



OPEN

## Production and purification of $^{43}\text{Sc}$ and $^{47}\text{Sc}$ from enriched $[^{46}\text{Ti}]\text{TiO}_2$ and $[^{50}\text{Ti}]\text{TiO}_2$ targets

Shelbie J. Cingoranelli<sup>1,2</sup>, Jennifer L. Bartels<sup>2</sup>, Pavithra H. A. Kankanamalage<sup>3</sup>, C. Shaun Loveless<sup>2</sup>, David A. Rotsch<sup>3,4</sup> & Suzanne E. Lapi<sup>1,2</sup>✉

The radioscandium isotopes,  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$ , compose a promising elementally matched theranostic pair that can be used for the development of imaging and therapeutic radiopharmaceuticals with identical structures. This study aimed to investigate the production of high radionuclidic purity  $^{43}\text{Sc}$  from enriched  $[^{46}\text{Ti}]\text{TiO}_2$  targets and  $^{47}\text{Sc}$  from enriched  $[^{50}\text{Ti}]\text{TiO}_2$  targets and establish a target recycling technique. Enriched  $[^{46}\text{Ti}]\text{TiO}_2$  targets were irradiated with 18 MeV protons, and enriched  $[^{50}\text{Ti}]\text{TiO}_2$  targets were bombarded with 24 MeV protons.  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  were purified using ion chromatography attaining recovery yields of  $91.7 \pm 7.4\%$  and  $89.9 \pm 3.9\%$ , respectively. The average radionuclidic purity for  $^{43}\text{Sc}$  was  $98.8 \pm 0.3\%$  and for  $^{47}\text{Sc}$   $91.5 \pm 0.6\%$ , while the average recovery of enriched titanium target material was  $96 \pm 4.0\%$ . The highest apparent molar activity for  $[^{43}\text{Sc}]\text{Sc-DOTA}$  was  $23.2 \text{ GBq}/\mu\text{mol}$  and  $3.39 \text{ GBq}/\mu\text{mol}$  for  $[^{47}\text{Sc}]\text{Sc-DOTA}$ . This work demonstrates the feasibility of using enriched recycled  $[^{46}\text{Ti}]\text{TiO}_2$  and  $[^{50}\text{Ti}]\text{TiO}_2$  targets to produce high purity  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  as an elementally matched theranostic isotope pair.

Theranostic techniques empower physicians to make decisions for targeted therapy based on imaging obtained using a similar or identical construct<sup>1–4</sup>. This approach involves the development of two chemically similar or identical radiopharmaceuticals, each serving a distinct purpose: one as a diagnostic agent and the other as a therapeutic agent. The key distinction lies in the incorporation of different radioisotopes. Diagnostic radiopharmaceuticals utilize radioisotopes with suitable decay properties, emitting detectable photons of energy that can escape the patient whereas therapeutic radiopharmaceuticals employ radioisotopes that emit  $\beta^-$ , Auger electrons, or  $\alpha$  particles, capable of causing DNA strand breaks.

Theranostic strategies leverage these distinct decay properties to create chemically similar drugs with different purposes<sup>1</sup>. The theranostic pairing in several FDA-approved radiopharmaceuticals employs  $^{68}\text{Ga}$  and  $^{177}\text{Lu}$  for targeted imaging and therapy in prostate and neuroendocrine cancers and has generated an increased interest in this technique. A disadvantage of using this pair is that  $^{68}\text{Ga}$  and  $^{177}\text{Lu}$  are chemically different, which may result in different pharmacokinetics of the resulting radiopharmaceuticals<sup>2–7</sup>. The ideal theranostic pair would include radioisotopes of the same element but with different emissions (i.e., one suitable for diagnosis and the other for therapy).

The radioisotopes of scandium can provide a true elementally matched pair for theranostic use<sup>8,9</sup>. Two positron ( $\beta^+$ ) emitting radioscandium isotopes,  $^{43}\text{Sc}$  ( $\beta^+ = 476 \text{ keV}$ ;  $t_{1/2} = 3.89 \text{ h}$ ) and  $^{44}\text{Sc}$  ( $\beta^+ = 632 \text{ keV}$ ,  $t_{1/2} = 3.97 \text{ h}$ ), are suitable for Positron Emission Tomography (PET) imaging of small molecules and peptides. These positron-emitting radioscandium nuclides serve as chemically identical radiopharmaceuticals to the ones radiolabeled with therapeutic  $^{47}\text{Sc}$ . The therapeutic  $^{47}\text{Sc}$  ( $\beta^- = 160 \text{ keV}$ ;  $t_{1/2} = 3.34 \text{ days}$ ) can also be employed in Single Photon Emission Computed Tomography (SPECT) due to the emission of a 159 keV  $\gamma$ -ray, which can be used to estimate dosimetry and visualize the biodistribution using SPECT imaging<sup>10</sup>. One of the main advantages of radioscandium nuclides is their ability to create a true elementally matched theranostic pair. This stems from their shared chemistry, including separation, purification, and radiolabeling conditions, resulting in identical in vivo kinetics and dynamics of these compounds. Utilizing the same element for either diagnostic ( $^{43}\text{Sc}$  and  $^{44}\text{Sc}$ ) or therapeutic ( $^{47}\text{Sc}$ ) compounds presents a significant advantage over the currently used  $^{68}\text{Ga}/^{177}\text{Lu}$  pair. Table S1 summarizes the relevant characteristics of  $^{68}\text{Ga}$ ,  $^{177}\text{Lu}$ , and  $^{43,44,47}\text{Sc}$ .

<sup>1</sup>Department of Chemistry, University of Alabama at Birmingham, 1924 6th Ave. S., WTI 310F, Birmingham, AL 35244, USA. <sup>2</sup>Department of Radiology, University of Alabama at Birmingham, Birmingham, USA. <sup>3</sup>Physics Division, Argonne National Laboratory, Lemont, USA. <sup>4</sup>Radioisotope Science and Technology Division, Oak Ridge National Laboratory, Oak Ridge, USA. ✉email: lapi@uab.edu

Recent literature has shown an increased interest in the production and purification of radioscandium isotopes<sup>5,11</sup>. Several production routes are being investigated, including proton, deuteron, alpha, neutron, and photon bombardment using calcium (Ca) and titanium (Ti) target material<sup>12–16</sup>. However, a significant challenge with these production routes is the co-production of other radioscandium isotopes from the multiple stable isotopes of Ca and Ti, with the long-lived <sup>46</sup>Sc being the primary contaminant of concern<sup>17</sup>.

Five naturally occurring stable isotopes of Ti can be exploited to produce radioscandium from proton irradiation. The four main proton-induced reactions to produce medically interesting radioscandium isotopes are <sup>46</sup>Ti(p,α)<sup>43</sup>Sc, <sup>47</sup>Ti(p,α)<sup>44</sup>Sc, <sup>48</sup>Ti(p,2p)<sup>47</sup>Sc, and <sup>50</sup>Ti(p,α)<sup>47</sup>Sc. A challenge behind these production routes is the natural abundances of Ti isotopes (<sup>46</sup>Ti: 8.25%; <sup>47</sup>Ti: 7.44%; <sup>48</sup>Ti: 73.72%; <sup>49</sup>Ti: 5.41%; <sup>50</sup>Ti: 5.18%) that lead to the co-production of multiple radioscandium nuclides when non-enriched material is used. Table S2 contains all the radioscandium nuclides produced via irradiation of natural abundance titanium targets, their respected nuclear reactions, half-lives, decay mode and characteristic γ-gamma rays used for their identification and activity quantification. Enriched material will result in lower production of the undesired radioscandium nuclides<sup>17–19</sup>. The impurities listed in Table S2 serve as a reminder of the challenges in achieving high purity radioisotopes, which can be partially addressed using enriched material. Target material that is highly enriched in one titanium isotope is a viable option, as the lower abundance of the titanium isotopes will decrease the production of the radioisotopic impurities.

This study aimed to investigate the production of <sup>43</sup>Sc from enriched [<sup>46</sup>Ti]TiO<sub>2</sub> and <sup>47</sup>Sc from enriched [<sup>50</sup>Ti]TiO<sub>2</sub> targets and to develop a reliable recycling method for the target material to offset the enriched target material cost. Quantification of yields and radionuclide purification utilized High Purity Germanium (HPGe) analysis and trace metal analysis using Inductively Coupled Plasma-Mass Spectrometry (ICP-MS). The quality of the products was further examined using a 1,4,7,10-tetraazacyclododecane-1,4,7,10-tetraacetic acid (DOTA) titration. PET imaging of a <sup>43</sup>Sc phantom was also carried out.

