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OPEN Studies on Isoniazid Derivatives through a Medicinal Chemistry **Approach for the Identification** of New Inhibitors of Urease and **Inflammatory Markers**

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A library of thiosemicarbazide derivatives of isoniazid 3–27, was synthesized and evaluated for their anti-inflammatory and urease inhibition activities, by using in vitro bioassays. Among these compounds 9, 10, 12, 21, and 26 were identified as new derivatives. Prolonged use of non-steroidal anti-inflammatory drugs (NSAIDs) and infections caused by Helicobacter pylori (ureolytic bacteria), are the two most significant causes of gastric and peptic ulcers. We focused on the identification of the dual inhibitors of inflammation and urease enzyme. Compound 23 was identified as the best dual inhibitor of inflammation (ROS; $IC_{50} = 12.3 \,\mu$ g/mL), and urease enzyme inhibition activity ($IC_{50} = 22.4 \,\mu$ M). Many of these compounds showed comparable activities to the standard anti-inflammatory drug (ibuprofen, $IC_{50} = 11.2 \,\mu$ g/mL) and urease inhibitor (thiourea/acetohydraoxamic acid, $IC_{50} = 21.1/20.3 \,\mu$ M). Compound 12 was found to be the most potent urease inhibitor ($IC_{s0} = 12.3 \,\mu M$) and good inhibitor of inflammation (IC₅₀ = 27.7 μ g/mL). Compounds 19, 11, 13, 9, 17, 10, and 16, were also found to be potent inhibitors of urease. Cytotoxicity was also evaluated and all the compounds were found to be noncytotoxic, except compound 18 and the parent drug isoniazid (IC_{so} = 29.5 and 28.5 μ M, respectively).

Urease (urea amidohydrolase; E.C. 3.5.1.5) is a nickel-containing enzyme produced by plants, bacteria, fungi and parasites. Urease catalyzes the hydrolysis of urea into ammonia and carbon dioxide which increases the pH of stomach of the host^{1,2}. The produced ammonia may cause several diseases, such as hepatic encephalopathy, gastric and peptic ulcers, atherosclerosis or rheumatoid arthritis. Urease produced by the Helicobacter pylori serve as a virulence factor through increasing the pH of the stomach, which helps the bacteria to colonize in the acidic environment of stomach and causes gastritis and peptic ulcers. Therefore, urease inhibitors serves as the anti-ulcer drugs³.

Inflammation is the host defense mechanism which protect the body from harmful stimuli and speeds up the restoration process⁴. The stimulus can be any microbial infection or chemicals. The inflammation is characterized with redness, pain, warmth, swelling and lack of function in the injured region⁵. The inadequate healing process of the wounds or any other dysfunction will result in a chronic inflammation which need to be treated⁶.

Currently Available Marketed Drugs and Their Side Effects. Globally used drugs for the treatment of inflammation and associated conditions, such as traumatic injuries, arthritis, fever, and pain, are non-steroidal anti-inflammatory drugs (NSAIDs), such as ketoprofen, ibuprofen, naproxin, diclofenac sodium, piroxicam, and etoricoxib⁷ (Fig. 1). These drugs are the selective inhibitors of cyclooxygenase-2 (COX-2) enzyme⁶. The major side effects caused by the NSAIDS are ulceration and gastrointestinal (GI) hemorrhage8. This has attracted the attention of the scientists towards the development of the new anti-inflammatory agents with no or less side effects⁹.

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Ketoprofen



Naproxin



Ibuprofen



Diclofenac Sodium



Etoricoxib



Piroxicam

Figure 1. Examples of Non-Steroidal Anti-inflammatory Drugs.



Figure 2. Examples of Urease Inhibitors Used as Anti-Ulcer Drugs.

The drugs currently available for the treatment of ulceration and gsastrointestinal (GI) hemorrhage include pantoprazole, lansoprazole, lithostat, and omeprazole¹⁰ (Fig. 2). A study by Saniee.*et al.* (2015) showed that the proton pump inhibitors (PPI), such as omeprazole and lansoprazole, are also the urease inhibitors¹¹. Due to the high prevalence of gastroesophageal reflux disease the use of PPIs has increased remarkably and now they are among the most frequently prescribed medicines globally¹². Adverse effects of PPIs has also been observed such as nutritional deficiencies, visual impairment, chronic kidney disease¹³, dementia, infections, nervous system related abnormalities. The most adverse effect is the decreased in the bacterial richness and alteration in the gut



Anti-cancer/Urease inhibitors

Figure 3. Some previously reported derivatives of isoniazid.



Figure 4. Component structural similarities between isoniazid, etoricoxib, and pantoprazole.

microbiome. Approximately 65% increase in the risk of enteric infections development especially *Clostridium difficile* infection has been reported¹⁴, which limit clinical applications of PPIs¹⁵.

A Medicinal Chemistry Approach of Drug Discovery. Drug development is a time-intensive, costly, and high-risk process. One approach that has attracted a lot of attention in modern drug discovery is drug repositioning or repurposing¹⁶. Drug repurposing include cases in which a current drug, endorsed by an administrative organization for a particular disease, is found to have effect against another illness. Conversely, drug repositioning also depicts a condition where a drug that is in use for a disease is utilized as a template for the synthesis of new analogs possessing activity against another disease¹⁷. Drug repositioning thus essentially shorten the drug development process and thus decrease the discovery cost¹⁸.

Current study describes the repositioning of isoniazid, an anti-bacterial agent. Isoniazid, was synthesized in 1952 for the treatment of tuberculosis¹⁹. The recommended daily dose of isoniazid is from 5–300 mg/day, which rarely causes side effects in individuals²⁰. The use of isoniazid as the main scaffold for the synthesis of medicinally important compounds is well known as reported in the literature^{21–24} (Fig. 3). Therefore, we have randomly synthesized the library of compounds (**3–27**) followed by random screening against various biological targets. It was observed that some of these compounds are the significant dual inhibitors of inflammation, and urease. The structural similarity of synthesized compounds with the pyridine based anti-ulcer drug pantoprazole²³, and anti-inflammatory drug etoricoxib²⁵ may be the reason for the activities of these compounds (Fig. 4).

During the current study, we have synthesized thiosemicarbazide derivatives of isoniazid (3–27) through modification at terminal NH₂ (Fig. 5) by reacting with different isothiocyanates. Thiosemicarbazide class of compounds possess the diverse biological activities, such as anti-cancer²⁶, anti-fungal²⁷, anti-helminthic²⁸, anti-bacterial²⁹ and anti-HIV³⁰. Among synthesized compounds, all were identified as previously known^{31–38}, except **9**, **10**, **12**, **21**, and **26**. However, these compounds have not been reported as the dual inhibitor of inflammation and urease. Cytotoxicity of these compounds were also evaluated against 3T3 mouse fibroblast cell line.



Figure 5. Isoniazid (1).



R= Ph-Alkyl or halogen, and alkyl

Figure 6. Synthesis of thiosemicarbazide derivatives of isoniazid (3-27).



Figure 7. NMR and Key 2D NOSEY correlations of *N*-(2, 3-Dichlorophenyl)–2-isonicotinoylhydrazinecarbot hioamide (**4**).

Results

Chemistry. Thiosemicarbazide derivatives of isoniazid (3–27) were synthesized by its reaction with various isothiocyanates using method reported by Yahyazadeh. *et al.* (2013) with slight modification³⁹ (Fig. 6). The resulting product was purified through solvent-solvent extraction with hexane and ethyl acetate, followed by recrystallization with methanol. Structures of synthesized compounds were elucidated by using different spectroscopic techniques, such as mass spectrometry, infrared spectrophotometry, and ¹H- and ¹³C-NMR spectroscopy. Among synthesized compounds, **9**, **10**, **12**, **21**, and **26** were found to be new.

Structure Elucidation of Representative Compound 4. The structure elucidation of the synthesized thiosemicarbazide derivatives (**3–27**) was performed through various spectroscopic techniques (MS, NMR, and IR). Structure elucidation of compound **4**, a representative member of the library, is presented in Fig. 7.

Singlets of protons attached to nitrogen were at δ 10.93, 10.02, and 9.82. A doublet at δ 8.76 showed two protons of pyridine ring *i.e.* H-3", H-5" which are coupled with H-2", H-6" ($J_{3",2"} = J_{5",6"} = 6$ Hz). A distorted singlet at δ 7.83 represented another two protons of pyridine ring *i.e.* H-2", and H-6". Doublet appeared at δ 7.56 showed H-4 of phenyl ring, coupled with the H-5, with *ortho* coupling ($J_{4,5} = 7.8$ Hz), while triplet at δ 7.36 showed a proton of phenyl ring *i.e.*H-5 which is coupled with H-4 and H-6 with coupling constant ($J_{5(4-6)} = 7.8$ Hz). H-6 proton of phenyl ring showed a distorted singlet at δ 7.32.

The conformation of the synthesized compound was deduced based on key 2D NOESY correlations (Fig. 7). NH (δ 9.82, singlet) showed correlations with the H-6 phenyl ring proton (δ 7.32, distorted singlet), and NH

(δ 10.93, singlet). On the other hand, proton attached to nitrogen NH (δ 10.02, singlet) showed correlation with pyridine ring H-6" (δ 7.83, distorted singlet).

