

SCIENTIFIC REPORTS



OPEN

Functional Recovery from Neural Stem/Progenitor Cell Transplantation Combined with Treadmill Training in Mice with Chronic Spinal Cord Injury

Syoichi Tashiro¹, Soraya Nishimura², Hiroki Iwai², Keiko Sugai², Liang Zhang^{1,3},
Munehisa Shinozaki^{3,4}, Akio Iwanami², Yoshiaki Toyama², Meigen Liu¹, Hideyuki Okano³ &
Masaya Nakamura²

Received: 01 March 2016

Accepted: 10 July 2016

Published: 03 August 2016

Most studies targeting chronic spinal cord injury (SCI) have concluded that neural stem/progenitor cell (NS/PC) transplantation exerts only a subclinical recovery; this in contrast to its remarkable effect on acute and subacute SCI. To determine whether the addition of rehabilitative intervention enhances the effect of NS/PC transplantation for chronic SCI, we used thoracic SCI mouse models to compare manifestations secondary to both transplantation and treadmill training, and the two therapies combined, with a control group. Significant locomotor recovery in comparison with the control group was only achieved in the combined therapy group. Further investigation revealed that NS/PC transplantation improved spinal conductivity and central pattern generator activity, and that treadmill training promoted the appropriate inhibitory motor control. The combined therapy enhanced these independent effects of each single therapy, and facilitated neuronal differentiation of transplanted cells and maturation of central pattern generator activity synergistically. Our data suggest that rehabilitative treatment represents a therapeutic option for locomotor recovery after NS/PC transplantation, even in chronic SCI.

Patients with spinal cord injury (SCI) experience various sequelae, such as motor paresis and spasticity, sensory disturbances, and bowel and rectal dysfunction. Although the injured central nervous system, including the spinal cord, shows only a small degree of plasticity, many studies on neural stem/progenitor cell (NS/PC) transplantation in the acute and subacute phase have shown significant recovery in locomotor and sensory function^{1,2}. The results of these studies collectively suggest that the critical time window for transplantation therapy in rodents is around 7–10 days post-injury (DPI). Most studies targeting chronically injured spinal cord have reported no significant recovery of function^{3–7}; however, there have been two exceptions to date^{8,9}. In both of these exceptional studies, the cell transplantation was performed relatively early, in the “early chronic phase” at around 21 or 30 DPI^{8,9}. This may indicate that the therapeutic window for transplantation closes by the end of the subacute phase or at the beginning of the early chronic phase. The majority of SCI patients are in the chronic phase, representing a major challenge for the clinical application of cell transplantation.

Recently, combined therapies for the treatment of chronically injured spinal cord have attracted the attention of researchers in regenerative medicine. One approach seeks to improve the viability or differentiation of NS/PC through the use of exogenous neurotrophic factors⁵. Another aims to degrade glial scarring or inhibit axonal growth inhibitors within the scar, by the use of chondroitinase ABC^{10,11} or semaphorin 3A inhibitor^{12,13}. It has also been reported that combination therapy with a neurotrophic factor, Neurotrophin-3 expressing NS/PCs and chondroitinase ABC, led to superior functional recovery⁴. Although the potential importance of combination

¹Department of Rehabilitation Medicine, Keio University School of Medicine, Tokyo, Japan. ²Department of Orthopaedic Surgery, Keio University School of Medicine, Tokyo, Japan. ³Department of Physiology, Keio University School of Medicine, Tokyo, Japan. ⁴Department of Neuroscience, City College of the City University of New York, NY, USA. Correspondence and requests for materials should be addressed to H.O. (email: hidokano@a2.keio.jp) or M.N. (email: masa@a8.keio.jp)

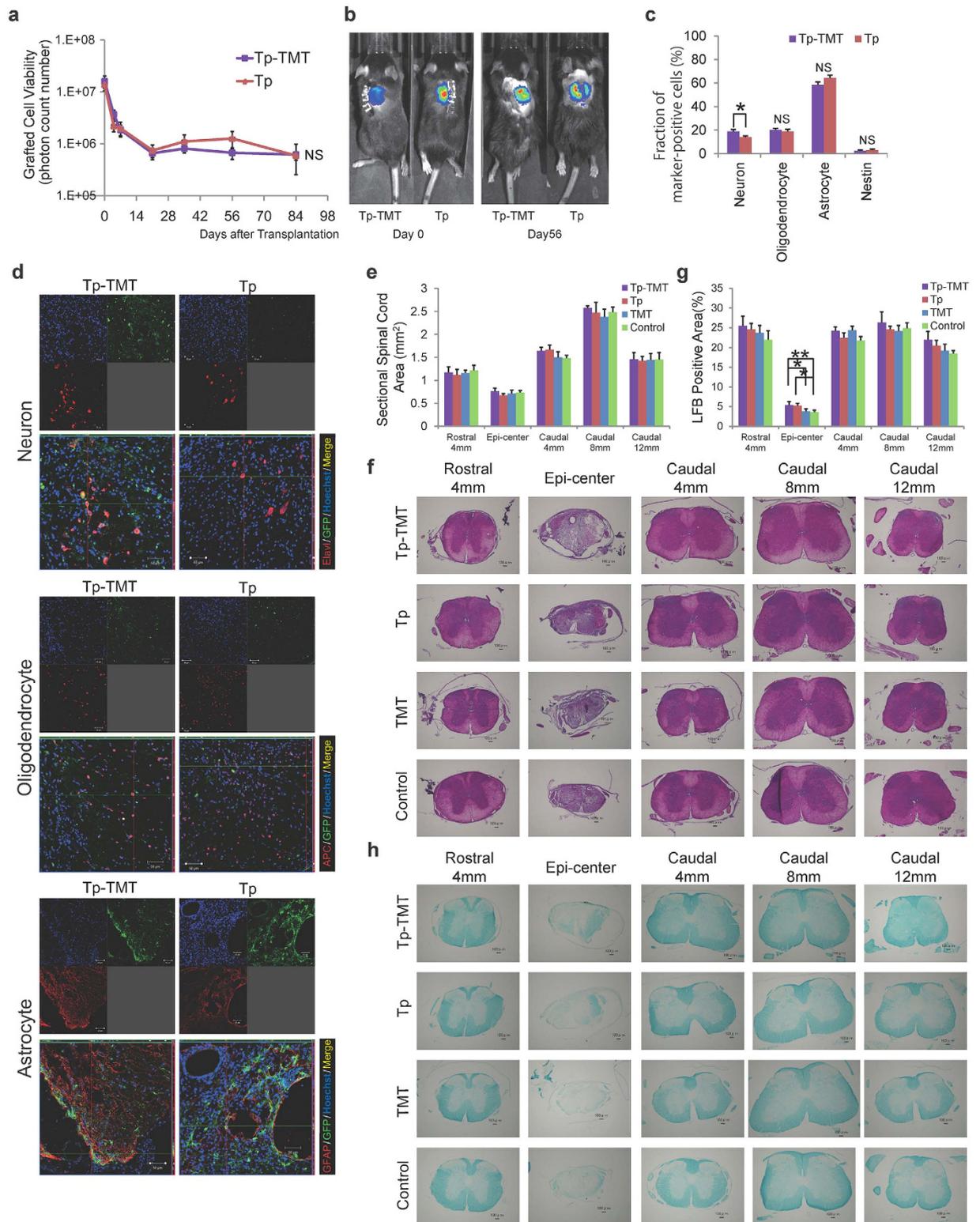


Figure 1. The effects of interventions for neural stem/progenitor cells. (a) Quantitative analysis using bioluminescence imaging (BLI) revealed that the survival rates of grafted cells were comparable between the Tp and Tp-TMT groups up to 84 days post-transplantation (the vertical axis has a logarithmic scale). Statistical analysis was performed using 2-way repeated measures ANOVA (Tp-TMT: $n = 18$, Tp: $n = 19$). (b) Representative BLI of animals in both groups. (c) The differentiation rate of grafted cells into the three neural cell lineages at 84 days post-transplantation. The proportion of Elavl⁺ cells is significantly higher in the Tp-TMT group than in the Tp group. No significant difference was observed in GFAP⁺ and APC⁺ cells. Only a small number of nestin⁺ and immature neural progenitor cells were observed in both groups. Statistical analysis was performed using paired *T*-tests ($n = 6$). (d) GFP positive grafted cells differentiated into Elavl⁺ neurons, GFAP⁺ astrocytes, and APC⁺ oligodendrocytes in Tp and Tp-TMT groups at the caudal site of the

lesion (from the lower part of T9 to T10 level). Scale bar: 50 μm . (e) The sectional spinal cord area at each spinal level, which was similar in all four groups assayed. Statistical analysis was performed using the Tukey-Kramer test ($n = 6$). (g) The myelinated area at each spinal level, which was significantly larger in the two transplanted groups (Tp and Tp-TMT groups) than in the two non-transplanted groups (TMT and Control groups). Statistical analysis was performed using the Tukey-Kramer test ($n = 6$). (f,h) Representative HE- and LFB-stained images of axial sections at each level in each group. Images of each group were obtained from identical animals. Scale bars: 100 μm . * $P < 0.05$, ** $P < 0.01$. Values are means \pm SEM.

therapies involving cell transplantation and rehabilitation is widely recognised, there have been very few studies to date, all of which were limited to the acute and subacute phase of SCI^{14,15}.

