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## Derivation of human decidua-like cells from amnion and menstrual blood

## SUBJECT AREAS:

STEM-CELL  
DIFFERENTIATION  
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We induced differentiation of human amnion-derived mesenchymal stem cells (AMCs) and menstrual blood-derived mesenchymal stem cells (MMCs) into endometrial stroma-like cells, which could be useful for cell therapy to support embryo implantation. Interestingly, the expression patterns of surface markers were similar among AMCs, MMCs, and endometrial stromal cells. In addition, whereas treatment with estrogen and progesterone was not very effective for decidualizing AMCs and MMCs, treatment with 8-Br-cAMP prompted remarkable morphological changes in these cells as well as increased expression of decidualization markers (prolactin and insulin-like growth factor binding protein-1) and attenuated expression of surface markers unique to mesenchymal stem cells. These results demonstrated that bone marrow-derived stem cells, which are considered a potential source of endometrial progenitor cells, as well as AMCs and MMCs show *in vitro* decidualization potential, which is characteristic of endometrial stromal cells.

Many infertile couples have achieved pregnancy by assisted reproductive technology (ART). However, despite having good-quality embryos for transfer, there are still women who experience repeated implantation failure even after several ART attempts<sup>1,2</sup>. Uterine receptivity is considered as another important factor for successful implantation and many studies have attempted to identify clinically useful markers of a receptive uterine state. However, such molecular markers related to repeated implantation failure have proven difficult to find, largely because uterine receptivity is regulated via the expression of various mediators, including cell adhesion molecules (e.g. cadherins and integrins), chemical mediators (e.g. prostaglandins), and cytokines (e.g. leukemia inhibitory factor and epithelial growth factor).

Thinning of endometrium has also been correlated with implantation failure, with causes including aging<sup>3</sup>, and repeated invasive procedures such as dilation and curettage for miscarriage or early-stage endometrial cancer. Some studies have reported clinical benefits from ascorbic acid (vitamin C), tocopherol (vitamin E), pentoxifylline (PTX), or sildenafil for repeated implantation failure in patients with thin endometrium, although the effectiveness and molecular mechanisms by which such agent could improve the implantation process is not well established<sup>4-6</sup>.

In studies to improve implantation rates, Landgren et al.<sup>7</sup> developed a coculture model of embryos with endometrial cells harvested from an endometrial biopsy taken at 4, 5, and 6 days after the luteinizing hormone (LH) peak in healthy women with normal menstrual cycles. The rate of pregnancy increased for embryos transferred after the coculture of embryo with endometrium than for embryos transferred after repeat ART<sup>8</sup>. However it is difficult to retrieve and culture endometrial cells from women whose endometrium has already become thin. Therefore we focused on mesenchymal stem cells (MSCs) as a source for this type of cell therapy using endometrial stroma-like cells. MSCs might also support embryo implantation by excreting cytokines and chemical mediators for adhesion, migration, or immunomodulation<sup>9-11</sup>. Indeed, fertility was restored in a patient with severe Asherman's syndrome treated using autologous bone marrow-derived MSC populations<sup>12</sup>.

MSCs are multipotent, adherent stem cells capable of differentiating into osteoblasts, adipocytes, and chondroblasts<sup>13</sup>, as well as endodermal lineages such as pancreatic islets<sup>14,15</sup> or hepatocytes<sup>16,17</sup>, and ectodermal lineage such as neurons<sup>14,18</sup>. An earlier study verified that human bone marrow-derived MSC (BMCs) are potential progenitors of endometrial stromal cells by demonstrating their ability to decidualize<sup>19</sup>. Decidualization is a remodeling process designed to prepare endometrium for pregnancy that is induced by progesterone secreted from the ovaries in the luteal phase. This remodeling is necessary in a successful pregnancy to regulate trophoblast invasion, resist oxidative stress, and protect the placental semi-allograft against maternal



immune responses. Endometrial stromal cells (ESCs) are reprogrammed into decidual cells, and display characteristic morphological changes from an elongated spindle-shaped fibroblastic appearance to a polygonal shape with large nuclei. Prolactin (PRL) and insulin-like growth factor binding protein-1 (IGFBP1) have been widely used as phenotypic markers of decidual cells. In the current study, we hypothesized that if MSCs could differentiate into ESC-like cells, their treatment with agents known to promote decidualization could induce the expression of PRL and IGFBP1 in the MSC-derived cells. Indeed, AMCs and MMCs successfully differentiate into endometrial stroma-like cells presenting decidualization potential to the same extent as bone marrow-derived cells<sup>19</sup>. In this context, human amnion in particular could be a powerful therapeutic cell source because not only is it an easily accessible tissue for cell harvesting, it does not require co-administration of immunosuppressive agent or matching of MHC typing for immunological tolerance.

## Results

**Characterization of human endometrial cells.** Immunofluorescence staining and flow cytometric analysis of cultured human endometrial cells and MRC5 fibroblasts revealed both to be positive for vimentin and negative for cytokeratin, whereas a human carcinoma cell line, HeLa cells, was positive for both vimentin and cytokeratin. (Fig. 1A–E). By flow cytometry, the endometrial cells and fibroblasts were negative for CD9 and positive for CD10, whereas HeLa cells were positive for CD9 and negative for CD10. Together, these results confirmed that the endometrial cells were stromal in origin, compared to the epithelial-derived HeLa cells.

We then investigated cell surface marker expressions of ESCs, AMCs, MMCs, BMCs, and MRC5 by flow cytometric analysis on the initial day of treatment (day 0) (Fig. 1F). Although there is no marker specific to MSCs, they are known to express CD73, CD90, and CD105<sup>20</sup>, and the AMCs and MMCs tested herein indeed expressed these cell surface markers as well as CD54, CD166 and HLA-ABC, but not HLA-DR (Supplementary Fig. S1). This result confirmed the earlier studies of our group demonstrating that both AMCs and MMCs express MSC markers such as CD29, CD44, CD59, CD73, CD105, and CD166, but not hematopoietic markers such as CD14, CD34, and CD45<sup>21–23</sup>. None of the cells analyzed in the present study expressed hematopoietic lineage markers such as CD34 or monocyte-macrophage antigens such as CD14 (a marker for macrophage and dendritic cells) and CD45 (leukocyte common antigen), suggesting no contaminating hematopoietic cells. Lastly, and interestingly, the surface marker expression patterns of AMCs and MMCs were very similar to those of ESCs.

**Decidual differentiation by treatment with  $E_2 + P_4$ .** Human ESCs showed distinct morphological changes following treatment with  $E_2 + P_4$ , from spindle-shaped to large round cells with large round nuclei, whereas AMCs, MMCs, and BMCs showed no remarkable morphological changes (Fig. 2A). Flow cytometric analysis of ESCs, AMCs, MMCs, BMCs, and MRC5 fibroblasts on day 14 of treatment with  $E_2 + P_4$  (Fig. 2B and Supplementary Fig. S2) also showed no remarkable changes in surface marker expressions compared to those on day 0.