¹³C NMR spectrum of compound 4 showed five characteristic peaks of quaternary carbons at δ 182.0 (*C*=S), 165.0 (C=O), 140.0 (C-1"), 139.3 (C-1) and 132.1, 131.0 (C-2, C-3). C-3", C-5" and C-2", C-6" of pyridine ring were resonated at δ 150.6 and 121.7, while C-4 and C-5 of phenyl ring were resonated at δ 128.6 and 127.5, respectively. C-6 of phenyl ring appeared at δ 129.9. The HRFAB-MS (+ve mode) was observed at *m*/*z* 341.0017 correlating with the molecular formula [(C₁₃H₁₀N₄O₁Cl₂S₁) + H]⁺ (341.0031).

Bioactivities. In Vitro Urease Inhibition Activity. Thiosemicarbazide derivatives, synthesized in the current study, were evaluated for their *in vitro* urease inhibitory activity. The standard inhibitor used in the assay was thiourea ($IC_{50} = 21.1 \pm 0.2 \,\mu$ M) and acetohydroxamic acid ($IC_{50} = 20.3 \pm 0.4 \,\mu$ M). Most of the synthesized thiosemicarbazides showed urease inhibition activity between the range of $63.2 \pm 2.0 \,\mu$ M to $12.3 \pm 1.04 \,\mu$ M (Table 1).

Compound **12** (IC₅₀ = 12.3 ± 1.04 µM), **19** (IC₅₀ = 12.7 ± 0.8 µM), **11** (IC₅₀ = 13.2 ± 1.64 µM), **13** (IC₅₀ 13.6 ± 1.61 µM), **9** (IC₅₀ = 14.2 ± 1.38 µM), **17** (IC₅₀ = 14.7 ± 1.01 µM), **10** (IC₅₀ = 15.7 ± 1.32 µM), and **16** (IC₅₀ = 21.5 ± 1.3 µM) were found more potent than the thiourea (Standard: IC₅₀ = 21.1 ± 0.2 µM)/ acetohydroxamic acid (Standard: IC₅₀ = 20.3 ± 0.4 µM). While compounds **20** (IC₅₀ = 22.0 ± 1.1 µM), **22** (IC₅₀ = 22.1 ± 1.91 µM), and **23** (IC₅₀ = 22.4 ± 1.83 µM) showed comparable activity with standard thiourea (IC₅₀ = 21.1 ± 0.2 µM)/acetohydroxamic acid (Standard: IC₅₀ = 20.3 ± 0.4 µM). Compounds **18** (IC₅₀ = 26.3 ± 1.59 µM), **21** (IC₅₀ = 28.2 ± 1.48 µM), **15** (IC₅₀ = 34.1 ± 0.45 µM), **14** (IC₅₀ = 35.8 ± 1.8 µM), **4** (IC₅₀ = 38.8 ± 0.35 µM) showed significant activity, while **24** (IC₅₀ = 63.2 ± 2.0 µM) showed moderate activity. Whereas, compound **3** and the parent drug isoniazid (**1**) was found to be inactive.

Indeed, the inhibition showed by all the synthesized analogues is due to the mutual participation of all parts of the molecule. However, it is also true that there are some characteristic features, which play an important role in the biological activity. As depicted in the Fig. 8 the current library has only aryl part (R) which is varying. Therefore, only the limited structure activity relationship can be drawn by comparing the position and nature of the substituents present in the aryl part of thiosemicarbazide derivatives (**3–27**).

The compound **24** with no substituents on aryl part showed the lowest activity ($IC_{50} = 63.2 \pm 2.0 \mu M$) as compared to other members of the library. Substitution of bromo group at the C-3 position as in compound **22** enhances the activity with $IC_{50} = 22.1 \pm 1.91 \mu M$. While changing the position of bromo group to the C-2 position as in compound **14**, slightly reduces the activity ($IC_{50} = 35.8 \pm 1.8 \mu M$).

Substitution of chloro group at the C-4 position as in compound **18** showed almost similar activity with the $(IC_{50} = 26.3 \pm 1.59 \,\mu\text{M})$. Compound **4** possess the two chloro groups at C-2 and C-3 positions and compound 5 with the chloro groups at C-2 and C-5 positions showed reduced activity with the IC_{50} values $38.8 \pm 0.35 \,\mu\text{M}$ and $36.5 \pm 0.14 \,\mu\text{M}$, respectively. The compound with bromo substitution at C-3 position was found to be more efficient in the inhibitory potential as compared to the chloro substituted analogues, similar effect has also been reported by ali. *et al.*⁴⁰.

Compound 12 ($IC_{50} = 12.3 \pm 1.04 \,\mu$ M), 19 ($IC_{50} = 12.7 \pm 0.8 \,\mu$ M), 16 ($IC_{50} = 21.5 \pm 1.3 \,\mu$ M), 15 ($IC_{50} = 34.1 \pm 0.45 \,\mu$ M), and 7 ($IC_{50} = 46.3 \pm 0.43 \,\mu$ M), possess fluoro group but they differ in the numbers and positions, compound 19 posses the fluoro group at C-4 positionis found to be the more active ($IC_{50} = 12.7 \pm 0.8 \,\mu$ M) than the compound 16 ($IC_{50} = 21.5 \pm 1.3 \,\mu$ M) having the fluoro group at C-3 position among, the disubstituted analogue 15 posses fluoro group at C-2 and C-4 showed inhibition with the IC₅₀ value $34.1 \pm 0.45 \,\mu$ M. Compound 7 posses the fluoro substituted compound 12 ($IC_{50} = 12.3 \pm 1.04 \,\mu$ M) showed the comparable activity as of mono substituted analogue 19. Similar SAR have been reported by taha. *et al.*⁴¹.

Incorporation of the chloro and trifluoromethyl group in compound 9 have further enhanced the activity with $IC_{50} = 14.2 \pm 1.38 \,\mu\text{M}$.

Compound **11** is a *para* substituted derivatives ($IC_{50} = 13.2 \pm 1.64 \mu M$), which posses an electron withdrawing trifluoromethoxy group, showed comparable activity with compound **10** ($IC_{50} = 15.7 \pm 1.32 \mu M$) having the cyano group, while compound **20** ($IC_{50} = 22.0 \pm 1.1 \mu M$) having the electron donating methoxy group showed less activity than the compounds **11** and **10**. It demonstrates that the electron withdrawing groups have the profound effect on inhibitory potential.

From the limited structure activity relationship, it can be summarized that the nature and position of the substituent both are equally responsible for demonstrating the inhibitory activity.

In Vitro Anti-inflammatory Activity. All synthesized derivatives of isoniazid (1) were also evaluated for their anti-inflammatory activity using oxidative burst assay. The standard inhibitor used was ibuprofen (IC₅₀ = $11.2 \pm 1.9 \mu$ g/mL). Among all the compounds, compounds **23**, **5**, **22**, **26**, **9**, **12**, **16**, and **17** showed promising anti-inflammatory activities with IC₅₀ value $12.3 \pm 1.2 \pm 0.36.9 \pm 3.0 \mu$ g/mL, while other compounds were found inactive (Table 1).

Among these compounds, compound **23** containing a cyclohexyl ring was found to be the most active inhibitor of ROS ($IC_{50} = 12.3 \pm 1.2 \mu g/mL$). Compound **5** bearing chloro groups at C-2 and C-5 positions showed a good activity with an IC_{50} value $18.5 \pm 1.0 \mu g/mL$, while with a bromo group at C-3, compound **22** was also found to be a good inhibitor but less active than compound **5** ($IC_{50} = 25.1 \pm 0.4 \mu g/mL$). On the other hand, compound **26** with a isothiocyanate group at C-3 showed the similar activity ($IC_{50} = 25.4 \pm 1.3 \mu g/mL$) as that of compound **5**. Compound **9** with a chloro group at C-4 and trifluoromethyl group at C-3 also showed a good activity ($IC_{50} = 26.7 \pm 2.5 \mu g/mL$), along with this pentaflouro containing phenyl ring analogue **12** which resulted in decrease of activity from IC_{50} values 26.7 ± 2.5 to $27.7 \pm 2.4 \mu g/mL$. Interestingly the mono substituted analogue of fluorine at C-3 (compound **16**) showed a decreased activity with IC_{50} value $29.7 \pm 1.7 \mu g/mL$, while the iodine containing analogue at C-3 (compound **17**) showed further decreased activity ($IC_{50} = 36.9 \pm 3.0 \mu g/mL$).