Treadmill training for SCI rodents is more established for rat models^{12,16,17}. Rats have certain advantages over mice, which include their calmer temperament, greater endurance, and larger body size, which allows for easier handling. Although training methods for mice are not as well established, methods including wheel running¹⁸ and bipedal¹⁹ or quadrupedal²⁰ treadmills have been introduced in a small number of trials conducted by a few research groups. Most studies of NS/PC transplantation have been performed using mice because of their amenability to genetic manipulation and the availability of bioluminescence imaging (BLI) for transplanted cells. The lack of an optimal well-suited model for both training and cell transplantation may be one reason why there have been very few studies of combination therapies and none in a chronic SCI animal model. In this study, we investigated how combination therapy with NS/PC transplantation and treadmill training affects behavioural function and histological manifestations in chronic SCI mice.

Results

Effects of treadmill training on survival rates and phenotype differentiation of grafted NS/PCs.

To investigate whether treadmill training changed the transplanted cell viability, BLI analysis was performed at 4, 7, 21, 35, 56, and 84 days post-transplantation. At 84 days post-transplantation, approximately 4% of the cells survived in both the combination transplantation and treadmill training (Tp-TMT) and the transplantation single therapy (Tp) groups ($3.93 \pm 2.33\%$ vs. $4.25 \pm 0.80\%$). The survival rates of the transplanted cells were comparable between the Tp-TMT group and the Tp only group (2-way repeated measures ANOVA; $P = 0.855$, Fig. 1a,b). Moreover, point-to-point analyses revealed no significant differences at each examined time point ($P < 0.05$, paired T -test).

To evaluate the differentiation phenotype of the grafted cells, immunohistochemical analyses for specific markers for each cell type were performed at 84 days post-transplantation for the Tp-TMT and the Tp only groups at the lesion epicentre and rostral and caudal sites. The following lineage specific markers were used; Elavl-like (Elavl) for neurons, adenomatous polyposis coli antigen (APC) for oligodendrocytes, and glial fibrillary acidic protein (GFAP) for astrocytes. Quantitative analyses revealed that the proportion of Elavl+ neurons was significantly higher in the Tp-TMT group than in the Tp group (Tp-TMT: 18.91%, Tp: 14.02%, $P = 0.0315$, paired T -test), whereas no significant inter-group difference was observed for APC + oligodendrocytes (Tp-TMT: $21.32 \pm 1.44\%$, Tp: $20.75 \pm 2.69\%$, $P = 0.855$) and GFAP+ astrocytes (Tp-TMT: $57.21 \pm 2.61\%$, Tp: $62.31 \pm 2.59\%$, $P = 0.195$). Nestin-positive immature cells represented around 2.5–3.0% of the grafted cells in both groups ($P = 0.799$; Fig. 1c,d).

To evaluate the effects of NS/PC transplantation and/or training on the sectional spinal cord area and myelinated area, in all the experimental groups, axial sections at the lesion epicentre, and 4 mm rostral and 4, 8, and 12 mm caudal to it, were assessed histologically with haematoxylin-eosin (HE) and Luxol Fast Blue (LFB) staining. Although LFB does not directly label the myelin (LFB cannot distinguish between myelin debris-filled macrophages, myelinated axons, remyelinated axons, or partially demyelinated axons), it is often used as a reliable index to reflect the overall myelination within the injured spinal cord^{5,7,21,22}. The experimental transverse area of the spinal cord did not significantly differ between any pair of groups, at any levels assessed, regardless of the transplantation, treadmill training, or combination of therapies (Fig. 1e,f). The LFB+ area at the epicentre was significantly higher in the two transplanted groups (Tp-TMT and Tp groups) than in the two phosphate-buffered saline (PBS) injected groups (TMT and Control groups; Tp-TMT: 5.4%, Tp: 5.3%, TMT: 3.8%, Control: 3.6%, $P = 0.048$, Tukey-Kramer test), whereas no significant difference was observed in the distant areas. There was no significant change induced by the TMT only therapy (Fig. 1g,h).

Effect of transplantation and treadmill training on the fibres in the epicentre and lumbar enlargement.

To quantify the fibres passing through the lesion related to the locomotor recovery, fibres positive to 200 kDa neurofilament (NF-H) were evaluated at the levels of the lesion epicentre, and 4 mm rostral and caudal to it, within all the experimental groups. Although it is known that increases in NF-H fibres occur secondary to subacute transplantation therapy²³, the NF-H positive areas were comparable within all four groups, at all tested levels within the chronically injured spinal cord ($P > 0.05$, Tukey-Kramer test, Fig. 2a–c). The regenerative fibres were further assayed using immunoreactivity of phosphorylated Growth Associated Protein-43 (pGAP43), which is specifically localised to regenerating, but not intact, axons²⁴. These assays were performed at the lesion epicentre and 4 mm rostral and 4, 8, and 12 mm caudal to it. In the transplanted groups, in sections 4 mm rostral and 4 mm caudal to the lesion, the pGAP43-positive area was significantly greater than in the non-transplanted groups. Although the increase of pGAP43 positive area in the Tp alone group was limited to the NS/PCs injected site, in the treadmill training combined group it was significantly greater than in the two non-transplanted groups, even in the lumbar enlargement distant from the epicentre (Tp vs Control: $P < 0.05$ at

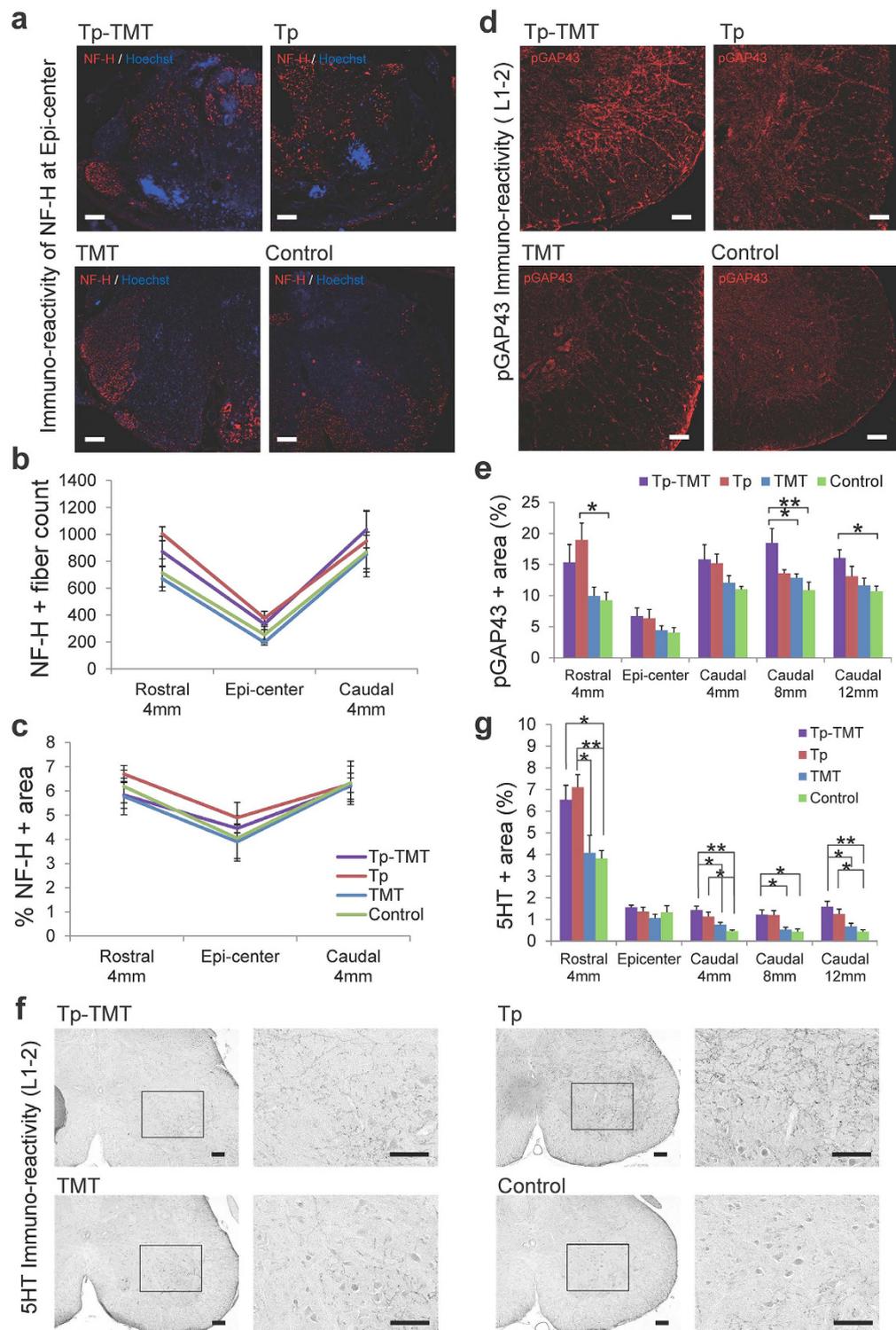


Figure 2. Fibres running through the lesion epicentre and lumbar enlargement. (a) Representative images of axial sections stained for NF-H around the lesion epicentre. (b,c) NF-H quantified with an NF-H+ dot count (b) and NF-H+ area (c). The values around the lesion epicentre were similar across the four groups for both assays. (d) Representative images of axial sections stained for pGAP43 at the lumbar enlargement. (e) Quantification of pGAP43+ area at the lesion epicentre and 4 mm rostral and 4, 8, and 12 mm caudal to the lesion across the four groups. Values increased in the two transplanted groups rostral to the lesion and were significantly greater only in the Tp-TMT group caudal to the lesion. (f) Representative images of axial sections stained for 5HT. (g) The areas of 5HT+ serotonergic fibres were significantly larger in the two transplanted groups, both rostral and caudal to the lesion epicentre, except for the epicentre itself. Statistical analyses were performed using the Tukey-Kramer test. Values are means \pm SEM ($n = 6$). * $P < 0.05$, ** $P < 0.01$. Scale bars: 100 μm in (a,d,f).