## Materials and methods

### Reagents

All chemicals used were analytical or trace metal grade unless otherwise stated. Water was obtained from a deionized 18.2 MΩ-cm Milli-Q System (Millipore, Billerica, MA). Hydrochloric acid (HCl, 37% by weight, 99.999%), ammonium bifluoride (NH<sub>4</sub>HF<sub>2</sub>, 99.999%), and 28% ammonia solution (NH<sub>4</sub>OH, 99.999%) were purchased from Sigma-Aldrich (St. Louis, MO). Nitric acid (HNO<sub>3</sub>, 67–70% by weight, 99.999%) was purchased from Fisher Scientific (Hampton, NH, USA). Enriched <sup>46</sup>Ti and <sup>50</sup>Ti were provided by the National Isotope Development Center, USA. The isotopic percentage, provided by the National Isotope Development Center, is given in Table S3 in supplemental information and the Certificates of Analysis in Table S4. The titanium cover foil with 4N5 purity and tantalum coin backing with 3N8 purity were purchased from ESPI Metals (Ashland, OR, USA). All periodic table analytical standards (mix 101, 103, 104, 5% HNO<sub>3</sub>, 10 mg/L) and SG-iTLC plates were purchased from Agilent Technologies (Santa Clara, CA, USA). Analytical-grade *N,N,N',N'*-tetra-2-ethylhexyldiglycolamide (branched DGA) resin was purchased from Eichrom Technologies (Lisle, IL) and empty 1 mL SPE fritted columns, 0.2 μm additional frits, and column adaptors were purchased from Sigma-Aldrich (Supelco, Sigma-Aldrich, St. Louis, MO). DOTA was purchased from Macrocylics (Plano, TX, USA). The Micro Deluxe Phantom™ with the Micro Deluxe Cold Rod Insert™ was purchased from Data Spectrum Corporation (Durham, NC, USA). Conical 15 mL screwcap perfluoroalkoxy alkane (PFA) digestion vessel, 6 mL Octagonal Body Vial, and PFA 10 mL volumetric flasks were purchased from Savillex (Eden Prairie, MN). Millipore Sigma MF-Millipore Mixed Cellulose Ester Membranes with 0.22 μm pore size were purchased from Fisher Scientific (Hampton, NH, USA). The <sup>68</sup>Ga was eluted from Eckert and Ziegler <sup>68</sup>Ge/<sup>68</sup>Ga generator using 0.1 M HCl and was concentrated on an Agilent Bond Elute SCX cartridge, where <sup>68</sup>Ga was recollected in 250 μL 5 M NaCl/0.1 M HCl. All glassware was cleaned in a 20% HNO<sub>3</sub> bath overnight before use.

### Target preparation and irradiation conditions

The 2-mm thick tantalum (Ta) target coins and target material preparation were carried out as previously reported by Loveless et al.<sup>17</sup>. Briefly, the TiO<sub>2</sub> material was kept at 250 °C in an oven for at least 24 h before bombardment. Approximately 100 mg targets were pressed in a FTIR pellet 10-mm evacuable die (Specac, Kent, UK) by increasing the applied pressure in 1-ton increments per minute up to 5 tons, then held at 5 tons for 15 min using a Carver model 3664 hydraulic press (Carver, INC, Wabash, IN, USA). The pellet was removed and placed into the divot of a 2 mm Ta coin. Targets were irradiated on a TR-24 cyclotron (Advanced Cyclotron Systems, Inc., Richmond, BC, Canada) using a 90° coin target holder (Advanced Cyclotron Systems, Inc., Richmond, BC, Canada)<sup>20</sup>. For <sup>43</sup>Sc production using [<sup>46</sup>Ti]TiO<sub>2</sub>, targets were bombarded with 18 MeV protons at 20 μA for 1.5 h. For <sup>47</sup>Sc production using [<sup>50</sup>Ti]TiO<sub>2</sub>, targets were bombarded with 24 MeV protons at 20 μA for 4 or 8 h. The theoretical predicted activity was calculated based on the cross section measurements retrieved from EXFOR shown in Fig. S1<sup>21–23</sup>. Cross sections for additional co-produced radiocontaminants are presented in the supplemental information Fig. S2.

### Target digestion and purification

#### *Digestion of TiO<sub>2</sub>*

The irradiated target material was removed from the Ta coin and added to a 15-mL screwcap PFA digestion vial with 300 mg of NH<sub>4</sub>HF<sub>2</sub>. The vial was capped and placed into a furnace at 250 °C for 2 h. After digestion, the vial was removed and allowed to cool before adding 5 mL of concentrated HCl. The vial was heated in a silicone oil bath at 160 °C for 45 min. The dissolved solution was transferred into a 10 mL PFA volumetric flask and diluted

to 10 mL, using 1 mL additions of 9 M HCl, rinsing the digestion vial in the process for a final concentration of 10.5 M HCl.

#### Purification

Approximately 150 mg of branched DGA resin was added to an empty SPE column with frits on either side of the resin bed. The column was conditioned with a syringe pump at 2 mL/min with the following solutions: 20 mL of 7 M HNO<sub>3</sub>, 20 mL of 1 M HNO<sub>3</sub>, 20 mL of Milli-Q water, 20 mL of 0.1 M HCl and 20 mL of 9 M HCl. Air was pushed through after each solution. Following column conditioning, the dissolved target was loaded onto the column through a 10 mL syringe, which was either pushed manually or by using a syringe pump with the flow rate set to 2 mL/min (collected in Flow-Through (FT) tube). The column was then washed with 20 mL of 9 M HCl (Elution 1 (E1)), followed by 10 mL of 7 M HNO<sub>3</sub> (Elution 2 (E2)), 3 mL of 1 M HNO<sub>3</sub> (Elution 3 (E3)), and three separate, 3 mL additions of 0.1 M HCl (Elution 4-6 (E4-6)) as shown in Fig. S3. Each of the eluents was collected in individual Falcon tubes. The E4 fraction was evaporated to dryness using a Smart Evaporator (BioChromato) before being reconstituted with 200 µL 0.1 M HCl and used for further studies. The evaporation conditions used a 5 mL PFA vial, heated at 100 °C in an Al bead bath under vacuum without the use of N<sub>2</sub>.

#### Target material recycling

The FT and E1 fractions were combined and diluted to 500 mL with Milli-Q water in a 1 L glass beaker. The beaker was wrapped with plastic wrap and the solution was heated to 80 °C for 1 h and then allowed to cool. While stirring, the solution was adjusted to pH 8 with 1 mL additions of 28% ammonium hydroxide, until the TiO<sub>2</sub> precipitated as a white cloudy solution. The solution/precipitate was left to settle overnight at room temperature and then vacuum filtered using a 0.22 µm mixed cellulose filter paper on a low vacuum. After the filter paper was dried, the TiO<sub>2</sub> was collected into a clean, dry beaker and placed into a vacuum furnace at 250 °C for at least 24 h before the next bombardment.

#### Gamma-ray spectroscopy

Gamma-ray spectroscopy was used for isotope identification, to determine radionuclide yields and radionuclidic purity using a Canberra GC2018 High Purity Germanium detector (HPGe), interfaced with a DSA = 100 multichannel analyzer (Meriden, CT, USA). Data acquisition and analysis were performed using Genie 2000 software (Canberra). A 1.5 mL microcentrifuge mixed, sealed, source, 8303-EG-SD (Eckert and Ziegler Analyticals, Atlanta, GA, USA) was used for energy and efficiency calibration. The radionuclides within the source were <sup>57</sup>Co(1.23 MBq), <sup>60</sup>Co(2.50 MBq), <sup>88</sup>Y(4.86 MBq), <sup>85</sup>Sr(3.61 MBq), <sup>109</sup>Cd (5.41 MBq), <sup>113</sup>Sn(3.09 MBq), <sup>137</sup>Cs(1.31 MBq), and <sup>139</sup>Ce(1.83 MBq). One and 10 µL aliquots of samples were diluted to 1 mL with MilliQ water and were analyzed at distances of 5 mm or 25 mm from the bottom of the tube to the face of the HPGe detector. Equations (1) and (2) below were used to determine end-of-bombardment yields:

$$A_1 = \frac{(N_p * \lambda)}{(\epsilon)(1 - e^{-\lambda t_1})(1 - D_r)(I_\gamma)} \quad (1)$$

$$A_o = \frac{A_1}{e^{-\lambda t_o}} \quad (2)$$

where A<sub>1</sub> is the activity calculated from the HPGe acquisition, N<sub>p</sub> is the net peak area in each photopeak, λ is the decay constant of radioisotope of interest, ε is the detector efficiency of the photopeak, t<sub>1</sub> is the real-time of HPGe acquisition, D<sub>r</sub> is the average dead time of the instrument, I<sub>γ</sub> is the branching ratio of the γ-ray of the radioisotope, A<sub>o</sub> is the activity at the end of bombardment, and t<sub>o</sub> is the time passed between the end of bombardment and the time of acquisition<sup>24</sup>. Measurements were taken for a minimum of 500 counts under each photopeak and a deadtime no larger than 5%.