Compounds	R	Urease Inhibition Activity IC ₅₀ ±SEM ^a µM	$\begin{array}{c} Anti-Inflammatory \ Activity \\ IC_{50} \pm SD^b \mu g/mL \end{array}$	Cytotoxicity (3T3 Cell line) $IC_{50} \pm SD^{b} \mu M$	
3	F	Inactive ^c	Inactive	>30	
4	CI	38.8±0.35	Inactive	>30	
5	CI	36.5±0.14	18.5±1.0	>30	
6	CI	Insoluble	Inactive	>30	
7	FFF	46.3±0.43	Inactive	>30	
8	F ₃ C Cl	44.4±0.43	Inactive	>30	
9	CI CF ₃	14.2±1.38	26.7±2.5	>30	
10	CN	15.7±1.32	Inactive	>30	
11	OCF ₃	13.2±1.64	Inactive	>30	
12	F F F F	12.3±1.04	27.7±2.4	>30	
13	\bigtriangledown	13.6±1.61	Inactive	>30	
Continued					

Compounds	R	Urease Inhibition Activity $IC_{50} \pm SEM^{a} \mu M$	$ \begin{array}{c} Anti-Inflammatory \ Activity \\ IC_{50} \pm SD^b \ \mu g/mL \end{array} $	Cytotoxicity (3T3 Cell line) $IC_{50} \pm SD^{b} \mu M$		
14	Br	35.8±1.89	Inactive	>30		
15	F	34.1±0.45	Inactive	>30		
16	F	21.5±1.3	29.7±1.7	>30		
17		14.7±1.01	36.9±3.0	>30		
18	CI	26.3±1.59	Inactive	29.5±1.9		
19	F	12.7±0.8	Inactive	>30		
20	OCH ₃	22.0±1.15	Inactive	>30		
21	CI	28.2±1.48	Inactive	>30		
22	Br	22.1±1.91	25.1±0.4	>30		
23	$\widehat{\mathbf{Q}}$	22.4±1.83	12.3±1.2	>30		
24		63.2±2.0	Inactive	>30		
25		Insoluble	Inactive	>30		
Continuea						

Compounds	R	Urease Inhibition Activity IC ₅₀ ± SEM ^a µM	$\begin{array}{c} Anti-Inflammatory \ Activity \\ IC_{50} \pm SD^b \mu g/mL \end{array}$	Cytotoxicity (3T3 Cell line) IC ₅₀ ±SD ^b µM
26	NCS	46.8±1.8	25.4±1.3	>30
27	NO ₂	Insoluble ^d	Inactive	>30
28	Isoniazid(1)	Inactive	Inactive	28.5 ± 1.2
29	Thiourea (Standard)	21.1 ± 0.2 (Observed)	-	-
30	Aetohydaoxamic acid (standard)	20.3 ± 0.4 (Observed) 17.2 ± 0.9 (Reported)		
31	Ibuprofen (Standard)	_		_
32	Cyclohexanamide (Standard)	-	-	$\begin{array}{c} 0.8 \pm 0.2 \text{ (observed)} \\ 0.26 \pm 0.04 \text{ (reported)} \end{array}$

Table 1. *In vitro* urease Inhibition, anti-inflammatory, and cytotoxicity of synthetic compounds 3–27. **SEM**^a is the standard error of the mean, **SD**^b is the standard deviation, and **Insoluble**^d refers to those analogue which are partially soluble in HPLC methanol used in the protocol of *in vitro* urease inhibition activity. Compounds showed <50% inhibition were considered as inactive. For Urease enzyme inhibition activity, screening concentration was 0.5 mM. For ROS inhibition assay screening concentration was 25 µg/mL. Note: All data were presented as mean ± standard deviation/standard error of the mean of three independent experiments where each sample was run in triplicate. The IC₅₀ values were obtained using three concentrations of test compound, and were calculated using Excel Based Program.



Figure 8. Rationale of the biological activity.

Evaluation of cytotoxicity on 3T3 normal cell line. Derivatives **3–27** and isoniazid (**1**) were evaluated for their cytotoxicity against 3T3 normal (mouse fibroblast) cell lines in which all derivatives were found to be inactive, except compound **18** and isoniazid (**1**) which showed IC₅₀ values of 29.5 ± 1.9 , and $28.5 \pm 1.2 \,\mu$ M, respectively, in comparison to the standard cyclohexamide (IC₅₀= $0.8 \pm 1.9 \,\mu$ M) (Table 1).

Discussion

The present research study indicates that thiosemicarbazides 3–27 possess promising urease inhibition activity, as well as significant anti-inflammatory activities. Compounds 5, 9, 12, 16, 17, 22, 23, and 26 were found to be the dual inhibitors of inflammation (ROS) and urease, few members of the current library have been reported as urease inhibitors by Ali. *et al.*³⁴ but here we report the dual inhibitors of urease and inflammation along with its cytoxicity profile against 3T3 mouse fibroblast cell line. The limited SAR concludes that the compound 23 with the cyclohexyl ring was found to be the most significant dual inhibitor of urease and inflammation ($IC_{50} = 22.4 \pm 1.83 \,\mu$ M and $12.3 \pm 1.2 \,\mu$ g/mL, respectively). This showed comparable activity to the clinically used anti-inflammatory drug ibuprofen ($IC_{50} = 11.2 \pm 1.9 \,\mu$ g/mL), and urease inhibitor acetohyroxamic acid ($IC_{50} = 20.3 \pm 1.9 \,\mu$ g/mL). Compound 12, is the pentafluoro substituted analogue. Compound 12 was identified as the most potent inhibitor of urease ($IC_{50} = 12.3 \pm 1.04 \,\mu$ M) and significant inhibitor of inflammation ($IC_{50} = 27.7 \pm 2.4 \,\mu$ g/mL).

Whereas compounds **10**, **11**, and **13** were identified as potent inhibitors of urease enzyme, however, these compounds were found to be inactive in anti-inflammatory assay. The SAR clearly shows that change in the substituents and its positions or changing the aryl ring by the alkyl ring greatly affects the biological activity of the synthesized compounds.

In conclusion, the compounds reported here may serve as the starting points for the designing and development of new and powerful dual inhibitors of inflammation and infections caused by ureolytic bacteria. **Experimental.** The experimental section describes various methods and technical aspects of the current study including synthesis, purification, characterization of the synthesized analogues through different spectroscopic techniques and evaluation of the biological activities, such as inhibition of urease enzyme and inflammation.

Chemicals. Isoniazid (1) was purchased from Sigma Aldrich, India; 2-bromophenyl isothiocyanate, 3-bromophenyl isothiocyanate, 4-fluorophenyl isothiocyanate, and 3-nitrophenyl isothiocyanate from Aldrich, Poland. Cyclopentylisothiocyanate, cyclohexylisothiocyanate, 2,5-dichlorophenyl isothiocyanate, 3-fluorophenyl isothiocyanate, and 4-methoxy phenyl isothiocyanate from Sigma Aldrich, USA; 2,4,5-trichloro phenyl isothiocyanate from Aldrich, USA. 4-Chlorophenyl isothiocyanate, 2,4-difluorophenyl isothiocyanate, and 2,6-dichlorophenyl isothiocyanate, 3,4-dichlorophenyl isothiocyanate were obtained from Alfa Aesar, USA. 2-Fluorophenyl isothiocyanate from Sigma Aldrich Chemie, Germany; 2-chloro-5-trifluoromethyl)phenyl isothiocyanate, 4-chloro-3-(trifluoromethyl)phenyl isothiocyanate, 4-cyanophenyl isothiocyanate, use bought from Fluka, Switzerland, phenyl isothiocyanate from Schuchardt, Hohenbrunn, Germany.

Precoated silica gel plates (ALUGRAM, SIL G/UV254) were used for thin layer chromatography (TLC). TLC chromatograms were viewed under the ultraviolet light of 254 and 365 nm. Electron impact mass spectroscopy (EI-MS) and Fast atom bombardment direct probe mass spectra (FAB-MS) were obtained through JEOLJMS-600H mass spectrometer (Japan). ¹H and ¹³C-NMR spectra were performed on a 300, 400, and 100 MHz Bruker Avance spectrometers (Switzerland). Buchi M-560 apparatus was used for recording the melting point (Japan). I. R. Spectrophotometry of the compounds was performed on FTIR-8900 (Shimadzu, Japan) through KBr disc.

General Procedure of the Synthesis of Compounds 3–27. Isoniazid (1) (2 mmol) was refluxed with methanol (10 mL) for 15 minutes. Corresponding isothiocyanate (2 mmol) was added, and kept on stirring for 8–10 hours at room temperature (25 °C). Reaction progress was examined by TLC (7.9: 2: 0.1) (ethyl acetate, methanol and acetic acid) analysis. Disappearance of starting material from reaction mixture indicated the completion of reaction. The reaction mixture was concentrated under reduced pressure, solid product thus obtained was purified through solvent-solvent extraction with the help of hexane and ethyl acetate, and then recrystallized from methanol. Among these compounds, 9, 10, 12, 21, and 26 were identified as new derivatives. The known compounds were found to be spectroscopically similar to that reported in the literature.

N-(2-Fluorophenyl)-2-isonicotinoylhydrazinecarbothioamide (**3**). **R**_f = 0.7, **Mp**: 241–242 °C; **IR** (**KBr**, **cm**⁻¹): 3285, 3128 (N−H), 1679 (C=O), 1596, 1545, 1509 (C=C), 1226 (C=S), 1149 (C− F); ¹**H**-**NMR(300 MHz**, **DMSO-d**₆): $\delta_{\rm H}$ 10.91 (s, 1 H, NH), 9.96 (s, 1 H, NH), 9.67 (s, 1 H, NH), 8.75 (d, $J_{3'',2''} = J_{5'',6''} = 6.0$ Hz, 2 H, H-3'', H-5''), 7.83 (d, $J_{2'',3''} = J_{6'',5''} = 5.1$ Hz, 2 H, H-2'', H-6''), 7.32–7.14 (m, 4 H, H-3, H-4, H-5, H-6); ¹³C-NMR (100 MHz, **DMSO-d**₆): $\delta_{\rm H}$ 26.0 (C=S), 165.0 (C=O), 159.0 (d, $J_{CF} = 248.8$ Hz, C-2), 140.0 (C-1''), 150.1 (C-3'',C-5''), 130.6 (C-5), 128.0 (C-4), 124.0 (C-6), 121.6 (C2'',C-6''), 116.1 (d, $J_{CF} = 19.9$ Hz, C-1), 115.6 (d, $J_{CF} = 19.8$ Hz,C-3); **Positive FAB-MS** *m/z* (**rel. int.** %): 291.1 [M+H]⁺ (56), 182.1 (100), 185.1 (27), 171.0 (16); HRFAB-MS (+ve mode) Calcd. for [(C₁₃H₁₁N₄O₁F₁S₁)+H]⁺: (*m/z* = 291.0716) Found 291.0712.