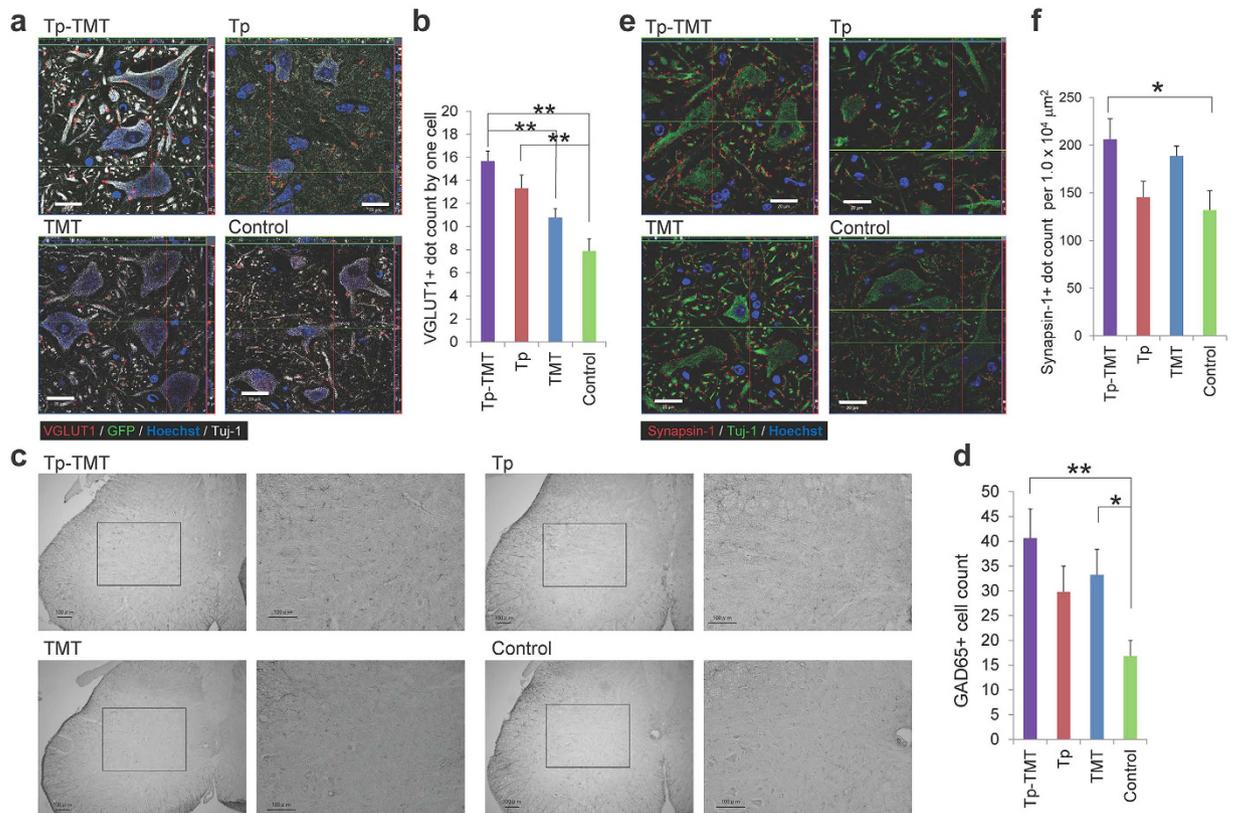


Figure 3. Effects of interventions on the neural circuit at the lumbar enlargement. (a) Representative images of axial sections stained for VGLUT1 at the anterior horn of the lumbar enlargement. (b) VGLUT1+ boutons are significantly increased in the two transplanted groups. Statistical analysis was performed using the Tukey-Kramer test. (c) Representative images of axial sections stained for GAD65 at the lumbar enlargement. (d) The number of GAD65+ cells is significantly higher in the Tp-TMT group than in the Control group. Statistical analysis was performed using the Tukey-Kramer test. (e) Representative images of axial sections stained for Synapsin-I at the anterior horn of the lumbar enlargement. (f) Synapsin-I+ boutons are significantly more numerous in the Tp-TMT group than in the Control group. Statistical analysis was performed using the Tukey-Kramer test. Values are means \pm SEM ($n=6$). * $P < 0.05$, ** $P < 0.01$. Scale bars: 20 μm in (a,e), and 50 μm in (c).

4 mm rostral to epicentre; Tp-TMT vs TMT: $P < 0.05$ at 8 mm caudal to epicentre; Tp-TMT vs Control: $P < 0.01$ at 8 mm caudal to epicentre; $P < 0.05$ at 12 mm caudal to epicentre; Tukey-Kramer test; Fig. 2d,e).

To investigate the effect of interventions on the serotonergic activity, which is also known to promote locomotor recovery after SCI through activation of the central pattern generator (CPG)^{25,26}, 5-hydroxytryptamine (5HT) positive fibres were immunohistologically assessed at the levels of the lesion epicentre, and 4 mm rostral and 4, 8, and 12 mm caudal to it. In the two transplanted groups, these fibres were significantly higher than in the two PBS injected groups, at all the levels assessed, except for the lesion epicentre. No significant effect was induced by the additional treatment with treadmill training (Tukey-Kramer test; Fig. 2f,g).

Effects of transplantation and treadmill training on the neural circuit at the lumbar enlargement. To evaluate the effects of NS/PCs transplantation and treadmill training on the CPG in the lumbar enlargement, molecules related to both its excitatory and inhibitory control were assessed immunohistologically. In rodents, the CPG elements are distributed throughout the entire lumbar region, although neurons located in L1 and L2 are shown to have a higher rhythmic capability^{27–29}. To investigate the excitatory activity in the lumbar enlargement, vesicular glutamate transporter 1 (VGLUT1) immunoreactive boutons around the motoneurons were quantified. These are known to provide an excitatory drive to the CPG^{30,31}. Although no significant difference was observed in the number of VGLUT1+ boutons between Tp-TMT and Tp groups ($P > 0.05$), their numbers were significantly increased in both Tp-TMT and Tp groups in comparison with the controls ($P < 0.01$, Tukey-Kramer test; Fig. 3a,b), indicating that transplantation had a positive effect on the excitatory drives to the CPG.

For improved gait performance, appropriate inhibitory control of the gait pattern and excitatory drive are indispensable. The number of glutamate decarboxylase-65 (GAD65) positive neurons was therefore quantified in lamina V–VII of the lumbar enlargement. These neurons are known to decrease in number after SCI, and are also known to be related to the rhythmic-coordinative inhibition of the gait and the manifestation of behavioural spasticity^{16,32–34}. Although no significant difference was observed in the number of GAD65+ neurons between the Tp-TMT and Tp groups, the number was significantly higher in the Tp-TMT group than in the Control group

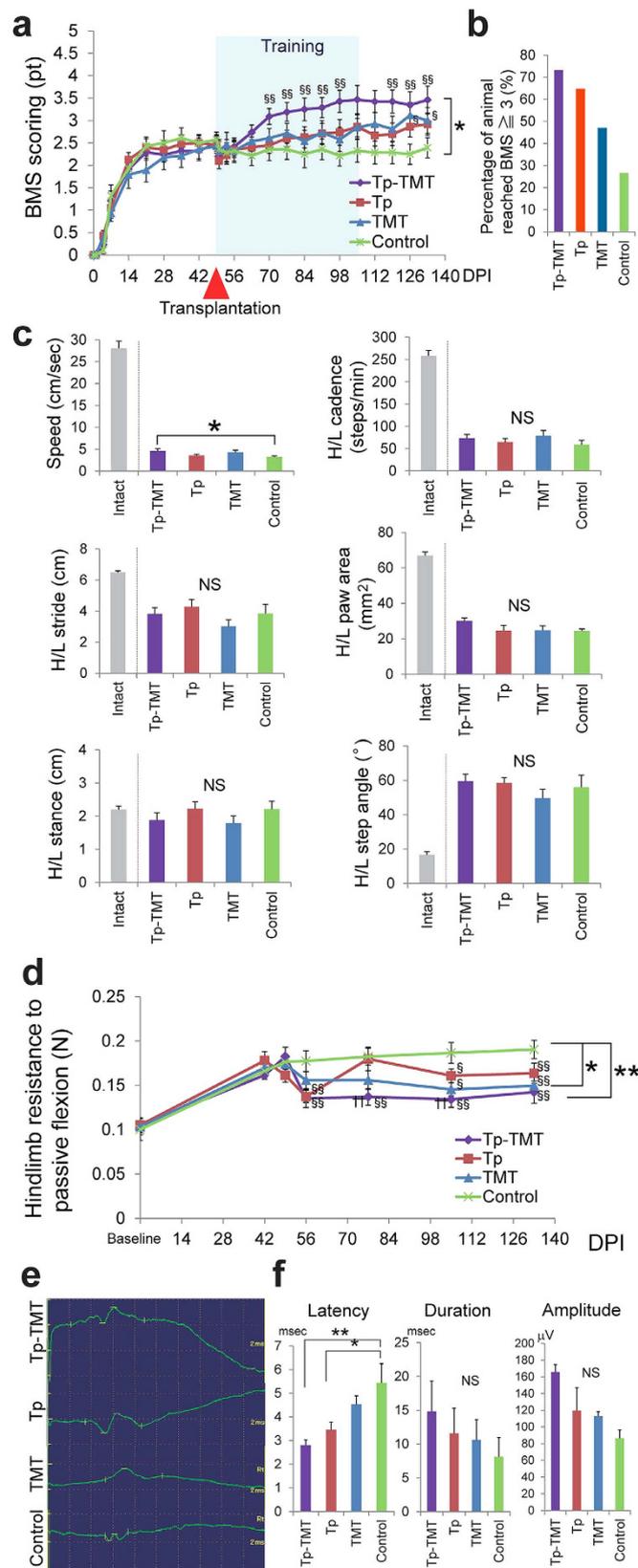


Figure 4. Behavioural and electrophysiological manifestations in each group. (a) The locomotor recovery represented by the Basso Mouse Score (BMS) in each of the four groups. The Tp-TMT group exhibited significantly better functional recovery than the control group. Statistical analyses were performed using repeated measures ANOVA followed by point to point comparisons (Tp-TMT: $n = 18$, Tp: $n = 19$, TMT: $n = 17$, Control: $n = 16$). * $P < 0.05$, in repeated measures ANOVA. $^{ss}P < 0.01$ and $^sP < 0.05$ compared to Control group in Tukey-Kramer tests. (b) The percentage of animals reaching a score of 3 or above in the BMS scoring. (c) Gait dynamics assessed