#### Inductively coupled plasma mass spectrometry (ICP-MS)

Elemental analysis was performed on Agilent Technologies 7800 ICP-MS (Santa Clara, CA, USA) with Agilent software, ICP-MS MassHunter v4.3. A 20 µL aliquot from each fraction from the separation was diluted to 10 mL with 2% HNO<sub>3</sub> to determine the elements present, in triplicate. The dissolved target solution and the flow-through collection had a secondary dilution step where 400 µL of the first ICP-MS sample was diluted to 10 mL in 2% HNO<sub>3</sub>. Multi-element standards in 2% HNO<sub>3</sub> were used for calibration of 0.1, 0.5, 1, 5, 10, 100, 200, 400, 600, 800, and 1000 ppb for the calibration curve and prepared in 10 mL volume from a 10 µg/mL stock. Elements selected for monitoring are given in the supplemental information Table S5.

#### Apparent molar activity

To determine apparent molar activity (AMA), a DOTA titration was performed using <sup>43</sup>Sc or <sup>47</sup>Sc, after which, the half-maximum effective concentration (EC<sub>50</sub>) for complete complexation was determined by taking the best-fit values of a transform of the log(µmol) versus percent radiolabeled, performed using Prism 8 software. Then, the average activity added was divided by EC<sub>50</sub> multiplied by 2, shown in Eq. (3) below.

$$AMA = \frac{\text{Average activity}}{(EC_{50} \times 2)} \quad (3)$$

Details of the titration are provided in the supplemental information; section i of the supplemental and Table S6. Analysis of the samples was performed via instant thin-layer chromatography by spotting 1  $\mu\text{L}$  of sample on an iTLC-SG paper and developing in 1 M citrate buffer. An Eckert & Ziegler AR2000 TLC scanner (Berlin, Germany) was used for TLC analysis.

### Phantom imaging

A phantom composed of poly(methyl methacrylate) with rod diameters between 1.2 and 4.8 mm was used for imaging studies. Complete details of the phantom dimensions are provided in supplemental information Fig. S5. The preparation process for the phantoms containing  $^{18}\text{F}$ ,  $^{43}\text{Sc}$ , or  $^{68}\text{Ga}$  involved diluting each radioactive solution to 20 mL with Milli-Q water (MQ). Subsequently, 18 mL of the diluted solution was added to the phantoms to achieve a radioactivity of  $3.72 \pm 0.06$  MBq ( $100.6 \pm 1.7$   $\mu\text{Ci}$ ). The phantoms were scanned for 30 min on the UAB small animal GNEXT PET/CT scanner (GNEXT PET/CT, Sofie Biosciences, CA, USA), with an energy window of 350–650 keV, followed by a 5-min CT scan at a voltage 80 kVp, current 150  $\mu\text{A}$ , and 720 projections. Images were reconstructed using a 3D-OSEM (Ordered Subset Expectation Maximization) algorithm (24 subsets and 3 iterations, with random, attenuation, and decay correction). Additionally, the  $^{18}\text{F}$  and  $^{43}\text{Sc}$  were reconstructed with the same number of total decays as the  $^{68}\text{Ga}$  scan (1693 s for  $^{18}\text{F}$  and 1614 s for  $^{43}\text{Sc}$ ). The images were processed using the description by Bunka et al.<sup>25</sup>. Image analysis was performed using VivoQuant software (VivoQuant 4.0, Invicro Imaging Service and Software, Boston USA), where one representative transversal section was used and analyzed at the different depths of the phantom. The resulting intensity plots of each phantom were exported to Origin<sup>®</sup> 2022 (OriginLab), where the full-width at half-maximum (FWHM) was processed for each slice, and an arithmetic mean and standard deviation were obtained<sup>25,26</sup>. All three radionuclides' FWHM were compared for all visibly distinguishable rods. The  $^{43}\text{Sc}$  FWHM were statistically compared to  $^{18}\text{F}$  and  $^{68}\text{Ga}$  FWHM using either one-way ANOVA, on rods 4.8, 4.0 and 3.2 mm, or to  $^{18}\text{F}$  only using t-test statistics for 2.4 and 1.6 mm rods.

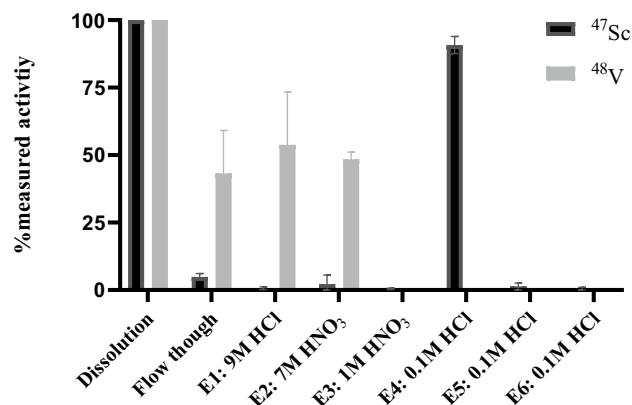
## Results

### Digestion and separation yields

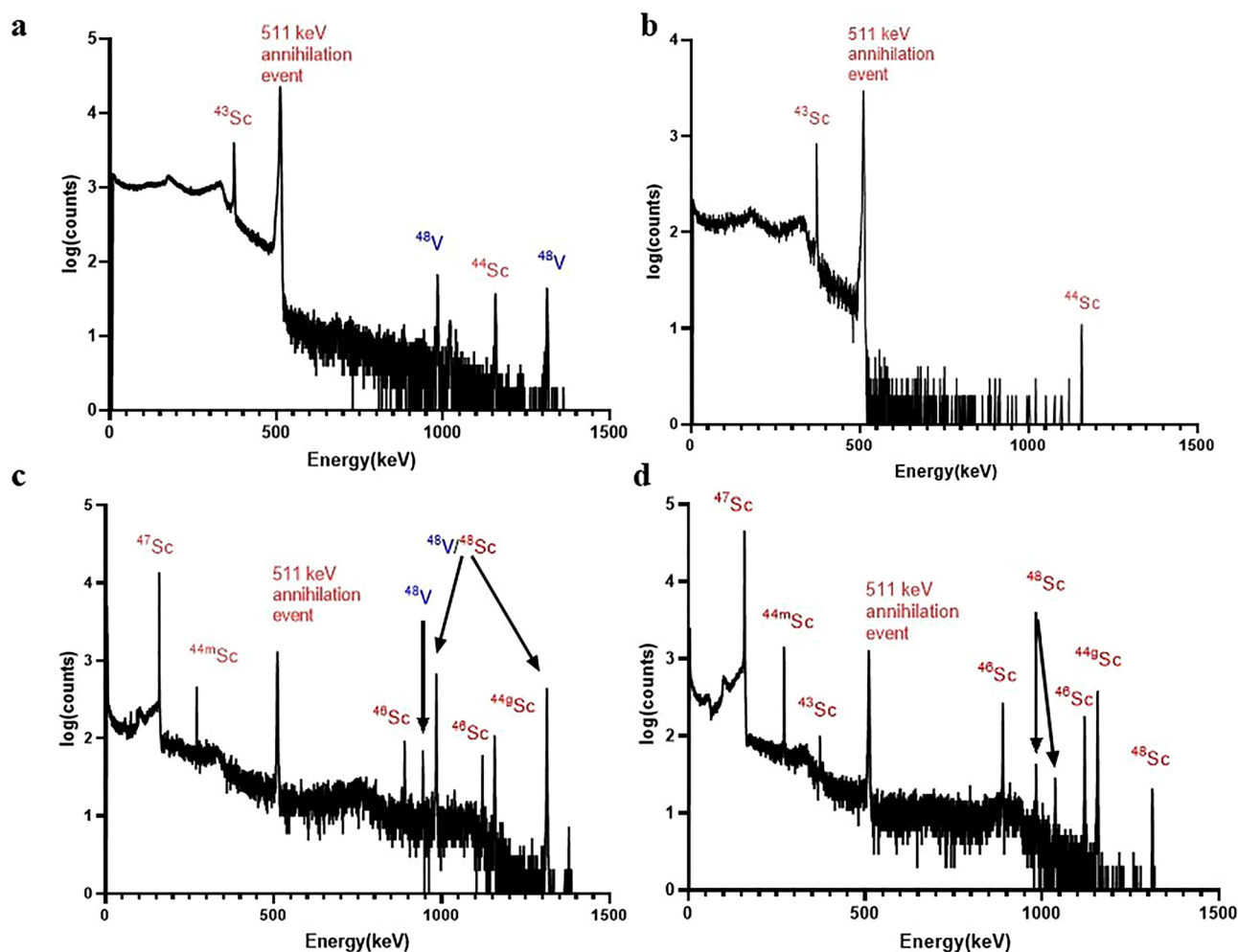
Targets were readily dissolved in a pressurized PFA digestion vial before isolating the radios scandium nuclides from co-produced radiovanadium and the titanium starting material. The two-step dissolution process took a total of 3 h. A representative separation profile for  $^{47}\text{Sc}$  is depicted in Fig. 1 and supplemental information Table S7 show the results of the separation, where the total average  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  recovery yields were  $91.7 \pm 7.4\%$  and  $89.9 \pm 3.9\%$ , respectively, with the majority in the E4 fraction. The co-produced  $^{48}\text{V}$  was removed before the elution of the  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  as shown in the gamma ray spectra in Fig. 2. The decay corrected activity at the end of bombardment for a 1.5 h run using  $^{46}\text{Ti}$  and a 4 h run using  $^{50}\text{Ti}$  was 510 MBq (13.8 mCi) for  $^{43}\text{Sc}$  and 52.17 MBq (1.42 mCi) for  $^{47}\text{Sc}$  (supplemental information Table S7).