N-(2, 3-*Dichlorophenyl*)-2-*isonicotinoylhydrazinecarbothioamide* (4). **R**_f=0.76, **Mp**: 198–199 °C; **IR** (**KBr**, **cm**⁻¹): 3289, 3137 (N–H), 1679 (C=O), 1546, 1512 (C=C), 1249 (C=S), 1058 (C–Cl). ¹H-NMR (600 MHz, **DMSO-***d*₆): $\delta_{\rm H}$ 10.93 (s, 1 H, NH), 10.02 (s, 1 H, NH), 9.82 (s, 1 H, NH), 8.76 (d, $J_{3'',2''} = J_{5'',6''} = 6.0$ Hz, 2 H, H-3", H-5"), 7.83 (distorted singlet, 2 H, H-2", H-6"), 7.56 (d, $J_{4,5} = 7.8$ Hz, 1 H, H-4), 7.36 (t, $J_{5(4,6)} = 7.8$ Hz, 1 H, H-5), 7.32 (distorted singlet, 1 H, H-6); ¹³C-NMR (100 MHz, **DMSO-***d*₆): δ 182.4 (C=S), 165.0(C=O), 150.6(C-3", C-5"), 139.9 (C-1"), 139.3(C-1), 132.1 (C-2), 131.0 (C-3), 129.9 (C-6), 128.6 (C-4), 127.5 (C-5), 121.7 (C-2", C-6"); **Positive FAB-MS** *m/z* (**rel. int.** %): 342.0 [M + 2]⁺ (3), 341.0 [M + H]⁺ (6), 185.1 (100), 277.1 (16), 219.1 (11); **HRFAB-MS** (+**ve mode**) Calcd. For [(C₁₃H₁₀N₄O₁Cl₂S₁) + H]⁺: (*m/z* = 341.0031) Found 341.0017.

 $\begin{array}{l} N-(2,\ 5-Dichlorophenyl)-2-isonicotinoylhydrazinecarbothioamide\ (5). \\ \mathbf{R_f}=0.73,\ \mathbf{Mp:}\ 172-173\ ^\circ\mathrm{C};\ \mathbf{IR}\ (\mathbf{KBr},\ \mathbf{cm^{-1}}):\ 3285,\ 3130\ (\mathrm{N-H}),\ 1679\ (\mathrm{C=O}),\ 1546,\ 1512\ (\mathrm{C=C}),\ 1252\ (\mathrm{C=S}),\ 1092\ (\mathrm{C-Cl}),\ ^1\mathbf{H-NMR}\ (\mathbf{400\ MHz},\ \mathbf{DMSO-d_6}):\ \delta_{\mathrm{H}}\ 10.93\ (\mathrm{s},\ 1\,\mathrm{H},\ \mathrm{NH}),\ 10.07\ (\mathrm{s},\ 1\,\mathrm{H},\ \mathrm{NH}),\ 9.76\ (\mathrm{s},\ 1\,\mathrm{H},\ \mathrm{NH}),\ 8.76\ (\mathrm{d},\ J_{3'',2''}=J_{5'',6''}=6.0\ \mathrm{Hz},\ 2\,\mathrm{H},\ \mathrm{H-3''},\ \mathrm{H-5''}),\ 7.84\ (\mathrm{d},\ J_{2'',3''}=J_{6'',5''}=5.0\ \mathrm{Hz},\ 2\,\mathrm{H},\ \mathrm{H-2''},\ \mathrm{H-6''}),\ 7.54\ (\mathrm{d},\ J_{5,4}=8.4\ \mathrm{Hz},\ 1\,\mathrm{H},\ \mathrm{H-6}),\ 7.38\ (\mathrm{distorted}\ \mathrm{t},\ 2\,\mathrm{H},\ \mathrm{H-3},\ \mathrm{H-4});\ \mathbf{NegativeFAB-MS}\ m/z\ (\mathrm{rel.\ int.\ \%}):\ 341.1\ [\mathrm{M+2}]^+\ (3),\ 339.1\ [\mathrm{M-H}]^+\ (6),\ 275.2\ (22)\ 183.0\ (100),\ 164.0\ (10),\ 150.9\ (11);\ \mathrm{HRFAB-MS}\ (+ve\ mode)\ \mathrm{Calcd.\ for}\ [(\mathrm{C}_{13}\mathrm{H}_{10}\mathrm{N}_4\mathrm{O}_1\mathrm{Cl}_2\mathrm{S}_1)+\mathrm{H}]^+:\ (m/z=341.0031)\ \mathrm{Found}\ 341.0010. \end{array}$

N-(2, 6-*Dichlorophenyl*)-2-*isonicotinoylhydrazinecarbothioamide* (6). **R**_f=0.73, **Mp**: 127–128 °C; **IR** (**KBr**, **cm**⁻): 3287, 3164 (N−H), 1681 (C=O), 1513 (C=C), 1244 (C=S), 1064 (C−Cl). ¹**H**-**NMR** (300 MHz, **DMSO**-*d*₆): $\delta_{\rm H}$ 10.91 (s, 1 H, NH), 10.0 (s, 1 H, NH), 9.75 (s, 1 H, NH), 8.76 (d, $J_{3'',2''} = J_{5'',6''} = 5.1$ Hz, 2 H, H-3'', H-5''), 7.83 (d, $J_{2'',3''} = J_{6'',5''} = 4.5$ Hz, 2 H, H-2'', H-6''), 7.51 (d, $J_{3,4} = J_{5,4} = 8.1$ Hz, 2 H, H-3, H-5), 7.39 (distorted t, 1 H, H-4); **Negative FAB-MS** *m/z* (rel. int. %): 341.0 [M + 2]⁺ (3), 339.2 [M-H]⁺ (8), 275.2 (18), 268.2 (14), 183.1(100), 176.1(44); **HRFAB-MS** (+ve mode) Calcd. for [(C₁₃H₁₀N₄O₁Cl₂S₁) + H]⁺: (*m/z* = 341.0031) Found 341.0010.

N-(2, 6-*Difluorophenyl*)-2-*isonicotinoylhydrazinecarbothioamide* (7). **R**_f=0.7, **Mp:** 248–249 °C; **IR** (**KBr, cm**⁻¹): 3292, 3131 (N–H), 1681 (C=O), 1595, 1546 (C=C), 1246 (C=), 1151 (C–F), ¹**H-NMR (400 MHz, DMSO-***d*₆):

 $δ_{\rm H}$ 10.95 (s, 1 H, NH), 10.11 (s, 1 H, NH), 9.47 (s, 1 H, NH), 8.75 (d, $J_{3'',2''} = J_{5'',6''} = 4.8$ Hz, 2 H, H-3'', H-5''), 7.84 (distorted singlet, 2 H, H-2'', H-6''), 7.34 (m, 1 H, H-4), 7.12 (t, $J_{3,2/3,4} = J_{5,4/5,6}$ 8.0 Hz, 2 H, H-3, H-5); ¹³C-NMR (125 MHz, DMSO-d₆): δ182.6 (C=S), 164.4 (C=O), 159.8 (d, $J_{CF} = 247.8$ Hz, C-2), 159.7 (d, $J_{CF} = 247$ Hz, C-6), 150.1 (C-3'',C-5''), 139.3 (C-1''), 128.8 (t, $J_{CF} = 10$ Hz, C-4), 121.7 (C2'',C-6''), 116.6 (t, $J_{CF} = 15.8$ Hz, C-1), 111.7 (d, $J_{CF} = 21.8$ Hz, C-3, C-5); Positive FAB-MS *m/z* (rel. int. %): 309.0 [M + H]⁺ (17), 277.0 (27), 219 (19), 185.0 (100), 171.1 (96), 157.0 (45); HRFAB-MS (+ve mode) Calcd. for [(C₁₃H₁₀F₂N₄OS) + H]⁺: (*m/z* = 309.0622) Found 309.0610.

