

SCIENTIFIC REPORTS



OPEN

Predicting chemotherapeutic drug combinations through gene network profiling

Received: 19 March 2015
Accepted: 23 November 2015
Published: 21 January 2016

Thi Thuy Trang Nguyen^{1,2}, Jacqueline Kia Kee Chua^{1,8}, Kwi Shan Seah^{1,2}, Seok Hwee Koo^{5,10}, Jie Yin Yee^{2,5}, Eugene Guorong Yang⁶, Kim Kiat Lim^{1,2}, Shermaine Yu Wen Pang¹, Audrey Yuen⁹, Louxin Zhang⁷, Wee Han Ang^{4,8}, Brian Dymock⁶, Edmund Jon Deoon Lee^{2,5} & Ee Sin Chen^{1,2,3,4}

Contemporary chemotherapeutic treatments incorporate the use of several agents in combination. However, selecting the most appropriate drugs for such therapy is not necessarily an easy or straightforward task. Here, we describe a targeted approach that can facilitate the reliable selection of chemotherapeutic drug combinations through the interrogation of drug-resistance gene networks. Our method employed single-cell eukaryote fission yeast (*Schizosaccharomyces pombe*) as a model of proliferating cells to delineate a drug resistance gene network using a synthetic lethality workflow. Using the results of a previous unbiased screen, we assessed the genetic overlap of doxorubicin with six other drugs harboring varied mechanisms of action. Using this fission yeast model, drug-specific ontological sub-classifications were identified through the computation of relative hypersensitivities. We found that human gastric adenocarcinoma cells can be sensitized to doxorubicin by concomitant treatment with cisplatin, an intra-DNA strand crosslinking agent, and suberoylanilide hydroxamic acid, a histone deacetylase inhibitor. Our findings point to the utility of fission yeast as a model and the differential targeting of a conserved gene interaction network when screening for successful chemotherapeutic drug combinations for human cells.

Cancer is a global health problem and there is a critical need for new advancements in its management and treatment if we are to meet the projected increase in new cases¹. One important challenge facing clinicians is how to determine the most appropriate treatment strategy for patients. Thus far, chemotherapy has remained the mainstay for cancer management². Yet, most of the widely employed anticancer drugs used in the clinic are cytotoxic, harbor a relatively narrow therapeutic window, and/or pose a risk for the development of severe side effects that can potentially jeopardize treatment outcomes³. Despite the toxicity experienced by patients, evidence shows the importance of maintaining a consistent dosage of chemotherapeutic agents within the body over the therapeutic period to kill cancer cells. This high, constant drug insult often induces selection pressure, which encourages the proliferation of certain cells that exhibit increased counter-chemotherapeutic responses, thus leading to chemoresistance⁴.

Drug responsiveness, cytotoxic side effects, and the onset of resistance are linked to the 'genetic makeup' of the patient but this concept itself also remains poorly defined⁵. Recent breakthroughs in genome-wide profiling have uncovered an intimate correlation between molecular signatures, such as gene expression, and patient drug responsiveness, and these findings provide support for the hypothesis that molecular processes are drivers of carcinogenesis that may be employed as discriminating criteria to further fine-tune treatment options².

¹Department of Biochemistry, National University of Singapore, Singapore. ²National University Health System (NUHS), Singapore. ³NUS Synthetic Biology for Clinical and Technological Innovation (SynCTI), Life Sciences Institute, National University of Singapore. ⁴NUS Graduate School for Integrative Sciences and Engineering, National University of Singapore, Singapore. ⁵Department of Pharmacology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore. ⁶Department of Pharmacy, Faculty of Science, National University of Singapore, Singapore. ⁷Department of Mathematics, Faculty of Science, National University of Singapore, Singapore. ⁸Department of Chemistry, Faculty of Science, National University of Singapore, Singapore. ⁹School of Chemical and Life Sciences, Singapore Polytechnic, Singapore. ¹⁰Changi General Hospital, Ministry of Health, Singapore. Correspondence and requests for materials should be addressed to E.S.C. (email: bchces@nus.edu.sg)

Numerous genes bearing significance in carcinogenesis or contributing significantly to various cancer signatures have been described as hallmarks of cancers⁶ and have been deemed as obvious ‘druggable’ targets for chemotherapy⁷. Genome-based profiling has also revealed a large collection of genes, many of which have, as yet, undefined connections to carcinogenesis. These genes do not function alone but, rather, act in synergy with other so-called partner genes to control heritable attributes⁸. Some of these interactions have been characterized through direct protein–protein binding studies but most have been defined through genetic means⁹. Although the functions of individual genes may not be resolved, it is clear that these synergistic relationships among genes form a ‘safety net’ to ensure cell survival, especially following environmental insults by agents such as cytotoxic chemotherapeutic compounds¹⁰. In recent years, considerable attention has been given to these genetic interactions, as they underpin the synthetic lethality approach, which is considered to be able to predict the precise combination of drugs that will improve the efficacy of chemotherapeutics¹¹. Despite the potential utility of this approach, elucidating genetic connectivity in human cells remains a technically and financially challenging task¹².

Clinical chemotherapeutic applications involve drug combinations of typically three to four drugs^{13,14}, and determining the most suitable drug combination profoundly affects chemotherapeutic success. Drugs that possess varied modes of action would be expected to counteract different mechanisms of cellular resistance, and, by that reasoning, should provide efficient or even better therapeutic outcomes when combined rather than applied singly. To this end, we herein describe an approach to predict the efficacy of chemotherapeutic drug combinations that will act to kill human cancer cells in a cooperative manner. This approach is based on an overlapping drug resistance network elucidated using fission yeast (*Schizosaccharomyces pombe*) as a surrogate model. Drugs with a smaller overlap in the resistance gene network are considered to have different mechanisms of action and thus, in view of our reasoning above, would be expected to demonstrate a stronger cooperative effect with the target drug (doxorubicin in our case).

To provide evidence for the utility of such a work flow, we interrogated our previously identified doxorubicin resistance (DXR) genetic network and the intersection of doxorubicin with six other drugs: hydroxyurea (HU), an inhibitor of the deoxyribonucleotide biosynthesis enzyme ribonucleotide reductase (RNR)¹⁵; methyl methanesulfonate (MMS), an alkylating agent that causes nucleotide base damage and impedes the progress of the DNA replication fork¹⁶; camptothecin (CPT), a topoisomerase I inhibitor¹⁷; thiabendazole (TBZ), a microtubule destabilizing agent¹⁸; cisplatin, an intra-DNA strand cross-linker and blocker of transcription^{19,20} and suberoylanilide hydroxamic acid (SAHA), a histone deacetylase inhibitor²¹. With the exception of TBZ, which targets mitotic spindle microtubules, the remaining drugs are reported to induce DNA aberrations that cause DNA damage^{15–23}.

Profiling performed in fission yeast, however, showed that the genes and networks that function to respond to HU, MMS and CPT—initially chosen based on differential mode of action—showed an unexpectedly high degree of overlap. SAHA and cisplatin, on the other hand, exhibited unique gene profiles. These drugs were further shown to act cooperatively with doxorubicin to result in preferential growth retardation in several DXR mutants and interestingly, also in human cancer cells. Thus, the degree of similarity in synthetic lethal drug hypersensitivity profiles and network overlap in fission yeast is useful to narrow down the potential drug candidates and select the best combination(s) that could be subsequently applied to a human cancer cell model.

Results

Study Design Overview. We recently performed a screen for genes that confer doxorubicin resistance (DXR) using fission yeast (*Schizosaccharomyces pombe*) as a model, and found that a network of genes protected cells against drug cytotoxicity^{24,25}. We noted that many cells with mutations in these DXR genes exhibited hypersensitivity to a range of drugs, not specifically doxorubicin^{23,26,27}, and this pointed to a potential overlap in drug resistance mechanisms and/or the existence of a genetic network counteracting multiple cytotoxic agents in fission yeast.

To assess this potential link between the DXR cellular network and that of other drugs, we tested the hypersensitivity of 90 DXR mutant strains with each of the following six drugs: HU, MMS, CPT, TBZ, cisplatin and SAHA. We chose again to use fission yeast cells to test the drug combinations in these 90 genes because of the time and cost efficiencies afforded by this model over human cells.