The radionuclides were measured by gamma-ray spectroscopy using their characteristic  $\gamma$ -rays presented previously in Table S2. The identification of the radionuclides is shown in the HPGe spectra in Fig. 2. Each photopeak is labeled with their corresponding radionuclide. The spectra shown in panels a and c in Fig. 2 represent the dissolved target solution of either production whereas panels b and d are for the purified material.

HPGe analysis was used to determine the final activity and the radionuclidic purity of  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  decay corrected to end of bombardment. Table 1 is the average activity produced of all radios scandium nuclides and the radionuclidic purity for  $[^{46}\text{Ti}]\text{TiO}_2$  and  $[^{50}\text{Ti}]\text{TiO}_2$  bombardments, decay corrected to end of bombardment. The quantification of the longer-lived  $^{46}\text{Sc}$ ,  $^{47}\text{Sc}$ ,  $^{48}\text{Sc}$  were measured at later time points ( $\geq 1$  day for  $^{47}\text{Sc}$  and  $^{48}\text{Sc}$  and  $\geq 4$  weeks for  $^{46}\text{Sc}$ ), after the shorter lived radioisotopes ( $^{43}\text{Sc}$  and  $^{44}\text{Sc}$ ) have decayed. The average radionuclidic purity for  $^{43}\text{Sc}$  from enriched targets was  $98.8 \pm 0.3\%$ . The average radionuclidic purity for  $^{47}\text{Sc}$  from enriched targets was  $91.5 \pm 0.6\%$ . ICP-MS was conducted to assess the elution and presence of trace metal



**Figure 1.** The elution profile of the  $^{47}\text{Sc}$  separation from enriched  $[^{50}\text{Ti}]\text{TiO}_2$  targets, shown as a percentage of activity in the eluted fraction in comparison to the total starting activity. The fractions representing the  $^{47}\text{Sc}$  are black and the fractions representing the  $^{48}\text{V}$  are represented in solid gray.



**Figure 2.** The gamma-ray spectra of the dissolved irradiated  $[^{46}\text{Ti}]\text{TiO}_2$  target (a). The gamma-ray spectra of the purified  $^{43}\text{Sc}$  elution (b). The gamma-ray spectra of the dissolved irradiated  $[^{50}\text{Ti}]\text{TiO}_2$  target (c). The gamma-ray spectra of the purified  $^{47}\text{Sc}$  elution (d). Each photopeak is labeled with the corresponding radionuclide.

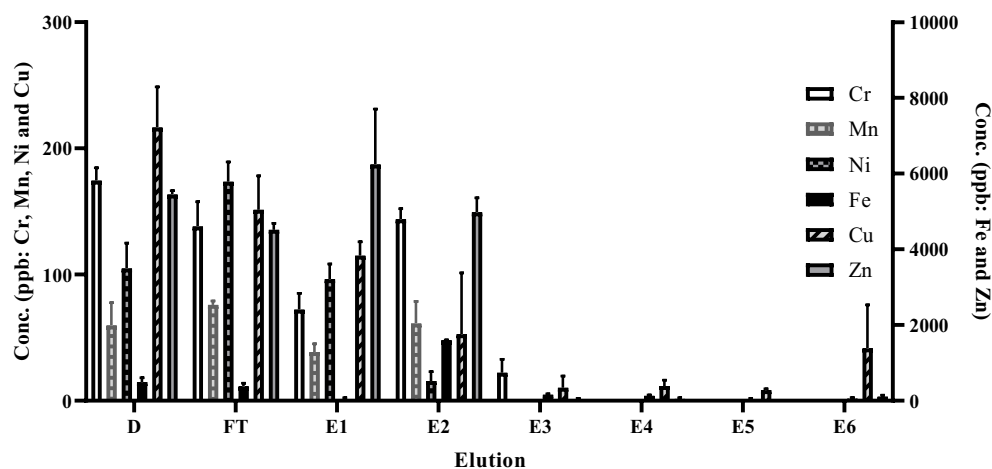
Activity corrected to end of bombardment for the collected $^{43}\text{Sc}$ : Fig. 2 (n = 3)							
Isotope	$^{43}\text{Sc}$	$^{44g}\text{Sc}$	$^{44m}\text{Sc}$	$^{46}\text{Sc}$	$^{47}\text{Sc}$	$^{48}\text{Sc}$	Total radioscandium
Activity MBq (mCi)	$510 \pm 22$ ( $13.7 \pm 0.7$ )	$5.19 \pm 0.30$ ( $0.14 \pm 0.01$ )	$0.11 \pm 0.01$ ( $< 0.01$ )	$0.02 \pm 0.01$ ( $< 0.01$ )	$0.35 \pm 0.04$ ( $0.01 \pm 0.01$ )	$< 0.01 \pm 0.01$ ( $< 0.01$ )	$517.6 \pm 25.5$ ( $13.9 \pm 0.7$ )
Percentage	$98.8 \pm 0.3$	$1.01 \pm 0.1$	$0.02 \pm < 0.01$	$< 0.01$	$0.07 \pm < 0.01$	$< 0.01$	100
Activity corrected to end of bombardment for the collected $^{47}\text{Sc}$ : Fig. 2 (n = 3)							
Isotope	$^{43}\text{Sc}$	$^{44g}\text{Sc}$	$^{44m}\text{Sc}$	$^{46}\text{Sc}$	$^{47}\text{Sc}$	$^{48}\text{Sc}$	Total radioscandium
Activity MBq (mCi)	Decayed	$1.63 \pm 0.3$ ( $0.04 \pm 0.01$ )	$1.63 \pm 0.3$ ( $0.04 \pm 0.01$ )	$1.33 \pm 0.05$ ( $0.04 \pm 0.01$ )	$52.2 \pm 2.3$ ( $1.4 \pm 0.17$ )	$0.24 \pm 0.08$ ( $< 0.01$ )	$57.4 \pm 6.3$ ( $1.5 \pm 0.1$ )
Percentage	0	$2.8 \pm 0.6$	$2.8 \pm 0.6$	$2.3 \pm 0.4$	$91.5 \pm 0.6$	$0.42 \pm 0.6$	100

**Table 1.** The calculated radionuclide purity of produced  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  at EOB.

contaminants including Cr, Mn, Ni, Fe, Cu, W, and Zn throughout the separation with results shown in Fig. 3. The following elements are shown to be removed before the elution of  $^{47}\text{Sc}$ : Cr, Mn, and Ni. Fe, Cu, W and Zn are shown to be significantly reduced before the product collection,  $< 15$  ppb. All other elements measured were below the limit of detection ( $< 15$  ppb for E3-6 and  $< 50$  ppb for other eluted fraction). The elements monitored during analysis were based on the Certificate of Analysis in supplemental information Table S4.

Additionally, continued recycling of the target resulted in improved purity as both rinse fractions E2 and E3 were not included during the recycling process, therefore any impurities in those elutions were not reintroduced into the next cycle. This is shown in Table 2, where the trace metal analysis of purified  $^{47}\text{Sc}$  is compared to





**Figure 3.** ICP-MS elemental analysis results for detectable trace metal contaminants found in the corresponding fractions (x-axis) during the purification of  $^{47}\text{Sc}$ . The element concentration range on the left y-axis is for Cr, Mn, Ni, and Cu. The element concentration range on the right y-axis is for Fe and Zn.

experiments where multiple-times recycled target material was used. All other elements monitored were below the limit of detection.

