## SCIENTIFIC REPORTS

natureresearch

### OPEN

Check for updates

# Five serum fatty acids are associated with subclinical hypothyroidism in a Chinese pregnant population

Ting Zhang<sup>1,2,7</sup>, Yinyin Xia<sup>3,7</sup>, Ting-Li Han<sup>1,4,5</sup>, Hua Zhang<sup>1,2</sup> & Philip N. Baker<sup>6</sup>

Subclinical hypothyroidism (SCH) is a common endocrine disorder affecting women of reproductive age. Although SCH and abnormal fatty acid composition are often associated with adverse pregnancy outcomes and metabolic syndrome later in maternal and fetal life, the longitudinal relationship between SCH and serum fatty acids during pregnancy has rarely been studied. Therefore, the aim of this study was to investigate the association between SCH and maternal serum fatty acids throughout gestation. A total of 240 women enrolled in the Complex Lipids in Mothers and Babies (CLIMB) study in Chongqing, China were included in our study. Clinical information and maternal serum samples were collected at three time points during pregnancy: 11–14<sup>th</sup>, 22–28<sup>th</sup>, and 32–34<sup>th</sup> weeks of gestation. Twenty serum fatty acids were quantified using gas chromatography-mass spectrometry (GC-MS) analysis. A majority of the 20 serum fatty acids increased as gestation progressed in women with a normal pregnancy and women experiencing SCH. Levels of arachidic acid, docosahexaenoic acid, and eicosenoic acid were significantly higher in the serum of women with SCH when compared to women with a normal pregnancy, in the second trimester. On the other hand, the levels of eicosadienoic acid and octadecanoic acid were significantly higher in SCH in the third trimester. Our findings demonstrate that serum fatty acid composition during the second and third trimesters was significantly associated with SCH in pregnant Chinese women.

Subclinical hypothyroidism (SCH) is a common endocrine disorder that can manifest in pregnant women. This complication of pregnancy is diagnosed when a mildly elevated serum thyroid-stimulating hormone (TSH) level above the upper limit of the trimester-specific reference is detected in combination with a normal free thyroxine (FT4) concentration during pregnancy<sup>1,2</sup>. SCH occurs in approximately 2–5% of all pregnant women<sup>3–5</sup>. In China, the incidence of SCH in pregnancy has reached epidemic proportions, affecting between 2.8% and 7.2% of all pregnancies<sup>6–8</sup>. SCH during pregnancy can lead to various maternal and fetal complications such as pregnancy loss, preterm birth, placental abruption, severe preeclampsia, gestational diabetes, and impaired offspring neurodevelopment<sup>9–13</sup>.

Several clinical studies have indicated that patients with SCH have altered serum lipid profiles and a higher prevalence of metabolic syndrome<sup>14-16</sup>. Elevated plasma free fatty acid (FFA) levels have been reported in patients with SCH regardless of age, sex, or menopausal status<sup>14,17</sup>. Fluctuations in fatty acid levels also have been associated with metabolic syndromes such as dyslipidemia, type-2 diabetes, and obesity<sup>18</sup>. Fatty acids play a crucial role in the support and regulation of a healthy pregnancy and normal infant development<sup>19</sup>. However, there is a lack of evidence directly relating the changes in serum fatty acids with SCH in pregnancy.

Considering the importance of fatty acids for both maternal and offspring health, as well as an increasing prevalence of SCH during pregnancy, our study aimed to investigate the changes in serum fatty acids in women with SCH, throughout pregnancy. We analyzed 20 fatty acids in maternal serum across the three trimesters of

<sup>1</sup>Department of Obstetrics and Gynaecology, The First Affiliated Hospital of Chongqing Medical University, Chongqing, 400016, China. <sup>2</sup>State Key Laboratory of Maternal and Fetal Medicine of Chongqing Municipality, Chongqing Medical University, Chongqing, 400016, China. <sup>3</sup>School of Public Health and Management, Chongqing Medical University, Chongqing, 400016, China. <sup>4</sup>Institute of Life Sciences, Chongqing Medical University, Chongqing, 400016, China. <sup>5</sup>Liggins Institute, The University of Auckland, Auckland, 1023, New Zealand. <sup>6</sup>College of Medicine, Biological Sciences and Psychology, University of Leicester, Leicester, UK. <sup>7</sup>These authors contributed equally: Ting Zhang and Yinyin Xia. <sup>Ex</sup>e-mail: t.han@auckland.ac.nz; zh2844@gmail.com