We serially diluted exponentially growing DXR mutant cells and manually spotted the cultures onto plates incorporated with various concentrations of each of the six drugs: 0, 2, and 4 mM of HU; 0, 6, 8, 10, and 12 μ M of CPT; 0%, 0.001%, 0.002%, 0.005%, 0.01%, and 0.02% of MMS; 0, 8, and 10 μ g/ml of TBZ; 0, 1.2, and 1.6 mM of cisplatin; 0 and 10 mM of SAHA. The following null mutants were employed as positive controls for our drug-hypersensitivity assays: for HU, MMS and CPT, we used the DNA damage checkpoint kinase *rad3* (Δ *rad3*) (Supplementary Fig. 1–3); for TBZ and cisplatin, α -tubulin *nda3* (*nda3-KM311*) (Supplementary Fig. 4, 5); and for SAHA, a type I histone deacetylase complex *alp13* (Δ *alp13*) (Supplementary Fig. 6)^{18,28–30}. In all cases, an isogenic, prototrophic wild-type (WT) strain was used as a negative control and also as an indicator that the drug concentrations used were within the physiological range and did not affect WT cell growth²⁴.

Classification of drug hypersensitivity mutants using sensitivity scores. The growth of the various strains was documented during the intermediate and stationary phases (days 3 and 7 after drug treatment, respectively (Supplementary Fig. 1–6)). Growth was quantified using a sensitivity score (*s*-score), which accounted for the relative fold sensitivity of a specific DXR mutant over the range of the drug concentrations tested as compared with that of untreated cells. Relative changes in fold sensitivity were normalized by comparing the WT strain on plates with drugs over those without. A negative *s*-score depicts drug sensitivity, whereas a positive *s*-score denotes drug resistance. An *s*-score of 0 was obtained when the growth of a particular strain resembled that of WT cells (Supplementary Fig. 7). Cells that showed less than 10-fold sensitivity at only one drug concentration were classified as weakly sensitive mutants (pink, Fig. 1b), whereas cells exhibiting 10-fold or greater sensitivity for all drug concentrations were classified as strongly sensitive (dark red, Fig. 1b); the values in between (from > 10-fold in one concentration up to 10-fold at all concentrations) were deemed to have a medium sensitivity (red, Fig. 1b).

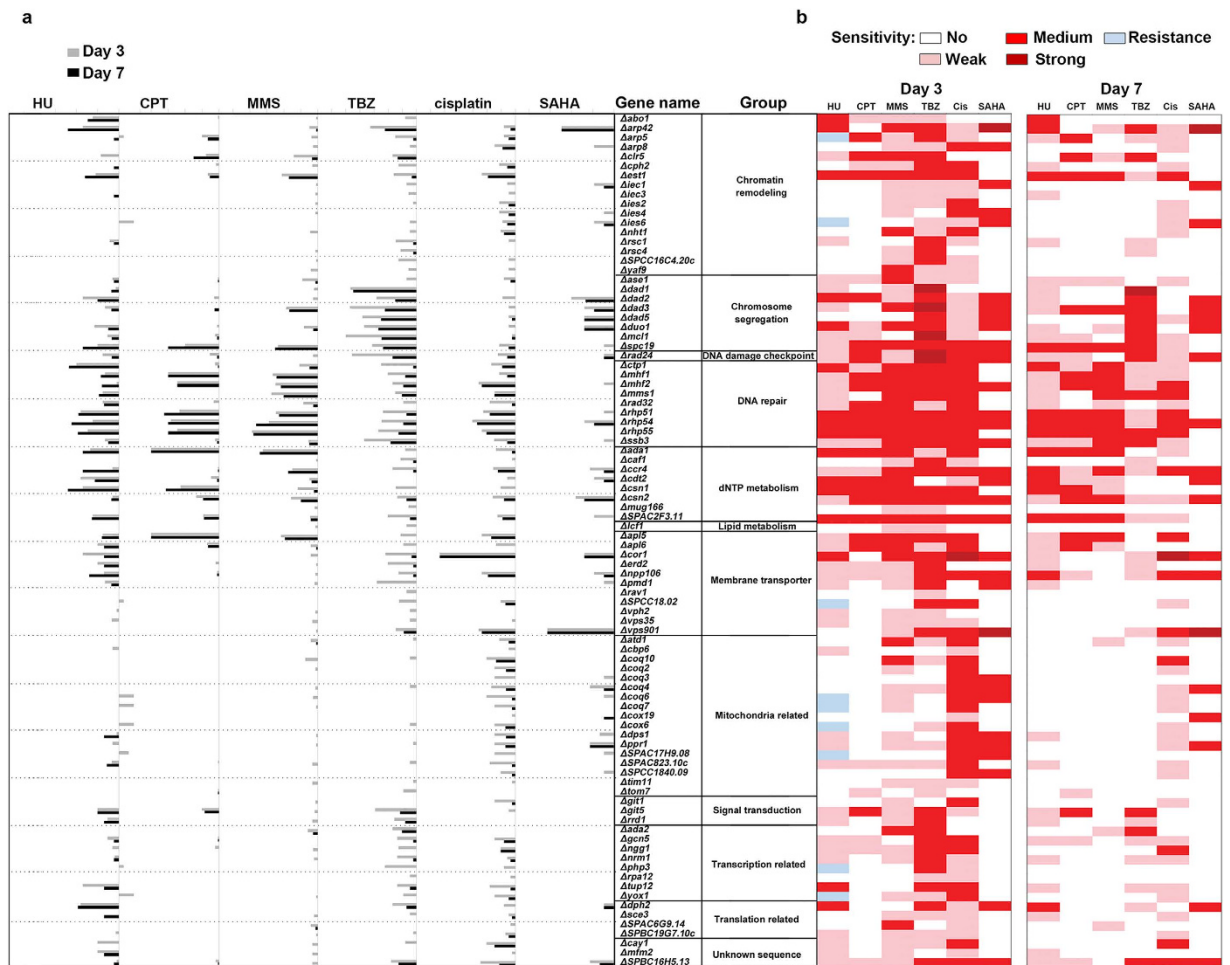


Figure 1. Sensitivity score (s-score) of the doxorubicin resistance (DXR) mutants obtained at different durations of drug exposure. (a) Grey and black bars indicate days 3 and 7, respectively, after drug exposure. (b) Level of hypersensitivity. Dark red, high; red, medium; pink, low; white, not sensitive; and light blue, resistant. DXR genes that were disrupted in the null mutants are listed. HU: hydroxyurea, CPT: camptothecin, MMS: Methyl methanesulfonate, TBZ: thiabendazole, Cis: cisplatin, SAHA: suberoylanilide hydroxamic acid.

Multidrug resistance is controlled by regulators of various DNA-related processes. We noted that almost all DXR mutants showed hypersensitivity to the drugs tested, with very few drug-resistant phenotypes (Fig. 1a). In strains showing drug resistance, this resistance was only transiently observed during the intermediate growth stage (day 3) when the strains grew faster relative to that of WT, reminiscent of a resistance-like growth pattern (Fig. 1a). However, upon further incubation, these ‘transiently resistant’ strains ($\Delta arp5$, $\Delta ies6$, $\Delta SPCC18.02$, $\Delta coq6$, $\Delta coq7$, $\Delta cox6$, $\Delta SPAC17H9.08$, $\Delta php3$, and $\Delta yox1$; Fig. 1) were shown to be hypersensitive, but only to HU.

A previous study used genome-wide synthetic lethality screens to test HU, CPT, and MMS in fission yeast²⁶. Consistent with this previous report, we observed that $\Delta rhp51$, $\Delta rhp54$, $\Delta rhp55$, $\Delta rad32$, $\Delta mhf1$, $\Delta rad24$, and $\Delta cdt2$ showed sensitivity towards HU, MMS and CPT (Fig. 1a)²⁶. Interestingly, these mutants also exhibited sensitivity towards doxorubicin, TBZ and SAHA, indicating that these genes may be involved in a fundamental mechanism to regulate multidrug resistance (Fig. 1b). Ontologically, these seven genes function in homologous recombination (HR) repair, checkpoint signaling and also nucleotide synthesis^{31,32}, suggesting that these mechanisms coordinate resistance against agents that induce chromosomal aberrations (Fig. 1b). The sensitivity towards HU, MMS, and CPT is consistent with the roles of these genes in the management of DNA damage. These mutants also exhibited strong hypersensitivity towards TBZ, a microtubule destabilizing agent that, conceptually, does not affect DNA integrity directly¹⁸. Although SAHA is a histone deacetylase inhibitor (HDACi), it has also been reported to induce DNA double-stranded breaks (DSBs) in humans^{22,33}, and this is probably due to the role of HDAC in chromatin compaction to safeguard cells against unwarranted access by DNA damaging agents²⁹.