### Recycling yields

The average recovery from the recycling process was  $96 \pm 4\%$  ( $n = 5$ ), showing a high recovery of the target material. The recovery over several run cycles is presented in supplemental information Fig. S6 for  $[^{46}\text{Ti}]\text{Ti}$  and  $[^{50}\text{Ti}]\text{Ti}$  by ICP-MS analysis. Furthermore, Table 3 demonstrates the high target material recovery as the production yields from the same recycled targets, either  $[^{46}\text{Ti}]\text{TiO}_2$  and  $[^{50}\text{Ti}]\text{TiO}_2$ , remain consistent.

### Apparent molar activity

The radioisotopes  $^{43}\text{Sc}$  or  $^{47}\text{Sc}$  were used to radiolabel DOTA at various concentrations with the % complex versus DOTA concentration represented in Table S6. An scan of an iTLC of a 100% radiochemical yield of  $[^{47}\text{Sc}]$

Element	Cycle 1 (ppb)	S.D.	Cycle 4 (ppb)	S.D.	Cycle 6 (ppb)	S.D.
Cr	311	33.1	< 15 <sup>a</sup>	–	< 15 <sup>a</sup>	–
Mn	340	48.6	< 15 <sup>a</sup>	–	< 15 <sup>a</sup>	–
Fe	1433	130	194	25.4	< 15 <sup>a</sup>	–
Ni	190	14.9	< 15 <sup>a</sup>	–	< 15 <sup>a</sup>	–
Cu	264	10.1	11.6	14.6	< 15 <sup>a</sup>	–
Zn	431	25.1	47.6	28.6	27.8	12.5
W	10,171	37.9	53.1	44.5	31.6	1.81
Pb	2172	216	< 15 <sup>a</sup>	–	< 15 <sup>a</sup>	–

**Table 2.** Trace metal analysis of collected  $^{47}\text{Sc}$  from recycled  $^{50}\text{Ti}$  targets. S.D., standard deviation. <sup>a</sup>Below the limit of detection.

Production of $^{43}\text{Sc}$ from a single enriched $^{46}\text{Ti}$ target bombarded for 1.5 h			
Activity	Cycle 1	Cycle 2	Cycle 3
MBq	499	540	529
mCi	13.5	14.6	14.3
Production of $^{47}\text{Sc}$ from a single enriched $^{50}\text{Ti}$ target bombarded for 8–9 h			
Activity	Cycle 1 <sup>a</sup>	Cycle 4	Cycle 8
MBq	110	85.1	84.7
mCi	2.97	2.3	2.29

**Table 3.** Production yields from single targets. <sup>a</sup>9 h bombardment.

Sc-DOTA and one scan with free [ $^{47}\text{Sc}$ ]Sc control is shown in Fig. S4. The apparent molar activity curve is shown in Fig. S7. The estimated apparent molar activity (AMA) was improved when the target material from later recycling cycles was used. The  $^{43}\text{Sc}$  AMA was 5.92 GBq/ $\mu\text{mol}$  (160.3 mCi/ $\mu\text{mol}$ ) for the second target cycle and improved to 23.2 GBq/ $\mu\text{mol}$  (628 mCi/ $\mu\text{mol}$ ) for the 6th cycle. The  $^{47}\text{Sc}$  AMA was 1.26 GBq/ $\mu\text{mol}$  (34.0 mCi/ $\mu\text{mol}$ ) for 3rd cycle and was 3.39 GBq/ $\mu\text{mol}$  (91.7 mCi/ $\mu\text{mol}$ ) for 6th cycle.

### Phantom imaging

A phantom comparison was performed for two purposes: as a proof-of-concept comparison to reported literature and to assess the image quality of our produced  $^{43}\text{Sc}$  compared to the clinically used  $^{18}\text{F}$  and  $^{68}\text{Ga}$  and reported literature. All phantoms were prepared and imaged in the same manner, the  $^{18}\text{F}$  and  $^{43}\text{Sc}$  were reconstructed under two parameters: where the first reconstruction included the entire 30 min scan and the second reconstruction used shorter time frame (1693 s for  $^{18}\text{F}$  and 1614 s for  $^{43}\text{Sc}$ ) for the same total decays of either isotope to match the total decays of the 30 min  $^{68}\text{Ga}$  scan. These images are shown in Fig. 4. A qualitative assessment of Fig. 4 suggests that  $^{43}\text{Sc}$  has a favorable imaging quality and slightly improved resolution over  $^{68}\text{Ga}$  for both reconstructions. Table 4 shows the numerical expression of the image difference using FWHM for all visibly distinguishable rods. The resulting FWHM corroborates the qualitative analysis of the resolution quality of these radionuclides, both for phantom scanned for the same time or reconstructed for the same total decays. The quantitative resolution results also illustrates that the order of resolution from highest to lowest is:  $^{18}\text{F} > ^{43}\text{Sc} > ^{68}\text{Ga}$ .

Both  $^{18}\text{F}$  and  $^{43}\text{Sc}$  transversal slice images were acquired from the same position. Both of the  $^{43}\text{Sc}$  scans were significantly different to the  $^{68}\text{Ga}$  scan at the 4.0 mm ( $p = 0.0004$  for full 30 min scan and 0.003 for normalized) and 3.2 mm ( $p = 0.0009$  for full 30 min scan and 0.001 for normalized). Both of the  $^{43}\text{Sc}$  scans were also significantly different from the  $^{18}\text{F}$  scan on the 2.4 mm rod ( $p = 0.034$  for the 30-min scan and  $p = < 0.0001$  for the normalized). All other comparisons were not statistically significant.

### Discussion

The production of elementally matched theranostic pairs is highly sought after but commonly requires the use of expensive enriched material. Developing a target recycling method is often essential to offset the cost of enriched material. The work presented here demonstrates a reproducible purification and recycling route for  $^{43}\text{Sc}$  and  $^{47}\text{Sc}$  production starting from enriched  $^{46}\text{Ti}$  and  $^{50}\text{Ti}$ , respectively.