	Normal (n=222)	SCH (n = 18)	P value			
Age, years	28 (26, 30)	28 (27, 31)	0.22			
Total years of schooling	16 (15, 16)	16 (15, 16)	0.64			
Gravidity	2 (1, 2)	2 (1, 2)	0.93			
Parity	0 (0, 0)	0 (0, 0)	0.74			
BMI (kg/m <sup>2</sup> )						
1 <sup>st</sup> trimester	20.6 (19.2, 22.4)	20.1 (18.5, 21.9)	0.35			
2 <sup>nd</sup> trimester	22.9 (21.2, 24.8)	22.2 (20.6, 23.7)	0.30			
3 <sup>rd</sup> trimester	24.5 (22.3, 26.2)	23.3 (22.2, 25.7)	0.51			
sBP (mmHg)						
1 <sup>st</sup> trimester	112 (106.3, 121.0)	112 (110.0, 118.5)	0.76			
2 <sup>nd</sup> trimester	116 (110.0, 122.0)	118 (112.0, 122.8)	0.65			
3 <sup>rd</sup> trimester	116 (108.0, 120.0)	120 (110.5, 121.0)	0.30			
dBP (mmHg)						
1 <sup>st</sup> trimester	70 (65.0, 76.0)	74.5 (70.0, 76.0)	0.17			
2 <sup>nd</sup> trimester	71 (68.0, 76.0)	69.5 (66.3, 74.8)	0.29			
3 <sup>rd</sup> trimester	72 (68.0, 76.0)	70.5 (63.5, 77.5)	0.84			
GA at sampling (weeks)						
1 <sup>st</sup> trimester	12.7 (12.1, 13.3)	12.3 (12.1, 13.1)	0.23			
2 <sup>nd</sup> trimester	24.3 (23.7, 24.6)	24.2 (23.9, 24.9)	0.47			
3 <sup>rd</sup> trimester	32.3 (31.7, 32.7)	32.1 (23.9, 24.9)	0.23			
s-TSH (uIU/ml)	1.19 (1.16, 1.39)	3.30 (3.16, 4.62)	1.41E-11***			
FT4 (ng/ml)	0.83 (0.81, 0.86)	0.79 (0.75, 0.85)	0.29			

**Table 1.** Clinical characteristics of the study participants (n = 240). All clinical data are summarized as median(25th percentile, 75th percentile) because each of the variables was not normally distributed. P values werecalculated using a Mann-Whitney test, \*\*\*p < 0.001. Abbreviations: SCH, subclinical hypothyroidism; BMI, body</td>mass index; BP, blood pressure; sBP, systolic blood pressure; dBP, diastolic blood pressure; GA, gestational age.

pregnancy in order to unravel their possible association with SCH for the identification of potential biomarkers for diagnosis or therapeutic intervention.

#### Results

**Clinical characteristics.** All clinical characteristics for the study population are listed in Table 1. Only the TSH levels were significantly higher in SCH women when compared to normal women (3.30 uIU/ml vs. 1.19 uIU/ml; p < 0.001). Other maternal characteristics including age, education, gravidity, parity, BMI, blood pressure, gestational age at sampling and FT4 exhibited no significant differences between case and control groups.

**Differences in the levels of serum fatty acids across trimesters.** A total of 20 serum fatty acids were absolutely quantified in the maternal serum of the participants in their first, second, and third trimesters, as shown in Tables 2–4 and Fig. 1. Fatty acids generally increased in both SCH women and normal women as gestation progressed, particularly from the first to second trimester. However, this trend did not occur between the second and third trimesters. In contrast, eicosapentaenoic acid (EPA) was reduced in both SCH and normal groups from the second to the third trimester, while docosapentenoic acid (DPA) and  $\gamma$ -linolenic acid were only reduced in the normal women from the second to the third trimester.

Differences in the levels of serum fatty acids between women diagnosed with SCH and women with a normal pregnancy. Although the majority of serum fatty acids were detected at higher concentrations in SCH women compared to the normal women, none of the fatty acids in the first trimester significantly discriminated women diagnosed with SCH from normal groups (Table 2). Interestingly, we observed a significantly higher level of three fatty acids (arachidic acid, docosahexaenoic acid (DHA), and eicosenoic acid) in the SCH group in the second trimester, and two fatty acids (eicosadienoic acid and octadecanoic acid) in the third trimester. Among them, three out of the five fatty acids displayed odds ratios and their 95% confidence interval (CI) greater than 1 (Tables 3 and 4); arachidic acid (OR = 1.29, 95% CI 1.01 to 1.65), eicosenoic acid (OR = 1.11, 95% CI 1.01 to 1.21), and eicosadienoic acid (OR = 1.15, 95% CI 1.03 to 1.28). Meanwhile, the odds ratios calculated for DHA (OR = 1.01, 95% CI 1.00 to 1.03) and octadecanoic acid (OR = 1.02, 95% CI 1.00 to 1.03) were very close to 1.0. All five fatty acids had an area under the ROC curve between 0.61–0.68, as illustrated in Fig. 2.

#### Discussion

To our knowledge, this is the first study to investigate the association between serum fatty acids and SCH among Chinese women across the three trimesters of pregnancy. In this study we observed that maternal serum fatty acid levels increased in both SCH and control groups as pregnancy progressed. In addition, we found a significantly higher concentration of three serum fatty acids (arachidic acid, DHA and eicosenoic acid) in women with SCH in