A loss of *est1*—a telomerase regulator that maintains the length of telomeric DNA at the chromosomal ends—showed sensitivity to most of the drugs tested, again, with the exception of SAHA (Fig. 1b). This hypersensitivity remained equally high on days 3 and 7, suggesting that the mutants may lose viability upon exposure to the drugs. This phenotype is highly reminiscent of that of DNA damage checkpoint mutants; for example $\Delta rad3$

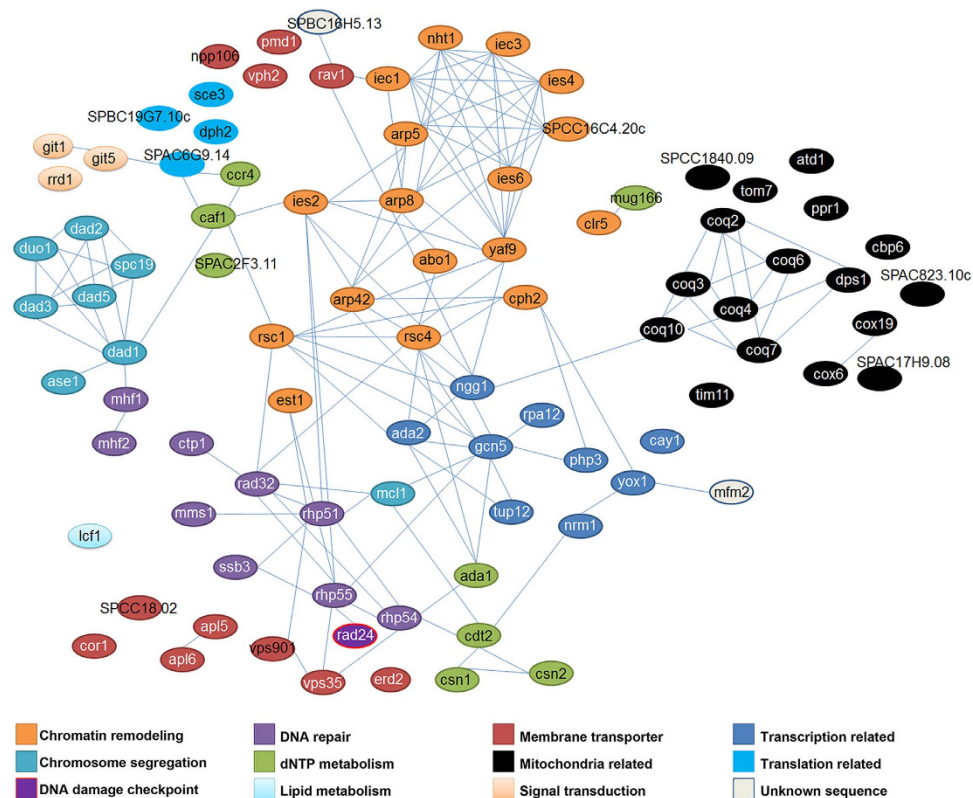


Figure 2. Gene network of doxorubicin resistance (DXR) genes. Linkages between the DXR genes were obtained using String ver. 9.1. The genes are color-coded according to their ontological/functional classification.

on HU plates^{28,30} (Supplementary Fig. 1). Overall, these findings are consistent with the previous hypothesis³⁴ that genomic instability at the telomere underlies cytotoxicity of chemotherapeutic drugs towards human cancer cells.

Overlap between gene sets regulating resistance toward multiple drugs. We next classified DXR genes by the number of drugs they showed sensitivity to at both the intermediate (day 3) and stationary (day 7) growth phases. Interestingly, in the intermediate growth phase, all mutants—aside from those that showed a resistance-like growth to HU (Fig. 1a)—showed sensitivity toward at least one drug (day 3, Fig. 1b), suggesting that DXR genes are part of a centralized network that confers resistance to cytotoxic agents. Furthermore, this differential sensitivity pointed toward the existence of sub-divisions within the centralized network.

We surmised that the genetic overlap seen for this subset of drugs may point to a similar mode of action for the drugs, despite choosing drugs with apparently disparate actions. We noted that at day 7, 13 genes showed no sensitivity toward all of the additional drugs tested aside from doxorubicin, whereas 26 genes remained hypersensitive to only one drug in addition to doxorubicin; this cohort comprised 43.3% (39/90) of the total number of DXR genes examined. It is possible that these genes are involved in a sub-network that responds to doxorubicin. Interestingly, many of the genes in this sub-cluster encode enzymes of the coenzyme Q10 synthesis pathway (*coq2*⁺, *coq4*⁺, *coq6*⁺, *coq7*⁺, *dps1*⁺)³⁵, suggesting that coenzyme Q10 may play an specific role in conferring resistance to doxorubicin.

Doxorubicin resistance gene network. We next investigated connectivity among the DXR genes by assessing published physical and genetic interactions using the online bioinformatics tool, String (version 9.1)³⁶. With the exception of *lcf1*⁺, *apl5*⁺, *apl6*⁺, *erd2*⁺, *sce3*⁺, *dph2*⁺, *npp106*⁺, *vph2*⁺, *pmd1*⁺, *mug166*⁺, *cor1*⁺, *clr5*⁺, *ppr1*⁺, *cay1*⁺, *atd1*⁺ and several uncharacterized coding sequences (*SPCC18.02*, *SPBC19G7.10c*, *SPAC17H9.08* and *SPAC823.10c*), most of the genes isolated in our DXR screen can be assimilated into an extensive network that conceptually depicts, at least in part, a centralized multidrug resistance (MDR) network (Fig. 2). These connections are functionally defined and mostly depict physical interactions as curated by String³⁶. However, they also comprise genetic interactions and catalysis mechanisms involved in similar biochemical pathways, particularly for ubiquinone³⁵.

The DXR gene network consists of several sub-clusters that can be grouped according to ontological pathways: mitochondrial function/coenzyme Q10 biosynthesis; chromatin remodeling, particularly highlighting Ino80 complex components; chromosomal segregation, primarily the DASH complex; membrane-associated transport; nucleotide metabolism; signal transducers (a small cluster); DNA damage response; transcriptional regulation; and protein translation (Fig. 2). We noticed that the DXR network was centrally connected by several ‘hubs’ encompassing the histone acetyltransferase SAGA complex and chromatin remodeling factors (but excluding the Ino80 complex) (Fig. 2). *S*-scores of $\Delta gcn5$ at day 3 indicated that it exhibited high sensitivity towards cisplatin and TBZ but was weakly affected by HU and CPT, and not sensitive to MMS and SAHA (Fig. 1b). By day 7, $\Delta gcn5$