Immunofluorescence staining was also performed to assess the expression of decidualization markers (PRL and IGFBP1) at day 0, 7, and 14 after treatment with  $E_2 + P_4$  (Fig. 3). None of these markers were expressed at day 0 in ESCs, AMCs, MMCs, and BMCs, but PRL and IGFBP1 were expressed in ESCs on days 7 and 14 following treatment with  $E_2 + P_4$  (Figs. 3A and 3B). AMCs also showed clear expression of PRL on days 7 and 14 following treatment, but only faint IGFBP1 expression (Figs. 3C and 3D), while the reverse pattern was observed in MMCs (Figs. 3E and 3F). Finally, both PRL and IGFBP1 were faintly expressed in BMCs (Figs. 3G and 3H).

The qRT-PCR analysis of expression changes in *PRL* and *IGFBP1* mRNAs with  $E_2 + P_4$  treatment (Figs. 2C and 2D) revealed

upregulation of *PRL* in each cell type on days 7 and 14 following treatment compared to day 0 levels, whereas *IGFBP1* was upregulated only in ESCs on days 7 and 14 compared to day 0, and not in AMCs, MMCs, and BMCs.

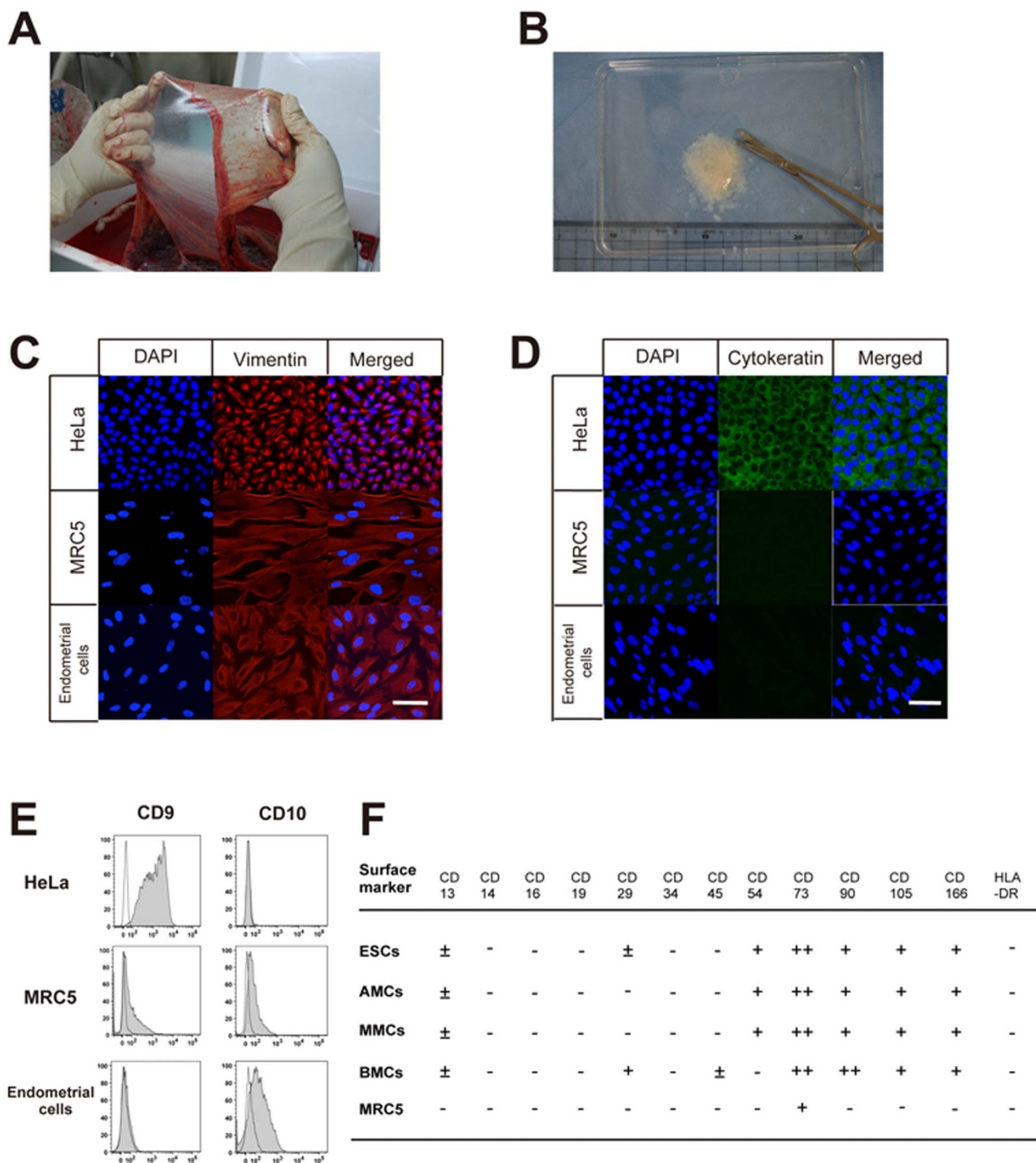
**Decidual differentiation by treatment with cAMP.** The human ESCs, AMCs, MMCs, and BMCs all showed morphological changes from a spindle-shaped appearance to large rounded cells with large round nuclei following treatment with cAMP (Fig. 4A). In addition, flow cytometry revealed that CD90, CD105, and CD166 were attenuated in the AMCs and ESCs on day 14 of the cAMP treatment (Fig. 4B and Supplementary Fig. S3) compared to day 0 levels. Together, these changes suggested that cAMP upregulates decidual differentiation in AMCs. On the other hand, CD105 and CD166 in MMCs as well as CD90 and CD166 in BMCs were unchanged, suggesting that AMCs could have more potential for differentiation to ESCs than MMCs and BMCs.

Immunofluorescence staining showed no expression of decidualization markers (PRL and IGFBP1) at day 0 in the four cell types, but positive staining for PRL on days 7 and 14 of the cAMP treatment (Fig. 5A, 5C, 5E and 5G). Although IGFBP1 was also expressed in all the types of cells on day 7 and day 14, AMCs and MMCs showed more remarkable IGFBP1 expression on day 7 compared to day 14.

Finally, the qRT-PCR analysis showed that *PRL* transcripts were more abundant in all the cell types on days 7 and 14 following cAMP treatment than on day 0 (Figs. 4C and 4D), whereas *IGFBP1* was upregulated in AMCs and ESCs, but not in MMCs and BMCs. These results suggested that AMCs treated with cAMP have the closest decidualization potential to ESCs.