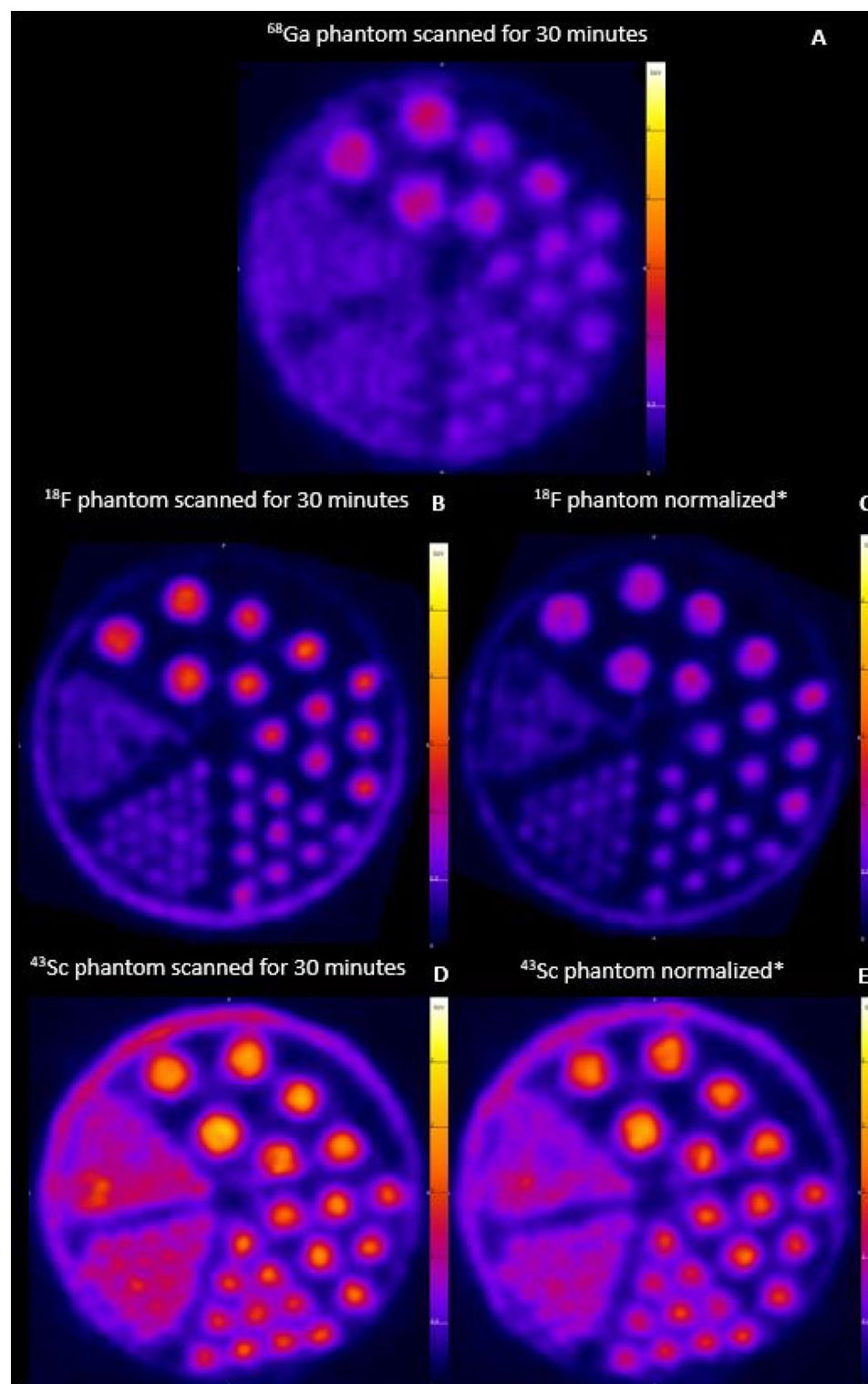
The radioactivity decay corrected to end of bombardment yields presented in Table S5 are in close agreement with the theoretical calculations and recent literature<sup>17,26</sup>. For  $^{43}\text{Sc}$ , 510.7  $\pm$  21.8 MBq (13.8  $\pm$  0.59 mCi) were produced, while the theoretical yield amount to 514.3 MBq (13.9 mCi) for a 1.5 h bombardment. In the case of  $^{47}\text{Sc}$ , 52.17  $\pm$  2.3 MBq (1.3  $\pm$  0.1 mCi) were produced versus 49.6 MBq (1.3 mCi) theoretical for a 4 h bombardment ( $n = 3$  for each). The optimization of the separation proposed by Loveless et al. was achieved by adopting three improvements<sup>17</sup>. First, an additional wash of 3 mL of 1 M  $\text{HNO}_3$  was added, which improved the radioscaandium purification by removing additional impurities (Cr, Fe, Cu and Zn are present in this elution as shown in Fig. 3) and reducing the final elution volume<sup>17,26</sup>. Next, the use of a SPE fritted tube instead of a BioRad column improved the time and reproducibility of the purification<sup>17,26</sup>. The SPE fritted tube columns were adapted for the use of a syringe pump that allowed for constant flow rates, which reduced flow rate variability. Lastly, the presence of HF in the dissolved target solution would decrease the radioscaandium affinity to BDGA resin; thus, the  $\text{TiO}_2$  with  $\text{NH}_4\text{HF}_2$  was heated for 2 h to ensure complete removal of the HF before addition of concentrated  $\text{HCl}$ <sup>24,27</sup>. These improvements allowed for shorter evaporation time as the collected volume was decreased from 10 to 3 mL for either the  $^{43}\text{Sc}$  or  $^{47}\text{Sc}$  while maintaining an effective separation of the radioscaandium from titanium target material and  $^{48}\text{V}$  species<sup>17</sup>.

The irradiated targets for  $^{43}\text{Sc}$  were dissolved 30 min after end of bombardment to allow decay of the short-lived radio contaminants (such as  $^{13}\text{N}$  and  $^{15}\text{O}$ ) to reduce the dose to personnel. The radiochemical purity of  $^{43}\text{Sc}$  was 98.8%, with  $^{44g}\text{Sc}$ , the other PET radionuclide, being the highest impurity at 1.01%. These results are in agreement with the reported radionuclidic purity from Domnanich et al.<sup>26</sup>.

The irradiated targets for  $^{47}\text{Sc}$  were dissolved the following day to allow for the decay of short-lived radiocontaminates and  $^{43}\text{Sc}$  and  $^{44g}\text{Sc}$  produced from other Ti isotopes present in the target material. The advantage of using enriched [ $^{50}\text{Ti}$ ]TiO<sub>2</sub> can be observed in the HPGe analysis as the percent of  $^{47}\text{Sc}$  at end of bombardment is 91.5  $\pm$  0.6% whereas reported radionuclidic  $^{47}\text{Sc}$  purity for  $^{nat}\text{Ti}$  targets, after which  $^{43}\text{Sc}$  and  $^{44g}\text{Sc}$  has substantially decayed, 28 h after end of bombardment, was 43%<sup>17</sup>.

The production of  $^{46}\text{Sc}$ , which is the radioscaandium contaminate of concern as its half-life is 83 days and the longest lived radioscaandium isotope, was determined to be  $< 2\%$  of all total radioscaandium for  $^{47}\text{Sc}$  production, an indication of the efficiency of using enriched target material. The production yields of  $^{46}\text{Sc}$  and other radioscaandium isotopes could be reduced with the use of higher purity enriched [ $^{50}\text{Ti}$ ]TiO<sub>2</sub>. Ideally, the percentages of [ $^{47}\text{Ti}$ ]Ti and [ $^{49}\text{Ti}$ ]Ti should remain consistently low, as both titanium isotopes are routes for  $^{46}\text{Sc}$  production, as shown in supplemental information Fig. S2.

The ICP-MS results indicated the presence of trace metal contaminants within the target material, which is in alignment with the certificate of analysis shown in Table S4, with Cr, Mn at  $< 100$  ppm and Fe, Cu, Zn, W, and Pb having some of the largest starting concentrations. The larger concentrations of Fe, Zn, and W are of concern as they will likely compete with radioscaandium for complexation sites<sup>28–30</sup>. The separation is shown to be effective at removing these metals before eluting the desired radioscaandium, as seen in Fig. 3. The majority of the contaminants were removed with the washes of 9 M  $\text{HCl}$ , 1 M and 7 M  $\text{HNO}_3$ . The overall separation process indicates the removal of the trace metal contaminants and high recovery of Ti species in flow through and E1 via ICP-MS analysis.



**Figure 4.** Phantom PET images of (A)  $^{68}\text{Ga}$ , (B, C)  $^{18}\text{F}$ , and (D, E)  $^{43}\text{Sc}$ . The  $^{68}\text{Ga}$  image contained 3.7 MBq (100  $\mu\text{Ci}$ ), imaged for 30 min for  $5.74\text{E}9$  total decays. The  $^{18}\text{F}$  and  $^{43}\text{Sc}$  phantoms contained  $3.7 \pm 0.07$  MBq ( $101 \pm 2$   $\mu\text{Ci}$ ) and imaged for 30 min (B and D) or reconstructed to be normalized to  $^{68}\text{Ga}$  for same total decays (C and E). Images were further processed using Vivoquant (VivoQuant 4.0, Invicro Imaging Service and Software, Boston USA, vivoquant.com).



Radionuclide	E $\beta^+$ average [keV]	Intensity	FWHM for 4.8 mm rod	FWHM for 4.0 mm rod	FWHM for 3.2 mm rod	FWHM for 2.4 mm rod	FWHM for 1.6 mm rod
<sup>18</sup> F (30 min scan)	250	96.7	4.55 $\pm$ 0.21	3.61 $\pm$ 0.18	2.61 $\pm$ 0.01	1.85 $\pm$ 0.01	1.43 $\pm$ 0.03
<sup>18</sup> F (Normalized to <sup>68</sup> Ga) <sup>a</sup>			4.71 $\pm$ 0.05	3.68 $\pm$ 0.08	2.69 $\pm$ 0.07	2.00 $\pm$ 0.05	1.57 $\pm$ 0.04
<sup>43</sup> Sc (30 min scan)	476	88.1	4.40 $\pm$ 0.29	3.57 $\pm$ 0.08	2.76 $\pm$ 0.02	2.34 $\pm$ 0.11	1.53 $\pm$ 0.09
<sup>43</sup> Sc (Normalized to <sup>68</sup> Ga) <sup>a</sup>			4.66 $\pm$ 0.03	3.59 $\pm$ 0.10	2.90 $\pm$ 0.07	2.44 $\pm$ 0.03	1.46 $\pm$ 0.04
<sup>68</sup> Ga (30 min scan)	830	88.9	4.84 $\pm$ 0.48	4.23 $\pm$ 0.10	3.84 $\pm$ 0.22	–	–

**Table 4.** The full-width half maximum for <sup>18</sup>F, <sup>43</sup>Sc and <sup>68</sup>Ga<sup>18</sup>. <sup>a</sup>Reconstructed to match the total decays of the <sup>68</sup>Ga scan (5.74E9 total decays).