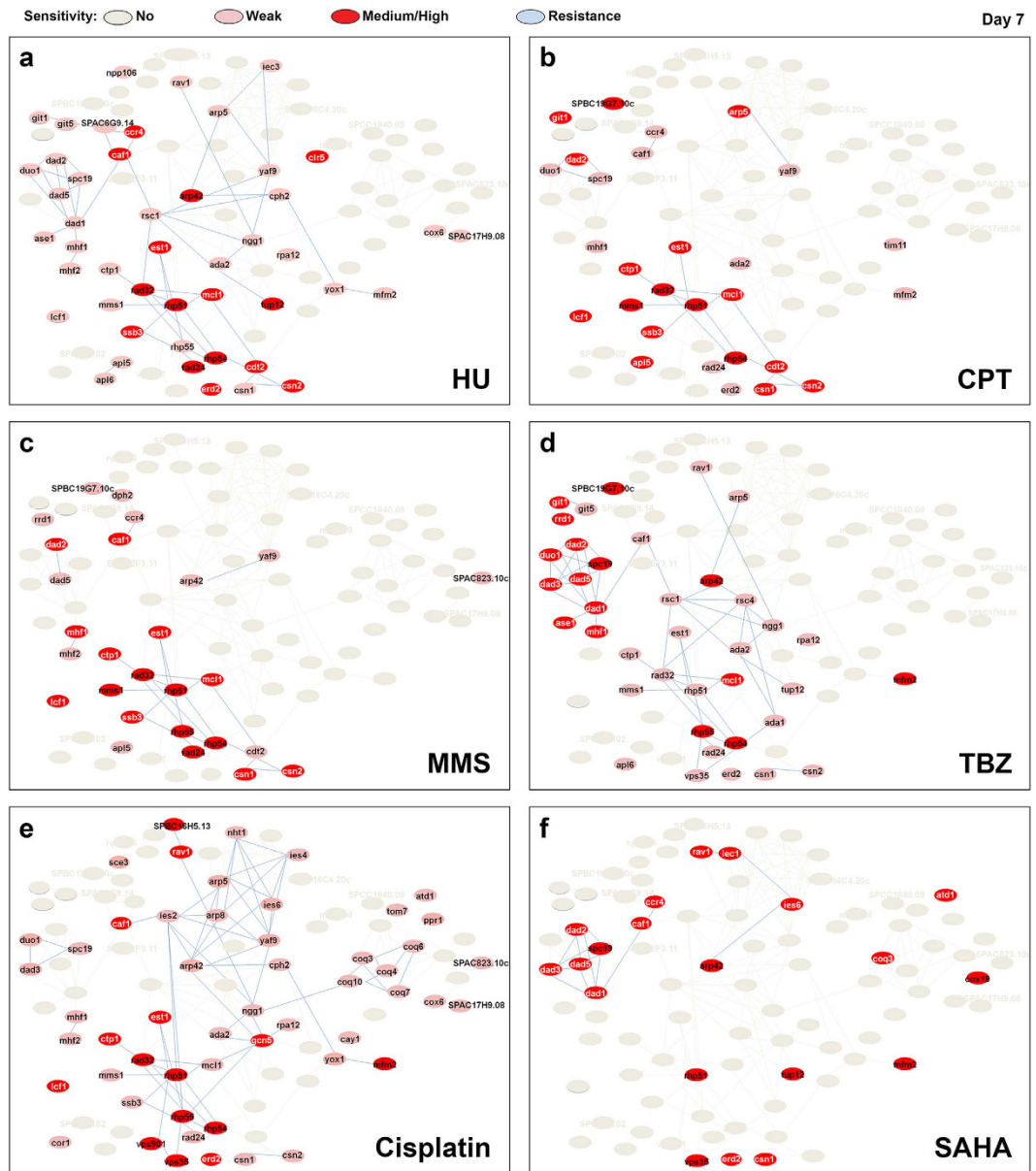


Figure 3. Overlap in drug resistance network between the tested drugs and doxorubicin. The DXR mutants that remained hypersensitivity on day 7 upon exposure to (a) hydroxyurea (HU), (b) camptothecin (CPT), (c) methyl methanesulfonate (MMS), (d) thiabendazole (TBZ), (e) cisplatin, and (f) suberoylanilide hydroxamic acid (SAHA). Strains that showed sensitivity across all drug concentrations tested or only on one of the tested concentrations: red, high to medium sensitivity; pink, weak sensitivity.

cells exhibited a weak sensitivity to the drugs (Fig. 1b). These findings suggest that $\Delta gcn5$ serves a supportive role that involves decondensing the chromatin to facilitate transcribed RNA-templated HR repair³⁷ or to regulate DNA damage checkpoint activation³⁸.

HR DNA repair factors also assumed a focal position, especially Rhp51 (Fig. 2). Unlike $\Delta gcn5$, $\Delta rhp51$ showed strong hypersensitivity towards all six drugs tested on both days 3 and 7 (Fig. 1b, 2b). Rhp51-like protein is required to repair DSBs and enforce DNA damage tolerance during S-phase of the cell cycle³⁹; this likely explains the importance of HR proteins in facilitating resistance to most drugs used in this study. Mutants with compromised HR integrity also showed hypersensitivity to TBZ, which disrupts microtubule assembly; this hypersensitivity is probably based on the connection between the DNA replication checkpoint and the spindle assembly checkpoint⁴⁰.

Sub-clusters of the doxorubicin gene network are also required to respond to other cytotoxic agents. Next, we sought to identify mutants that showed concomitant hypersensitivity to doxorubicin and one or more of the other tested drugs. This was achieved by determining any overlap in the drug-resistance network. Null mutants that were hypersensitive (red: medium-strong; pink: weak sensitivity, Fig. 3a–f) to HU (Fig. 3a), CPT (Fig. 3b), MMS (Fig. 3c), TBZ (Fig. 3d), cisplatin (Fig. 3e), or SAHA (Fig. 3f) were overlaid with the DXR gene

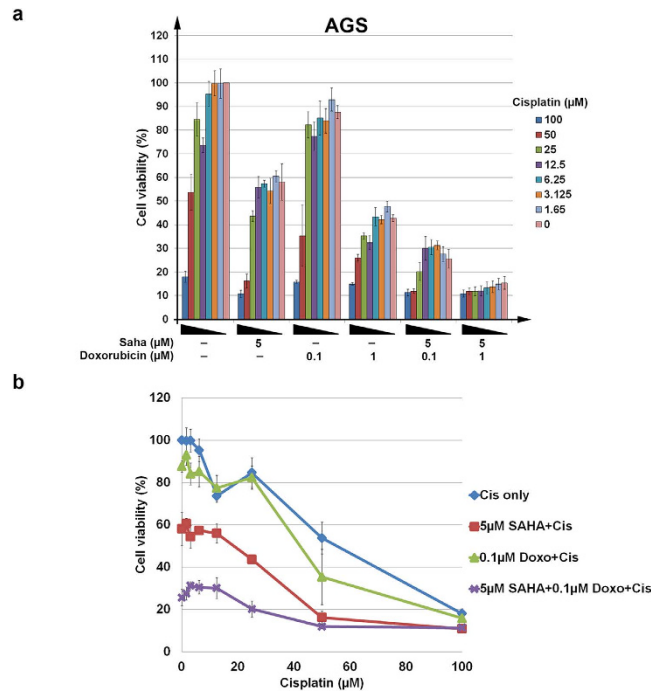


Figure 4. Sensitization of human gastric adenocarcinoma (AGS) cells to doxorubicin via concurrent treatment with cisplatin and SAHA. (a) Cells were co-treated with varying concentrations of cisplatin in the presence of 5 μM SAHA, or 0.1 or 1 μM doxorubicin or with a triple combination of cisplatin, 5 μM SAHA and 0.1 or 1 μM doxorubicin. (b) Dose response effect on the viability of AGS cells was analyzed. Cells were treated with varying concentrations of cisplatin alone (blue), in combination with 5 μM SAHA (red) or 0.1 μM doxorubicin (green), or both 0.1 μM doxorubicin and 5 μM SAHA (purple).

network to highlight a genetic overlap. We found a significant overlap of DXR nodes for HU, CPT and MMS—each of which disrupt DNA replication ($p < 0.0001$)^{15–17,41} (Fig. 3a–c). This overlap lends further support to the theory that DXR genes belong to a centralized MDR network⁴². We noted that cisplatin also shared considerable overlap with the HU/CPT/MMS cluster; albeit, with more variation (Fig. 3f).

Cooperative effect among doxorubicin, SAHA and cisplatin in fission yeast. Given the significant gene overlap noted among several of the drugs, we further interrogated the MDR drug response network to predict an efficient drug combination that may be used to sensitize human cancer cells toward doxorubicin. We first spotted serially diluted WT fission yeast cells on media incorporated with increasing concentrations of doxorubicin and sub-lethal doses of MMS (0.01%), TBZ (8 $\mu\text{g}/\text{ml}$), HU (2 mM), CPT (8 μM), cisplatin (1.2 mM), or SAHA (5 mM) as a control (Supplementary Fig. 8). Overall, WT cells were not affected by the drug treatments. We observed a slight growth reduction with doxorubicin, CPT and SAHA on day 3 (Supplementary Fig. 8a, day 3) but cells were generally unaffected, as growth on day 7 was similar between treated and untreated cells (Supplementary Fig. 8b, day 7). No change was observed for cells treated with HU, CPT, MMS or TBZ.

We further assessed the synergism noted for the combination of doxorubicin, SAHA and cisplatin by repeating the serial dilution and spotting assays using several DXR null mutants: $\Delta rav1$, $\Delta rhp51$, $\Delta vps35$, $\Delta caf1$ and $\Delta tim11$. These genes encode for a vacuolar ATPase assembly factor ($\Delta rav1$), a HR factor ($\Delta rhp51$), a subunit of endosome sorting ($\Delta vps35$), a protein of the CCR/NOT deadenylase complex ($\Delta caf1$), and a mitochondrial ATPase complex protein ($\Delta tim11$)^{32,43–45}. In human cancers, these proteins have been linked with drug resistance and/or cancer cell proliferation^{25,46–49}. We found that $\Delta rhp51$ cells showed exceedingly high sensitivity to doxorubicin as well as to that of other drugs, and therefore any synergistic effect was likely masked. The $\Delta rav1$ null mutant, in comparison, showed particular hypersensitivity to cisplatin, again preventing the identification of any cumulative drug effects. Interestingly, the remaining three mutants ($\Delta vps35$, $\Delta caf1$ and $\Delta tim11$) showed prominent attenuation of a growth defect when the cells were treated with the triple combination of doxorubicin, cisplatin and SAHA, thus demonstrating a synergistic effect for the drugs and a role for these genes (Supplementary Fig. 9).

Sensitization of human cancer cells to doxorubicin by SAHA and cisplatin. Next we tested whether the cooperative effect of doxorubicin, cisplatin and SAHA could be recapitulated in human cancer cell models and thus confirm whether fission yeast serves as a suitable model for the screening of chemotherapeutic drug combinations. Others have reported in the past that mutations in genes of the vacuolar sorting pathway are connected to gastric cancer occurrence⁴⁶, so we tested our drug combination first on gastric adenocarcinoma (AGS) cells. Indeed, gastric cancers are treated clinically with a chemotherapeutic regime containing doxorubicin and cisplatin⁵⁰.