$$\begin{split} & N-(2-Chloro-5-(trifluoromethyl)phenyl)-2-isonicotinoylhydrazinecarbothioamide ~(8). ~~~ R_{\rm f}=0.8, ~~ Mp: \\ & 183-184~^{\rm C}; ~~ IR~(KBr, cm^{-1}): ~~ 3304, ~~ 3134~(N-H), ~~ 1679~(C=O), ~~ 1548, ~~ 1513~(C=C), ~~ 1253~(C=S), ~~ 1080~(C-Cl). ~~ ^{\rm H-NMR}~(400~MHz, DMSO-d_6): ~~ \delta_{\rm H}~10.96~({\rm s}, ~~ 1\,{\rm H}, {\rm NH}), ~~ 10.14~({\rm s}, ~~ 1\,{\rm H}, {\rm NH}), ~~ 9.83~({\rm s}, ~~ 1\,{\rm H}, {\rm NH}), ~~ 8.76~({\rm d}, ~~ J_{3'',2''}=J_{5'',6''}=5.6~{\rm Hz}, ~~ 2\,{\rm H}, ~~ H-5''), ~~ 7.84~({\rm br}~{\rm s}, ~2\,{\rm H}, ~~ H-6''), ~~ 7.76~({\rm d}, ~~ J_{6,4}=8.8~{\rm Hz}, ~~ 1\,{\rm H}, {\rm H-6}), ~~ 7.67~({\rm br}~{\rm s}, ~~ 2\,{\rm H}, ~~ H-3'', ~~ H-5''), ~~ 7.84~({\rm br}~{\rm s}, ~~ 2\,{\rm H}, ~~ H-6''), ~~ 7.76~({\rm d}, ~~ J_{6,4}=8.8~{\rm Hz}, ~~ 1\,{\rm H}, ~~ H-6), ~~ 7.67~({\rm br}~{\rm s}, ~~ 2\,{\rm H}, ~~ H-3, ~~ H-4); ~~ Positive FAB-MS~(rel.~~ int.~~ \%): ~~ 376.1[M+2]^+~(13), 375.1~[M+H]^+~(19), ~~ 277.1~(8), ~~ 255.0~(16), ~~ 185.2~(83), ~~ 157.1~(100); ~~ HRFAB-MS~(+ve~mode)~Calcd.~~ for~[(C_{14}H_{10}N_4O_1Cl_1F_3S_1)+H]^+:~(m/z=375.0294)~~ Found~ 375.0307. \end{split}$$

N-(4-*Chloro-3*-(*trifluoromethyl*)*phenyl*)-2 *isonicotinoylhydrazinecarbothioamide* (9). **R**_f = 0.76, **Mp**: 231–232 °C;**IR** (**KBr**, **cm**⁻¹): 3279, 3125 (N–H), 1675 (C=O), 1551, 1516 (C=C), 1256 (C=S), 1065 (C–C). ¹H-NMR (400 MHz, DMSO-d₆): $\delta_{\rm H}$ 10.92 (s, 1 H, NH), 10.14 (s, 1 H, NH), 10.02 (s, 1 H, NH), 8.77 (d, $J_{3'',2''} = J_{5'',6''} = 5.6$ Hz, 2 H, H-3'', H-5''), 7.99 (br s, 1 H, H-2), 7.85 (br.s, 3 H, H-6'', H-2'', H-5), 7.67 (d, $J_{3,2} = 8.4$ Hz, 1 H, H-6), ¹³C-NMR (125 MHz, DMSO-d₆): δ 180.8 (C=S), 164.4 (C=O), 150.2 (C-3'',C-5''), 139.2 (C-1''), 138.6 (C-1), 131.2 (C-6), 130.5 (C-5), 125.9 (m, CF₃), 124.2 (C-2), 123.7 (C-3), 121.6 (C2'', C-6''), 119.4 (C-4); **Positive FAB-MS m/z (rel. int.** %): 377.2 [M + 2]⁺ (22), 375.1 [M + H]⁺ (32), 277.0 (27), 219 (19), 185.0 (100), 171.1 (96), 157.0 (45); **HRFAB-MS** (+**ve mode**) Calcd. for [(C₁₄H₁₀N₄O₁Cl₁F₃S₁) + H]: (m/z = 375.0294) Found 375.0295.

N-(4-*Cyanophenyl*)-2-*isonicotinoylhydrazinecarbothioamide* (**10**). **R**_f=0.66, **Mp**: 162–163 °C; **IR** (**KBr**, **cm**⁻¹): 3314, 3216 (N–H), 2224 (C≡N), 1675 (C=O), 1543, 1510 (C=C), 1217 (C=S). ¹H-NMR (**400 MHz**, **CD**₃**OD**): $\delta_{\rm H}$ 8.72 (d, $J_{3'',2''} = J_{5'',6''} = 6.0$ Hz, 2 H, H-3", H-5"), 7.88 (d, $J_{2'',3''} = J_{6'',5''} = 6.0$ Hz, 2 H, H-2", H-6"), 7.82 (d, $J_{2,3} = J_{6,5} = 8.4$ Hz, 2 H, H-2, H-6), 7.67 (d, $J_{3,2} = J_{5,6} = 8.4$ Hz, 2 H, H-3, H-5), ¹³C-NMR (**150 MHz**, **DMSO-d**₆): $\delta_{\rm I80.6}$ (C=S), 164.4 (C=O), 150.2 (C-3",C-5"), 143.6 (C-1"), 139.3 (C-1), 132.5 (C-3), 132.1 (C-5), 125.4 (C-2), 122.6 (C-6), 118.9 (C-4), 106.8 (CN); **Positive FAB-MS** *m/z* (**rel. int.** %): 297.1 [M+H]⁺ (4), 277.1 (15), 219.2 (8), 185.0 (100), 171.0 (23), 157.0 (8); **HRFAB-MS** (+**ve mode**) Calcd. for [(C₁₄H₁₁N₅O₁S₁) + H]⁺: (*m/z* = 298.0763) Found 298.0788.

2-Isonicotinoyl-N-(4-(trifluoromethoxy) phenyl) hydrazine carbothioamide (11). $\mathbf{R}_{f} = 0.76$, **Mp**: 192–193 °C; **IR** (**KBr**, **cm**⁻¹): 3271, 3125 (N–H), 1678 (C=O), 1513 (C=C), 1267 (C=S). ¹H-NMR (400 MHz, DMSO- d_{6}): δ_{H} 10.86 (s, 1 H, NH), 9.92 (s, 2 H, NH), 8.76 (d, $J_{3'',2''} = J_{5'',6''} = 5.6$ Hz, 2 H, H-3'', H-5''), 7.83 (d, $J_{2'',3''} = J_{6'',5''} = 5.6$ Hz, 2 H, H-3'', H-5''), 7.84 (d, $J_{2'',3''} = J_{6'',5''} = 5.6$ Hz, 2 H, H-3'', H-5''), 7.83 (d, $J_{2'',3''} = J_{6'',5''} = 5.6$ Hz, 2 H, H-2'', H-6''), 7.54 (br s, 2 H, H-2, H-6), 7.32 (d, $J_{3,2} = J_{5,6} = 8.4$ Hz, 2 H, H-3, H-5), **Positive FAB-MS** *m/z* (rel. int. %): 357.1 [M + H]⁺ (38), 329 (8), 277.1 (15), 236.9 (58), 185.0 (100), 171.0 (19), 157.0 (8); HRFAB-MS (+ve mode) Calcd. for [(C₁₄H₁₁N₄O₂F₃S₁) + H]⁺: (*m/z* = 357.0633) Found 357.0619.

2-Isonicotinoyl-N-(perfluorophenyl) hydrazinecarbothioamide (12). \mathbf{R}_{f} =0.7, Mp: 255–256 °C; IR (KBr, cm⁻¹): 3303, 3150 (N–H), 1682 (C=O), 1528, 1505 (C=C), 1249 (C=S), 1153 (C–F). ¹H-NMR (300 MHz, DMSO-d_6): δ_{H} 11.06 (s, 1 H, NH), 10.44 (s, 1 H, NH), 9.72 (s, 1 H, NH), 8.77 (d, $J_{3'',2''} = J_{5'',6''} = 5.1$ Hz, 2 H, H-3'', H-5''), 7.84 (distorted singlet, 2 H, H-2'', H-6''), ¹³C-NMR (125 MHz, DMSO-d_6): $\delta_{182.6}$ (C=S), 164.4 (C=O), 150.2 (C-3'',C-5''), 144.7 (d, J = 206 Hz, C-2, C-6), 140.5 (t, J = 11.4 Hz, C-1), 139.5 (C-1''), 138.9 (m, C-4), 137.8 (t, J = 11.9 Hz, C-3), 136.2 (t, J = 14.3 Hz, C-3), Positive FAB-MS m/z (rel. int. %): 363.0 [M + H]⁺ (15), 335.0 (4), 277.0 (17), 243.0 (7), 185.2 (100); HRFAB-MS (+ve mode) Calcd. for $[(C_{13}H_7N_4O_1F_5S_1) + H]^+$ (m/z = 363.0339) Found 363.0323.

N-*Cyclopentyl*-2-*isonicotinoylhydrazinecarbothioamide* (**13**). **R**_f = 0.7, **Mp**: 223–224 °C; **IR** (**KBr**, **cm**⁻¹): 3268, 3142 (N–H), 1679 (C=O), 1552, 1527 (C=C), 1451 (CH₂ bending), 1263 (C=S stretching). ¹**H**-**NMR** (**300 MHz**, **DMSO-***d*₆): $\delta_{\rm H}$ 10.54 (s, 1 H, NH), 9.26 (s, 1 H, NH), 8.74 (d, $J_{3'',2''} = J_{5'',6''} = 5.7$ Hz, 2 H, H-3'', H-5''), 7.80 (d, $J_{2'',3''} = J_{6'',5''} = J_{3',4'} = 5.7$ Hz, 3 H, H-2'', H-6'', NH), 4.11 (br s, 1 H, H-1), 1.77–1.55 (m, 4 H, H-2a, H-2b, H-5a, H-5b), 1.27–1.03 (m, 4 H, H-3a, H-3b, H-4a, H-4b), **Negative FAB-MS** *m*/*z* (rel. int. %): 265.2 [M-H]⁺ (4), 219.9 (17), 207.0 (18), 183.0 (100), 164.0 (4); **HRFAB-MS** (+ve mode) Calcd. for. [C₁₂H₁₆N₄OS + H]⁺: (*m*/*z* = 265.1045) Found 265.1043.