	Fatty acids concentrations (mg/L)				
Fatty acids	Normal (n = 222)	SCH (n = 18)	AUC	OR (95%CI)	P value
Arachidic acid	6.14±1.73	$6.35 \pm 1.93$	0.48	1.07 (0.80,1.37)	0.63
Arachidonic acid	$176.31 \pm 52.16$	$175.96 \pm 66.18$	0.57	1.00 (0.99,1.01)	0.98
Docosahexaenoic acid	$70.55 \pm 22.35$	$80.98 \pm 38.71$	0.45	1.01 (1.00,1.03)	0.08
Docosanoic acid	$14.45 \pm 4.41$	$15.12 \pm 5.53$	0.49	1.03 (0.93,1.13)	0.54
Docosapentenoic acid	$11.25 \pm 4.43$	$12.45 \pm 6.81$	0.48	1.05 (0.95,1.14)	0.30
Docosatetraenoic acid	$6.55 \pm 2.31$	$6.52 \pm 3.42$	0.57	0.99 (0.79,1.19)	0.96
Eicosadienoic acid	$10.08 \pm 3.84$	$9.87 \pm 4.58$	0.54	0.99 (0.85,1.11)	0.82
Eicosapentaenoic acid	$10.26 \pm 8.51$	$12.42 \pm 9.61$	0.56	1.02 (0.97,1.06)	0.32
Eicosatrienoic acid	$74.49 \pm 33.57$	$78.80\pm69.3$	0.59	1.00 (0.99,1.01)	0.64
Eicosenoic acid	$5.50\pm2.41$	$5.00\pm2.00$	0.56	0.90 (0.68,1.11)	0.39
Hexadecanoic acid	$469.15 \pm 113.21$	$493.8 \pm 167.37$	0.47	1.00 (1.00,1.01)	0.39
Hexadecenoic acid	$24.7 \pm 12.69$	$31.63 \pm 35.53$	0.51	1.02 (0.99,1.04)	0.10
Lignoceric acid	$10.68 \pm 3.64$	$10.78 \pm 5.06$	0.54	1.01 (0.88,1.13)	0.92
Linoleic acid	$752.88 \pm 177.04$	$756.70 \pm 187.03$	0.53	1.00 (1.00,1.00)	0.93
Octadecanoic acid	$150.47 \pm 33.29$	$161.49 \pm 42.01$	0.57	1.01 (1.00,1.02)	0.19
Octadecenoic acid	$375.63 \pm 92.41$	$382.86 \pm 134.38$	0.53	1.00 (1.00,1.01)	0.76
Tetracosenoic acid	$36.07 \pm 13.14$	$38.28 \pm 17.58$	0.50	1.01 (0.98,1.04)	0.50
Tetradecanoic acid	$12.96 \pm 8.08$	$14.59 \pm 14.60$	0.53	1.02 (0.96,1.06)	0.45
$\alpha\text{-Linolenic}$ acid	$29.09 \pm 14.25$	$26.44 \pm 13.41$	0.58	0.98 (0.94,1.02)	0.44
γ-Linolenic acid	$8.69 \pm 5.96$	$9.39 \pm 11.18$	0.58	1.02 (0.94,1.08)	0.66

Table 2. The concentrations, AUC, OR and statistical tests of maternal serum fatty acids between SCH and normal pregnancies in the first trimester. Data are presented as mean  $\pm$  SD, \*p < 0.05. Abbreviation: SCH, subclinical hypothyroidism; AUC, area under the curve; OR, odds ratios; CI, confidence intervals.

	Fatty acids concentrations (mg/L)				
Fatty acids	Normal (n = 222)	SCH (n = 18)	AUC	OR (95%CI)	P value
Arachidic acid	$6.92 \pm 1.74$	$7.85 \pm 2.16$	0.63	1.29 (1.01,1.65)	0.04 *
Arachidonic acid	$177.67 \pm 50.69$	$181.01 \pm 53.4$	0.50	1.00 (0.99,1.01)	0.79
Docosahexaenoic acid	85.31±27.73	$100.12 \pm 38.51$	0.61	1.01 (1.00,1.03)	0.04*
Docosanoic acid	$16.69 \pm 4.61$	$17.37 \pm 4.76$	0.53	1.03 (0.93,1.13)	0.55
Docosapentenoic acid	$12.79 \pm 4.74$	$13.45 \pm 5.57$	0.49	1.03 (0.93,1.13)	0.58
Docosatetraenoic acid	7.4±2.33	$7.04 \pm 2.27$	0.46	0.93 (0.73,1.15)	0.53
Eicosadienoic acid	$12.65 \pm 3.81$	$13.84 \pm 4.22$	0.57	1.07 (0.95,1.19)	0.21
Eicosapentaenoic acid	$13.18 \pm 8.84$	$18.07 \pm 18.87$	0.56	1.03 (1.00,1.07)	0.06
Eicosatrienoic acid	96.6±35.11	$107.19 \pm 48.88$	0.55	1.01 (0.99,1.02)	0.24
Eicosenoic acid	$7.12 \pm 3.72$	9.54±5.3	0.64	1.11 (1.01,1.21)	0.02*
Hexadecanoic acid	$565.06 \pm 127.01$	598.47±151.20	0.55	1.00 (1.00,1.01)	0.29
Hexadecenoic acid	$35.64 \pm 20.49$	$41.70 \pm 28.66$	0.56	1.01 (0.99,1.03)	0.25
Lignoceric acid	12.3±4.22	$12.07 \pm 3.93$	0.51	0.99 (0.87,1.10)	0.82
Linoleic acid	$820.02 \pm 168.63$	838.34±196.93	0.49	1.00 (1.00,1.00)	0.66
Octadecanoic acid	$164.99 \pm 32.94$	$178.49 \pm 36.34$	0.59	1.01 (1.00,1.02)	0.10
Octadecenoic acid	435.73±99.58	475.67±111.84	0.61	1.00 (1.00,1.01)	0.11
Tetracosenoic acid	$41.34 \pm 14.71$	48.17±17.93	0.60	1.02 (1.00,1.05)	0.07
Tetradecanoic acid	$22.64 \pm 11.28$	$23.93 \pm 12.57$	0.52	1.01 (0.97,1.05)	0.64
α-Linolenic acid	39.21±19.59	48.67±22.33	0.65	1.02 (1.00,1.04)	0.07
γ-Linolenic acid	9.75±5.83	$9.98 \pm 8.94$	0.54	1.01 (0.92,1.08)	0.88

Table 3. The concentrations, AUC, OR and statistical tests of maternal serum fatty acids between SCH and normal pregnancies in the second trimester. Data are presented as mean  $\pm$  SD, \*p < 0.05. Abbreviation: SCH, subclinical hypothyroidism; AUC, area under the curve; OR, odds ratios; CI, confidence intervals.

the second trimester and higher concentration of another two serum fatty acids (eicosadienoic acid and octadecanoic acid) in the third trimester, when compared to normal pregnant women.

