovery of novel antibacterial agents. *Biochemistry* **54**, 6514–6524, doi: 10.1021/acs.biochem.5b00966 (2015).
13. Fang, M. *et al.* Succinylphosphonate esters are competitive inhibitors of MenD that show active-site discrimination between homologous alpha-ketoglutarate-decarboxylating enzymes. *Biochemistry* **49**, 2672–2679, doi: 10.1021/bi901432d (2010).
14. Pulaganti, M., Banaganapalli, B., Mulakayala, C., Chitta, S. K. & C., M. A. Molecular modeling and docking studies of O-succinylbenzoate synthase of M. tuberculosis—a potential target for antituberculosis drug design. *Appl Biochem Biotechnol* **172**, 1407–1432, doi: 10.1007/s12010-013-0569-4 (2014).
15. Debnath, J. *et al.* Discovery of selective menaquinone biosynthesis inhibitors against Mycobacterium tuberculosis. *J Med Chem* **55**, 3739–3755 (2012).
16. Kurosu, M., Narayanasamy, P., Biswas, K., Dhiman, R. & Crick, D. C. Discovery of 1,4-Dihydroxy-2-naphthoate Prenyltransferase Inhibitors: New Drug Leads for Multidrug-Resistant Gram-Positive Pathogens. *J. Med. Chem.* **50**, 3973–3975, doi: 10.1021/jm070638m (2007).
17. Kurosu, M. & Crick, D. C. MenA is a promising drug target for developing novel lead molecules to combat Mycobacterium tuberculosis. *Med. Chem.* **5**, 197–207, doi: 10.2174/157340609787582882 (2009).
18. Dhiman, R. K. *et al.* Menaquinone synthesis is critical for maintaining mycobacterial viability during exponential growth and recovery from nonreplicating persistence. *Mol. Microbiol.* **72**, 85–97, doi: 10.1111/j.1365-2958.2009.06625.x (2009).
19. Matsumoto, K. *et al.* Cytotoxic benzophenone derivatives from Garcinia species display a strong apoptosis-inducing effect against human leukemia cell lines. *Biol. Pharm. Bull.* **26**, 569–571, doi: 10.1248/bpb.26.569 (2003).
20. Kuete, V. *et al.* Cytotoxicity and modes of action of four naturally occurring benzophenones: 2,2',5,6'-Tetrahydroxybenzophenone, guttiferone E, isogarcinol and isoxanthochymol. *Phytomedicine* **20**, 528–536, doi: 10.1016/j.phymed.2013.02.003 (2013).
21. Baggett, S., Mazzola, E. P. & Kennelly, E. J. The benzophenones: isolation structural elucidation and biological activities. *Stud. Nat. Prod. Chem.* **32**, 721–771 (2005).
22. Proctor, R. A. *et al.* Small colony variants: a pathogenic form of bacteria that facilitates persistent and recurrent infections. *Nat Rev Microbiol* **4**, 295–305, doi: 10.1038/nrmicro1384 (2006).
23. Proctor, R. A. *et al.* Staphylococcus aureus Small Colony Variants (SCVs): a road map for the metabolic pathways involved in persistent infections. *Front Cell Infect Microbiol* **4**, 99, doi: 10.3389/fcimb.2014.00099 (2014).
24. Haussler, S. *et al.* Highly adherent small-colony variants of Pseudomonas aeruginosa in cystic fibrosis lung infection. *J Med Microbiol* **52**, 295–301, doi: 10.1099/jmm.0.05069-0 (2003).
25. Wellinghausen, N. *et al.* Characterization of clinical Enterococcus faecalis small-colony variants. *J Clin Microbiol* **47**, 2802–2811, doi: 10.1128/jcm.00485-09 (2009).
26. Proctor, R. A. *et al.* Staphylococcal small colony variants have novel mechanisms for antibiotic resistance. *Clin Infect Dis* **27 Suppl 1**, S68–74 (1998).
27. Olson, M. E., Ceri, H., Morck, D. W., Buret, A. G. & Read, R. R. Biofilm bacteria: formation and comparative susceptibility to antibiotics. *Can J Vet Res* **66**, 86–92 (2002).
28. Scheller, J., Chalaris, A., Schmidt-Arras, D. & Rose-John, S. The pro- and anti-inflammatory properties of the cytokine interleukin-6. *Biochim Biophys Acta* **1813**, 878–888, doi: 10.1016/j.bbasmcr.2011.01.034 (2011).
29. Fei, Y. *et al.* The combination of a tumor necrosis factor inhibitor and antibiotic alleviates staphylococcal arthritis and sepsis in mice. *J Infect Dis* **204**, 348–357, doi: 10.1093/infdis/jir266 (2011).
30. Lee, J., Hartman, M. & Kornfeld, H. Macrophage apoptosis in tuberculosis. *Yonsei Med J* **50**, 1–11 (2009).
31. Keane, J., Remold, H. G. & Kornfeld, H. Virulent Mycobacterium tuberculosis strains evade apoptosis of infected alveolar macrophages. *J Immunol* **164**, 2016–2020 (2000).
32. Balcewicz-Sablinska, M. K., Keane, J., Kornfeld, H. & Remold, H. G. Pathogenic Mycobacterium tuberculosis evades apoptosis of host macrophages by release of TNF-R2, resulting in inactivation of TNF- α . *J Immunol.* **161**, 2636–2641 (1998).
33. Kreckler, L. M., Wan, T. C., Ge, Z. D. & Auchampach, J. A. Adenosine inhibits tumor necrosis factor-alpha release from mouse peritoneal macrophages via A2A and A2B but not the A3 adenosine receptor. *J Pharmacol Exp Ther* **317**, 172–180, doi: 10.1124/jpet.105.096016 (2006).
34. Kreckler, L. M., Gizewski, E., Wan, T. C. & Auchampach, J. A. Adenosine suppresses lipopolysaccharide-induced tumor necrosis factor-alpha production by murine macrophages through a protein kinase A- and exchange protein activated by cAMP-independent signaling pathway. *J Pharmacol Exp Ther* **331**, 1051–1061, doi: 10.1124/jpet.109.157651 (2009).
35. Lopez-Castejon, G. & Brough, D. Understanding the mechanism of IL-1beta secretion. *Cytokine Growth Factor Rev* **22**, 189–195, doi: 10.1016/j.cytogfr.2011.10.001 (2011).
36. Bose, J. L., Lehman, M. K. K., Fey, P. D. & Bayles, K. W. In *PLoS One* Vol. 7 (ed H. Ton-That) (2012).
37. Narayanasamy, P., Switzer, B. L. & Britigan, B. E. Prolonged-acting, Multi-targeting Gallium Nanoparticles Potently Inhibit Growth of Both HIV and Mycobacteria in Co-Infected Human Macrophages. *Scientific reports* **5**, 8824, doi: 10.1038/srep08824 (2015).

Acknowledgements

We would like to thank Nebraska Research Initiative Faculty Award and UNMC College of Medicine for funding. We would like to thank Nebraska Medical Center Clinical Microbiology laboratory for providing clinical MRSA strains.

Author Contributions

S.R.C. designed experiment, performed all experiments, analyzed results and wrote the manuscript. J.F. performed MIC determinations and edited the manuscript. P.N. conceived the idea for the project, designed the experiment, coordinated the project, analyzed the results and edited the manuscript. All authors have given approval to the final version of the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Choi, S.-r. *et al.* Novel long-chain compounds with both Immunomodulatory and MenA Inhibitory activities against *Staphylococcus aureus* and its biofilm. *Sci. Rep.* 7, 40077; doi: 10.1038/srep40077 (2017).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2017