$$\begin{split} &N-(2\text{-}Bromophenyl)\text{-}2\text{-}isonicotinoylhydrazinecarbothioamide (14).} \quad \mathbf{R_f}=0.73, \, \mathbf{Mp}\text{: }156-157\ ^\circ\text{C}\text{; IR}\ (\mathbf{KBr}, \mathbf{cm}^{-1})\text{:} \\ &3275, \,3131\ (N-H), \,1679\ (C=O), \,1543, \,1474\ (C=C), \,1251\ (C=S), \,1059\ (C-Br).\ ^1\text{H-NMR}\ (300\ MHz, \, DMSO-d_6)\text{:} \\ &\delta_{H}\ 10.89\ (s, \,1\,\,H, \,\,NH), \,9.90\ (s, \,1\,\,H, \,\,NH), \,9.70\ (s, \,1\,\,H, \,\,NH), \,8.75\ (d, J_{3''2''}=J_{5'',5''}=6.3\ Hz, \,2\,\,H, \,H-3'', \,H-5''), \,7.84\ (d, J_{2''3''}=J_{6'',5''}=5.1\ Hz, \,2\,\,H, \,H-2'', \,H-6''), \,7.65\ (d, \,J_{3,4}=7.8\ Hz, \,1\,\,H, \,H-3), \,7.38\ (br\,s, \,2\,\,H, \,H-4, \,H-6), \,7.19\ (m, \,1\,\,H, \,H-5), \, \text{Positive FAB-MS m/z}\ (rel.\ int.\ \%)\text{: }351.9\ [(M+2)]^+\ (7), \,350.9\ [M+H]^+\ (8), \,277.1\ (15), \,185.2\ (100), \,171.2\ (17), \,\text{HRFAB-MS}\ (+ve\ mode)\ Calcd.\ for\ [(C_{13}H_{11}N_4O_1Br_1S_1)+H]^+\ (m/z=351.9993)\ Found\ 351.9999. \end{split}$$

N-(2, 4-*Difluorophenyl*)-2-*isonicotinoylhydrazinecarbothioamide* (15). **R**_f=0.43, **Mp** 191–192 °C; **IR** (**KBr**, **cm**⁻): 3300, 3141 (N−H), 1682 (C=O), 1550, 1514 (C=C), 1251 (C=S stretching), 1142 (C−F), ¹**H-NMR** (400 MHz,

DMSO- d_6): δ_H 10.91 (s, 1 H, NH), 10.0 (s, 1 H, NH), 9.61 (s,1 H, NH), 8.75 (d, $J_{3'',2''} = J_{5'',6''} = 4.8$ Hz, 2 H, H-3'', H-5''), 7.83 (s, 2 H, H-2'', H-6''), 7.26 (t, $J_{5,6} = J_{6,5} = 8.4$ Hz, 2 H, H-6, H-5), 7.00 (t, $J_{3(4,2)} = 8.2$ Hz, 1 H, H-3); **Electron Ionization Mass spectrometry (direct probe)** m/z 308.1 [M]⁺ (2.2), 274.1 (11.5), 171.0 (100), 106.0 (45.6), 78.0 (43.1); **HREI-MS** Calcd. For [C₁₃H₉N₄O₁F₂S₁]: (m/z = 308.0543) Found 308.0548.

$$\begin{split} &N-(3\text{-}Fluorophenyl)\text{-}2\text{-}isonicotinoylhydrazinecarbothioamide (16).} \quad \mathbf{R_f} = 0.73, \, \mathbf{Mp} \, 174-175 \,^\circ \mathrm{C}; \, \mathbf{IR} \, (\mathbf{KBr}, \, \mathbf{cm}^{-1})\text{:} \\ &3266, 3112 \, (\mathrm{N-H}), 1674 \, (\mathrm{C=O}), 1601, 1515 \, (\mathrm{C=C}), 1238 \, (\mathrm{C=S}), 1142 \, (\mathrm{C-F})\text{;} \, ^1\mathbf{H}\text{-}\mathbf{NMR} \, (\mathbf{400 \, MHz}, \, \mathbf{DMSO-d_6})\text{:} \\ &\delta_{\mathrm{H}} \, 10.86 \, (\mathrm{s}, 1\,\mathrm{H}, \, \mathrm{NH}), 9.93 \, (\mathrm{s}, 2\,\mathrm{H}, \, \mathrm{NH}), 8.76 \, (\mathrm{d}, J_{3'',2''} = J_{5'',6''} = 6\,\mathrm{Hz}, 2\,\mathrm{H}, \, \mathrm{H-3''}, \, \mathrm{H-5''}), 7.83 \, (\mathrm{d}, J_{2'',3''} = J_{6'',5''} = 5.6\,\mathrm{Hz}, \\ &2\,\mathrm{H}, \, \mathrm{H-2''}, \, \mathrm{H-6''}), 7.45 \, (\mathrm{br.s}, 1\,\mathrm{H}, \, \mathrm{H-2}), 7.35 \, (\mathrm{q}, J_{5(4,6)} = 8.0\,\mathrm{Hz}, 1\,\mathrm{H}, \, \mathrm{H-5}), 7.27 \, (\mathrm{d}, J_{6,5} = 8.0\,\mathrm{Hz}, 1\,\mathrm{H}, \, \mathrm{H-6}), 6.98 \, (\mathrm{t}, \\ &J_{4(5,6)} = 7.6\,\mathrm{Hz}, 1\,\mathrm{H}, \, \mathrm{H-4}); \, \mathbf{Positive FAB-MS} \, \mathbf{m/z} \, (\mathrm{rel. int.} \, \%): 291.0 \, [\mathrm{M} + \mathrm{H}]^+ \, (5), 185.1 \, (100), 277.1 \, (15), 219.1 \, (12); \, \mathrm{HRFAB-MS} \, (+\mathrm{ve \ mode}) \, \mathrm{Calcd. for} \, [(\mathrm{C}_{13}\mathrm{H}_{11}\mathrm{FN}_4\mathrm{OS}) + \mathrm{H}]^+ \, (\mathbf{m/z} = 291.0716) \, \mathrm{Found} \, 291.0721. \end{split}$$

 $\begin{array}{l} N-(3-Iodophenyl)-2-isonicotinoylhydrazinecarbothioamide (17). \quad \mathbf{R_f}=0.73, \ \mathbf{Mp}\ 192-193\ ^\circ C; \ \mathbf{IR}\ (\mathbf{KBr}, \mathbf{cm}^{-1}): \\ 3266, \ 3112.3\ (N-H), \ 1674.5\ (C=O), \ 1601, \ 1515\ (C=C), \ 1237.9\ (C=S), \ 851.8\ (C-I). \ ^1\mathbf{H}-\mathbf{NMR}(\mathbf{400\ MHz}, \mathbf{DMSO-d_6}): \\ \delta_{\mathrm{H}}\ 10.85\ (s,\ 1\,\mathrm{H},\ \mathrm{NH}), \ 9.93\ (s,\ 1\,\mathrm{H},\ \mathrm{NH}), \ 9.84\ (s,\ 1\,\mathrm{H},\ \mathrm{NH}), \ 8.76\ (d,\ J_{3'',2''}=J_{5'',6''}=5.6\ \mathrm{Hz},\ 2\,\mathrm{H},\ \mathrm{H}^{-3''}, \\ \mathrm{H}^{-5''}), \ 7.83\ (d,\ J_{2'',3''}=J_{6'',5''}=J_{4,5}=5.2\ \mathrm{Hz},\ 3\,\mathrm{H},\ \mathrm{H}^{-2''},\ \mathrm{H}^{-6''},\ \mathrm{H}^{-4}), \ 7.51\ (t,\ J_{5,4/5,6)}=J_{2,4/2,6)}=6.8\ \mathrm{Hz},\ 2\,\mathrm{H},\ \mathrm{H}^{-2}, \\ 7.12\ (t,\ J_{6,5}=8.0\ \mathrm{Hz},\ 1\,\mathrm{H},\ \mathrm{H}^{-6}); \ \textbf{Positive\ FAB-MS\ m/z\ (rel.\ int.\ \%):\ 399.0\ [\mathrm{M}+\mathrm{H}]^+\ (16),\ 183.2\ (100),\ 275.2\ (19), \\ 243.0\ (7);\ \mathrm{HFAB-MS\ (+ve\ mode)\ Calcd.\ for\ [(C_{13}\mathrm{H}_{12}\mathrm{N}_4\mathrm{O}_1\mathrm{I}_{5}_1)+\mathrm{H}]^+\ (m/z=398.9776)\ Found\ 398.9759. \end{array}$

N-(4-Chlorophenyl)-2-isonicotinoylhydrazine carbothioamide (**18**). **R**_f = 0.73, **Mp** 150−151 °C; **IR** (**KBr**, **cm**⁻¹): 3251.8, 3130.7 (N–H), 1676 (C=O), 1596, 1545 (C=C), 1256 (C=S), 1093.7 (C–Cl). ¹H-NMR (**300** MHz, **DMSO-***d*₆): $\delta_{\rm H}$ 10.86 (s, 1 H, NH), 9.90 (s, 2 H, NH), 8.76 (dd, $J_{3'',2''} = J_{5'',6''} = J_1 = 1.8$, $J_2 = 1.5$ Hz, 2 H, H-3",H-5"), 7.83 (dd, $J_{2'',3''} = J_{6'',5''} = J_1 = 1.5$ Hz, $J_2 = 1.2$ Hz, 2 H, H-2", H-6"), 7.70 (s, 1 H, H-2), 7.47 (appear d, $J_{3,2} = J_{5,6} = 7.3$ Hz, 2 H, H-3, H-5), 7.37 (m, 2 H, H-2, H-6); **Positive FAB-MS** *m/z* (rel. int. %): 307.0 [M+2]⁺ (5), 305 [M⁺] (15), 185.1(100), 277.1 (14), 219.2 (10); **HRFAB-MS** (+ve mode) Calcd. for [(C₁₃H₁₁N₄O₁Cl₁S₁) + H]⁺ (*m/z* = 307.0420) Found 307.0450.