Our observations of increasing levels of serum fatty acids across the first two trimesters and to a lesser extent in the third trimester in both SCH and normal pregnant women, have been found previously in studies based on normal pregnancy<sup>20-23</sup>: Al et al.<sup>20</sup> analyzed plasma essential fatty acids (EFA) at eleven different time points



**Figure 1.** The concentrations of serum fatty acids in the first, second, and third trimesters collected from normal pregnancies and pregnancies with SCH. Red triangles represent serum fatty acid levels collected from women with SCH. Blue circles represent serum fatty acid levels from normal pregnancies. Red asterisks (\*) indicate fatty acids with significantly different levels in SCH and normal pregnancies (*p*-values less than 0.05).

during pregnancy (at 10, 14, 18, 22, 26, 30, 32, 34, 36, 38 and 40 weeks) in 110 healthy pregnant women in the Netherlands; Otto *et al.*<sup>21</sup> evaluated plasma EFA at three different periods (before 18 weeks and at 22 and 32 weeks of gestation) in 239 healthy pregnant women from the Netherlands, Hungary, Finland, England, and Ecuador; Stewart *et al.*<sup>22</sup> assessed the fatty acid composition of the erythrocyte membranes in each trimester of pregnancy (mean gestational weeks of 12.5, 26.1 and 35.5) in 47 pregnant Australian women without any metabolic complications; and Pinto *et al.*<sup>23</sup> examined serum fatty acid concentrations in the 5–13, 20–26 and 30–36 weeks of gestation in 146 healthy pregnancies in Brazil. The possible underlying reason for all of the described studies to have observed a similar increasing trend of fatty acid levels from the first to the second trimester, may be due to the accumulation of maternal fat reserves that occurs to meet the anabolic storage phase in the early and middle periods of gestation. This phenomenon is mediated by the progressive increase in insulin level, promoting lipogenesis and suppression of lipolysis via progesterone and cortisol<sup>24,25</sup>. Meanwhile, the observed trend of a lesser increase or sustained levels of fatty acids in the third trimester may occur as a result of upregulated lipolysis in maternal fat

	Fatty acids concentrations (mg/L)				
Fatty acids	Normal (n = 222)	SCH (n = 18)	AUC	OR (95%CI)	P value
Arachidic acid	$7.37 \pm 2.03$	$8.20 \pm 1.79$	0.64	1.19 (0.96,1.47)	0.10
Arachidonic acid	$180.20 \pm 52.59$	$189.09 \pm 51.28$	0.46	1.00 (0.99,1.01)	0.49
Docosahexaenoic acid	$89.92 \pm 28.57$	$103.73 \pm 32.45$	0.64	1.01 (1.00,1.03)	0.06
Docosanoic acid	$17.27 \pm 4.77$	$18.83 \pm 5.92$	0.57	1.06 (0.97,1.16)	0.19
Docosapentenoic acid	$12.72 \pm 4.70$	$13.89\pm5.35$	0.56	1.05 (0.95,1.15)	0.32
Docosatetraenoic acid	$7.53 \pm 2.39$	$7.96 \pm 3.11$	0.53	1.07 (0.88,1.27)	0.47
Eicosadienoic acid	$13.05 \pm 3.71$	$15.48\pm5.34$	0.64	1.15 (1.03,1.28)	0.01*
Eicosapentaenoic acid	$13.17 \pm 9.37$	$15.60 \pm 10.57$	0.59	1.02 (0.97,1.06)	0.30
Eicosatrienoic acid	$101.58 \pm 36.18$	$116.28 \pm 45.63$	0.58	1.01 (1.00,1.02)	0.11
Eicosenoic acid	$8.06 \pm 4.18$	$10.06\pm4.03$	0.67	1.08 (0.98,1.17)	0.08
Hexadecanoic acid	$622.33 \pm 139.05$	$661.60 \pm 150.11$	0.60	1.00 (1.00,1.01)	0.25
Hexadecenoic acid	$39.69 \pm 18.71$	$43.36 \pm 20.89$	0.56	1.01 (0.98,1.03)	0.43
Lignoceric acid	$12.31 \pm 4.05$	$12.57 \pm 4.77$	0.50	1.02 (0.90,1.13)	0.79
Linoleic acid	$854.95 \pm 173.94$	$933.57 \pm 237.68$	0.61	1.00 (1.00,1.01)	0.08
Octadecanoic acid	$172.86 \pm 33.15$	$194.13 \pm 40.54$	0.68	1.02 (1.00,1.03)	0.01*
Octadecenoic acid	$489.96 \pm 108.26$	$544.08 \pm 137.12$	0.61	1.00 (1.00,1.01)	0.05
Tetracosenoic acid	$43.98 \pm 17.17$	$48.20 \pm 15.01$	0.59	1.01 (0.99,1.04)	0.31
Tetradecanoic acid	$25.52 \pm 11.96$	$24.57 \pm 11.16$	0.52	0.99 (0.95,1.03)	0.74
$\alpha\text{-Linolenic}$ acid	$45.39 \pm 22.07$	$49.81 \pm 20.76$	0.57	1.01 (0.99,1.03)	0.41
γ-Linolenic acid	$9.68\pm5.90$	$10.90\pm9.40$	0.52	1.03 (0.95,1.10)	0.43