## Discussion

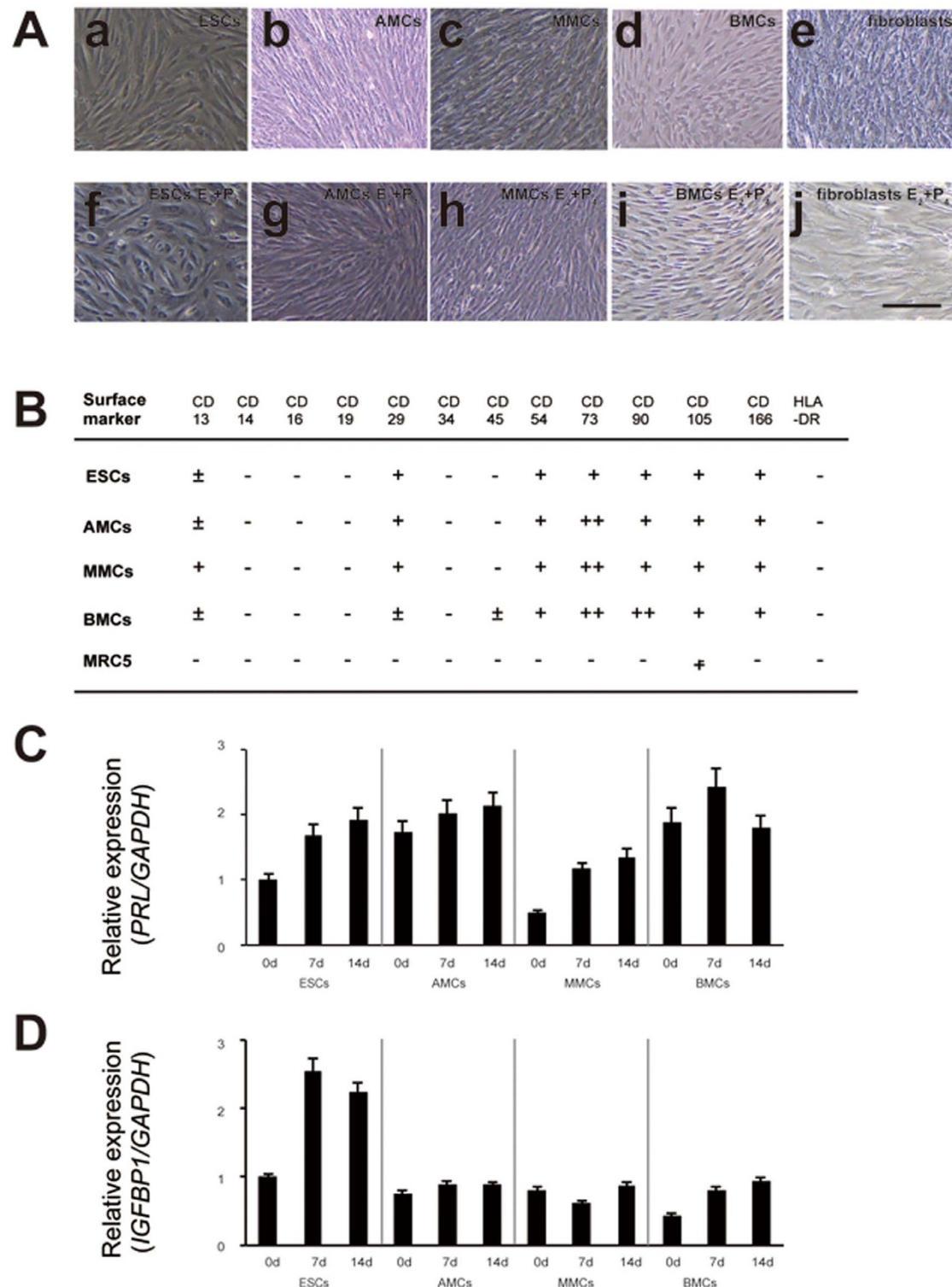
**Insufficient response of MSCs to  $E_2 + P_4$ .** In this study, we investigated whether human MSCs such as AMCs and MMCs could be differentiated *in vitro* into endometrial stroma-like cells using decidualization stimuli and analysis of cellular morphology and expression of decidualization markers (PRL and IGFBP1). The MSCs and ESCs were treated with  $E_2 + P_4$  to mimic the normal endocrinological condition in the secretory phase of a human menstrual cycle. ESCs and BMCs were considered as positive controls. Unexpectedly, not only the AMCs and MMCs, but also the BMCs, failed to show remarkable decidual changes with  $E_2 + P_4$  treatment. Sustained expression of cell surface markers, no marked morphological changes, and little expression of IGFBP1 in MSCs treated with  $E_2 + P_4$  indicated that these cells are not sufficiently responsive to such stimulation to induce differentiation, supporting previous results by Aghajanova et al.<sup>19</sup> in BMCs treated with  $E_2 + P_4$ . The qRT-PCR in this study showed significant expression of progesterone receptors (PR) and estrogen receptors (ESR1 and ESR2) in AMCs, MMCs, and ESCs (Supplementary Fig. S5). Aghajanova et al. also demonstrated *PR*, *ESR1*, and *ESR2* mRNA expression in BMCs and ESCs, but this expression was not significantly influenced by cAMP treatment of the cells<sup>19</sup>. Furthermore, with respect to secretory-phase endometrium *in vivo*, decidualization of the superficial endometrial layers is only apparent ~10 days after the postovulatory increase in progesterone levels. All these observations indicate that additional signals other than the progesterone signaling are required to initiate decidualization. Interestingly, trichostatin A (TSA), a specific histone deacetylase (HDACs) inhibitor, enhanced the upregulation of PRL and IGFBP1 in a dose-dependent manner in cultured endometrial stromal cells following treatment with  $E_2 + P_4$ <sup>24</sup>. In addition, IGFBP1 expression induced by cAMP was associated with the histone acetylation status of the promoter region in human endometrial stromal cells<sup>25</sup>. Based on these results, we speculated that TSA might be effective in enhancing IGFBP1 expression in AMCs and MMCs treated with  $E_2 + P_4$ ; however, AMCs and MMCs treated with  $E_2 + P_4$  and several concentrations



**Figure 1 | Characterization of endometrial cells, amnion-derived cells, and menstrual blood-derived cells.** (A), Macroscopic view of human amniotic membrane. (B), The amniotic membrane was cut into pieces of approximately 2 mm<sup>3</sup> in size. (C and D), Laser confocal microscopic view of immunofluorescence staining of endometrial cells, HeLa cells, and MRC5 fibroblasts in culture with anti-vimentin (red) (C) and anti-cytokeratin antibodies (green) (D), DAPI staining (blue). Scale bars, 100  $\mu$ m. (E), Surface marker (CD9 or CD10) expression of endometrial cells, HeLa cells, and MRC5 fibroblasts. White and grey areas indicate reactivity of antibodies for isotype controls and that for CD9 or CD10, respectively. (F), Summary of flow cytometric analysis of endometrial stromal cells (ESCs), amnion-derived mesenchymal stem cells (AMCs), menstrual blood-derived mesenchymal stem cells (MMCs), bone marrow-derived mesenchymal stem cells (BMCs), and MRC5 fibroblasts before treatment. The shift in peak histogram value of flow cytometric data was measured and classified into four groups: -, from negative to 2 log shift;  $\pm$ , 2 log shift from negative control; +, 3 log shift from negative control; ++, 4 log shift from negative control.

of TSA showed no enhancement of IGFBP1 expression (Supplemental Fig. S4). We therefore further investigated the decidualization potential of MSCs in the current study using cAMP, a well-known and common facilitator of decidualization<sup>19,25</sup>.