$$\begin{split} &N-(4\text{-}Fluorophenyl)\text{-}2\text{-}isonicotinoylhydrazinecarbothioamide (19).} \quad \mathbf{R_f}=0.73, \, \mathbf{Mp} \, 191-192\,^\circ \mathrm{C}; \, \mathbf{IR} \, (\mathbf{KBr}, \mathbf{cm}^{-1})\text{:} \\ &3201, 3155 \, (\mathrm{N-H}), \, 1684 \, (\mathrm{C=O}), \, 1544, \, 1512 \, (\mathrm{C=C}), \, 1263 \, (\mathrm{C=S}), \, 1223 \, (\mathrm{C-F}). \, ^1 \mathrm{H-NMR} \, (\mathbf{400 \, MHz}, \, \mathbf{DMSO-d_6})\text{:} \\ &\delta_{\mathrm{H}} \, 10.84 \, (\mathrm{s}, 1\, \mathrm{H}, \mathrm{NH}), \, 9.82 \, (\mathrm{s}, 2\, \mathrm{H}, \mathrm{NH}), \, 8.76 \, (\mathrm{d}, J_{3'',2''}=J_{5'',6''}=6\, \mathrm{Hz}, \, 2\, \mathrm{H}, \, \mathrm{H-3''}, \, \mathrm{H5''}), \, 7.83 \, (\mathrm{d}, J_{2'',3''}=J_{6'',5''}=5.6\, \mathrm{Hz}, \\ &2\, \mathrm{H}, \, \mathrm{H-2''}, \, \mathrm{H-6''}), \, 7.39 \, (\mathrm{br}, \mathrm{s}, 2\, \mathrm{H}, \, \mathrm{H-2}, \, \mathrm{H-6}), \, 7.16 \, (\mathrm{t}, \, J_{3,2/3,4}=J_{5.6/5,4}=8.8\, \mathrm{Hz}, \, 2\, \mathrm{H}, \, \mathrm{H-3}), \, \mathrm{Positive} \, \mathrm{FAB-MS} \\ &m/z \, (\mathrm{rel. \, int} \, \, \%) \text{:} \, 291.0 \, [\mathrm{M}+\mathrm{H}]^+ \, (\mathrm{6}), \, 185.1 \, (100), \, 277.1 \, (14), \, 219.2 \, (10); \, \mathrm{HRFAB-MS} \, (+\mathrm{ve \ mode}) \, \mathrm{Calcd. \ for} \\ & [(\mathrm{C}_{13}\mathrm{H}_{11}\mathrm{N}_4\mathrm{O}_1\mathrm{F}_1\mathrm{S}_1) + \mathrm{H}]^+ \, (m/z = 291.0716) \, \mathrm{Found} \, 291.0730. \end{split}$$

2-Isonicotinoyl-N-(4-methoxyphenyl) hydrazinecarbothioamide (20). $\mathbf{R}_{f} = 0.66$, Mp: 171–172 °C; IR (KBr, cm⁻¹): 3255, 3124 (N–H), 1675 (C=O), 1545, 1514 (C=C), 1253 (C=S). ¹H-NMR (400 MHz, DMSO- d_{6}): δ_{H} 10.79 (s,1 H, NH), 9.72 (s,1 H, NH), 9.67 (s,1 H, NH), 8.75 (d, $J_{3'',2''} = J_{5'',5''} = 5.6$ Hz, 2 H, H-3'', H-5''), 7.83 (d, $J_{2'',3''} = J_{6'',5''} = 5.6$ Hz, 2 H, H-2'', H-6''), 7.25 (d, $J_{3,2} = J_{5,6} = 7.6$ Hz, 2 H, H-2, H-6), 6.88 (d, $J_{2,3} = J_{6,5} = 9.2$ Hz, 2 H, H-3, H-5), 3.11 (s, 3 H, H-7); Positive FAB-MS *m/z* (rel. int. %): 303.0 [M + H]⁺ (4), 185.1(100), 277.1(12), 219.1(8); HRFAB-MS (+ve mode) Calcd. for [(C₁₄H₁₄N₄O₂S₁) + H]⁺ (*m/z* = 303.0916) Found 303.0924.

Isonicotinoyl-N-(2, 4, 5-trichlorophenyl) hydrazinecarbothioamide (21). $\mathbf{R}_{\mathbf{f}}$ = 0.73, Mp: 241–242°C; IR (KBr, cm⁻¹): 3282, 3127 (N–H), 1678 (C=O), 1561, 1510, (C=C), 1259 (C=S), 1079 (C–Cl). ¹H-NMR (300 MHz, DMSO-d_6): 10.94 (s, 1 H, NH), 10.14 (s, 1 H, NH), 9.79 (s, 1 H, NH), 8.76 (d, $J_{3'',2''} = J_{5'',6''} = 6$ Hz, 2 H, H-3'', H-5''), 7.92 (s, 1 H, H-2), 7.83 (br s, 2 H, H-2'', H-6''), 7.61 (br s, 1 H, H-5); ¹³C-NMR (125 MHz, DMSO-d_6): $\delta 181.9$ (C=S), 164.5 (C=O), 150.2 (C-3'',C-5''), 139.3 (C-1''), 137.0 (C-1), 132.0 (C-2), 131.4 (C-4), 130.4 (C-5), 129.9 (C-6), 129.2 (C-3)121.7 (C2'',C-6''); Positive FAB-MS *m/z* (rel. int. %): 377.0 [M + 2]⁺ (17),375.2 [M + H]⁺, (19), 275.3 (25), 183.1 (100), 136.1 (14), 127.1 (10). HRFAB-MS (+ve mode) Calcd. for [(C₁₃H₉N₄O₁Cl₃S₁) + H]⁺ (*m/z* = 374.9641) Found 374.9633.

$$\begin{split} &N-(3\text{-}Bromophenyl)\text{-}2\text{-}isonicotinoylhydrazinecarbothioamide (22).} \quad R_f = 0.73, \text{ Mp: } 192-193 \ ^\circ\text{C}; \text{ IR (KBr, cm}^{-1})\text{:} \\ &3301, 3143 \ (N-H), 1683 \ (C=O), 1589, 1548 \ (C=C), 1252 \ (C=S), 1066 \ (C-Br). \ ^1\text{H-NMR} \ (400 \ \text{MHz}, \text{DMSO-d}_6)\text{:} \\ & \delta_H \ 10.86 \ (s, 1\ H, \ NH), 9.95 \ (s, 1\ H, \ NH), 9.88 \ (s, 1\ H, \ NH), 8.76 \ (d, \ J_{3'',2''} = J_{5'',6''} = 5.6 \ \text{Hz}, 2\ H, \ H^{-3''}, \ H^{-5''}), 7.83 \\ & (d, J_{2'',3''} = J_{6'',5''} = 4.8 \ \text{Hz}, 2\ H, \ H^{-2''}, \ H^{-6''}), 7.70 \ (s, 1\ H, \ H^{-2}), 7.49 \ (d, \ J_{4,5} = 7.6 \ \text{Hz}, 1\ H, \ H^{-4}), 7.32 \ (d, J_{6,5} = 7.6 \ \text{Hz}, 1\ H, \ H^{-4}), 7.32 \ (d, J_{6,5} = 7.6 \ \text{Hz}, 1\ H, \ H^{-6}), 7.28 \ (t, \ J_{5,4/5,6} = 8.0 \ \text{Hz}, 1\ H, \ H^{-5}); \ \text{Positive FAB-MS m/z (rel. int. \ \%): } 353.0[(M+2]^+ \ (14), \ 351.0 \ [M+H]^+ \ (15), 185.0 \ (100), 277.2 \ (14), 219.1 \ (4); \ \text{HRFAB-MS} \ (+ve\ mode) \ Calcd. \ for \ [(C_{13}H_{11}N_4O_1S_1Br_1) + H]^+ \ (m/z = 350.9915) \ \text{Found } 350.9895. \end{split}$$

N-*Cyclohexyl-2-isonicotinoylhydrazinecarbothioamide* (23). $\mathbf{R}_{\mathbf{f}} = 0.71$, **Mp:** 212–213 °C; **IR** (**KBr**, **cm**⁻¹): 3268, 3141 (N–H), 1680 (C=O), 1551, 1530 (C=C), 1452 (CH₂ bending), 1266 (C=S). ¹H-NMR (300 MHz, **DMSO-***d*₆): $\delta_{\mathbf{H}}$ 10.56 (s, 1 H, NH), 9.28 (s, 1 H, NH), 8.76 (dd, $J_{3'',2''} = J_{5'',6'} = 1.5$ Hz, 2 H, H-3", H-5"), 7.81 (d, $J_{2''-3''} = J_{6''-5''} = 6.0$ Hz, 3 H, H-2", H-6", NH), 4.11 (m, 1 H, H-1), 1.78 (m, 4 H, H-2a, H-2b, H-6a, H-6b), 1.60 (d, $J_{5a,6} = 12.3$ Hz, 1 H, H-5a), 1.28 (m, 4 H, H-3a, H-3b, H-4a, H-4b), 1.07 (m, 1 H, H-5b); **Positive FAB-MS** *m/z* (rel. int. %): 279.1 [M + H]⁺ (25), 185.1 (100), 219.2 (6), 251.2 (4); **HRFAB-MS** (+ve mode) Calcd. for [(C₁₃H₁₈N₄O₁S₁) + H]⁺ (*m/z*=279.1280) Found 279.1273.