**Table 4.** The concentrations, AUC, OR and statistical tests of maternal serum fatty acids between SCH and normal pregnancies in the third trimester. Data are presented as mean  $\pm$  SD, \*p < 0.05. Abbreviation: SCH, subclinical hypothyroidism; AUC, area under the curve; OR, odds ratios; CI, confidence intervals.

depots. The subsequent stimulation of the lipid transportation across the placenta to meet the increased energy demands of the mother and the increased fetal growth rate<sup>26–28</sup>.

Not only were the majority of the fatty acid levels raised as pregnancy progressed in both SCH and normal participants, we found that women diagnosed with SCH generally demonstrated a higher concentration of fatty acids than the normal pregnancies. An earlier study reported that TSH was the dominating lipolytic hormone in vitro during the neonatal period, when serum TSH levels surge dramatically<sup>29</sup>. FFAs have been found previously to be higher in SCH patients and the higher blood glycerol levels observed suggested that this increase in circulating FFAs was, at least in part, caused by enhanced lipolysis<sup>17</sup>. Similarly, another study showed that TSH raised serum FFA levels in vivo by stimulating adipocyte lipolysis, managed by phosphorylation of perilipin and hormone-sensitive lipase in a protein kinase A-dependent manner in differentiated adipocytes<sup>30</sup>. These studies suggest that TSH may exert extrathyroidal effects to increase FFAs by elevating lipolysis, therefore playing an important regulatory role in lipolysis during pregnancy. Furthermore, there were five long-chain fatty acids, namely arachidic acid, DHA, eicosenoic acid, eicosadienoic acid, and octadecanoic acid, which were significantly higher in the serum of women diagnosed with SCH in pregnancy, in either the second or third trimester. It has previously been reported that patients with hypothyroidism had a higher level of octadecanoic acid in their serum lipid profile<sup>31</sup>. In a rat model of hypothyroidism, an increased level of DHA was found in the liver<sup>32</sup>. Therefore, these five long-chain fatty acids may have the potential to discriminate SCH from normal pregnancy in the middle and later stages of pregnancy. However, caution is advised when interpreting the significance of DHA and octadecanoic acid in relation to SCH because their odds ratios were very close to 1.

Our research has several limitations that deserve mention. Firstly, the sample size of the SCH group was small in this study. There were only 18 pregnant women who met the selection criteria for SCH, without other pregnancy complications. Secondly, maternal dietary intake data should be included to evaluate how maternal diets were linked to the serum fatty acid levels. Lastly, all our subjects diagnosed with SCH were administrated levothyroxine (LT4) treatment, which may interfere with the serum fatty acid outcomes. Nevertheless, some meta-analyses investigating the effect of LT4 treatment on lipid profiles in SCH patients concluded that LT4 replacement did not significantly affect triglyceride levels<sup>33–35</sup>.

In conclusion, our study has highlighted the association between SCH and maternal serum fatty acid profiles and shortlisted five fatty acids (arachidic acid, DHA, eicosenoic acid, eicosadienoic acid, and octadecanoic acid) that were significantly increased in women diagnosed with SCH during pregnancy. Future studies should consider validating our findings using a larger SCH sample size as well as animal models to further understand the pathophysiological mechanisms underlying the link between fatty acids and SCH during pregnancy.

#### Methods

**Study participants.** All participants were selected from the Complex Lipids in Mothers and Babies (CLIMB) study. The CLIMB study was conducted at the First Affiliated Hospital of Chongqing Medical University and Chongqing Health Centre for Women and Children in China from September 2015 to June 2017. The details of the CLIMB study have been published previously<sup>36</sup>. A total of 1,500 women were recruited into the CLIMB study.





.....

For fatty acids analysis, 750 women were selected at random using randomly generated numbers. We excluded women lost to follow up at birth (n = 24) and those who had other adverse pregnancy outcomes (n = 486) such as gestational diabetes mellitus (GDM), and pregnancy-induced hypertension (PIH). This resulted in 240 women who were eligible for inclusion in the study; 1) 222 normal women without any pregnancy complications and 2) 18 women who were diagnosed with SCH based on the criteria used in China (Supplementary Table 1) which included having TSH > the upper limit of reference during pregnancy (97.5th percentiles) and a normal FT4. A flowchart of the study participants included in this study is shown in Fig. 3. All procedures performed in this study were in accordance with the principles in the Declaration of Helsinki 1964 and the International Conference on Harmonisation Good Clinical Practice E6 (ICH-GCP). The study was approved by the Ethics Committee of Chongqing Medical University (2014034). Written informed consent was obtained from all participants included in the study at enrollment.