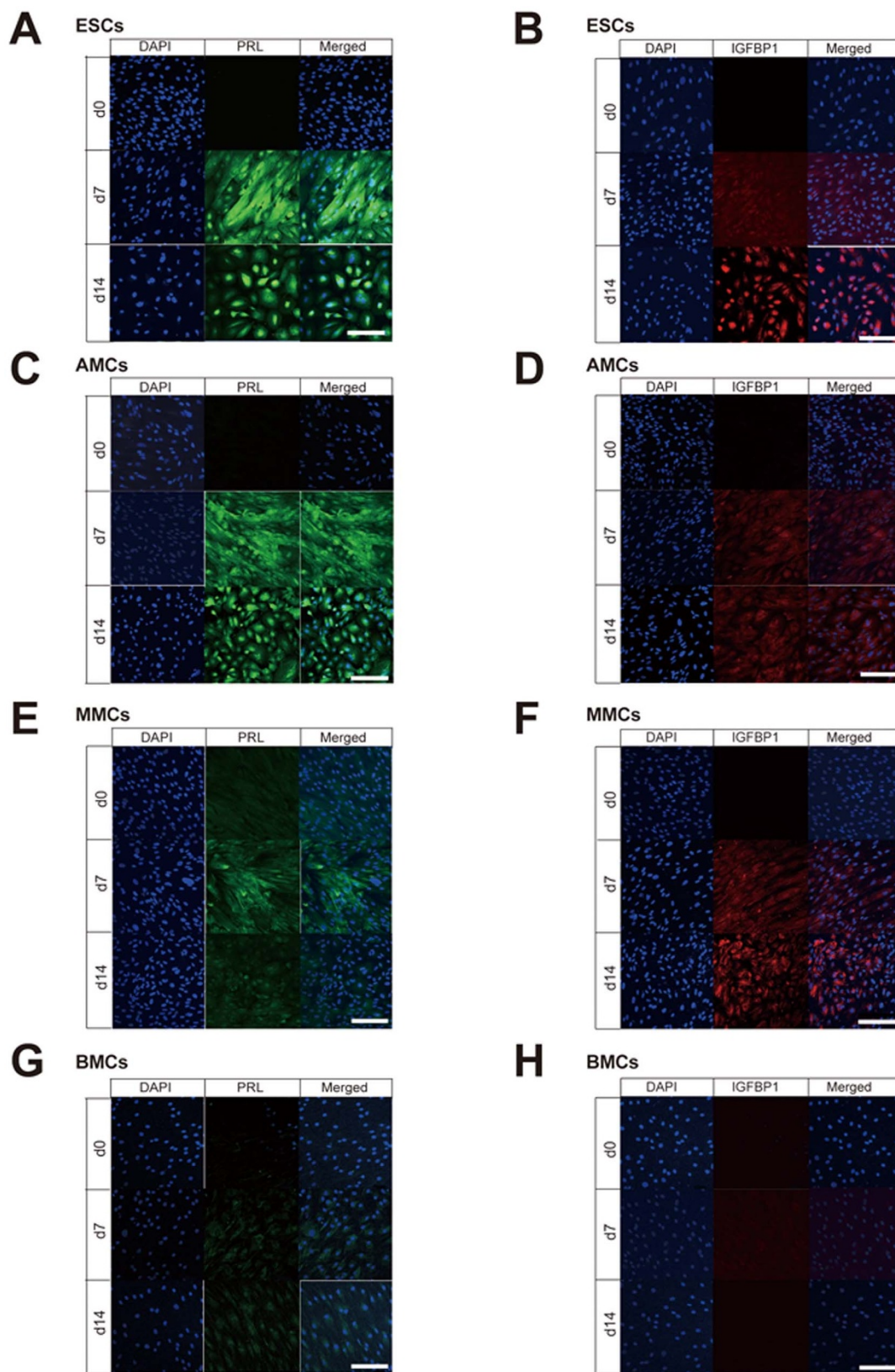
**Attenuation of cell surface markers for MSCs during successful decidualization by cAMP.** Estrogen and progesterone bind to nuclear receptor proteins for nuclear importing, where they bind DNA directly to regulate transcription, whereas cAMP activates the protein



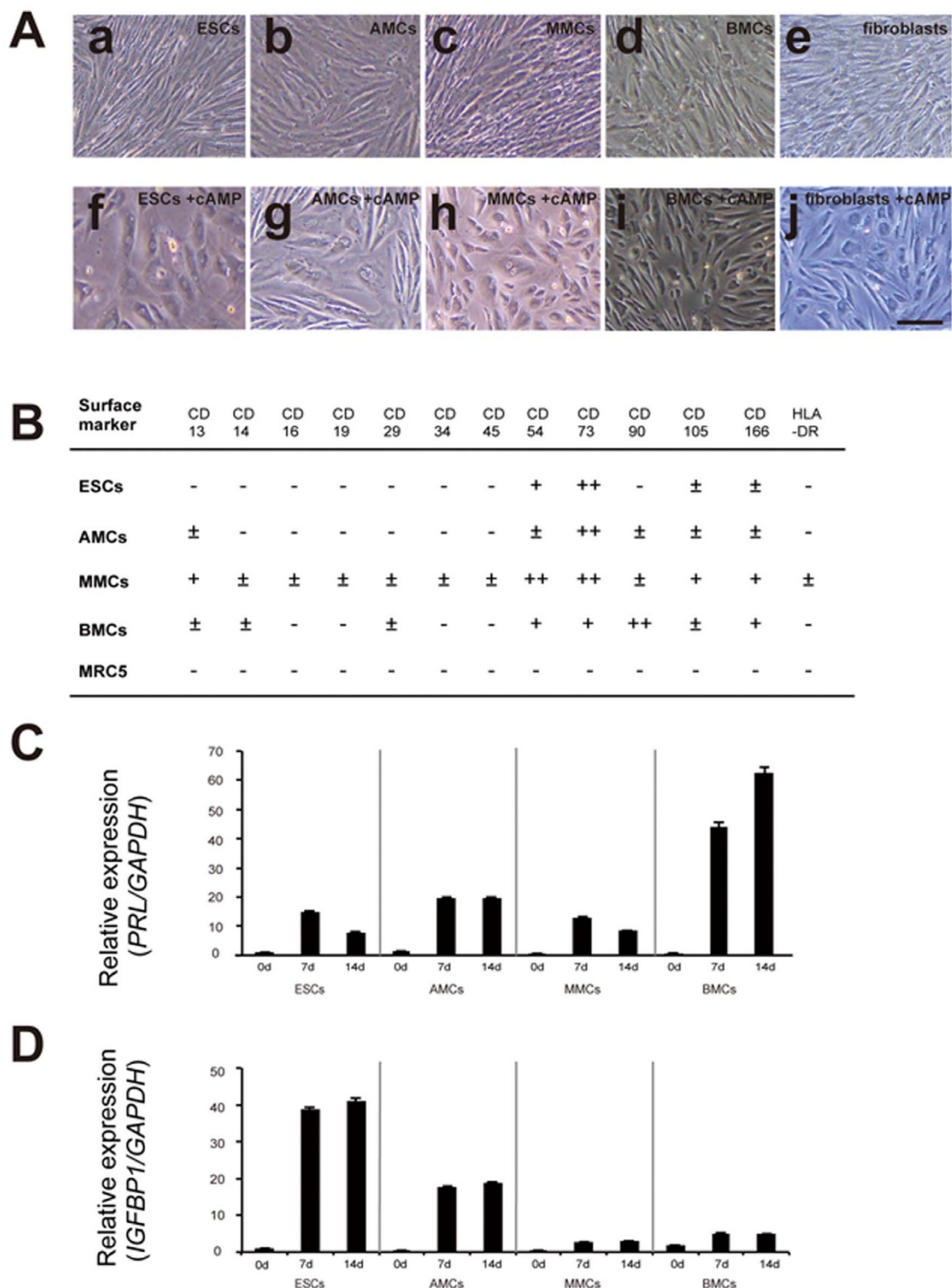
**Figure 2 | Morphological changes and alteration of flow cytometric profiles in mesenchymal stem cells treated with  $E_2 + P_4$ .** (A), Morphological changes in endometrial stromal cells (ESCs) (a, f), amnion-derived mesenchymal stem cells (AMCs) (b, g), menstrual blood-derived mesenchymal stem cells (MMCs) (c, h), bone marrow-derived mesenchymal stem cells (BMCs) (d, i), and MRC5 fibroblasts (e, j) before (a–c) and after (f–j) treatment with 10 nM  $E_2$  and 1  $\mu$ M  $P_4$  ( $E_2 + P_4$ ) for 14 days. Scale bars, 200  $\mu$ m. (B), Summary of flow cytometric analysis of ESCs, AMCs, MMCs, BMCs, and MRC5 fibroblasts treated with  $E_2 + P_4$  for 14 days. We measured the shift of peak histogram value for the flow cytometric data and defined each result by the aforementioned criteria. (C), PRL mRNA expression in ESCs, AMCs, MMCs, and BMCs treated with  $E_2 + P_4$  for 0, 7, and 14 days. (D), IGFBP1 mRNA expression in ESCs, AMCs, MMCs, and BMCs treated with  $E_2 + P_4$  for 14 days.