as gait speed, hindlimb (H/L) step pitch, H/L stride length, H/L stance width, and H/L step angle in each of the four groups at 133 days post-injury (DPI). The values for intact mice are cited from previous studies^{74,75}. Statistical analyses were performed using Tukey-Kramer tests (Tp-TMT: $n = 15$, Tp: $n = 15$, TMT: $n = 12$, Control: $n = 11$). * $P < 0.05$, NS: Not significant. (d) The transition over time of the resistance to flexing of animals' hindlimbs, which were passively measured using a strain-gauge in each of the four groups. The spasticity in the Tp-TMT and TMT groups was significantly reduced compared with the Control group. Statistical analyses were performed using repeated measures ANOVA followed by point to point comparisons (Tp-TMT: $n = 16$, Tp: $n = 15$, TMT: $n = 15$, Control: $n = 13$). * $P < 0.05$ and ** $P < 0.01$, in repeated measures ANOVA. ^{§§} $P < 0.01$, [§] $P < 0.05$ compared to Control group, and $H P < 0.01$ compared to Tp group in Tukey-Kramer tests. (e) Representative MEPs in the four groups at 133 DPI. (f) The latency, duration, and amplitude of the MEP are shown. The latency was significantly shortened in Tp-TMT and Tp groups. Statistical analyses were performed using Tukey-Kramer tests (Tp-TMT: $n = 10$, Tp: $n = 10$, TMT: $n = 10$, Control: $n = 9$). * $P < 0.05$. Values are means \pm SEM.

($P < 0.05$, Tukey-Kramer test; Fig. 3c,d), indicating that the combination of treadmill training with transplantation had an additional beneficial effect on coordinative control.

To further clarify the effects of the combination therapy on the maturation of the neural circuit, the number of synapses was further assessed according to the extent of Synapsin-I immuno-reactive boutons in the axial sections of the lumbar enlargement. Synapsin-1 is widely used as a presynaptic marker to examine activity-dependent synaptic plasticity and synaptic function¹². It was significantly higher in the Tp-TMT group than in the Control group ($P < 0.05$, Tukey-Kramer test; Fig. 3e,f), indicating that a combination of NS/PC transplantation and treadmill training promoted the maturation of synapses in the chronic post-transplantation spinal cord.

Functional recovery was enhanced by the combination of NS/PC transplantation and treadmill training.

Locomotor recovery was behaviourally assessed with respect to open-field locomotor function, footprint gait analysis, and spasticity. The open-field locomotor function was assessed with Basso-Mouse-Scale (BMS) scoring up to 133 DPI. It was significantly improved in the Tp-TMT group compared with the Control group (repeated measures ANOVA, multiple comparison, $P = 0.035$), whereas no significant change was detected between the control group and each of the single therapies. Point-to-point comparisons further demonstrated a significant benefit of the combination therapy compared with each of the single therapies. In this assessment, significant differences between Tp and Control groups were also revealed at 126 and 133 DPI (Steel-Dwass multiple comparison tests; Fig. 4a). The percentage of animals reaching a score of 3 or above in the BMS scoring (which corresponds to weight-bearing stepping with the hindlimb or more) was 73.3% for Tp-TMT, 64.7% for Tp, 47.1% for TMT, and 26.7% for the Control group (Fig. 4b).

Gait performance was further analysed using the DigiGait system, which revealed gait speed in the Tp-TMT group to be significantly faster than in the Control group ($P = 0.026$). No other significant changes were observed in gait performance, as measured by step cadence, stride length, hindlimb paw area, hindlimb step angle, or hindlimb step width (Steel-Dwass multiple comparison tests, $P > 0.05$ for all; Fig. 4c).

Hindlimb spasticity was assessed as the resistance to full flexion from full extension using strain-gauge testing. Spasticity was significantly suppressed in the Tp-TMT and TMT groups relative to the Control group (Fig. 4d; Tp-TMT vs Control: $P < 0.01$, TMT vs Control: $P < 0.05$, repeated measures ANOVA multiple comparison). Whereas significant increases in the spasticity were observed between the baseline time point and the last time point in the Tp only (baseline: 0.105 ± 0.007 vs 133 DPI: 0.164 ± 0.011 , $P < 0.01$), TMT only (baseline: 0.103 ± 0.009 vs 133 DPI: 0.150 ± 0.009 , $P < 0.01$), and Control groups (baseline: 0.100 ± 0.012 vs 133 DPI: 0.190 ± 0.010 , $P < 0.01$), no significant difference was observed in the Tp-TMT group (baseline: 0.105 ± 0.008 vs 133 DPI: 0.142 ± 0.012 , $P = 0.089$). A point-to-point comparison revealed that spasticity in the two transplanted groups was significantly suppressed in the early phase after transplantation therapy compared with the Control group. In the late phase, spasticity was significantly suppressed in each of the single therapies and the combination therapy, in comparison with the Control group (Fig. 4d; Tukey-Kramer tests).

The motor evoked potential (MEP) was assessed to clarify the effect of interventions on the spinal cord conduction capability. The latency of the evoked compound motor action potential was significantly shorter in the two transplanted groups than in the Control group, whereas no significant difference was induced by the addition of treadmill training (Tp-TMT vs TMT: $P < 0.05$, Tp-TMT vs Control: $P < 0.01$, Tp vs Control: $P < 0.05$, Tukey-Kramer test). Both the duration and the amplitude of the MEP were comparable between the four groups ($P > 0.05$; Fig. 4e,f).

Discussion

In this study, we show for the first time that a combination therapy with NS/PC transplantation and treadmill training can promote functional recovery, even in chronic SCI animal models. Although a paradigm for meaningful recovery has been established, which requires therapy combining neurotrophic factors, suppression of axonal growth inhibitors, rehabilitation toward functional reorganisation, and regenerative cell replacement therapy^{4,5,7}, to our knowledge there have been no reports on the effects of a combined therapy with treadmill training and transplantation in chronic animal models.

In the present study, no significant locomotor improvement was detected in the Tp only or TMT only groups, in comparison with the Control group. However, the following results indicate that each single therapy promoted potentially beneficial changes in chronically injured spinal cord. Firstly, NS/PC transplantation enhanced electrophysiological recovery, such as the shortening of MEP latency, compared with the Control group (Fig. 4e,f), which is consistent with the histological finding of an increase in the area of LFB myelin labelling at the lesion epicentre (Fig. 1g,h). Although there is controversy over whether re-myelination enhancement is secondary to cell

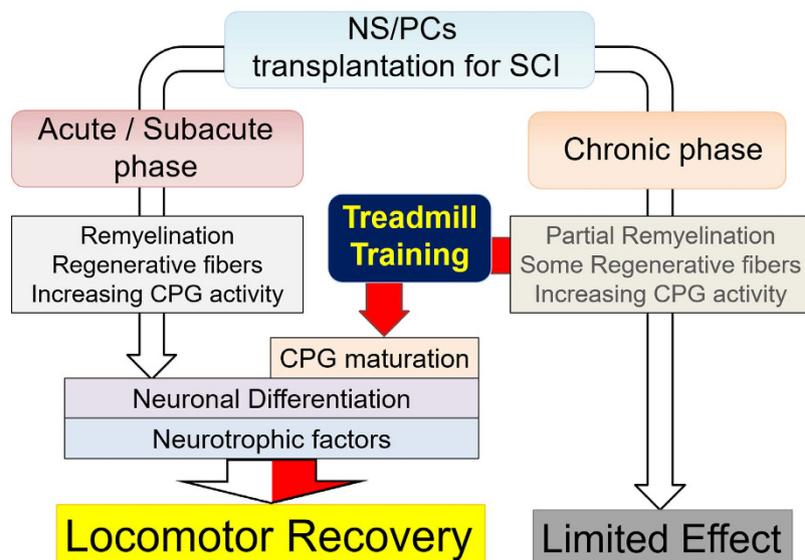


Figure 5. A scheme summarizing the beneficial mechanisms brought about by each intervention. The effects of neural stem/progenitor cell transplantation and treadmill training therapies towards functional recovery in the chronic and acute/subacute SCI model animals. Although transplantation single therapy induces only a limited effect on partial remyelination, 5HT fibre regeneration, and increasing CPG activity without appropriate inhibition, the addition of treadmill training further facilitates neuronal differentiation and CPG maturation, together with trophic support, leading to significant locomotor recovery.