**Clinical information and sample collection.** Clinical information, demographic factors (maternal age, total years of schooling, last menstrual period (LMP), gravidity, parity), gynecologic and obstetric history (gravidity, delivery, abortion, infertility) were collected at enrollment. Maternal anthropometry (body mass index (BMI) and blood pressure (BP)) and maternal blood samples were collected at three visits during pregnancy (11–14, 22–28, and 32–34 gestational weeks) by trained nurses. Maternal blood samples were collected from participants in the morning after an overnight fast. Fasted blood samples were collected from the antecubital vein into vacutainer tubes containing separator gel and separated by centrifugation twice (3000 rpm at 4 °C for 10 min, then 4000 rpm at 4 °C for another 10 min). The supernatant was transferred into cryotubes (Micronic, Lelystad, The Netherlands) then stored at –80 °C until further processing. Pregnancy outcomes were diagnosed and managed by experienced obstetricians. Throughout the pregnancy, maternal gestational week was determined by LMP and mid-trimester ultrasound information (if the two estimates differed by more than seven days, gestational age was based on the obstetric ultrasound data).



Figure 3. Flowchart of study participants.

**Reagents and fatty acid standards.** Twenty one fatty acid standards were purchased from Sigma-Aldrich (St. Louis, MO, USA) (Supplementary Table 2). Methanol (chromatography grade) and n-hexane (chromatography grade) were purchased from Merck-Chemicals, KGaA (Germany). Hydrochloric acid and Milli-Q pure water were purchased from Guangzhou Chemical Reagent Factory (Guangzhou, China). Stock solutions of the 20 fatty acids and the internal standard (heptadecanoic acid) were prepared at 250 mg/mL in n-hexane. Working solutions were made up with methanol at concentrations of 0.10, 0.50, 2.5, 25, 100, 250 mg/L. All standard solutions were stored at -20 °C until required. The internal standard working solution was a 20 mg/L heptadecanoic acid n-hexane solution.

**Sample preparation and gas chromatography-mass spectrometry (GC-MS) analysis.** A 250 uL aliquot of thawed serum from each participant was spiked with 250 uL of the internal standard (IS) working solution, followed by the addition of 1 mL hydrochloric acid/methanol and a mix using a vortex mixer. The sample was then incubated at 90 °C for 3 h using an electric blast drying oven (Shanghai Yiheng Scientific Instrument Co., Ltd.). After cooling, 2 mL of n-hexane was added to extract the fatty acid methyl ester products and the sample was vortexed. The supernatant was isolated after centrifugation and then evaporated to dryness under nitrogen. Following the addition of 400 uL n-hexane to dissolve the fatty acid methyl esterification product, the suspension was mixed and then transferred to a sample vial (Agilent Technologies, USA) prior to mass spectrometry analysis.

Gas Chromatography-Mass Spectrometry (GC-MS) was performed using an Agilent 7890B gas chromatograph coupled to a 5977 A mass spectrometer (Agilent Technologies, USA). The fatty acid separation was performed on the DB-23 capillary column ( $20 \text{ m} \times 0.18 \text{ mm} \times 0.20 \mu\text{m}$ , Agilent Technologies, USA). Helium was used as the carrier gas with a constant flow rate of 0.78 ml/min. The 1  $\mu$ L of derivatized sample was injected into the inlet set at 230 °C, using the splitless inlet mode. Free fatty acid methyl esters were separated using the following oven temperature program: (1) 50 °C hold for 1 min; (2) increased to 175 °C at 25 °C/min; (3) reached 223 °C at 4 °C/min; and (4) hold at 223 °C for 8 min. The ion-trap mass spectrometer was operated in full scan (Scan) and selected ion scan (SIM) monitoring mode (mass range: 60.00-450.00 m/z). The transfer line was maintained at 230 °C, the source temperature was set at 220 °C, and solvent delay time was set at 10 min.

**Fatty acid quantification and statistical analysis.** The chromatographic height of each of the fatty acids was extracted using Agilent ChemStation (version 2.6). Levels of individual serum fatty acids were first normalized by the internal standard and then quantified to absolute concentration using calibration curves derived from the corresponding chemical standard. A student's t-test and non-parametric Mann-Whitney U test were executed in R to compare clinical characteristics between women with a normal pregnancy and women diagnosed with SCH. Prior to metabolomic statistical analysis, all fatty acid concentrations were corrected to a Gaussian distribution through logarithmic transformation and Pareto scaling. The area under the receiver operating characteristic (ROC) curve was analysed using the pROC R-package<sup>37</sup>. Odds ratios (OR) were calculated to assess associations between the levels of fatty acids and the occurrence of SCH. Tukey's HSD test was implicated to account for multiple comparisons. A *p*-value <0.05 was considered statistically significant. Data were described as mean  $\pm$  SD or median (IQR) for continuous variables. Boxplots were illustrated using ggplot2 R-based packages<sup>38</sup>.

#### Data availability

The datasets generated and/or analyzed during the current study are available from the corresponding author on reasonable request.