kinase A (PKA) pathway and a catalytic subunit of PKA translocates into the cell nucleus to phosphorylate cAMP response element-binding protein (CREB). Activated CREB then enhances gene transcription via cAMP response element (CRE)<sup>26</sup>. In this study, cAMP treatment of

AMCs and MMCs produced morphological changes characteristic of decidual cells, and PRL and IGFBP1 expression increases consistent with those seen in ESCs. AMCs and MMCs were thus demonstrated to have decidualization potential.



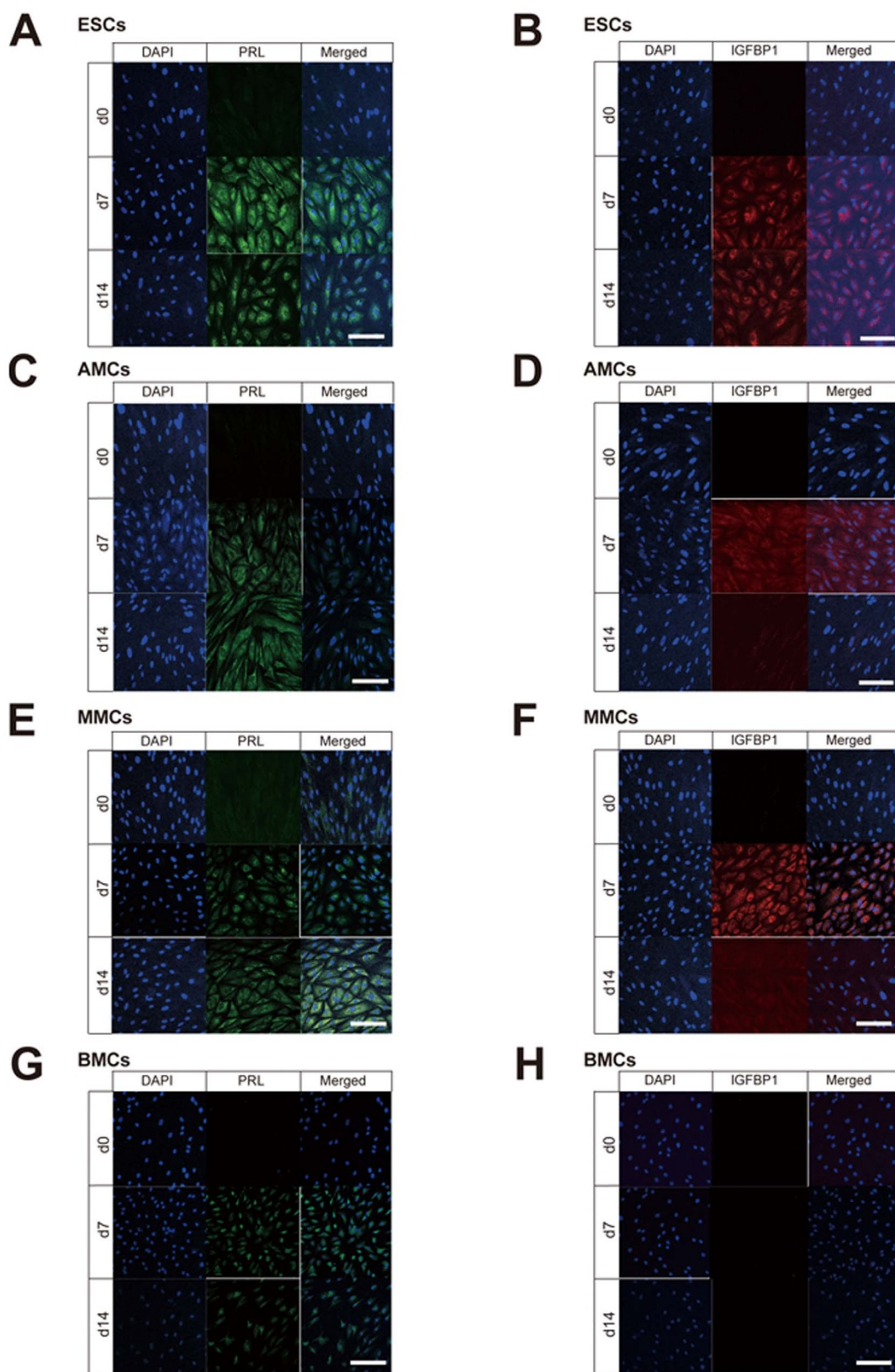
**Figure 3** | Immunofluorescence staining of decidualization markers (PRL and IGFBP1) in mesenchymal stem cells treated with  $E_2 + P_4$ . Laser confocal microscopy of PRL immunofluorescence staining (green signals in A, C, E, and G) and IGFBP1 (red signals in B, D, F, H) in endometrial stromal cells (ESCs) (A and B), amnion-derived mesenchymal stem cells (AMCs) (C and D), menstrual blood-derived mesenchymal stem cells (MMCs) (E and F), and bone marrow-derived mesenchymal stem cells BMCs (G and H) treated with  $E_2 + P_4$  for 0, 7, or 14 days. Scale bars, 200  $\mu$ m.



**Figure 4 | Morphological changes and alteration of flow cytometric profiles in mesenchymal stem cells treated with cAMP.** (A), Morphological changes of endometrial stromal cells (ESCs) (a, f), amnion-derived mesenchymal stem cells (AMCs) (b, g), menstrual blood-derived mesenchymal stem cells (MMCs) (c, h), bone marrow-derived mesenchymal stem cells (BMCs) (d, i), and MRC5 fibroblasts (e, j) before (a–e) and (f–j) after treatment with 1 mM 8-Br-cAMP for 14 days. Scale bars, 200  $\mu$ m. (B), Summary of flow cytometric analysis of ESCs, AMCs, MMCs, BMCs, and MRC5 fibroblasts treated with 1 mM 8-Br-cAMP for 14 days. We measured the shift of peak histogram value for the flow cytometric data and defined each result by the aforementioned criteria. (C), PRL mRNA expression in ESCs, AMCs, MMCs, and BMCs treated with 1 mM 8-Br-cAMP for 0, 7, and 14 days. (D), IGFBP1 mRNA expression in ESCs, AMCs, MMCs, and BMCs treated with 1 mM 8-Br-cAMP for 0, 7, and 14 days.