AGS cells were treated in single, double or triple combinations of cisplatin (0 μ M to 100 μ M), SAHA (5 μ M), and doxorubicin (0.1 or 1 μ M) (Fig. 4). Cisplatin alone induced a concentration-dependent cytotoxicity in AGS cells and showed an additive effect with doxorubicin, particularly at higher concentrations of doxorubicin (1 μ M); lower doxorubicin concentrations (0.1 μ M) had little to no effect (Fig. 4, beige bars). SAHA treatment alone reduced AGS cell viability, and this was further decreased with the addition of doxorubicin. The triple combination killed the cells and affected the entire dose-response curve (Fig. 4b). This synergism among the three drugs was particularly apparent at 25–50 μ M cisplatin, 0.1 μ M doxorubicin and 5 μ M SAHA (Fig. 4b). Thus, as with fission yeast cells, the three drugs worked cooperatively to kill gastric adenocarcinoma cells (Fig. 4). We also observed a similar cooperative effect in human cervical carcinoma (HeLa) cells (Supplementary Fig. 10a,b) but not in the non-cancerous embryonic kidney HEK293 cells (Supplementary Fig. 11a,b). This observation is reminiscent of the lack of effect of the three-drug combination on WT fission yeast cells (Supplementary Fig. 8). Taken together, our results show that fission yeast can be used as a surrogate model to derive effective chemotherapeutic combinations to target human cancer cells.

Discussion

In this report, we show that large-scale genetic synthetic lethality screening in fission yeast can be used to increase the reliability of predicting the synergistic effect of multiple drugs when targeting human cancer cells. This study provides a proof-of-concept that three drugs—doxorubicin, cisplatin and SAHA, which show a low degree of overlap among gene networks in fission yeast cells—can synergistically work to affect cell growth.

With the completion of sequencing for many organisms, especially that of yeast, the ability to interrogate the functional implications of individual genes to a phenotype has revolutionized genome-based studies^{25,51}. Large-scale genetic interaction assays have made it possible to model the lives of eukaryotic organisms in order to identify the complicated connectivity among genes and study how this interactivity constitutes a network that determines the phenotype^{52,53}; in this case, chemotherapeutic resistance. The study of genetic interactions has gained momentum, and the revelation of the ‘interactome’⁵⁴ has shed light on how multiple components can be regulated concurrently. It has also (fortunately) revealed that there are rules that govern how the structure and topology of genetic networks operate, and studies show that genes within the network often form a relatively restricted set of positive and negative interactions¹². Cross-species studies have also identified conservation of a fundamental network that is universal in higher eukaryotes, including humans, and it has been determined that this network shares, unexpectedly, great overlap with that of single cellular eukaryotes, such as yeast^{25,55}. Given that toxin responses are fundamental to all cells, it is likely that this forms the basis for the conserved mechanism of drug resistance observed between fission yeast and human cells. Thus, it is plausible to take advantage of the commonality amongst eukaryotic cells to understand the molecular mechanism of resistance to chemotherapeutic agents.

The results presented here support our expectation that large-scale synthetic lethality network elucidation in unicellular eukaryotes (such as fission yeast) would be a suitable model with which to predict a pharmacological regimen for sensitizing human cancer cells to chemotherapeutic agents. Gene knockout screening in human cell models is typically costly and time consuming. By comparison, our synthetic lethality workflow with fission yeast demonstrates the power and possibility to improve chemotherapeutic combinatorial therapeutics. The positive outcomes arising from this study suggest that lower eukaryotic systems, such as the fission yeast, will offer a cheaper and faster alternative to screen for drug combinations that are more likely to offer a better therapy. Fission yeast is also highly amendable to genetic manipulation, and thus useful for scaling up assay sizes to test many drugs and genes simultaneously.

In this work, we employed a serial dilution spotting assay, a commonly used method, which can be varied by streaking log-phase cells or by measuring changes in cell density^{23,30,56–58}. Unlike previous studies, we have uniquely incorporated a computation of dose response, which, in our view, increases the accuracy of the assay. Furthermore, studying dose-response changes also helps to emphasize the pharmacological behavior of each compound over a range concentrations. However, the accuracy of cell growth can be further improved and incorporated into future experiments, such as the use of flow cytometric measurements of fluorescently labeled strains⁵⁹ and the measurement of yeast colony sizes in conjunction with image-analysis software⁶⁰.

One paradigm highlighted in this work is that cells possess a centralized, responsive network to counteract cytotoxic agents; this finding is consistent with a large-scale chemogenomic fitness screen in budding yeast, which uncovered a 45-gene cluster that coordinated the response of cells to small pharmacological molecules⁴². Mutants of the DXR genes showed differential sensitivity towards multiple DNA damaging agents, indicating an unequal contribution of each DXR gene toward this hypothesized centralized multidrug resistance (MDR) network; this unequal contribution may be partially explained by the differences in connectivity among partners in the network^{61,62} or specificity for the central target of the drugs, such as that demonstrated for topoisomerase II in response to doxorubicin⁶³.

The clustering of drug hypersensitivity exhibited by the DXR genes suggests a hierarchical architecture in which smaller networks are integrated into progressively more extensive networks that are directed against an increasingly larger number of drugs. This model depicts that a targeted destabilization of widely separated sub-networks would result in stronger synergistic effects as compared to a disruption of closely related ones. Hence, a drug combination that can destabilize a more widely separated nexus within the MDR network would be expected to triumph as an effective combination. An understanding of the unique network architecture specific for cancer cells would help shed light on how drug resistance is coordinated in cancer cells, and this knowledge would facilitate the targeted killing of cancer cells over that of normal cells.

We recently showed that two DXR mutants, $\Delta mcl1$ and $\Delta mhf1$, disrupt chromatin integrity at the fission yeast centromere, and that this activity could be suppressed by a point mutation in topoisomerase II (Top2) (*top2-191*)⁶³. The $\Delta mcl1$ mutant showed sensitivity to four of the six (not MMS and SAHA) drugs tested, whereas $\Delta mhf2$ was sensitive to all but SAHA (Fig. 1b). This hypersensitivity toward multiple cytotoxic agents suggests that disruption

to centromeric integrity may be a more widespread mechanism underlying susceptibility to cytotoxic chemotherapeutic agents besides doxorubicin. The disruption of centromeric integrity may also underlie the rampant hypersensitivity of mutants in DASH complex components to a DNA damaging agent that does not directly undermine microtubule stability⁶⁴ (Figs 1b, 3).

Pmd1 is the fission yeast homolog of the human ATP-binding cassette (ABC) transporter permeability glycoprotein (P-gp/MDR1/ABCB1)²⁵ and we have previously shown that Pmd1 acts synergistically with the vacuolar-ATPase pathway to govern doxorubicin hypersensitivity in human cervical carcinoma cells, probably via modulation of the intracellular accumulation of doxorubicin²⁵. In humans, P-gp has been implicated in resistance against multiple drugs⁴ and it is surprising that $\Delta pmd1$ was among the mutants that showed only limited sensitivity towards the drugs tested²⁵. This limited sensitivity may be due to peculiar substrate specificities or functional differences between the fission yeast Pmd1 and human P-gp. However, it is also possible that P-gp in human cells does not work alone but in conjunction with other membrane-associated transporters, resembling the functional cooperation between Pmd1/P-gp and V-ATPase in counteracting doxorubicin cytotoxicity²⁵. In this context, it is possible that a network of different transporters may be involved in controlling drug resistance by regulating the levels of drug accumulation, which may be further modulated by the metabolism of these drugs within the cells.

A potential complication in using a cell-based assay to test drug cooperativity is the difference in bioavailability of the drugs due to the differences in transporter profiles among different cell types, which has been reported with different human cells and between *in vitro* cell culture models and *in vivo* animal models^{65,66}. The results from cell-based models must therefore be interpreted with caution. Based on a similar extrapolation, the transporter expression profiles would also be different between the yeast systems and human cells. Hence additional experiments using whole animal testing and/or xenografts would be needed to validate the results from cell-based models. In this aspect, despite the perceived difference between yeast and human cultured cells, our previous results still suggest the value of the yeast system in elucidating plausible, cooperatively acting drug combinations²⁵. Furthermore, screening yeast cells offers advantages of speed and cost efficiency over that in human cells, and provides the opportunity to target fewer drugs in a more specific manner.