Received: 30 August 2019; Accepted: 23 March 2020; Published online: 21 April 2020

#### References

- Stagnaro-Green, A. et al. Guidelines of the American Thyroid Association for the diagnosis and management of thyroid disease during pregnancy and postpartum. Thyroid 21, 1081–1125, https://doi.org/10.1089/thy.2011.0087 (2011).
- Bulletins, Co. P. Practice Bulletin: Thyroid Disease in Pregnancy. The American College of Obstetricians and Gynecologists 125, 996–1005, https://doi.org/10.1097/01.AOG.0000462945.27539.93 (2015).
- 3. Casey, B. M. *et al.* Subclinical hypothyroidism and pregnancy outcomes. *Obstet Gynecol* **105**, 239–245, https://doi.org/10.1097/01. AOG.0000152345.99421.22 (2005).
- Jane Cleary-Goldman et al. Maternal Thyroid Hypofunction and Pregnancy Outcome. OBSTETRICS & GYNECOLOGY 112, 85–92, https://doi.org/10.1097/AOG.0b013e3181788dd7 (2008).
- Fitzpatrick, D. L. & Russell, M. A. Diagnosis and management of thyroid disease in pregnancy. Obstet Gynecol Clin North Am 37, 173–193, https://doi.org/10.1016/j.ogc.2010.02.007 (2010).
- 6. Wang, W. *et al.* The prevalence of thyroid disorders during early pregnancy in China: the benefits of universal screening in the first trimester of pregnancy. *Eur J Endocrinol* **164**, 263–268, https://doi.org/10.1530/EJE-10-0660 (2011).
- Shi, X. et al. Optimal and safe upper limits of iodine intake for early pregnancy in iodine-sufficient regions: a cross-sectional study of 7190 pregnant women in China. J Clin Endocrinol Metab 100, 1630–1638, https://doi.org/10.1210/jc.2014-3704 (2015).
- Liu, H. *et al.* Maternal subclinical hypothyroidism, thyroid autoimmunity, and the risk of miscarriage: a prospective cohort study. *Thyroid* 24, 1642–1649, https://doi.org/10.1089/thy.2014.0029 (2014).
- Tudela, C. M., Casey, B. M., McIntire, D. D. & Cunningham, F. G. Relationship of subclinical thyroid disease to the incidence of gestational diabetes. *Obstet Gynecol* 119, 983–988, https://doi.org/10.1097/AOG.0b013e318250aeeb (2012).
- Wilson, K. L., Casey, B. M., McIntire, D. D., Halvorson, L. M. & Cunningham, F. G. Subclinical thyroid disease and the incidence of hypertension in pregnancy. *Obstet Gynecol* 119, 315–320, https://doi.org/10.1097/AOG.0b013e318240de6a (2012).
- 11. Chan, S. & Boelaert, K. Optimal management of hypothyroidism, hypothyroxinaemia and euthyroid TPO antibody positivity preconception and in pregnancy. *Clin Endocrinol (Oxf)* **82**, 313–326, https://doi.org/10.1111/cen.12605 (2015).
- 12. Su, P. Y. *et al.* Maternal thyroid function in the first twenty weeks of pregnancy and subsequent fetal and infant development: a prospective population-based cohort study in China. *J Clin Endocrinol Metab* **96**, 3234–3241, https://doi.org/10.1210/jc.2011-0274 (2011).
- Negro, R. *et al.* Increased pregnancy loss rate in thyroid antibody negative women with TSH levels between 2.5 and 5.0 in the first trimester of pregnancy. *J Clin Endocrinol Metab* 95, E44–48, https://doi.org/10.1210/jc.2010-0340 (2010).
- Lee, Y. K. et al. Serum TSH level in healthy Koreans and the association of TSH with serum lipid concentration and metabolic syndrome. Korean J Intern Med 26, 432–439, https://doi.org/10.3904/kjim.2011.26.4.432 (2011).
- Park, S. B., Choi, H. C. & Joo, N. S. The relation of thyroid function to components of the metabolic syndrome in Korean men and women. J Korean Med Sci 26, 540–545, https://doi.org/10.3346/jkms.2011.26.4.540 (2011).
- Fazaeli, M., Khoshdel, A., Shafiepour, M. & Rohban, M. The influence of subclinical hypothyroidism on serum lipid profile, PCSK9 levels and CD36 expression on monocytes. *Diabetes Metab Syndrome* 13, 312–316, https://doi.org/10.1016/j.dsx.2018.08.021 (2018).
- Caraccio, N. et al. Muscle metabolism and exercise tolerance in subclinical hypothyroidism: a controlled trial of levothyroxine. J Clin Endocrinol Metab 90, 4057–4062, https://doi.org/10.1210/jc.2004-2344 (2005).
- Lu Wang, Aaron R Folsom, Zhi-Jie Zheng, James S Pankow & Investigators, J. H. E. t. A. S. Plasma fatty acid composition and incidence of diabetes in middle-aged adults: the Atherosclerosis Risk in Communities (ARIC) Study. Am J Clin Nutr 78, 91–98, doi:10.1016/S0939-4753(03)80029-7 (2003).
- 19. Koletzko, B. *et al.* The roles of long-chain polyunsaturated fatty acids in pregnancy, lactation and infancy: review of current knowledge and consensus recommendations. *J Perinat Med* **36**, 5–14, https://doi.org/10.1515/JPM.2008.001 (2008).
- Al, M. D. M. et al. Maternal essential fatty acid patterns during normal pregnancy and their relationship to the neonatal essential fatty acid status. British Journal of Nutrition 74, 55–68, https://doi.org/10.1079/bjn19950106 (1995).
- Otto, S. J. et al. Maternal and neonatal essential fatty acid status in phospholipids: an international comparative study. European Journal of Clinical Nutrition 51, 232–242, https://doi.org/10.1038/sj.ejcn.1600390 (1997).
- Stewart, F. et al. Longitudinal assessment of crythrocyte fatty acid composition throughout pregnancy and post partum. Lipids 42, 335–344, https://doi.org/10.1007/s11745-006-3005-5 (2007).
- Pinto, T. J. *et al.* Lower inter-partum interval and unhealthy life-style factors are inversely associated with n-3 essential fatty acids changes during pregnancy: a prospective cohort with Brazilian women. *PLoS One* 10, e0121151, https://doi.org/10.1371/journal. pone.0121151 (2015).
- 24. Baird, J. D. Some aspects of the metabolic and hormonal adaptation to pregnancy. Acta Endocrinol, 11-18, https://doi.org/10.1530/ acta.0.111S0011\0 (1986).
- Hadden, D. R. & McLaughlin, C. Normal and abnormal maternal metabolism during pregnancy. Semin Fetal Neonatal Med 14, 66–71, https://doi.org/10.1016/j.siny.2008.09.004 (2009).
- Haggarty, P. Placental regulation of fatty acid delivery and its effect on fetal growth-a review. Placenta 23(Suppl A), S28-38, https:// doi.org/10.1053/plac.2002.0791 (2002).
- Haggarty, P. Effect of placental function on fatty acid requirements during pregnancy. Eur J Clin Nutr 58, 1559–1570, https://doi. org/10.1038/sj.ejcn.1602016 (2004).
- Herrera, E., Amusquivar, E., Lopez-Soldado, I. & Ortega, H. Maternal lipid metabolism and placental lipid transfer. Horm Res 65(Suppl 3), 59–64, https://doi.org/10.1159/000091507 (2006).
- Marcus, C., Ehren, H., Bolme, P. & Amer, P. Regulation of lipolysis during the neonatal period. *Importance of thyrotropin. J Clin Invest* 82, 1793–1797, https://doi.org/10.1172/JCI113793 (1988).
- Gagnon, A. et al. Thyroid-stimulating hormone stimulates lipolysis in adipocytes in culture and raises serum free fatty acid levels in vivo. Metabolism 59, 547–553, https://doi.org/10.1016/j.metabol.2009.08.018 (2010).
- Jurand, J. & Oliver, M. F. Effect of thyroid activity on fatty acid composition of serum lipids. *Atherosclerosis* 11, 125–140, https://doi. org/10.1016/0021-9150(70)90011-0 (1970).
- 32. H. Faas, F., Q. Dang, A. & J. Carter, W. 681-684 (1986).
- Anagnostis, P. et al. The effect of L-thyroxine substitution on lipid profile, glucose homeostasis, inflammation and coagulation in patients with subclinical hypothyroidism. Int J Clin Pract 68, 857–863, https://doi.org/10.1111/ijcp.12394 (2014).
- HCCE., V., H., S., O., V. & AN., A. Thyroid hormone replacement for subclinical hypothyroidism (Review). The Cochrane Library, doi:10.1002/14651858.CD003419.pub2 (2007).
- Ineck, B. A. & Ng, T. M. Effects of subclinical hypothyroidism and its treatment on serum lipids. Ann Pharmacother 37, 725–730, https://doi.org/10.1345/aph.1C376 (2003).