The high target recovery yield of 96% provides a steady life cycle of the target material to help balance the cost of the enriched material. The percent recovery is also in accordance with the reported Dommanich et al.<sup>26</sup> target recycling for <sup>nat</sup>TiO<sub>2</sub> (97.6%). Furthermore, the yields of activity produced in subsequent irradiations of the same target remained consistent, validating the constant quality of the target material up to eight target cycles. The target recycling method here also resulted in a purification of the target material. As shown in the trace metal analysis of the purified <sup>47</sup>Sc and the increasing AMA of both <sup>43</sup>Sc and <sup>47</sup>Sc, the target material was gradually purified during each cycle. This procedure shows high reproducibility of the target life cycle, from target collection to <sup>43</sup>Sc and <sup>47</sup>Sc purification and increased purity of the recycled target material after each use.

An additional characterization of the produced <sup>43</sup>Sc and <sup>47</sup>Sc is the measured AMA of the complex formation with DOTA. The separation and target purification from repeated cycles is shown to be effective for the removal of trace contaminants that may compete with <sup>43</sup>Sc or <sup>47</sup>Sc and is shown with the increased AMA after each target recycling<sup>11</sup>.

The PET images of the <sup>18</sup>F, <sup>43</sup>Sc and <sup>68</sup>Ga were employed to validate <sup>43</sup>Sc PET imaging resolution, considering two scenarios: 30 min static scan and scans reconstructed and normalized to the total decays of the <sup>68</sup>Ga scan. The image resolution of <sup>43</sup>Sc was quantified and compared to that of <sup>68</sup>Ga, revealing a smaller FWHM at all visibly distinguishable rods. This suggests a higher resolution, particularly evident at the 3.2 mm rod diameter, which represents the smallest distinguishable rod on the <sup>68</sup>Ga scan. Additionally, the resolution order remained consistent, with <sup>18</sup>F having the highest resolution and the smallest FWHM value at each rod, while <sup>68</sup>Ga exhibited the lowest resolution and largest FWHM values. The significant differences between the FWHM data confirms our hypothesis of decreasing resolution order of <sup>18</sup>F > <sup>43</sup>Sc > <sup>68</sup>Ga. These results are in line with expectations, as higher positron energies result in a decreased resolution, which can already be observed by qualitative evaluation of the PET phantom images. The resulting 1.85 FWHM of <sup>18</sup>F for the 2.4 mm rod is in agreement with literature values of 1.9 mm<sup>31</sup>. Comparing the FWHM of <sup>18</sup>F and <sup>43</sup>Sc on the next smaller rod size also corroborates that <sup>18</sup>F has a higher resolution in comparison to <sup>43</sup>Sc but as this resolution difference is demonstrated using a small animal PET scanner, it may have little impact in clinical practice using human scanners. These results indicate that <sup>43</sup>Sc has favorable characteristics for PET imaging.

Future studies will continue the characterization of the proton induced nuclear reactions on enriched <sup>47</sup>Ti for production of <sup>44</sup>Sc and on enriched <sup>48</sup>Ti for production of <sup>47</sup>Sc. Although there are limited reports on the production of <sup>44</sup>Sc from titanium targets, the major challenge lies in the co-production of the metastable state <sup>44m</sup>Sc (IT: t<sub>1/2</sub>: 58.6 h) during this production process<sup>19</sup>. However, there has been interest in utilizing <sup>44m</sup>Sc as an in vivo generator for longer-lived targeting moieties like antibodies<sup>32–34</sup>.

Additionally, further analysis of the recycled TiO<sub>2</sub> targets and proof-of-concept studies using <sup>43</sup>Sc and <sup>47</sup>Sc as a theranostic matched pair in targeted imaging and treatment will be explored. Further enhancement of the target design to increase production is desirable. The first improvement in the design revolves around augmenting the target thickness, thereby ensuring the capture of the entire excitation function under the energy of 24 MeV. The second design modification involves tailoring the target to accommodate higher energy cyclotrons (30 MeV). Theoretical predictions regarding <sup>47</sup>Sc production, based on the 24 MeV design and the same enriched [<sup>50</sup>Ti]TiO<sub>2</sub> and 4 h bombardment parameters in this work, would yield 5.8 mCi of <sup>47</sup>Sc. The predicted radionuclidic purity for both <sup>46</sup>Sc and <sup>47</sup>Sc is 4.1% and 90.2%, respectively. The 30 MeV target design would increase the <sup>47</sup>Sc yields to 11.6 mCi with a radionuclidic percentages of <sup>46</sup>Sc and <sup>47</sup>Sc at 6.2% and 90.1%, respectively. Although the percentage of <sup>46</sup>Sc increases, it's noteworthy that the <sup>47</sup>Sc yields nearly double while maintaining a comparable level of purity. Investigation of the cross sectional data for the <sup>50</sup>Ti(p,p+ $\alpha$ )<sup>46</sup>Sc at 30 MeV can help guide the ideal bombardment parameters, such as irradiating a 28 or 29 MeV, to decrease the <sup>46</sup>Sc production. It's important to acknowledge that the utilization of higher enrichment levels (> 83%) of [<sup>50</sup>Ti]TiO<sub>2</sub> also holds potential. This is contingent upon the condition that the percentages of <sup>47</sup>Ti and <sup>49</sup>Ti remain equal or lower to reduce <sup>46</sup>Sc and <sup>48</sup>Sc when operating within energies of 30 MeV or below.

## Conclusions

This work demonstrates the feasibility of utilizing enriched [<sup>46</sup>Ti]TiO<sub>2</sub> and [<sup>50</sup>Ti]TiO<sub>2</sub> for the production of high purity <sup>43</sup>Sc and <sup>47</sup>Sc using 18 or 24 MeV protons. The enrichment of 83% [<sup>50</sup>Ti]TiO<sub>2</sub> yielded 91% radionuclidic purity of <sup>47</sup>Sc at the end of bombardment, after the decay of short lived <sup>43,44</sup>Sc while the 96% enriched [<sup>46</sup>Ti]TiO<sub>2</sub> yielded 98% pure <sup>43</sup>Sc at the end of bombardment. The reported separation technique removed radio and trace

metal contaminates that would compete with complexation studies, results of which were verified by HPGe and ICP-MS analysis. The DOTA titration illustrates high molar activity with produced radioscandium that would allow the radiolabeling of other targeting compounds, encompassing a DOTA chelator, followed by subsequent in vivo and in vitro studies. The recycling method resulted in high recovery yields that would compensate for the high cost of target material and demonstrated purification of target material after each cycle.