Doxorubicin is widely employed in chemotherapeutic drug combinations along with cisplatin, for example, in the cells of urothelial cancers or with cisplatin, vinblastine and methotrexate in gastric cancer^{51,65}. Here, we discovered the HDACi SAHA as a new partnering drug for the doxorubicin–cisplatin combination in gastric adenocarcinoma and cervical carcinoma cells. There is a recent resurgence in the utility of HDACis for drug sensitization. Initial studies emphasized the use of HDACis in the pretreatment of cancer cells to increase the cytotoxicity of the drugs, presumably by decondensing the chromatin for improved accessibility of other cytotoxic DNA-adduct-causing drugs^{66,67}. However, here we show that SAHA exerts a similar effect to elicit an increase in the cytotoxicity of doxorubicin and further accentuates the effect of cisplatin. The cooperative effect of the three drugs was applied at a level where each separately did not show much effect (Fig. 4).

While we were preparing this manuscript, a phase I/II clinical trial was reported, demonstrating that the addition of belinostat, a HDACi, can improve a drug regimen containing cisplatin and doxorubicin (also with cyclophosphamide) in thymic epithelial tumors⁶⁸. This report increases our confidence that drug combinations that target resistance can be determined from yeast synthetic lethal network analyses and provides evidence for the utility of SAHA in the triple combination. Future large-scale profiling of drugs and genetic interactions are expected to revolutionize the application of drug combination chemotherapy.

In conclusion, our work provides a novel approach for determining potential chemotherapeutic drug combinations for the sensitization of human cancer cells. Interfacing yeast synthetic lethality and human studies may provide a cheap and more targeted workflow with which to address drug resistance.

Methods

Drugs. Methyl methanesulfonate (MMS) and thiabendazole (TBZ) were purchased from Sigma-Aldrich (St Louis, MO). Camptothecin (CPT), hydroxyurea (HU), doxorubicin and cisplatin were from Wako Pure Chemical Industries Ltd (Osaka, Japan). Vorinostat/suberoylanilide hydroxamic acid (SAHA) was synthesized in-house (see Supplementary Materials)⁶⁹. Doxorubicin was dissolved in water, SAHA (refer supplementary procedure) in DMSO, and cisplatin (Wako, Pure Chemical Industries, Ltd, Osaka, Japan) in 10% NaCl. All drugs were further diluted with their respective solvents prior to use according to manufacturers' recommendations.

Fission yeast techniques. A standard procedure for the treatment of fission yeast was followed⁷⁰. Fission yeast strains were serially diluted and then spotted onto drug plates, as previously reported^{24,25}. Cell growth was analyzed at two time points, at days 3 and 7, after spotting on the agar media. We chose these two time points in order to better estimate the effects of the drugs on cell growth as compared with cell viability. Specifically, during the growing phase and before maximum growth is attained (on day 3), cells will exhibit growth retardation if both their growth and viability are affected. However, by day 7 and when the strains have reached maximum growth, only cells that have lost viability in response to the drug will exhibit growth defects. Cells that show reduced growth but no loss in viability will show growth that is near or equivalent to that of WT cells. All DXR strains were obtained in a previous screen that tested for the hypersensitivity of fission yeast single-gene knockout strains from commercial libraries (versions 1.0 and 2.0) (Bioneer, Daejeon, Korea) and subsequently converted into the prototrophic genetic background by backcrossing with prototrophic WT strains.

Calculation of sensitivity score (s-score). The step-wise derivation of the s-score is described in Supplementary Fig. 7. Growth fitness of the strains was obtained by comparing the growth of strains on drug-treated plates relative to that on non-drug-treated plates. The fitness values obtained for mutants were normalized against

that of the WT cells before a logarithmic transformation was performed. The *s*-score was obtained from the mean of the sum of the relative fitness measurement resultant values obtained.

Interaction network construction tool. The online network construction tool, String version 9.1³⁶, was employed to discover interaction links among the previously identified DXR genes.

Human cell culture and drug treatment. Human gastric adenocarcinoma (AGS) cells (ATCC, Manassas, VA) were grown in RPMI media (Sigma-Aldrich) supplemented with 10% fetal bovine serum (FBS) (Life Technologies), whereas human cervical carcinoma (HeLa) (ATCC) and human embryonic kidney cells (HEK293) (ATCC) were maintained in EMEM media (Sigma-Aldrich) supplemented with 10% FBS. Drug concentrations were determined by dose-response assay, where cells were seeded into 96-well culture plates and treated in triplicate with various drug concentrations of doxorubicin: (0, 0.001, 0.01, 0.1, 1, 5, 10, 25, 100 μ M) and SAHA (0, 0.39, 0.78, 1.56, 3.13, 6.25, 12.5, 25, 50 μ M). The doxorubicin and SAHA concentrations that elicited approximately 30% cell death were selected for cytotoxicity tests, in which the cells were then exposed to varying concentrations of cisplatin (0, 1.65, 3.125, 6.25, 12.5, 25, 50, 100 μ M; Fig. 4). Cell viability was determined with cck-8 assay, according to the manufacturer's protocol (Dojindo, Kumamoto, Japan). Triplicates of each treatment were performed over five separate sets of experiments.

Statistical analyses. Dose-response curves of the following treatments were plotted with standard deviations: varying cisplatin only; 5 μ M SAHA with varying cisplatin; 5 μ M doxorubicin with varying cisplatin; and the triple treatment of 5 μ M SAHA with 5 μ M doxorubicin in varying cisplatin. Cell viability at each concentration of cisplatin was compared against the 0 μ M cisplatin for the respective treatment group. The statistical significance of the synergistic effects was examined with a two-tailed Student's *t*-test and performed using Microsoft Excel (2007). A simulation was performed to ascertain similarity between CPT, HU and MMS, as previously reported⁴¹. For each pair of drugs *x* and *y*, two sets (S_x and S_y) each of 10,000 random profiles were generated. The significance of the similarity $s(x, y)$ between *x* and *y* was assessed by the comparison of $s(x, y)$ with the correlation between two random profiles taken respectively from S_x and S_y .