2-Isonicotinoyl-N-phenylhydrazinecarbothioamide (24). R_f=0.7, Mp: 188–189 °C; IR (KBr, cm⁻¹): 3262, 3120 (N–H), 1675 (C=O), 1547, 1512 (C=C), 1254 (C=S), ¹H-NMR (300 MHz, DMSO-d₆): δ_H 10.84 (s, 1 H, NH),

9.84 (s, 1 H, NH), 9.78 (s,1 H, NH), 8.76 (m, 2 H, H-3", H-5"), 7.83 (d, $J_{2",3"} = J_{6",5"} = 6.0$ Hz, 2 H, H-2", H-6"), 7.41 (d, $J_{2,3} = J_{6,5} = 7.2$ Hz, 2 H, H-2, H-6), 7.32 (t, $J_{3,2} = J_{4,5} = J_{4,6} = 7.8$ Hz, 2 H, H-3, H-5), 7.15 (t, $J_{4,3} = J_{4,6} = 7.2$ Hz, 1 H, H-4); **Positive FAB-MS** *m/z* (rel. int. %): 273.1 [M + H]⁺ (15), 185.1 (100), 219.1 (8), 263.1 (5), 191.1 (4); **HRFAB-MS** (+ve mode) Calcd. for [(C₁₃H₁₂N₄O₁S₁) + H]⁺ (*m/z* = 273.0810) Found 273.0799.

N-*Hexyl*-2-*isonicotinoylhydrazinecarbothioamide* (25). **R**_f = 0.46, **Mp** 215–216 °C; **IR** (**KBr**, **cm**⁻¹): 3300.5, 3170.6 (N–H), 1677.4 (C=O), 1555, 1527 (C=C), 1245 (C=S), 753 (long chain band). ¹H-NMR (400 MHz, **DMSO-***d*₆): $\delta_{\rm H}$ 10.57 (s, 1 H, NH), 9.29 (s, 1 H, NH), 8.74 (d, $J_{3'',2''} = J_{5'',6''} = 6.0$ Hz 2 H, H-3", H-5"), 8.12 (s, 1 H, NH), 7.79 (d, $J_{2'',3''} = J_{6'',5''} = 5.6$ Hz, 2 H, H-2", H-6"), 3.42 (m, 2 H, H-1a, H-1b), 1.46 (m, 2 H, H-2a, H-2b), 1.23 (m, 6 H, H-3a, H-3b, H-4a, H-4b, H5a, H-5b), 0.84 (m, 3 H, H-6a, H-6b, H-6c), **Negative FAB-MS** *m/z* (rel. int. %): 279 0 [M-H]⁺ (23), 275 (19), 255.1 (8), 183 (100), **HRFAB-MS** (–ve mode) Calcd. for [(C₁₃H₂₀N₄O₁S₁) – H]: (*m/z* = 279.0811) Found 279.0810.

 $\begin{array}{l} N-(3-Thiocyanophenyl)-2-isonicotinoylhydrazinecarbothioamide (26). R_{\rm f}=0.39, Mp: 195-196 \ {}^\circ C; IR (KBr, cm^{-1}): 3283, 3139 (N-H), 2118 (N=C=S), 1678 (C=O), 1595, 1548 (C=C), 1241 (C=S). {}^1 H-NMR (300 MHz, DMSO-d_6): \delta_{\rm H} 10.89 (s, 1 H, NH), 9.91 (s, 2 H, NH), 8.76 (m, 2 H, H-3'', H-5''), 7.83 (d, <math>J_{2'',3''}=J_{6'',5''}=5.7$ Hz, 2 H, H-2'', H-6''), 7.56 (s, 1 H, H-2), 7.47 (d, $J_{6,5}=8.4$ Hz, 1 H, H-6), 7.39 (m, 1 H, H-5), 7.21 (d, $J_{4,5}=7.8$ Hz, 1 H, H-4), 1 C-NMR (125 MHz, DMSO-d_6): $\delta 180.8 (C=S)$, 164.4 (C=O), 150.2 (C-3'',C-5''), 140.4 (C-1''), 139.5 (C-1), 139.4 (C-3), 134.1 (NCS), 129.4 (C-5), 125.2 (C-6), 122.8 (C-4), 122.4 (C-2) 121.6 (C2'', C-6''); Negative FAB-MS *m/z* (rel. int. %): 328.1 [M-H]^+ (15), 275.1 (44), 183 (100), HRFAB-MS (-ve mode) Calcd. for [(C₁₄H₁₁N₅OS₂) - H]: (*m/z* = 328.0320) Found 328.0315. \\ \end{array}

2-Isonicotinoyl-N-(3-nitrophenyl) hydrazinecarbothioamide (27). $\mathbf{R}_{\mathbf{f}} = 0.7$, **Mp**: 159–160 °C; **IR** (**KBr**, **cm**⁻¹): 3284, 3124 (N–H), 1676 (C=O), 1597, 1531 (C=C), 1477 (N=O), 1247 (C=S). ¹H-NMR (300 MHz, DMSO-*d*₆): $\delta_{\mathbf{H}} 10.94$ (s, 1 H, NH), 10.15 (s, 2 H, NH), 8.78 (d, $J_{3'',2''} = J_{5'',6''} = 6.0$ Hz, 2 H, H-3'', H5''), 8.42 (bs, 1 H, H-2), 7.99 (t, $J_{4,5} = J_{6,5} = 6.6$ Hz, 2 H, H-4, H-6), 7.85 (d, $J_{2'',3''} = J_{6'',5''} = 5.7$ Hz, 2 H, H-2'', H-6''), 7.61 (t, $J_{5,4/5,6)} = 8.1$ Hz, 1 H, H-5), **Negative FAB-MS** *m/z* (**rel. int.** %): 316 [M-H]⁺ (13), 275 (23), 183 (100), **HRFAB-MS** (–**ve mode**) Calcd. for [(C₁₃H₁₁N₅O₃S) – H]: (*m/z* = 316.1281) Found 316.1280.

Bioassays. Protocol for In Vitro Urease Inhibition Assay. The urease inhibition activity of compounds 3–27 was evaluated by using the method reported by Weatherburn. *et al.* (1967). Thiourea and acetohydraoxamic acid were used as standard compounds⁴²⁻⁴⁴. During the experiments all the compounds were evaluated at 0.5 mM each in triplicate. The compounds with >50% (greater than 50%) inhibition were further studied to determine their IC₅₀ value in a separate experiment where, different concentration of the compound from 0.5–0.0078125 mM were tested in triplicates. On the other hand if compound showed <50% (less than 50%) inhibition at 0.5 mM then the compound was considered inactive.

Protocol for In Vitro Anti-Inflammatory Assay. Oxidative Burst Assay: Anti-inflammatory activity of the thiosemicarbazide derivatives **3–27**, and isoniazid (**1**) was evaluated by following the method reported in Helfand. *et al.* (1982)^{45,46}. In this experiment all the compounds were evaluated at $25 \mu g/mL$, each in triplicate. To determine the IC₅₀ values, the compounds with >50% inhibition were further evaluated on three different concentrations (1, 10 and 100 $\mu g/mL$). While, the compound failed to inhibit the production of ROS from zymosan activated whole blood cells at highest used dose (100 $\mu g/mL$) was considered as inactive.

All studies on human blood cells was carried out after an approval from independent ethics committee (Prof. Dr. Ghazala H. Rizwani Chair, IEC, Prof. Qamar Amin, Prof, Dr. Muddasir Uddin, Dr. Shahnaz Ghazi, Dr. SamiuzZaman, Prof. Dr. Ahsana Dar Farooq, Dr. M. Raza Shah member IEC), International Center for Chemical and Biological Sciences, University of Karachi, No: ICCBS/IEC-008-BC-2015/Protocol/1.0. Informed consents were obtained from the volunteers before drawing the blood. All the experiments were performed in accordance with relevant guidelines and regulations.

Protocol for Cytotoxicity (MTT assay) Assay. 3T3 Cytotoxicity Assay: Cytotoxicity of the thiosemicarbazide derivatives **3–27**, and isoniazid (1) was evaluated by the method reported by Pauwels. *et al.* (1988)^{47,48}. In this experiment all the compounds were evaluated at 30 μ M each in triplicate. If the compound showed >50% inhibition then for the determination of IC₅₀ value different concentration of the compound from 30–0.9375 μ M were tested. While if the compound showed <50% inhibition at 30 μ M then the compound was considered to be inactive.

Data Availability

All the supplementary mateial is available.

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Author Contributions

The concept of presented research was developed, and designed by M.I.C. and H.S.. All the data were analysed and interpreted by M.I.C., H.S. and F.R., M.K., A.J. F.R., M.K. and A.J. performed the experiments. M.I.C., H.S., F.R. wrote the manuscript. All the authors checked and revised the manuscript. The final version of manuscript was also approved by M.I.C. before submission.

Additional Information

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