## Data availability

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

Received: 11 May 2023; Accepted: 7 December 2023

Published online: 19 December 2023

## References

- Langbein, T., Weber, W. A. & Eiber, M. Future of theranostics: An outlook on precision oncology in nuclear medicine. *J. Nucl. Med.* **60**, 13s–19s. <https://doi.org/10.2967/jnumed.118.220566> (2019).
- Hennrich, U. & Kopka, K. Lutathera(®): The first FDA- and EMA-approved radiopharmaceutical for peptide receptor radionuclide therapy. *Pharmaceut. Basel* **12**, 114. <https://doi.org/10.3390/ph12030114> (2019).
- Waldmann, C. M., Stuparu, A. D., van Dam, R. M. & Slavik, R. The Search for an alternative to [(68)Ga]Ga-DOTA-TATE in neuroendocrine tumor theranostics: Current state of (18)F-labeled somatostatin analog development. *Theranostics* **9**, 1336–1347. <https://doi.org/10.7150/thno.31806> (2019).
- Fang, P., Jacobson, M. S. & Hung, J. C. Influential factors in the preparation of (68)Ga-DOTATATE. *J. Nucl. Med. Technol.* **48**, 263–268. <https://doi.org/10.2967/jnmt.119.241224> (2020).
- Misiak, R. *et al.* (47)Sc production development by cyclotron irradiation of (48)Ca. *J. Radioanal. Nucl. Chem.* **313**, 429–434. <https://doi.org/10.1007/s10967-017-5321-z> (2017).
- Deilami-nezhad, L., Moghaddam-Banaem, L., Sadeghi, M. & Asgari, M. Production and purification of Scandium-47: A potential radioisotope for cancer theranostics. *Appl. Radiat. Isotopes* **118**, 124–130. <https://doi.org/10.1016/j.apradiso.2016.09.004> (2016).
- Miller, C. *et al.* Implications of physics, chemistry and biology for dosimetry calculations using theranostic pairs. *Theranostics* **12**, 232–259. <https://doi.org/10.7150/thno.62851> (2022).
- Müller, C. *et al.* Promising prospects for 44Sc-/47Sc-based theragnostics: Application of 47Sc for radionuclide tumor therapy in mice. *J. Nucl. Med.* **55**, 1658–1664. <https://doi.org/10.2967/jnumed.114.141614> (2014).
- Huclier-Markai, S. *et al.* Promising scandium radionuclides for nuclear medicine: A review on the production and chemistry up to in vivo proofs of concept. *Cancer Biother. Radiopharm.* **33**, 316–329. <https://doi.org/10.1089/cbr.2018.2485> (2018).
- Siwowska, K. *et al.* Therapeutic potential of (47)Sc in comparison to (177)Lu and (90)Y: Preclinical investigations. *Pharmaceutics* **11**, 424. <https://doi.org/10.3390/pharmaceutics11080424> (2019).
- Chernysheva, M. *et al.* Accelerator production of scandium radionuclides: Sc-43, Sc-44, and Sc-47. *Curr. Radiopharm.* <https://doi.org/10.2174/1874471014999210112205535> (2021).
- Valdovinos, H. F. *et al.* Separation of cyclotron-produced (44)Sc from a natural calcium target using a dipentyl phosphonate functionalized extraction resin. *Appl. Radiat. Isot.* **95**, 23–29. <https://doi.org/10.1016/j.apradiso.2014.09.020> (2015).
- Duchemin, C., Guertin, A., Haddad, F., Michel, N. & Métivier, V. Production of scandium-44 m and scandium-44 g with deuterons on calcium-44: Cross section measurements and production yield calculations. *Phys. Med. Biol.* **60**, 6847–6864. <https://doi.org/10.1088/0031-9155/60/17/6847> (2015).
- Krajewski, S. *et al.* Cyclotron production of 44Sc for clinical application. *Radiochim. Acta* **101**, 333–338. <https://doi.org/10.1524/ract.2013.2032> (2013).
- Szkliniarz, K. *et al.* Production of medical Sc radionuclides with an alpha particle beam. *Appl. Radiat. Isotopes* **118**, 182–189. <https://doi.org/10.1016/j.apradiso.2016.07.001> (2016).
- Rotsch, D. A. *et al.* Electron linear accelerator production and purification of scandium-47 from titanium dioxide targets. *Appl. Radiat. Isot.* **131**, 77–82. <https://doi.org/10.1016/j.apradiso.2017.11.007> (2018).
- Loveless, C. S. *et al.* Cyclotron production and separation of scandium radionuclides from natural titanium metal and titanium dioxide targets. *J. Nucl. Med.* **62**, 131–136. <https://doi.org/10.2967/jnumed.120.242941> (2021).
- NuDat 2.8. Brookhaven National Laboratory website (2021, accessed 2 Jun 2021). <https://nndc.bnl.gov/nudat2/>.
- Chernysheva, M. *et al.* Accelerator production of scandium radionuclides: Sc-43, Sc-44, and Sc-47. *Curr. Radiopharm.* **14**, 359–373. <https://doi.org/10.2174/1874471014999210112205535> (2021).
- Queern, S. L. *et al.* Production of Zr-89 using sputtered yttrium coin targets. *Nucl. Med. Biol.* **50**, 11–16. <https://doi.org/10.1016/j.nucmedbio.2017.03.004> (2017).
- Levkovski, V. N. Cross sections of medium mass nuclide activation (A=40–100) by medium energy protons and alpha-particles (E=10–50 MeV). *Atomnaya Energ.* **69**, 180 (1991).
- Gadioli, E., Gadioli-Erba, E., Hogan, J. J. & Burns, K. I. Emission of alpha particles in the interaction of 10–85 MeV protons with 48,50Ti. *Zeitschr. Phys. A Atoms Nuclei* **301**, 289–300. <https://doi.org/10.1007/BF01421692> (1981).
- Carzaniga, T. S. *et al.* Measurement of (43)Sc and (44)Sc production cross-section with an 18MeV medical PET cyclotron. *Appl. Radiat. Isot.* **129**, 96–102. <https://doi.org/10.1016/j.apradiso.2017.08.013> (2017).
- Loveless, C. S. *et al.* Photonuclear production, chemistry, and in vitro evaluation of the theranostic radionuclide 47Sc. *EJNMMI Res.* **9**, 42. <https://doi.org/10.1186/s13550-019-0515-8> (2019).
- Bunka, M. *et al.* Imaging quality of (44)Sc in comparison with five other PET radionuclides using Derenzo phantoms and preclinical PET. *Appl. Radiat. Isot.* **110**, 129–133. <https://doi.org/10.1016/j.apradiso.2016.01.006> (2016).
- Domnanich, K. A. *et al.* Production and separation of 43Sc for radiopharmaceutical purposes. *EJNMMI Radiopharm. Chem.* **2**, 14. <https://doi.org/10.1186/s41181-017-0033-9> (2017).
- McLain, D. R., Brossard, T. W., De Kruijff, R., Kankanamalage, P. H. A. & Rotsch, D. A. Evaluation of two extraction chromatography resins for scandium and titanium separation for medical isotope production. *J. Radioanal. Nucl. Chem.* <https://doi.org/10.1007/s10967-023-08783-x> (2023).
- Šimeček, J., Hermann, P., Wester, H.-J. & Notni, J. How is 68Ga labeling of macrocyclic chelators influenced by metal ion contaminants in 68Ge/68Ga generator eluates?. *Chem. Med. Chem.* **8**, 95–103. <https://doi.org/10.1002/cmdc.201200471> (2013).
- Mueller, D. *et al.* Radiolabeling of DOTA-like conjugated peptides with generator-produced (68)Ga and using NaCl-based cationic elution method. *Nat. Protoc.* **11**, 1057–1066. <https://doi.org/10.1038/nprot.2016.060> (2016).
- Walczak, R., Gawęda, W., Dudek, J., Choiński, J. & Bilewicz, A. Influence of metal ions on the 44Sc-labeling of DOTATATE. *J. Radioanal. Nucl. Chem.* **322**, 249–254. <https://doi.org/10.1007/s10967-019-06700-9> (2019).
- Chaple, I. F. *et al.* Optimized methods for production and purification of Titanium-45. *Appl. Radiat. Isotopes* **166**, 109398. <https://doi.org/10.1016/j.apradiso.2020.109398> (2020).

32. Alliot, C., Kerdjoudj, R., Michel, N., Haddad, F. & Huclier-Markai, S. Cyclotron production of high purity 44m,44Sc with deuterons from 44CaCO<sub>3</sub> targets. *Nucl. Med. Biol.* **42**, 524–529. <https://doi.org/10.1016/j.nucmedbio.2015.03.002> (2015).
33. Edem, P. E., Fonslet, J., Kjær, A., Herth, M. & Severin, G. In vivo radionuclide generators for diagnostics and therapy. *Biomorg. Chem. Appl.* **2016**, 6148357. <https://doi.org/10.1155/2016/6148357> (2016).
34. Huclier-Markai, S. *et al.* Optimization of reaction conditions for the radiolabeling of DOTA and DOTA-peptide with 44m/44Sc and experimental evidence of the feasibility of an in vivo PET generator. *Nucl. Med. Biol.* **41**, e36–e43. <https://doi.org/10.1016/j.nucmedbio.2013.11.004> (2014).

## Acknowledgements

This work was supported by the DOE Isotope Program through grant DESC0020197 (PI: Lapi). Imaging studies were supported by the Small Animal Imaging Core through O'Neal Cancer Center P30CA013148. The authors acknowledge financial support from the Alabama Graduate Research Scholars Program (GRSP) funded through the Alabama Commission for Higher Education and administered by the Alabama EPSCoR. The authors would like to acknowledge the UAB Cyclotron Facility team and UAB Machine shop.

## Author contributions

S.J.C. conducted all experiments, wrote the main manuscript, and prepared all Figures and Tables. J.L.B. assisted in experiments, data analysis and provided feedback on editing the manuscript. C.S.L. provided initial radio-chemical separation and targetry, feedback on experimental design and edited the manuscript. P.H.A.K. provided feedback on experimental design and analysis and edited the manuscript. D.A.R. provided feedback on experimental design and analysis and edited the manuscript. S.E.L. provided concepts, feedback on experimental design and analysis, and supervision of this work and edited the manuscript. All authors reviewed the manuscript.

## Competing interests

The authors declare no competing interests.

## Additional information

**Supplementary Information** The online version contains supplementary material available at <https://doi.org/10.1038/s41598-023-49377-7>.

**Correspondence** and requests for materials should be addressed to S.E.L.

**Reprints and permissions information** is available at [www.nature.com/reprints](http://www.nature.com/reprints).

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2023