References

- Bray, F., Jemal, A., Grey, N., Ferlay, J. & Forman, D. Global cancer transitions according to the Human Development Index (2008–2030): a population-based study. *Lancet Oncol.* **13**, 790–801, doi: 10.1016/S1470-2045(12)70211-5 (2012).
- Munoz, J., Swanton, C. & Kurzrock, R. Molecular profiling and the reclassification of cancer: divide and conquer. *Am. Soc. Clin. Oncol. Educ. Book.* 127–134, doi: 10.1200/EdBook_AM.2013.33.127 (2013).
- Fujita, K. & Sasaki, Y. Optimization of cancer chemotherapy on the basis of pharmacokinetics and pharmacodynamics: from patients enrolled in clinical trials to those in the 'real world'. *Drug Metab. Pharmacokinet.* **29**, 20–28 (2014).
- Szakacs, G., Paterson, J. K., Ludwig, J. A., Booth-Genthe, C. & Gottesman, M. M. Targeting multidrug resistance in cancer. *Nat. Rev. Drug Discov.* **5**, 219–234, doi: 10.1038/nrd1984 (2006).
- Wiehche, E. & Hansen, L. L. The effect of genetic variability on drug response in conventional breast cancer treatment. *Eur. J. Pharmacol.* **625**, 122–130, doi: 10.1016/j.ejphar.2009.08.045 (2009).
- Hanahan, D. & Weinberg, R. A. Hallmarks of cancer: the next generation. *Cell* **144**, 646–674, doi: 10.1016/j.cell.2011.02.013 (2011).
- Bailon-Moscoso, N., Romero-Benavides, J. C. & Ostrosky-Wegman, P. Development of anticancer drugs based on the hallmarks of tumor cells. *Tumour Biol.* **35**, 3981–3995, doi: 10.1007/s13277-014-1649-y (2014).
- Zuk, O., Hechter, E., Sunyaev, S. R. & Lander, E. S. The mystery of missing heritability: Genetic interactions create phantom heritability. *Proc. Natl. Acad. Sci. USA* **109**, 1193–1198, doi: 10.1073/pnas.1119675109 (2012).
- Ghera, D. & Singh, M. Disentangling function from topology to infer the network properties of disease genes. *BMC Syst. Biol.* **7**, 5, doi: 10.1186/1752-0509-7-5 (2013).
- Vizeacoumar, F. J. *et al.* A negative genetic interaction map in isogenic cancer cell lines reveals cancer cell vulnerabilities. *Mol. Syst. Biol.* **9**, 696, doi: 10.1038/msb.2013.54 (2013).
- Sajesh, B. V., Guppy, B. J. & McManus, K. J. Synthetic genetic targeting of genome instability in cancer. *Cancers (Basel)* **5**, 739–761, doi: 10.3390/cancers5030739 (2013).
- Boone, C. Yeast systems biology: our best shot at modeling a cell. *Genetics* **198**, 435–437, doi: 10.1534/genetics.114.169128 (2014).
- Greenhalf, W. & Thomas, A. Combination therapy for the treatment of pancreatic cancer. *Anticancer Agents Med. Chem.* **11**, 418–426 (2011).
- Watanabe, K. Current chemotherapeutic approaches for hepatoblastoma. *Int. J. Clin. Oncol.* **18**, 955–961, doi: 10.1007/s10147-013-0616-8 (2013).
- Saban, N. & Bujak, M. Hydroxyurea and hydroxamic acid derivatives as antitumor drugs. *Cancer Chemother. Pharmacol.* **64**, 213–221, doi: 10.1007/s00280-009-0991-z (2009).
- Wyatt, M. D. & Pittman, D. L. Methylating agents and DNA repair responses: Methylated bases and sources of strand breaks. *Chem. Res. Toxicol.* **19**, 1580–1594, doi: 10.1021/tx060164e (2006).
- Beretta, G. L., Gatti, L., Perego, P. & Zaffaroni, N. Camptothecin resistance in cancer: insights into the molecular mechanisms of a DNA-damaging drug. *Curr. Med. Chem.* **20**, 1541–1565 (2013).
- Hiraoka, Y., Toda, T. & Yanagida, M. The NDA3 gene of fission yeast encodes beta-tubulin: a cold-sensitive *nda3* mutation reversibly blocks spindle formation and chromosome movement in mitosis. *Cell* **39**, 349–358 (1984).
- Malinge, J. M., Giraud-Panis, M. J. & Leng, M. Interstrand cross-links of cisplatin induce striking distortions in DNA. *J. Inorg. Biochem.* **77**, 23–29 (1999).
- Zhu, G., Myint, M., Ang, W. H., Song, L. & Lippard, S. J. Monofunctional platinum-DNA adducts are strong inhibitors of transcription and substrates for nucleotide excision repair in live mammalian cells. *Cancer Res.* **72**, 790–800, doi: 10.1158/0008-5472.CAN-11-3151 (2012).
- Marks, P. A. Discovery and development of SAHA as an anticancer agent. *Oncogene* **26**, 1351–1356, doi: 10.1038/sj.onc.1210204 (2007).
- Lee, J. H., Choy, M. L., Ngo, L., Foster, S. S. & Marks, P. A. Histone deacetylase inhibitor induces DNA damage, which normal but not transformed cells can repair. *Proc. Natl. Acad. Sci. USA* **107**, 14639–14644, doi: 10.1073/pnas.1008522107 (2010).
- Mojardin, L., Botet, J., Moreno, S. & Salas, M. Chromosome segregation and organization are targets of 5'-Fluorouracil in eukaryotic cells. *Cell Cycle* **14**, 206–218, doi: 10.4161/15384101.2014.974425 (2015).
- Tay, Z. *et al.* Cellular robustness conferred by genetic crosstalk underlies resistance against chemotherapeutic drug doxorubicin in fission yeast. *PLoS One* **8**, e55041, doi: 10.1371/journal.pone.0055041 (2013).

25. Tay, Z. *et al.* P-glycoprotein and vacuolar ATPase synergistically confer anthracycline resistance to fission yeast and human cells. *Curr. Med. Chem.* **21**, 251–260 (2014).
26. Deshpande, G. P. *et al.* Screening a genome-wide *S. pombe* deletion library identifies novel genes and pathways involved in genome stability maintenance. *DNA Repair (Amst)* **8**, 672–679, doi: 10.1016/j.dnarep.2009.01.016 (2009).
27. Han, T. X., Xu, X. Y., Zhang, M. J., Peng, X. & Du, L. L. Global fitness profiling of fission yeast deletion strains by barcode sequencing. *Genome Biol.* **11**, R60, doi: 10.1186/gb-2010-11-6-r60 (2010).
28. Chu, Z. *et al.* Modulation of cell cycle-specific gene expressions at the onset of S phase arrest contributes to the robust DNA replication checkpoint response in fission yeast. *Mol. Biol. Cell* **18**, 1756–1767, doi: 10.1091/mbc.E06-10-0928 (2007).
29. Nicolas, E. *et al.* Distinct roles of HDAC complexes in promoter silencing, antisense suppression and DNA damage protection. *Nat. Struct. Mol. Biol.* **14**, 372–380, doi: 10.1038/nsmb1239 (2007).
30. Zhao, H., Tanaka, K., Nogochi, E., Nogochi, C. & Russell, P. Replication checkpoint protein Mrc1 is regulated by Rad3 and Tel1 in fission yeast. *Mol. Cell. Biol.* **23**, 8395–8403 (2003).
31. Blaikley, E. J. *et al.* The DNA damage checkpoint pathway promotes extensive resection and nucleotide synthesis to facilitate homologous recombination repair and genome stability in fission yeast. *Nucleic Acids Res.* **42**, 5644–5656, doi: 10.1093/nar/gku190 (2014).
32. Takahashi, S., Kontani, K., Araki, Y. & Katada, T. Caf1 regulates translocation of ribonucleotide reductase by releasing nucleoplasmic Spd1–Suc22 assembly. *Nucleic Acids Res.* **35**, 1187–1197, doi: 10.1093/nar/gkm015 (2007).
33. Singh, S., Le, H., Shih, S. J., Ho, B. & Vaughan, A. T. Suberoylanilide hydroxyamic acid modification of chromatin architecture affects DNA break formation and repair. *Int. J. Radiat. Oncol. Biol. Phys.* **76**, 566–573, doi: 10.1016/j.ijrobp.2009.08.031 (2010).
34. Lu, Y., Leong, W., Guerin, O., Gilson, E. & Ye, J. Telomeric impact of conventional chemotherapy. *Front. Med.* **7**, 411–417, doi: 10.1007/s11684-013-0293-z (2013).
35. Kawamukai, M. Biosynthesis and bioproduction of coenzyme Q10 by yeasts and other organisms. *Biotechnol. Appl. Biochem.* **53**, 217–226, doi: 10.1042/BA20090035 (2009).
36. Franceschini, A. *et al.* STRING v9.1: protein-protein interaction networks, with increased coverage and integration. *Nucleic Acids Res.* **41**, D808–815, doi: 10.1093/nar/gks1094 (2013).
37. Keskin, H. *et al.* Transcript-RNA-templated DNA recombination and repair. *Nature* **515**, 436–439, doi: 10.1038/nature13682 (2014).
38. Wang, Y. *et al.* Histone H3 lysine 14 acetylation is required for activation of a DNA damage checkpoint in fission yeast. *J. Biol. Chem.* **287**, 4386–4393, doi: 10.1074/jbc.M111.329417 (2012).
39. Carr, A. M. & Lambert, S. Replication stress-induced genome instability: the dark side of replication maintenance by homologous recombination. *J. Mol. Biol.* **425**, 4733–4744, doi: 10.1016/j.jmb.2013.04.023 (2013).
40. Sugimoto, I., Murakami, H., Tonami, Y., Moriyama, A. & Nakanishi, M. DNA replication checkpoint control mediated by the spindle checkpoint protein Mad2p in fission yeast. *J. Biol. Chem.* **279**, 47372–47378, doi: 10.1074/jbc.M403231200 (2004).
41. Nguyen, C. T., Tay, Y. C. & Zhang L. Divide-and-conquer approach for the exemplar breakpoint distance. *Bioinformatics* **21**, 2171–2176, doi: 10.1093/bioinformatics/bti327 (2005).
42. Lee, A. Y. *et al.* Mapping the cellular response to small molecules using chemogenomic fitness signatures. *Science* **344**, 208–211, doi: 10.1126/science.1250217 (2014).
43. Arnold, I., Bauer, M. F., Brunner, M., Neupert, W. & Stuart, R. A. Yeast mitochondrial F1F0-ATPase: the novel subunit e is identical to Tim11. *FEBS Lett.* **411**, 195–200 (1997).
44. Iwaki, T. *et al.* Vacuolar protein sorting receptor in *Schizosaccharomyces pombe*. *Microbiology* **152**, 1523–1532, doi: 10.1099/mic.0.28627-0 (2006).
45. Khasanov, F. K. *et al.* A new recombinational DNA repair gene from *Schizosaccharomyces pombe* with homology to *Escherichia coli* RecA. *Genetics* **152**, 1557–1572 (1999).
46. An, C. H. *et al.* Frameshift mutations of vacuolar protein sorting genes in gastric and colorectal cancers with microsatellite instability. *Hum. Pathol.* **43**, 40–47, doi: 10.1016/j.humpath.2010.03.015 (2012).
47. Aslam, A., Mittal, S., Koch, F., Andrau, J. C. & Winkler, G. S. The Ccr4-NOT deadenylase subunits CNOT7 and CNOT8 have overlapping roles and modulate cell proliferation. *Mol. Biol. Cell* **20**, 3840–3850, doi: 10.1091/mbc.E09-02-0146 (2009).
48. Spencer, D. M. *et al.* DNA repair in response to anthracycline-DNA adducts: a role for both homologous recombination and nucleotide excision repair. *Mutat. Res.* **638**, 110–121, doi: 10.1016/j.mrfmmm.2007.09.005 (2008).
49. Ying, H., Yu, Y. & Xu, Y. Antisense of ATP synthase subunit e inhibits the growth of human hepatocellular carcinoma cells. *Oncol. Res.* **12**, 485–490 (2001).
50. Chang, T. L. *et al.* Claudin-1 has tumor suppressive activity and is a direct target of RUNX3 in gastric epithelial cells. *Gastroenterology* **138**, 255–265 e251–253, doi: 10.1053/j.gastro.2009.08.044 (2010).
51. Necchi, A. *et al.* Long-term efficacy and safety outcomes of modified (simplified) MVAC (methotrexate/vinblastine/doxorubicin/cisplatin) as frontline therapy for unresectable or metastatic urothelial cancer. *Clin. Genitourin. Cancer* **12**, 203–209 e201, doi: 10.1016/j.jclgc.2013.11.022 (2014).
52. Baryshnikova, A., Costanzo, M., Myers, C. L., Andrews, B. & Boone, C. Genetic interaction networks: toward an understanding of heritability. *Annu Rev Genomics Hum Genet* **14**, 111–133, doi: 10.1146/annurev-genom-082509-141730 (2013).
53. Braberg, H., Moehle, E. A., Shales, M., Guthrie, C. & Krogan, N. J. Genetic interaction analysis of point mutations enables interrogation of gene function at a residue-level resolution: exploring the applications of high-resolution genetic interaction mapping of point mutations. *Bioessays* **36**, 706–713, doi: 10.1002/bies.201400044 (2014).
54. Schwartz, A. S., Yu, J., Gardenour, K. R., Finley, R. L., Jr. & Ideker, T. Cost-effective strategies for completing the interactome. *Nat. Methods* **6**, 55–61, doi: 10.1038/nmeth.1283 (2009).
55. van Pel, D. M., Stirling, P. C., Minaker, S. W., Sipahimalani, P. & Hieter, P. *Saccharomyces cerevisiae* genetics predicts candidate therapeutic genetic interactions at the mammalian replication fork. *G3 (Bethesda)* **3**, 273–282, doi: 10.1534/g3.112.004754 (2013).
56. Alao, J. P., Weber, A. M., Shabro, A. & Sunnerhagen, P. Suppression of sensitivity to drugs and antibiotics by high external cation concentrations in fission yeast. *PLoS One* **10**, e0119297, doi: 10.1371/journal.pone.0119297 (2015).
57. Ohtsuka, H. *et al.* Screening for long-lived genes identifies Oga1, a guanine-quadruplex associated protein that affects the chronological lifespan of the fission yeast *Schizosaccharomyces pombe*. *Mol. Genet. Genomics* **288**, 285–295, doi: 10.1007/s00438-013-0748-6 (2013).
58. Zhang, L., Ma, N., Liu, Q. & Ma, Y. Genome-wide screening for genes associated with valproic acid sensitivity in fission yeast. *PLoS One* **8**, e68738, doi: 10.1371/journal.pone.0068738 (2013).
59. Breslow, D. K. *et al.* A comprehensive strategy enabling high-resolution functional analysis of the yeast genome. *Nat. Methods* **5**, 711–718, doi: 10.1038/nmeth.1234 (2008).
60. Baryshnikova, A. *et al.* Quantitative analysis of fitness and genetic interactions in yeast on a genome scale. *Nat. Methods* **7**, 1017–1024, doi: 10.1038/nmeth.1534 (2010).
61. Beyer, A., Bandyopadhyay, S. & Ideker, T. Integrating physical and genetic maps: from genomes to interaction networks. *Nat. Rev. Genet.* **8**, 699–710, doi: 10.1038/nrg2144 (2007).
62. Guenole, A. *et al.* Dissection of DNA damage responses using multiconditional genetic interaction maps. *Mol. Cell* **49**, 346–358, doi: 10.1016/j.molcel.2012.11.023 (2013).
63. Nguyen, T. T., Lim, J. S., Tang, R. M., Zhang, L. & Chen, E. S. Fitness profiling links topoisomerase II regulation of centromeric integrity to doxorubicin resistance in fission yeast. *Sci. Rep.* **5**, 8400, doi: 10.1038/srep08400 (2015).