In the cell surface marker analysis, the attenuation of CD90, CD105, and CD166 during decidualization was notable. Such a response by these well-defined human MSCs markers could indicate the direction of differentiation in MSCs treated with cAMP.

Furthermore, CD90 is known to regulate adhesion to T cells in MSCs, and a decrease in CD90-positive cells might be a novel predictive marker of immunosuppressive activity<sup>27</sup>. Based on these observations, the decreased rate of CD90-positive cells with cAMP



**Figure 5 | Immunofluorescence staining of decidualization markers (PRL and IGFBP1) in mesenchymal stem cells treated with cAMP.** Laser confocal microscopic view of immunofluorescence staining for PRL (green signals in A, C, E, and G) and IGFBP1 (red signals in B, D, F, and H) in endometrial stromal cells (ESCs) (A and B), amnion-derived mesenchymal stem cells (AMCs) (C and D), menstrual blood-derived mesenchymal stem cells (MMCs) (E and F), bone marrow-derived mesenchymal stem cells (BMCs) (G and H) treated with 1 mM 8-Br-cAMP for 0, 7, and 14 days. Scale bars, 200  $\mu$ m.



observed in the current study may indicate immunosuppression in AMCs and MMCs that is advantageous in terms of their potential clinical application. CD105 (endoglin) is a reliable marker for highly multipotent human MSCs, with CD105-expressing MSCs showing high efficiency in repairing the infarcted heart<sup>28</sup>. Therefore, the disappearance of CD105 with cAMP treatment would ensure differentiation of MSCs<sup>29,30</sup>. Collectively, our findings indicated that administration of cAMP effectively leads to differentiation and decidualization of AMCs and MMCs.

**Advantages of AMCs and MMCs.** It has been reported that endometrial stem cells are CD45-positive cells around blood vessels and likely to be derived from BMCs<sup>31</sup>. A previous study also indicated that BMCs might be a source of endometrial stem/progenitor cells based on successful decidualization and gene expression profiles of BMCs treated with cAMP<sup>19</sup>. If BMCs are indeed a physiological source of endometrial progenitor cells, BMCs could be ideal for use in cell-based therapies for infertile women who have thinning of the endometrium. However, our findings revealed that not only BMCs, but AMCs and MMCs, also showed decidualization potential, and that all three cell types showed a similar profile of cell surface marker expression to that of ESCs. Therefore, the current study suggested that BMCs is not the only such candidate for cell therapy.

In general, the collection of bone marrow is an invasive technique performed with a biopsy needle that requires local or even general anesthesia, and occasionally raises an ethical problem. On the other hand, human amnion is non-invasively and adequately obtained at delivery. Therefore, human amnion is more simply accessed with informed consent than other potential cell sources. Human primary menstrual blood-derived cells are also obtained by a simple, safe, and painless procedure, and efficiently expanded in vitro. In particular, the MMC-derived cells can be used for autologous transplantation. Accordingly, if culture condition for MMCs could be more successfully optimized for differentiation to ESCs, MMCs would be definitely useful for eventual cell-based therapies for infertility.

Furthermore, previous studies suggested that human amnion has the potential for immunological tolerance<sup>23,32</sup>. AMCs are known to express less MHC compared to other MSCs, which is important in host tolerance. Cells that do not express MHC are potentially recognized by the immune system as non-self cells and accordingly, attacked by natural killer (NK) cells. A soluble factor such as the non-classic MHC class I antigen, HLA-G, is also critical for the immunosuppressive properties of MSCs<sup>33</sup>. HLA-G is expressed on the extravillous cytotrophoblast cells at the fetomaternal interface and plays a major role in protecting the fetus from maternal rejection by NK cells<sup>34,35</sup>. HLA-G thus blocks the immunological response of NK cells<sup>36</sup> and induces regulatory T cells<sup>37</sup>, which is a requisite condition of immunological tolerance<sup>35,38</sup>. We previously showed by western blotting that HLA-G is also expressed in placenta-derived tissues including amnion<sup>23</sup>. Thus, AMC-derived cells have several advantages for application to cell-based therapy.

Based on the results of our current study, AMCs and MMCs in addition to BMCs show potential for decidualization, and are therefore candidate cell sources for therapy to support implantation and placentation. The current study provides novel and important information on AMCs and MMCs treated with cAMP as a model of decidualization.

## Methods

**Tissue isolation and cell culture.** All experiments involving human cells and tissues were performed according to the Tenets of the Declaration of Helsinki and were approved by the Ethics Committee of the National Institute for Child and Health Development, Tokyo. Signed informed consent was obtained from all donors, and all specimens were irreversibly de-identified. Human endometrium was obtained from a patient undergoing hysterectomy due to myoma uteri and human placenta was collected after selective caesarean section delivery.

Human amnion-derived cells were isolated using the explant culture method, in which the cells were outgrown from pieces of amnion. Briefly, the amnion was cut into pieces of approximately 2 mm<sup>3</sup> in size, which were then washed in Dulbecco's modified Eagle's medium (DMEM) (high glucose; Sigma) supplemented with 100 U/ml penicillin-streptomycin (Gibco). Some pieces were attached to the substratum in a 10-cm dish. The cells migrated out from the cut ends after approximately 20 days of incubation at 37°C in 5% CO<sub>2</sub>. The migrated cells were harvested with Dulbecco's phosphate-buffered saline containing 0.1% trypsin and 0.25 mM EDTA for 5 min at 37°C and counted. The harvested cells were then re-seeded at a density of 3 × 10<sup>5</sup> cells in 10-cm dishes. Once confluent, the cell monolayers were subcultured at a 1:8 split ratio onto new 10-cm dishes, and the culture medium was replaced with fresh DMEM supplemented with 10% fetal bovine serum (FBS) and 100 U/ml penicillin-streptomycin every 3 or 4 days<sup>22</sup>.

Menstrual blood samples (n = 21) were collected in DMEM containing 100 U/ml penicillin-streptomycin and 2% FBS, and processed within 24 h. The centrifuged pellets containing endometrium-derived cells were resuspended in high-glucose DMEM with 10% FBS and penicillin-streptomycin, and then maintained at 37°C in a humidified atmosphere containing 5% CO<sub>2</sub> to attach for 48 h. Non-adherent cells were removed by changing the medium. Once sub-confluent, the cells were harvested with 0.25% trypsin and 1 mM EDTA, and plated to new dishes. After 2–3 passages, the attached cells were devoid of blood cells<sup>21</sup>.

A human bone marrow-derived mesenchymal cell line, 3F0664 was purchased from Lonza (PT-2501, Basel, Switzerland) and cultured in the mesenchymal stem-cell-basal-medium (MSCBM)-Medium-BulletKit (PT-3238, Lonza). The normal human fetal fibroblast cell line MRC5 was also passaged in DMEM supplemented with 10% (v/v) FBS and 100 U/ml penicillin-streptomycin. Cultures were maintained at 37°C with 5% CO<sub>2</sub> in a humidified atmosphere.