	Tp	TMT	Tp-TMT
Locomotor function	±	±	+
Spasticity	–	+	+
Recovery within Lesion epicentre	–	–	–
Improvement in transplanted cell survival	NA	NA	–
Neuronal differentiation of transplanted cells	+	NA	++
Remyelination	+	–	+
Increasing excitability of CPG	+	–	+
Appropriate inhibition of CPG	–	+	+
Maturation of CPG	–	–	+

Table 1. Effects of each of the single therapies and the combination therapy on specific aspects of recovery that may lead to functional recovery.

therapy in chronic SCI, this finding is compatible with those of previous reports^{35,36}. Secondly, our results indicate that NS/PC transplantation up-regulates CPG activity, which is supported by the histological findings that 5-HT positive fibres (Fig. 2f,g) and VGLUT1 positive boutons around the motoneuron (Fig. 3a,b) are significantly increased in the transplanted groups. This reflects the excitatory activity of the CPG^{25,26,30,31}.

Treadmill training facilitates suppressive regulation related to coordinated rhythmic motor control within the lumbar enlargement, even in the chronically injured spinal cord, as shown in previous studies on subacute SCI^{16,37}. This is supported by the finding that treadmill training improved behavioural manifestations of spasticity (Fig. 4d) and increased GABAergic activity within the lumbar enlargement (Fig. 3c,d). Furthermore, neuronal differentiation and the phenomenon of appropriate inhibition recovery and/or synaptic regeneration, which we have termed “CPG maturation”, are promoted by the addition of rehabilitative therapy. The case for CPG maturation is supported by the finding of increased pGAP43 immunoreactivity (Fig. 2d,e) and synapsin-I+ synapse numbers (Fig. 3e,f). In Fig. 5 and Table 1, we have summarised the mechanisms contributing towards functional recovery that were brought about by either the transplantation or treadmill training therapies separately, or by their synergistic effects. However, it is notable that these observations provide only limited support for each aspect of recovery; this study outlines only the profiles of recovery secondary to the combination therapy of transplantation and treadmill training. Therefore, further investigations on each aspect of the findings are needed for verification.

Trophic support is one of the key factors behind the effect of combination therapy; both NS/PC transplantation and treadmill training have the potential to supply neurotrophic factors to the injured spinal cord, and the effects of combined NS/PC transplantation and treadmill training are facilitated by the combined secretion

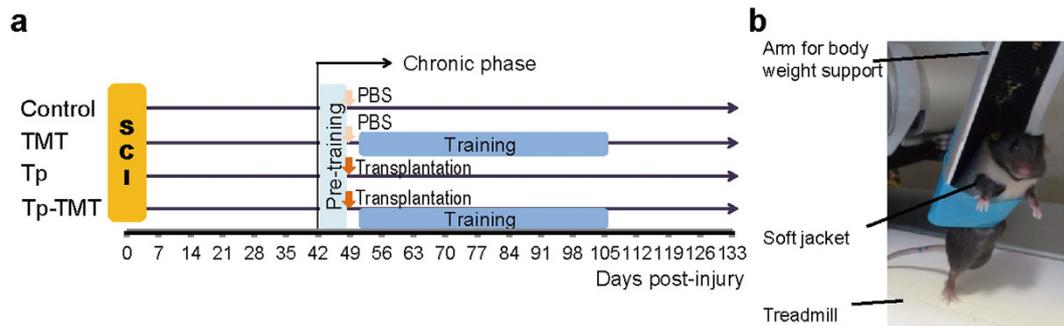


Figure 6. Experimental design and training device. (a) The complete experimental schedule. With the beginning of the chronic phase (42 DPI), all of the experimental animals are pre-trained for 1 week. Neural stem/progenitor cell transplantation or PBS injection is performed 49 days post-injury, and is then followed by 8 weeks of training period. (b) Mice are trained with a partial body weight supported bipedal gait using a commercially available robotic device.

of more neurotrophic factors than in the single conditions. It is known that various neurotrophic factors are up-regulated in subacute transplantation models^{23,38,39}, and this up-regulation is even maintained in chronically injured spinal cord⁶. When neurotrophic factors are combined with treadmill training, functional recovery is enhanced, regardless of the phase after injury^{40–43}. Similarly, treadmill training up-regulates various neurotrophic factors in both subacute and chronic SCI^{15,17,44–46}. Previous studies have demonstrated that the combination of NGF or IGF-1 with NS/PC transplantation exerts a neuroprotective effect^{5,47}. BDNF and NGF promote neuronal differentiation^{48,49}, and BDNF and NT-3 both enhance locomotor functional recovery in subacute SCI^{50–53}. Furthermore, transplantation of NS/PCs expressing NT-3 induces significant locomotor recovery, even after chronic SCI⁵. Taken together, we suggest that such trophic support enhances the intrinsic ability of these interventions for inducing locomotor recovery.

The refractory state of chronically injured spinal cord interferes with the functional recovery secondary to NS/PC transplantation^{6,7}. We suggest that the following rehabilitation-specific effects may contribute to the functional recovery observed in our study: 1) activity-dependent neuronal plasticity and modification of neural circuit(s), and 2) the treatment of learned non-use. Both task-specific and use-dependent neuronal plasticity and neural circuit modification involve the formation of new neuronal circuits⁵⁴, reinforcement of locomotor networks in a more selective and stable manner⁵⁵, changes in the synapse strength with long-term structural change⁵⁶, limiting maladaptive plasticity through training⁵⁷, spinal fixation through the peripheral sensory input from the use of paretic limbs⁵⁸, and reorganisation of the cortical network⁵⁹. We consider the principle of our rehabilitative intervention, in which we encouraged the voluntary gait, should have resulted in specific sensorimotor input into the lumbar spinal cord. This would be suitable for promoting these beneficial effects. Researchers have reported that specific training paradigms to encourage voluntary stepping are more effective than completely passive stepping^{16,60}. Instrumental training promotes activity-based learning and suppresses maladaptive plasticity⁵⁷, whereas inappropriate training intervention can reduce the capacity for motor learning⁶¹. These plasticity changes in the circumstances where NS/PCs were transplanted and survived may affect their fate and function. Furthermore, in the chronic SCI animals, “learned non-use”⁶² and disuse may trigger dysfunction, which could further suppress the impaired hindlimb activity, thereby masking the beneficial effects of transplantation. Animals in the Tp group consistently showed a gradual functional recovery, and even a significant difference compared with the control animals, when given pre-training before NS/PC transplantation in this study, as distinct from previous studies on chronic SCI transplantation^{3–7}. This concept would further support the idea that even one week of voluntary or instrumental training can induce the up-regulation of significant amounts of neurotrophic factors like BDNF, which promote spinal learning^{42,63}. Thus, even short-term conditioning training before NS/PC transplantation may induce beneficial changes by partly releasing animals from undesirable behavioural states.

No significant difference was observed in locomotor function between the Tp-TMT and TMT groups. Although the recovery in the Tp-TMT group was significant compared with the Control group, this does not necessarily indicate that combination therapy is effective for chronic SCI. Our combination therapy was not sufficiently effective to repair the lesion epicentre, as there were no significant changes in residual spinal volume, spared fibres, 5HT-positive fibres, regenerative fibres, or synaptic generation. Previous studies have shown that glial scarring composed of chondroitin-sulfate-proteoglycans physically impedes regenerative activity, and chemically impedes it by the generation of various effectors, such as semaphorin 3A. This is especially the case in chronically injured spinal cord¹³. In this regard, many researchers have investigated the efficacy of combinatorial drug treatments for inhibition of these factors in chronic SCI^{4,11,64}. Further combined treatments targeting glial scarring and/or axonal growth inhibitors using chondroitinase ABC or semaphorin3A inhibitors in combination with NS/PC transplantation and rehabilitation represent a promising future strategy.

In conclusion, rehabilitative treatments represent a third therapeutic option to facilitate locomotor recovery after NS/PC transplantation. Together with the effect of transplantation itself, treadmill training promotes functional recovery through neuronal differentiation and CPG maturation, even in chronic SCI animal models. Comprehensive treatment, including cell replacement, medication, and rehabilitation, may therefore be useful in the treatment of chronically injured spinal cord, refractory to each of these interventions alone.

Materials and Methods

Study approval. All experiments were approved by the Animal Ethics Committee of Keio University (Tokyo, Japan; No. 12082-1) and were performed in full compliance with the Guide for the Care and Use of Laboratory Animals (National Institutes of Health, Bethesda, MD).

Experimental design. To investigate the profiles of the change induced by each of the treadmill training and transplantation therapies, a four-armed design with groups composed of combination therapy (Tp-TMT) transplantation single therapy (Tp), treadmill training single therapy (TMT), and a control group (Control) was applied. As shown in Fig. 6, the NS/PCs were transplanted at 49 DPI. Treadmill training was performed for two independent periods following different strategies: pre-training for all of the SCI mice at 42–48 DPI, and intervention training for mice in the TMT and Tp-TMT groups at 52–105 DPI (Fig. 6a).

Animals. Eighty C57BL/6J mice (8–9 weeks old, female, 18–22 g; Clea, Tokyo, Japan) were used in this study. Because some animals died in the process of the experiment, a total of seventy were included in the results. The sample size for each experiment was determined according to averages and standard deviations were calculated from preliminary experiments. The animals were housed 3–5 per cage in an accredited facility. The animals were maintained on a 12 h light/dark cycle with access to food and water ad libitum.