64. Li, Y. *et al.* The mitotic spindle is required for loading of the DASH complex onto the kinetochore. *Genes Dev.* **16**, 183–197, doi: 10.1101/gad.959402 (2002).
65. Erdei, Z. *et al.* Expression pattern of the human ABC transporters in pluripotent embryonic stem cells and in their derivatives. *Cytometry B Clin. Cytom.* **86**, 299–310, doi: 10.1002/cyto.b.21168 (2014).
66. Calcagno, A. M., Ludwig, J. A., Fostel, J. M., Gottesman, M. M. & Ambudkar, S. V. Comparison of drug transporter levels in normal colon, colon cancer, and Caco-2 cells: impact on drug disposition and discovery. *Mol. Pharm.* **3**, 87–93 (2006).
67. Cascinu, S. *et al.* Pegylated liposomal doxorubicin, 5-fluorouracil and cisplatin versus mitomycin-C, 5-fluorouracil and cisplatin for advanced gastric cancer: a randomized phase II trial. *Cancer Chemother. Pharmacol.* **68**, 37–43, doi: 10.1007/s00280-010-1424-8 (2011).
68. Hacker, S. *et al.* Histone deacetylase inhibitors prime medulloblastoma cells for chemotherapy-induced apoptosis by enhancing p53-dependent Bax activation. *Oncogene* **30**, 2275–2281, doi: 10.1038/onc.2010.599 (2011).
69. Kim, M. S. *et al.* Inhibition of histone deacetylase increases cytotoxicity to anticancer drugs targeting DNA. *Cancer Res.* **63**, 7291–7300 (2003).
70. Thomas, A. *et al.* A phase I/II trial of belinostat in combination with cisplatin, doxorubicin, and cyclophosphamide in thymic epithelial tumors: a clinical and translational study. *Clin. Cancer Res.* **20**, 5392–5402, doi: 10.1158/1078-0432.CCR-14-0968 (2014).
71. Wang, H. *et al.* Discovery of (2E)-3-[2-butyl-1-[2-(diethylamino)ethyl]-1H-benzimidazol-5-yl]-N-hydroxyacrylamide (SB939), an orally active histone deacetylase inhibitor with a superior preclinical profile. *J. Med. Chem.* **54**, 4694–4720, doi: 10.1021/jm2003552 (2011).
72. Moreno, S., Klar, A. & Nurse, P. Molecular genetic analysis of fission yeast *Schizosaccharomyces pombe*. *Methods Enzymol.* **194**, 795–823 (1991).

Acknowledgements

We thank Rebecca A. Jackson for critically editing the manuscript, Kenneth Hon-Kim Ban, Yoshiaki Ito, Dominic Chih-Cheng Voon, Motomi Osato and Jean Paul Thiery for comments and reagents, and Yen Ling Tan, Huifang Guo, Siew Lee Chan and Yue Rong Tan for technical assistance. The work is supported by a Singapore Ministry of Education Tier II Academic Research Fund (MOE2013-T2-1-112) and NUS DPRT Start-up Grant on Synthetic Biology awarded to E.S.C.

Author Contributions

T.T.T.N., J.K.K.C., K.K.L. and S.Y.W.P. performed fission yeast spottings. T.T.T.N. derived the s-scores and analyzed all data. K.S.S., S.H.K., J.Y.Y. and E.J.D.L. performed all drug treatments in human cancer cell cultures. E.G.Y. and B.D. synthesized SAHA. W.H.A. provided cell lines, and synthesized and advised on cisplatin usage. L.Z. performed simulation, A.Y. and E.S.C. co-supervised the work by J.K.K.C. as a final year project. E.S.C. conceived the study, and coordinated the team. E.S.C., T.T.T.N. and S.H.K. wrote the manuscript. All authors were involved in editing and checking of the manuscript.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Nguyen, T. T. T. *et al.* Predicting chemotherapeutic drug combinations through gene network profiling. *Sci. Rep.* **6**, 18658; doi: 10.1038/srep18658 (2016).



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>