For decidual differentiation, human endometrial cells, amnion-derived cells, menstrual blood-derived cells, bone marrow-derived cells, and MRC5 were first cultured as described above, then trypsinized and plated in 10-cm plates. Nearly confluent cells were cultured for 0, 7, and 14 days in low-serum medium (2% FBS) supplemented with: 1) 10 nM estrogen (E<sub>2</sub>) and 1 μM progesterone (P<sub>4</sub>) or 2) 1 mM 8-Br-cAMP (cAMP) (Sigma-Aldrich, Saint Louis, MO, USA).

**Flow cytometric analysis.** Cells were incubated for 20 min at 4°C with primary antibodies or isotype-matched control antibodies, followed by immunofluorescent secondary antibodies. The cells were then analyzed on a FACS Aria™ IIIu Cellsorter (Becton Dickinson, Inc.) using FlowJo Ver.7 (Tree Star, Inc.). Antibodies against human CD9, CD10, CD13, CD14, CD16, CD19, CD29, CD31, CD34, CD44, CD45, CD54, CD55, CD59, CD73, CD90, CD105, CD133, CD146, CD166, HLA-ABC, and HLA-DR were purchased from Beckman Coulter, Immunotech (Marseille, France), BD Pharmingen and BioLegend (San Diego, CA, USA)<sup>21,22</sup>. Supplementary table S1 shows the list of CD antigens investigated in this study<sup>39–49</sup>.

**Immunofluorescence staining.** The immunofluorescence staining was performed as previously described<sup>50</sup>. Briefly, cells were fixed with 4% paraformaldehyde for 10 min at 4°C, treated with 0.1% Triton X-100 (Sigma) in PBS for 15 min at RT, and then incubated for 30 min at RT in protein-blocking solution consisting of PBS supplemented with 5% FBS. The samples were then incubated overnight with the following primary antibodies diluted in PBS: an anti-prolactin mouse monoclonal antibody, (QED Bioscience, San Diego, CA, USA), anti-IGFBP1 mouse monoclonal antibody (Santa Cruz Biotechnology, Dallas, Texas, CA, USA), anti-vimentin mouse monoclonal antibody, V9, (DAKO, Tokyo, Japan), and an anti-cytokeratin mouse monoclonal antibody, AE1 + AE3, (DAKO). Tissues were washed twice for 5 min in PBS and incubated with the appropriate secondary antibodies (Alexa Fluor 488 goat anti-mouse IgG, 1:600 and Alexa Fluor 546 goat anti-mouse IgG, 1:600; Life Technologies, Grand Island, NY). Nuclei were counterstained with 4', 6-diamidino-2-phenylindole (DAPI; Biotium, CA, USA) for 30 min at RT followed by 10 final 5-min PBS washes before being cover slipped using mounting medium. Omission of primary antibodies served as a negative control.

**Quantitative reverse transcriptase-polymerase chain reaction (qRT-PCR) analysis.** Total cellular RNA was isolated from cells using an RNeasy Plus Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. Total RNA (2.5 μg each) for quantitative RT-PCR was converted to cDNA using an Oligo (dT) primer with Superscript VILO™ cDNA Synthesis Kit (Life Technologies Corp., Carlsbad, CA, USA), according to the manufacturer's manual. The cDNA template was amplified by thermal cycle reactions (Quant Studio™ 12 K Flex Real-Time PCR System) using the Platinum SYBR Green qPCR SuperMix-UDG (Life Technologies Corp.) under the following reaction conditions: 40 cycles of PCR (95°C for 15 s and 60°C for 1 min) after an initial denaturation (95°C for 2 min). Fluorescence was monitored during every PCR cycle at the annealing step. The authenticity and size of the PCR products were confirmed by a melting curve analysis using software provided by Applied Biosystems. mRNA levels were normalized using *GAPDH* as a housekeeping gene. Primers used to amplify a cDNA fragment for human PRL and IGFBP1 were the following: PRL, 5'-CATCAACAGCTGCCACACTT-3' (F) and 5'-CGTTTGGTTGCTCCTCAAT-3' (R); IGFBP1, 5'-CTATGATGGCTCGAAG-GTCTC-3' (F) and 5'-TTCTTGTTCAGTTTGGCAG-3' (R).

- Lee, T. H. *et al.* Embryo quality is more important for younger women whereas age is more important for older women with regard to in vitro fertilization outcome and multiple pregnancy. *Fertil Steril* **86**, 64–69 (2006).