NS/PC culture. NS/PCs were cultured and expanded as previously described⁶⁵. The source of NS/PCs was selected according to previous reports^{1,66,67}. Briefly, the cells were harvested from the striata of transgenic mice established from C57BL/6J on embryonic day 14. These mice ubiquitously express fLuc-cp156, which is a fusion protein of a yellow variant of Aequorea GFP and firefly luciferase⁵¹. Dissociated cells were collected and re-suspended in culture medium composed of Dulbecco's modified Eagle medium/F12 (Sigma-Aldrich, St. Louis, MO, USA) with hormone mixture. Human recombinant FGF-2 (Peprotech, Rocky hill, NJ, USA) and EGF (Peprotech; 20 ng/ml each) were added every 2 days. Cells were expanded for three passages and the neurospheres were used for cell transplantation.

SCI model and grouping. Severe lower thoracic level contusive SCI was performed as described previously⁷. Briefly, all mice were anaesthetised with an intraperitoneal (i.p.) injection of ketamine (100 mg/kg) and xylazine (10 mg/kg). Following T9 laminectomy, a 70 kilodyne contusive injury was applied to the exposed dura mater using a commercially available SCI device (IH Impactor, Precision Systems and Instrumentation, Lexington, KY, USA). For 3 days after the injury, 12.5 mg/kg ampicillin was administered intramuscularly. All injured mice received twice-daily manual bladder evacuations until recovery of function. The SCI animals were randomly assigned to each group (20 mice per group). As some mice died in the process of the interventions, the following numbers of animals were included in the analyses: Tp-TMT, $n = 18$; Tp, $n = 19$; TMT, $n = 17$; and Control, $n = 16$. All the information regarding the groups was handled separately and investigators performing behavioural assessments were blind to this information.

NS/PC transplantation. NS/PCs (approximately 5×10^5 cells/2 μ l) were transplanted separately into regions 1 mm rostral and caudal to the rim of the lesion epicentre at 49 DPI, following the method reported in a previous study⁶⁸. In brief, NS/PCs were injected with a glass micropipette at a rate of 1 μ l/min using a Hamilton syringe (25 μ l, Hamilton, Bonaduz, Switzerland) and a stereotaxic microinjector (KDS 310, Muromachi-kikai Co. Ltd., Tokyo, Japan). In the TMT and Control groups, PBS was injected in the same manner, instead of the NS/PCs.

Bioluminescence imaging. *In vivo* BLI analysis was performed with a Xenogeny-IVIS spectrum CCD optical macroscopic imaging system (Caliper LifeSciences, Hopkinton, MA, USA) as previously reported^{1,7,68}. Briefly, the signal intensity from the transplanted cells was recorded as the maximum 5 min integration of bioluminescence over the period from 15 to 45 min post i.p. injection of D-luciferin (0.3 mg/g). Images were analysed with Living Image software (Caliper LifeSciences), and the signal intensity was measured as photon flux.

Treadmill training. SCI animals underwent partial body weight supported, voluntary bipedal gait training using a commercially available treadmill training device (Rodent Robot 3000; Robomedica Inc., Irvine, CA; Fig. 6b)¹⁶. The training intervention was performed for two purposes in two different periods; the conditioning training was conducted for all animals before the injection of NS/PCs or PBS from 42 to 48 DPI, and the intervention training was conducted post-injection for animals in TMT and Tp-TMT groups from 52 to 105 DPI. The training was performed for 20 minutes per day, 5 days per week, with approximately 80% to 90% of the animals' body weight supported. The speed of the treadmill was set between 0.5 to 1.5 cm/s. Weight-support and speed were adjusted on every intervention day to optimally induce a voluntary gait.

Behavioural analyses. Hindlimb locomotor function was evaluated weekly up to 133 DPI using the Basso-Mouse-Scale (BMS)⁶⁹. Quadrupedal gait dynamics regarding gait speed, cadence, stride length, paw area, stance width, and step angle of hindlimbs were evaluated from the footprints of the mice using a DigiGait imaging and analysis system (Mouse Specifics, Boston, MA)^{21,70}. Hindlimb spasticity was assessed as the resistance to full flexion from full extension⁷¹. The maximum force required was measured using a handheld strain-gauge (model FGP-0.5; Nidec-Shimpo, Kyoto, Japan) and the average value of six measurements was recorded¹⁶.

Electrophysiology. MEPs were recorded using a Neuropack S1 MEB-9402 (Nihon Kohden, Tokyo, Japan) at 133 DPI, as previously described²¹. Briefly, a C1 laminectomy was performed after anaesthetisation with an i.p. injection of ketamine (60 mg/kg) and xylazine (6 mg/kg). Stimulation was applied from the C0 level of the spinal

cord with a wired electrode, and the MEP was picked up from the quadriceps muscle of the hindlimbs by a needle electrode. The intensity of the stimulus was set to trigger a supra-maximum response (approximately 1.0–2.0 mA intensity), and the duration and the stimulus frequency were set to 0.2 ms and 1 Hz, respectively. The latency, amplitude, and duration of the induced potential were recorded.

Histological analyses. At 133 DPI, anaesthetised animals were transcardially perfused with PBS, followed by fixation with 4% paraformaldehyde (in 0.1 M PBS ($n = 6$), each group). The spinal cords were removed, postfixed overnight in 4% paraformaldehyde, soaked overnight in 10% sucrose, followed by 30% sucrose, embedded in Optimal Cutting Temperature compound (Sakura Finetechnical Co., Ltd., Tokyo, Japan), and frozen as previously described⁷. A cryostat (CM3050 S; Leica Microsystems, Wetzlar, Germany) was used to dissect sections (20 μ m thick) out of the injured spinal cords from 4 mm rostral to the lesion epicentre to 12 mm caudal to the lesion epicentre. The sections of transplanted spinal cord were then subjected to histological analyses. For assessment of NS/PC differentiation, and quantification of NF-H, pGAP43, VGLUT1, Synapsin-I, 5HT, and GAD65, the sections were incubated at 4 °C overnight with primary antibodies, and then incubated with appropriate secondary antibodies after washing. Supplementary Table 1 lists the primary antibodies used in the study. Nuclei were stained with Hoechst 33258 (10 μ g/ml, Sigma-Aldrich). All images except for HE, LFB, 5HT and GAD65 were obtained using a confocal laser scanning microscope (LSM 780; Carl Zeiss, Munich, Germany). Images of HE, LFB, 5HT and GAD65 staining were captured using fluorescence microscopy (BZ-9000; Keyence, Tokyo, Japan). Immunoreactivity of 5HT and GAD65 was analysed using 3,3'-diaminobenzidine immunohistochemistry. A biotinylated secondary antibody (Jackson ImmunoResearch Laboratories Inc., West Grove, PA, USA) was used after exposing the sections to 0.3% H₂O₂ to inactivate endogenous peroxidases. Signals were enhanced with the Vectastain ABC kit (Vector Laboratories, Inc., Burlingame, CA, USA). Diaminobenzidine (Wako; 0.005%) was used as a chromogen, and the reactions with 0.0075% hydrogen peroxide in water with Tris buffer were sustained for 3 min. Threshold values were maintained at constant levels for all analyses.

Quantitative analyses. To quantify the proportion of each differentiated cell phenotype among the *in vivo* grafted cells, five regions were captured within axial sections at 200 \times magnification. GFP and phenotypic marker double-positive cells were counted in five sections, for six animals per group, at the lesion epicentre and at the rostral and caudal sites (T8–T10 level), as previously described^{7,68}. The sectional spinal area was determined using HE- and LFB- stained images of axial sections from the lesion epicentre and 4.0 mm rostral and 4.0, 8.0, and 12.0 mm caudal to the epicentre, captured at 100 \times magnification ($n = 6$, each group). The myelinated areas were measured as for LFB+, in the manner following previous studies^{7,68,72}. The NF-H+ fibres were quantified using axial sections from the lesion epicentre and 4.0 mm rostral and 4.0 mm caudal to the epicentre, captured at 200 \times magnification. The whole axial sections were evaluated for immunoreactive area and the number of fibres running through the section. The pGAP43+ and the 5HT+ fibres were evaluated using axial sections from the lesion epicentre and 4.0 mm rostral and 4.0, 8.0 and 12.0 mm caudal to the epicentre, captured at 200 \times magnification. The pGAP43+ fibres were quantified for immunoreactive area within the whole axial section, and the 5HT+ fibres were quantified for immunoreactive area within the automatically captured images of spinal grey matter. ImageJ software was used for these analyses (version 1.47; National Institutes of Health, USA). The average value of the neighbouring three sections was recorded ($n = 6$, each group) following the methods of previous studies^{7,12,26,68}. VGLUT1 and synapsin-1 immunoreactive synapses were assessed using the axial sections of the lumbar enlargement captured at 630 \times magnification, and following procedures described in previous studies ($n = 6$, each group)^{12,30,31}. Briefly, the numbers of VGLUT1 or synapsin-1 immunoreactive boutons around a motoneuron labelled with Tuj-1 were counted from three optical sections for one cell. Twelve randomly selected motoneurons from six optical sections within lamina IX at L1–2 were assayed in this manner for one animal. GAD65+ cells were determined using the axial sections of the lumbar enlargement captured at 100 \times , according to the methods in previous studies ($n = 6$, each group)^{33,34}. The average number of GAD65+ neurons within lamina V–VII was counted, and then the average value from three different levels at L1–2 of the lumbar enlargement was calculated. The threshold values were maintained at a constant level for all analyses.

Statistical analysis. All data are presented as mean \pm SEM. Two-way repeated measures ANOVAs, along with point-by-point comparisons with Steel-Dwass tests or Tukey-Kramer tests, were used to examine the differences between groups in assessments of the BMS score, strain-gauge test, and grafted cell viability, following methods in a previous study⁷³. The comparisons between the four groups were performed using Tukey-Kramer tests (*i.e.*, the parameters from the behavioural DigiGait and electrophysiological MEP analyses, histological assessments regarding sectional spinal area with HE staining, myelinated area with LFB staining, NF-H+ dot count and area, pGAP43+ area, 5-HT+ area, VGLUT1+ dot count, GAD65+ area, and synapsin-1+ dot count). The comparisons between two groups (*i.e.*, assessments on each lineage in the cell differentiation assay) were performed using *T*-tests.