- 36. Huang, S. et al. The CLIMB (Complex Lipids In Mothers and Babies) study: protocol for a multicentre, three-group, parallel randomised controlled trial to investigate the effect of supplementation of complex lipids in pregnancy, on maternal ganglioside status and subsequent cognitive outcomes in the offspring. BMJ Open 7, e016637, https://doi.org/10.1136/bmjopen-2017-016637 (2017).
- Robin, X. et al. pROC: an open-source package for R and S+ to analyze and compare ROC curves. BMC Bioinformatics 12, 77, https://doi.org/10.1186/1471-2105-12-77 (2011).
- 38. Wickham, H. ggplot2: elegant graphics for data analysis. (Springer New York, 2009).

#### Acknowledgements

The authors are grateful to the women who participated in this study and the health-care staff who were involved in data and sample collection at the First Affiliated Hospital of Chongqing Medical University and the Chongqing Health Center for Women and Children. This work was supported by the National Natural Science Foundation of China (No.81571453, 81771607, 81871185, 81701477, 81961128004), The 111 Project (Yuwaizhuan (2016)32), The National Key Research and Development Program of Reproductive Health & Major Birth Defects Control and Prevention (2016YFC1000407), Chongqing Health Commission (2017ZDXM008, 2018ZDXM024), and Chongqing Science & Technology Commission (cstc2017jcyjBX0062). This study was also supported by the New Zealand Primary Growth Partnership post farm gate dairy programme, funded by Fonterra Co-operative Group Ltd, New Zealand and the New Zealand Ministry for Primary Industries.

#### **Author contributions**

Y.Y.X., T.L.H., H.Z. and P.B. conceived and designed research; T.Z., Y.Y.X. and H.Z. recruited the patients and collected the samples. Y.Y.X., T.L.H. and H.Z. performed research and analyzed data. T.Z. wrote the manuscript text and prepared the figures. Y.Y.X., T.H., H.Z and P.B. revised the manuscript. T.L.H. and H.Z. provided funding resources. All authors reviewed the manuscript.

#### **Competing interests**

The authors declare no competing interests.

#### Additional information

Supplementary information is available for this paper at https://doi.org/10.1038/s41598-020-63513-7.

Correspondence and requests for materials should be addressed to T.-L.H. or H.Z.

**Reprints and permissions information** is available at www.nature.com/reprints.

**Publisher's note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2020