2. Navot, D. *et al.* Poor oocyte quality rather than implantation failure as a cause of age-related decline in female fertility. *Lancet* **337**, 1375–1377 (1991).
3. Amir, W. *et al.* Predicting factors for endometrial thickness during treatment with assisted reproductive technology. *Fertil Steril* **87**, 799–804 (2007).
4. Senturk, L. M. & Erel, C. T. Thin endometrium in assisted reproductive technology. *Curr Opin Obstet Gynecol* **20**, 221–228 (2008).
5. Sallam, H. N. Embryo transfer: factors involved in optimizing the success. *Curr Opin Obstet Gynecol* **17**, 289–298 (2005).
6. Check, J. H. *et al.* Neither sildenafil nor vaginal estradiol improves endometrial thickness in women with thin endometria after taking oral estradiol in graduating dosages. *Clin Exp Obstet Gynecol* **31**, 99–102 (2004).
7. Landgren, B. M., Johannisson, E., Stavreus-Evers, A., Hamberger, L. & Eriksson, H. A new method to study the process of implantation of a human blastocyst in vitro. *Fertil Steril* **65**, 1067–1070 (1996).
8. Eyheremendy, V. *et al.* Beneficial effect of autologous endometrial cell coculture in patients with repeated implantation failure. *Fertil Steril* **93**, 769–773 (2010).
9. Lee, E. J. *et al.* Spherical bullet formation via E-cadherin promotes therapeutic potency of mesenchymal stem cells derived from human umbilical cord blood for myocardial infarction. *Mol Ther* **20**, 1424–1433 (2012).
10. Qiu, Y., Marquez-Curtis, L. & Janowska-Wieczorek, A. Mesenchymal stromal cells derived from umbilical cord blood migrate in response to complement C1q. *Cytotherapy* **14**, 285–295 (2012).
11. Moorefield, E. C. *et al.* Cloned, CD117 selected human amniotic fluid stem cells are capable of modulating the immune response. *PLoS One* **6**, e26535 (2011).
12. Gargett, C. E. & Healy, D. L. Generating receptive endometrium in Asherman's syndrome. *J Hum Reprod Sci* **4**, 49–52 (2011).
13. Pittenger, M. F. *et al.* Multilineage Potential of Adult Human Mesenchymal Stem Cells. *Science* **284**, 143–147 (1999).
14. Miki, T., Lehmann, T., Cai, H., Stolz, D. B. & Strom, S. C. Stem cell characteristics of amniotic epithelial cells. *Stem Cells* **23**, 1549–1559 (2005).
15. Santos, T. M. *et al.* Expression of pancreatic endocrine markers by mesenchymal stem cells from human umbilical cord vein. *Transplant Proc* **42**, 563–565 (2010).
16. Lee, K. D. *et al.* In vitro hepatic differentiation of human mesenchymal stem cells. *Hepatology* **40**, 1275–1284 (2004).
17. Pournasr, B. *et al.* In vitro differentiation of human bone marrow mesenchymal stem cells into hepatocyte-like cells. *Arc Iran Med* **14**, 244–249 (2011).
18. Dezawa, M. *et al.* Specific induction of neuronal cells from bone marrow stromal cells and application for autologous transplantation. *J Clin Invest* **113**, 1701–1710 (2004).
19. Aghajanova, L., Horcajadas, J. A., Esteban, F. J. & Giudice, L. C. The bone marrow-derived human mesenchymal stem cell: potential progenitor of the endometrial stromal fibroblast. *Biol Reprod* **82**, 1076–1087 (2010).
20. Diaz-Prado, S. *et al.* Human amniotic membrane as an alternative source of stem cells for regenerative medicine. *Differentiation* **81**, 162–171 (2011).
21. Cui, C. H. *et al.* Menstrual blood-derived cells confer human dystrophin expression in the murine model of Duchenne muscular dystrophy via cell fusion and myogenic transdifferentiation. *Mol Biol Cell* **18**, 1586–1594 (2007).
22. Kawamichi, Y. *et al.* Cells of extraembryonic mesodermal origin confer human dystrophin in the mdx model of Duchenne muscular dystrophy. *J Cell Physiol* **223**, 695–702 (2010).
23. Tsuji, H. *et al.* Xenografted human amniotic membrane-derived mesenchymal stem cells are immunologically tolerated and transdifferentiated into cardiomyocytes. *Circ Res* **106**, 1613–1623 (2010).
24. Sakai, N. *et al.* Involvement of histone acetylation in ovarian steroid-induced decidualization of human endometrial stromal cells. *J Biol Chem* **278**, 16675–16682 (2003).
25. Tamura, I. *et al.* Induction of IGFBP-1 expression by cAMP is associated with histone acetylation status of the promoter region in human endometrial stromal cells. *Endocrinology* **153**, 5612–5621 (2012).
26. Malumbres, M. & Pellicer, A. RAS pathways to cell cycle control and cell transformation. *Front Biosci* **3**, d887–912 (1998).
27. Campioni, D. *et al.* A decreased positivity for CD90 on human mesenchymal stromal cells (MSCs) is associated with a loss of immunosuppressive activity by MSCs. *Cytometry B Clin Cytom* **76**, 225–230 (2009).
28. Gaebel, R. *et al.* Cell origin of human mesenchymal stem cells determines a different healing performance in cardiac regeneration. *PLoS One* **6**, e15652 (2011).
29. Rosu-Myles, M., Fair, J., Pearce, N. & Mehic, J. Non-multipotent stroma inhibit the proliferation and differentiation of mesenchymal stromal cells in vitro. *Cytotherapy* **12**, 818–830 (2010).
30. Jin, H. J. *et al.* Down-regulation of CD105 is associated with multi-lineage differentiation in human umbilical cord blood-derived mesenchymal stem cells. *Biochem Biophys Res Commun* **381**, 676–681 (2009).
31. Bratincák, A. *et al.* CD45-positive blood cells give rise to uterine epithelial cells in mice. *Stem Cells* **25**, 2820–2826 (2007).
32. Cui, C. H. *et al.* Dystrophin conferral using human endothelium expressing HLA-E in the non-immunosuppressive murine model of Duchenne muscular dystrophy. *Hum Mol Genet* **20**, 235–244 (2011).
33. Selmani, Z. *et al.* HLA-G is a crucial immunosuppressive molecule secreted by adult human mesenchymal stem cells. *Transplantation* **87**, S62–S66 (2009).
34. Rouas-Freiss, N., Goncalves, R. M., Menier, C., Dausset, J. & Carosella, E. D. Direct evidence to support the role of HLA-G in protecting the fetus from maternal uterine natural killer cytotoxicity. *Proc Natl Acad Sci USA* **94**, 11520–11525 (1997).
35. Fernandez, N. *et al.* A critical review of the role of the major histocompatibility complex in fertilization, preimplantation development and fetomaternal interactions. *Hum Reprod Update* **5**, 234–248 (1999).
36. Pazmany, L. *et al.* Protection from natural killer cell-mediated lysis by HLA-G expression on target cells. *Science* **274**, 792–795 (1996).
37. Selmani, Z. *et al.* Human leukocyte antigen-G5 secretion by human mesenchymal stem cells is required to suppress T lymphocyte and natural killer function and to induce CD4<sup>+</sup> CD25<sup>high</sup> FOXP3<sup>+</sup> regulatory T cells. *Stem Cells* **26**, 212–222 (2008).
38. Hutter, H. *et al.* Expression of HLA class I molecules in human first trimester and term placenta trophoblast. *Cell Tissue Res* **286**, 439–447 (1996).
39. Paietta, E. Expression of cell-surface antigens in acute promyelocytic leukaemia. *Best Practice & Research Clinical Haematology* **16**, 369–385 (2003).
40. Gayoso, I. *et al.* Immunosenescence of human natural killer cells. *J Innate Immun* **3**, 337–343 (2011).
41. Fujimoto, M. & Sato, S. B cell signaling and autoimmune diseases: CD19/CD22 loop as a B cell signaling device to regulate the balance of autoimmunity. *J Dermatol Sci* **46**, 1–9 (2007).
42. Mafi, P., Hindocha, S., Mafi, R., Griffin, M. & Khan, W. S. Adult mesenchymal stem cells and cell surface characterization - a systematic review of the literature. *The Open Orthopaedics Journal* **5**, 253–260 (2011).
43. Han, K. *et al.* Human amnion-derived mesenchymal stem cells are a potential source for uterine stem cell therapy. *Cell Prolif* **41**, 709–725 (2008).
44. Nery, A. A. *et al.* Human mesenchymal stem cells: from immunophenotyping by flow cytometry to clinical applications. *Cytometry Part A* **83**, 48–61 (2013).
45. Kasprzak, A., Surdacka, A., Tomczak, M. & Konkol, M. Role of high endothelial postcapillary venules and selected adhesion molecules in periodontal diseases: a review. *J Periodontol Res* **48**, 1–21 (2013).
46. Elghetany, M. T. & Lacombe, F. Physiologic variations in granulocytic surface antigen expression: impact of age, gender, pregnancy, race, and stress. *J Leukoc Biol* **75**, 157–162 (2004).
47. Crisan, M., Corselli, M., Chen, W. C. W. & Péault, B. Perivascular cells for regenerative medicine. *J Cell Mol Med* **16**, 2851–2860 (2012).
48. Alakel, N. *et al.* Direct contact with mesenchymal stromal cells affects migratory behavior and gene expression profile of CD133<sup>+</sup> hematopoietic stem cells during ex vivo expansion. *Exp Hematol* **37**, 504–513 (2009).
49. Dvorakova, J., Hrubá, A., Velebný, V. & Kubala, L. Isolation and characterization of mesenchymal stem cell population entrapped in bone marrow collection sets. *Cell Biol Int* **32**, 1116–1125 (2008).
50. Mori, T. *et al.* Combination of hTERT and bmi-1, E6, or E7 induces prolongation of the life span of bone marrow stromal cells from an elderly donor without affecting their neurogenic potential. *Mol Cell Biol* **25**, 5183–5195 (2005).

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## Author contributions

K.S. and T.H. contributed to the experimental design, data acquisition, analysis and interpretation, and drafting of the article. M.Y., S.K. and S.O. provided technical support and assisted with the data analysis and interpretation. N.K., H.A., K.M., Y.Y. and A.U. assisted with experimental design as well as data analysis and interpretation. All authors examined the data and approved the final manuscript.

## Additional information

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