References

- Okada, S. *et al.* *In vivo* imaging of engrafted neural stem cells: its application in evaluating the optimal timing of transplantation for spinal cord injury. *FASEB J* **19**, 1839–1841 (2005).
- Ogawa, Y. *et al.* Transplantation of *in vitro*-expanded fetal neural progenitor cells results in neurogenesis and functional recovery after spinal cord contusion injury in adult rats. *J Neurosci Res* **69**, 925–933 (2002).
- Parr, A. M., Kulbatski, I. & Tator, C. H. Transplantation of adult rat spinal cord stem/progenitor cells for spinal cord injury. *J Neurotrauma* **24**, 835–845 (2007).
- Karimi-Abdolrezaee, S., Eftekharpour, E., Wang, J., Schut, D. & Fehlings, M. G. Synergistic effects of transplanted adult neural stem/progenitor cells, chondroitinase, and growth factors promote functional repair and plasticity of the chronically injured spinal cord. *J Neurosci* **30**, 1657–1676 (2010).

5. Kusano, K. *et al.* Transplanted neural progenitor cells expressing mutant NT3 promote myelination and partial hindlimb recovery in the chronic phase after spinal cord injury. *Biochem Biophys Res Commun* **393**, 812–817 (2010).
6. Kumamaru, H. *et al.* Therapeutic activities of engrafted neural stem/precursor cells are not dormant in the chronically injured spinal cord. *Stem Cells* **31**, 1535–1547 (2013).
7. Nishimura, S. *et al.* Time-dependent changes in the microenvironment of injured spinal cord affects the therapeutic potential of neural stem cell transplantation for spinal cord injury. *Mol Brain* **6**, 3 (2013).
8. Salazar, D. L., Uchida, N., Hamers, F. P., Cummings, B. J. & Anderson, A. J. Human neural stem cells differentiate and promote locomotor recovery in an early chronic spinal cord injury NOD-scid mouse model. *PLoS One* **5**, e12272 (2010).
9. Cusimano, M. *et al.* Transplanted neural stem/precursor cells instruct phagocytes and reduce secondary tissue damage in the injured spinal cord. *Brain* **135**, 447–460 (2012).
10. Bradbury, E. J. *et al.* Chondroitinase ABC promotes functional recovery after spinal cord injury. *Nature* **416**, 636–640 (2002).
11. Garcia-Alias, G. *et al.* Chondroitinase ABC combined with neurotrophin NT-3 secretion and NR2D expression promotes axonal plasticity and functional recovery in rats with lateral hemisection of the spinal cord. *J Neurosci* **31**, 17788–17799 (2011).
12. Zhang, L. *et al.* Rewiring of regenerated axons by combining treadmill training with semaphorin3A inhibition. *Mol Brain* **7**, 14 (2014).
13. Kaneko, S. *et al.* A selective Sema3A inhibitor enhances regenerative responses and functional recovery of the injured spinal cord. *Nat Med* **12**, 1380–1389 (2006).
14. Sun, T., Ye, C., Zhang, Z., Wu, J. & Huang, H. Cotransplantation of olfactory ensheathing cells and Schwann cells combined with treadmill training promotes functional recovery in rats with contused spinal cords. *Cell Transplant* **22** Suppl 1, S27–38 (2013).
15. Hwang, D. H. *et al.* Survival of neural stem cell grafts in the lesioned spinal cord is enhanced by a combination of treadmill locomotor training via insulin-like growth factor-1 signaling. *J Neurosci* **34**, 12788–12800 (2014).
16. Tashiro, S. *et al.* BDNF Induced by Treadmill Training Contributes to the Suppression of Spasticity and Allodynia After Spinal Cord Injury via Upregulation of KCC2. *Neurorehabil Neural Repair* **29**, 677–689 (2015).
17. Ying, Z., Roy, R. R., Edgerton, V. R. & Gomez-Pinilla, F. Exercise restores levels of neurotrophins and synaptic plasticity following spinal cord injury. *Exp Neurol* **193**, 411–419 (2005).
18. Engesser-Cesar, C. *et al.* Wheel running following spinal cord injury improves locomotor recovery and stimulates serotonergic fiber growth. *Eur J Neurosci* **25**, 1931–1939 (2007).
19. Fong, A. J. *et al.* Spinal cord-transected mice learn to step in response to quipazine treatment and robotic training. *J Neurosci* **25**, 11738–11747 (2005).
20. Goldshmit, Y., Lythgo, N., Galea, M. P. & Turnley, A. M. Treadmill training after spinal cord hemisection in mice promotes axonal sprouting and synapse formation and improves motor recovery. *J Neurotrauma* **25**, 449–465 (2008).
21. Nori, S. *et al.* Grafted human-induced pluripotent stem-cell-derived neurospheres promote motor functional recovery after spinal cord injury in mice. *Proc Natl Acad Sci USA* **108**, 16825–16830 (2011).
22. Kobayashi, Y. *et al.* Pre-evaluated safe human iPSC-derived neural stem cells promote functional recovery after spinal cord injury in common marmoset without tumorigenicity. *PLoS One* **7**, e52787 (2012).
23. Kumagai, G. *et al.* Roles of ES cell-derived gliogenic neural stem/progenitor cells in functional recovery after spinal cord injury. *PLoS One* **4**, e7706 (2009).
24. Oyamatsu, H., Koga, D., Igarashi, M., Shibata, M. & Ushiki, T. Morphological assessment of early axonal regeneration in end-to-side nerve coaptation models. *J Plast Surg Hand Surg* **46**, 299–307 (2012).
25. Ghosh, M. & Pearse, D. D. The role of the serotonergic system in locomotor recovery after spinal cord injury. *Front Neural Circuits* **8**, 151 (2014).
26. Kim, D. *et al.* Direct agonists for serotonin receptors enhance locomotor function in rats that received neural transplants after neonatal spinal transection. *J Neurosci* **19**, 6213–6224 (1999).
27. Nishimaru, H., Takizawa, H. & Kudo, N. 5-Hydroxytryptamine-induced locomotor rhythm in the neonatal mouse spinal cord *in vitro*. *Neurosci Lett* **280**, 187–190 (2000).
28. Kjaerulff, O. & Kiehn, O. Distribution of networks generating and coordinating locomotor activity in the neonatal rat spinal cord *in vitro*: a lesion study. *J Neurosci* **16**, 5777–5794 (1996).
29. Guertin, P. A. Central Pattern Generator for Locomotion: Anatomical, Physiological, and Pathophysiological Considerations. *Frontiers in Neurology* **3**, doi: 10.3389/fneur.2012.00183 (2013).
30. Kitzman, P. VGLUT1 and GLYT2 labeling of sacrocaudal motoneurons in the spinal cord injured spastic rat. *Exp Neurol* **204**, 195–204 (2007).
31. Cantoria, M. J., See, P. A., Singh, H. & de Leon, R. D. Adaptations in glutamate and glycine content within the lumbar spinal cord are associated with the generation of novel gait patterns in rats following neonatal spinal cord transection. *J Neurosci* **31**, 18598–18605 (2011).
32. Nishimaru, H., Sakagami, H., Kakizaki, M. & Yanagawa, Y. Locomotor-related activity of GABAergic interneurons localized in the ventrolateral region in the isolated spinal cord of neonatal mice. *J Neurophysiol* **106**, 1782–1792 (2011).
33. Kaptza, S. *et al.* Tail spasms in rat spinal cord injury: changes in interneuronal connectivity. *Exp Neurol* **236**, 179–189 (2012).
34. Wilson, J. M., Blagovetchenski, E. & Brownstone, R. M. Genetically defined inhibitory neurons in the mouse spinal cord dorsal horn: a possible source of rhythmic inhibition of motoneurons during fictive locomotion. *J Neurosci* **30**, 1137–1148 (2010).
35. Zurita, M., Aguayo, C., Bonilla, C., Rodriguez, A. & Vaquero, J. Perilesional intrathecal administration of autologous bone marrow stromal cells achieves functional improvement in pigs with chronic paraplegia. *Cytotherapy* **15**, 1218–1227 (2013).
36. Nakano, N. *et al.* Effects of bone marrow stromal cell transplantation through CSF on the subacute and chronic spinal cord injury in rats. *PLoS One* **8**, e73494 (2013).
37. Tillakaratne, N. J. *et al.* Use-dependent modulation of inhibitory capacity in the feline lumbar spinal cord. *J Neurosci* **22**, 3130–3143 (2002).
38. Hawryluk, G. W. *et al.* An *in vivo* characterization of trophic factor production following neural precursor cell or bone marrow stromal cell transplantation for spinal cord injury. *Stem Cells Dev* **21**, 2222–2238 (2012).
39. Kamei, N. *et al.* BDNF, NT-3, and NGF released from transplanted neural progenitor cells promote corticospinal axon growth in organotypic cocultures. *Spine (Phila Pa 1976)* **32**, 1272–1278 (2007).
40. Ying, Z. *et al.* BDNF-exercise interactions in the recovery of symmetrical stepping after a cervical hemisection in rats. *Neuroscience* **155**, 1070–1078 (2008).
41. Kao, C. H., Chen, S. H., Chio, C. C., Chang, C. K. & Lin, M. T. Exogenous administration of glial cell line-derived neurotrophic factor improves recovery after spinal cord injury. *Resuscitation* **77**, 395–400 (2008).
42. Gomez-Pinilla, F. *et al.* BDNF and learning: Evidence that instrumental training promotes learning within the spinal cord by up-regulating BDNF expression. *Neuroscience* **148**, 893–906 (2007).
43. Boyce, V. S., Tumolo, M., Fischer, I., Murray, M. & Lemay, M. A. Neurotrophic factors promote and enhance locomotor recovery in untrained spinalized cats. *J Neurophysiol* **98**, 1988–1996 (2007).
44. Cote, M. P., Azzam, G. A., Lemay, M. A., Zhukareva, V. & Houle, J. D. Activity-dependent increase in neurotrophic factors is associated with an enhanced modulation of spinal reflexes after spinal cord injury. *J Neurotrauma* **28**, 299–309 (2011).

45. Skup, M. *et al.* Long-Term Locomotor Training Up-Regulates TrkBFL Receptor-like Proteins, Brain-Derived Neurotrophic Factor, and Neurotrophin 4 with Different Topographies of Expression in Oligodendroglia and Neurons in the Spinal Cord. *Experimental Neurology* **176**, 289–307 (2002).
46. Houle, J. D. & Cote, M. P. Axon regeneration and exercise-dependent plasticity after spinal cord injury. *Ann N Y Acad Sci* **1279**, 154–163 (2013).
47. Lunn, J. S. *et al.* Autocrine production of IGF-I increases stem cell-mediated neuroprotection. *Stem Cells* **33**, 1480–1489 (2015).
48. Liu, F. *et al.* Combined effect of nerve growth factor and brain-derived neurotrophic factor on neuronal differentiation of neural stem cells and the potential molecular mechanisms. *Mol Med Rep* **10**, 1739–1745 (2014).
49. Chen, S. Q., Cai, Q., Shen, Y. Y., Cai, X. Y. & Lei, H. Y. Combined use of NGF/BDNF/bFGF promotes proliferation and differentiation of neural stem cells *in vitro*. *Int J Dev Neurosci* **38**, 74–78 (2014).
50. He, B. L. *et al.* BDNF expression with functional improvement in transected spinal cord treated with neural stem cells in adult rats. *Neuropeptides* **47**, 1–7 (2013).
51. Kitagawa, A. *et al.* Lectinized brain-derived neurotrophic factor promotes the differentiation of embryonic stem cells *in vitro* and *in vivo*. *Biochem Biophys Res Commun* **328**, 1051–1057 (2005).
52. Gu, Y. L. *et al.* Neurotrophin expression in neural stem cells grafted acutely to transected spinal cord of adult rats linked to functional improvement. *Cell Mol Neurobiol* **32**, 1089–1097 (2012).
53. Lu, H. *et al.* Retrovirus delivered neurotrophin-3 promotes survival, proliferation and neuronal differentiation of human fetal neural stem cells *in vitro*. *Brain Res Bull* **77**, 158–164 (2008).
54. Girgis, J. *et al.* Reaching training in rats with spinal cord injury promotes plasticity and task specific recovery. *Brain* **130**, 2993–3003 (2007).
55. Ichiyama, R. M. *et al.* Step training reinforces specific spinal locomotor circuitry in adult spinal rats. *J Neurosci* **28**, 7370–7375 (2008).
56. Tahayori, B. & Kocaja, D. M. Activity-dependent plasticity of spinal circuits in the developing and mature spinal cord. *Neural Plast* **2012**, 964843 (2012).
57. Hansen, C. N. *et al.* Sparing of Descending Axons Rescues Interneuron Plasticity in the Lumbar Cord to Allow Adaptive Learning After Thoracic Spinal Cord Injury. *Front Neural Circuits* **10**, 11 (2016).
58. Steinmetz, J. E., Cervenka, J., Robinson, C., Romano, A. G. & Patterson, M. M. Fixation of spinal reflexes in rats by central and peripheral sensory input. *J Comp Physiol Psychol* **95**, 548–555 (1981).
59. Winchester, P. *et al.* Changes in supraspinal activation patterns following robotic locomotor therapy in motor-incomplete spinal cord injury. *Neurorehabil Neural Repair* **19**, 313–324 (2005).
60. Lee, C., Won, D., Cantoria, M. J., Hamlin, M. & de Leon, R. D. Robotic assistance that encourages the generation of stepping rather than fully assisting movements is best for learning to step in spinally contused rats. *J Neurophysiol* **105**, 2764–2771 (2011).
61. Bigbee, A. J. *et al.* Two chronic motor training paradigms differentially influence acute instrumental learning in spinally transected rats. *Behav Brain Res* **180**, 95–101 (2007).
62. Taub, E., Williams, M., Barro, G. & Steiner, S. S. Comparison of the performance of deafferented and intact monkeys on continuous and fixed ration schedules of reinforcement. *Exp Neurol* **58**, 1–13 (1978).
63. Gomez-Pinilla, F., Ying, Z., Roy, R. R., Molteni, R. & Edgerton, V. R. Voluntary exercise induces a BDNF-mediated mechanism that promotes neuroplasticity. *J Neurophysiol* **88**, 2187–2195 (2002).
64. Kanno, H. *et al.* Combination of engineered Schwann cell grafts to secrete neurotrophin and chondroitinase promotes axonal regeneration and locomotion after spinal cord injury. *J Neurosci* **34**, 1838–1855 (2014).
65. Reynolds, B. A., Tetzlaff, W. & Weiss, S. A multipotent EGF-responsive striatal embryonic progenitor cell produces neurons and astrocytes. *J Neurosci* **12**, 4565–4574 (1992).
66. Tsuji, O. *et al.* Cell therapy for spinal cord injury by neural stem/progenitor cells derived from iPS/ES cells. *Neurotherapeutics* **8**, 668–676 (2011).
67. Watanabe, K. *et al.* Comparison between fetal spinal-cord- and forebrain-derived neural stem/progenitor cells as a source of transplantation for spinal cord injury. *Dev Neurosci* **26**, 275–287 (2004).
68. Iwai, H. *et al.* Transplantation of neural stem/progenitor cells at different locations in mice with spinal cord injury. *Cell Transplant* **23**, 1451–1464 (2014).
69. Basso, D. M. *et al.* Basso Mouse Scale for locomotion detects differences in recovery after spinal cord injury in five common mouse strains. *J Neurotrauma* **23**, 635–659 (2006).
70. Yasuda, A. *et al.* Significance of remyelination by neural stem/progenitor cells transplanted into the injured spinal cord. *Stem Cells* **29**, 1983–1994 (2011).
71. Baker, D. *et al.* Cannabinoids control spasticity and tremor in a multiple sclerosis model. *Nature* **404**, 84–87 (2000).
72. Tsuji, O. *et al.* Therapeutic potential of appropriately evaluated safe-induced pluripotent stem cells for spinal cord injury. *Proc Natl Acad Sci USA* **107**, 12704–12709 (2010).
73. Renault-Mihara, F. *et al.* Beneficial compaction of spinal cord lesion by migrating astrocytes through glycogen synthase kinase-3 inhibition. *EMBO Mol Med* **3**, 682–696 (2011).
74. Vincette, J. *et al.* Gait analysis in a murine model of collagen-induced arthritis. *Arthritis Res Ther* **9**, R123 (2007).
75. Kale, A., Amende, I., Meyer, G. P., Crabbe, J. C. & Hampton, T. G. Ethanol's Effects on Gait Dynamics in Mice Investigated by Ventral Plane Videography. *Alcoholism: Clinical & Experimental Research* **28**, 1839–1848 (2004).

Acknowledgements

We appreciate the help of all the members of the spinal cord research team at the Department of Orthopaedic Surgery, Physiology, and Rehabilitation Medicine at Keio University's School of Medicine, and Dr H. Nishimaru at University of Toyama Faculty of Medicine. We also thank Dr D. Sipp at RIKEN Center for Developmental Biology for the critical reading of the manuscript. The authors disclose receipt of the following financial support for the research, authorship, and/or publication of this article. This work was supported by grants from the following organisations: the Japan Science and Technology-California Institute for Regenerative Medicine collaborative program; the Grants-in-Aid for Scientific Research from the Japan Society for the Promotion of Science (SPS), and the Ministry of Education, Culture, Sports, Science, and Technology of Japan (MEXT). This work was also supported by the Research Center Network for Realization of Regenerative Medicine by the Japan Science and Technology Agency (JST) and the Japan Agency for Medical Research and Development (AMED; to H.O. and M.N.). H.O. is a paid scientific advisory board member for SanBio Co., Ltd.; the Keio Gijuku Academic by a Grant-in-Aid for Scientific Research on Innovative Areas (Comprehensive Brain Science Network) from MEXT; and the General Insurance Association of Japan.

Author Contributions

S.T., Y.T., M.L., H.O. and M.N. designed the research; S.T., S.N. and L.Z. conducted research; S.N. and K.S. provided NS/PCs, S.T., H.I. and M.S. acquired data; S.T. and A.I. analysed the data; S.T., H.O. and M.N. wrote the manuscript; and M.N. and H.O. supervised all the experiments. All authors read and approved the final manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: H. Okano is a scientific consultant for SanBio Co., Ltd (Japan), Eisai Co., Ltd (Japan), and Daiichi Sankyo Co., Ltd (Japan). The remaining authors report no conflicts of interest.

How to cite this article: Tashiro, S. *et al.* Functional Recovery from Neural Stem/Progenitor Cell Transplantation Combined with Treadmill Training in Mice with Chronic Spinal Cord Injury. *Sci. Rep.* **6**, 30898; doi: 10.1038/srep30898 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2016