

# SCIENTIFIC REPORTS

OPEN

## Application of spin-crossover water soluble nanoparticles for use as MRI contrast agents

Asami Tsukiashi<sup>1</sup>, Kil Sik Min<sup>2</sup>, HIKARU Kitayama<sup>1</sup>, Hiroaki Terasawa<sup>3</sup>, Sosuke Yoshinaga<sup>3</sup>, Mitsuhiro Takeda<sup>3</sup>, Leonard F. Lindoy<sup>4</sup> & Shinya Hayami<sup>1,5</sup>

Water soluble spin-crossover (SCO) iron(II) nanoparticles (NPs) were synthesized by the polyethylene glycol (PEG) coating of  $[\text{Fe}(\text{Htrz})_{3-3x}(\text{NH}_2\text{trz})_{3x}](\text{BF}_4)_2$  ( $x = 0, 0.1, 0.5$  and  $1$ ). The NPs with  $x = 0.1$  show gradual SCO behavior over 280–330 K in water. The relaxation times,  $T_1$  and  $T_2$ , were determined and the thermally-responsive  $T_2$  values making these NPs a candidate for use as a MRI contrast agent.

Magnetic resonance imaging (MRI) is a useful diagnostic medical tool for the non-invasive visualization of internal body parts. For the imaging, the hydrogen nucleus is used because of its abundance in water and fat. The time taken for the protons to fully relax is measured in two ways. The first is the time taken for the magnetic vector to return to its resting state and the second is the time needed for the axial spin to return to its resting state. The first is called  $T_1$  relaxation, the second is called  $T_2$  relaxation. A MRI contrast agent is normally employed to enhance the MRI image. Commonly, Gd-chelate compounds<sup>1</sup> and iron oxide materials<sup>2,3</sup> are used as MRI contrast agents. The Gd-chelate compounds are especially effective owing to the large spin quantum number of Gd(III) ( $S = 7/2$ ). Nevertheless, nephrogenic systemic fibrosis (NSF) is known as a serious kidney disease that can result from the use of such Gd-chelate contrast agents<sup>4-6</sup>. This provides a motivation for the development of new contrast agents not based on Gd(III).

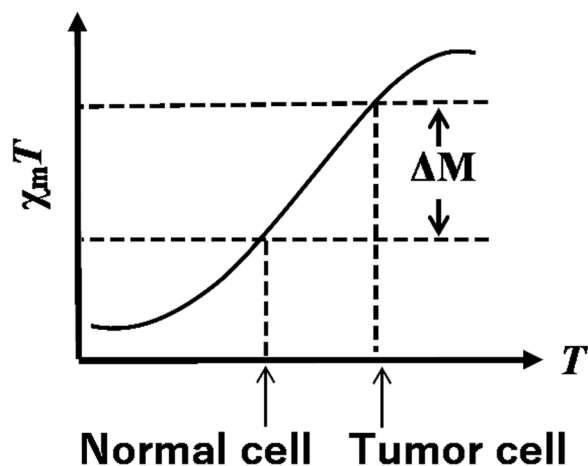
Recent studies have demonstrated that SCO materials can potentially be employed as MRI contrast agents<sup>7,8</sup>. Spin crossover (SCO) materials undergo a change in their spin state between high-spin (HS) and low-spin (LS) on exposure to external stimuli, i.e., temperature, pressure, magnetic field, and light irradiation. Such materials have been intensively investigated for use in applications such as information storage, sensor development, and display technologies<sup>9-11</sup>. SCO iron(II) complexes undergo a spin state change ( $\Delta M$  in Fig. 1) between LS ( $S = 0$ ) and HS ( $S = 2$ ) that is temperature dependent. Generally, tumor cells are more thermally-sensitive than normal cells, and the temperature of tumor cells can be readily increased by means of hyperthermia treatment<sup>12,13</sup>. In view of this, SCO materials can in principle be employed as MRI contrast agents by using the change of their magnetism with temperature when the temperature of tumor cells is higher than normal cells.

In this work, we focused on the use of iron(II) SCO materials of type  $[\text{Fe}(\text{Htrz})_{3-3x}(\text{NH}_2\text{trz})_{3x}](\text{BF}_4)_2$  ( $\text{Htrz} = 1,2,4\text{-IH-triazole}$ ,  $\text{NH}_2\text{trz} = 4\text{-NH}_2\text{-1,2,4-triazole}$ , and  $x = 0, 0.1, 0.5$  and  $1$ ) as candidates for use as new contrast agents<sup>14-16</sup>. For practical application in MRI, water dispersion is important and to this end we synthesized the SCO nanoparticles (NPs) with a PEG coating. Here, we report a study of these thermally-responsive MRI contrast agents based on the SCO NPs.

Magnetic susceptibility measurements for the NPs with  $x = 0, 0.1, 0.5$  and  $1$  in aqueous dispersion were performed by NMR measurements. The  $\chi_m T$  values ( $\chi_m$ , molar magnetic susceptibility;  $T$ , temperature) were estimated by the Evans method employing eq. (1) below (Fig. S5)<sup>17-19</sup>; the calculation parameters are given in Table S2. All NPs showed gradual SCO behaviour over the temperature range 283–323 K.

$$\chi_M^P = \frac{3\Delta f M^P}{4\pi f m} + \chi^0 M^P + \frac{\chi^0 M^P (d_0 - d_s)}{m} - \chi_M^{dia} \quad (1)$$

<sup>1</sup>Department of Chemistry, Graduate School of Science and Technology, Kumamoto University, 2-39-1 Kurokami, Chuo-ku, Kumamoto, 860-8555, Japan. <sup>2</sup>Department of Chemistry Education and Green-Nano Materials Research Center, Kyungpook National University, Daegu, 41566, Republic of Korea. <sup>3</sup>Department of Structural BiImaging, Faculty of Life Sciences, Kumamoto University, 5-1 Oe-honmachi, Chuo-ku, Kumamoto, 862-0973, Japan. <sup>4</sup>School of Chemistry, The University of Sydney, Sydney, NSW, 2006, Australia. <sup>5</sup>Institute of Pulsed Power Science (IPPS), Kumamoto University, 2-39-1 Kurokami, Chuo-ku, Kumamoto, 860-8555, Japan. Correspondence and requests for materials should be addressed to S.H. (email: hayami@kumamoto-u.ac.jp)



**Figure 1.** Application of SCO to MRI.

- $\chi_M^P$ : Molar paramagnetic susceptibility in  $\text{cm}^3 \text{mol}^{-1}$ .  
 $\Delta f$ : Frequency difference between the two peaks of inner and outer tube in Hz.  
 $M^P$ : Molecular weight of the substance in  $\text{g mol}^{-1}$ .  
 $f$ : Frequency of NMR instruments in Hz.  
 $m$ : Mass of the substance in 1 mL of solution in  $\text{g mL}^{-1}$ .  
 $\chi^0$ : Mass susceptibility of the solvent in  $\text{cm}^3 \text{mol}^{-1}$ .  
 $d_0$ : Density of the solvent in  $\text{g cm}^{-3}$ .  
 $d_s$ : Density of the solution in  $\text{g cm}^{-3}$ .  
 $\chi_M^{dia}$ : Diamagnetic correction for the magnetic susceptibility in  $\text{cm}^3 \text{mol}^{-1}$ .

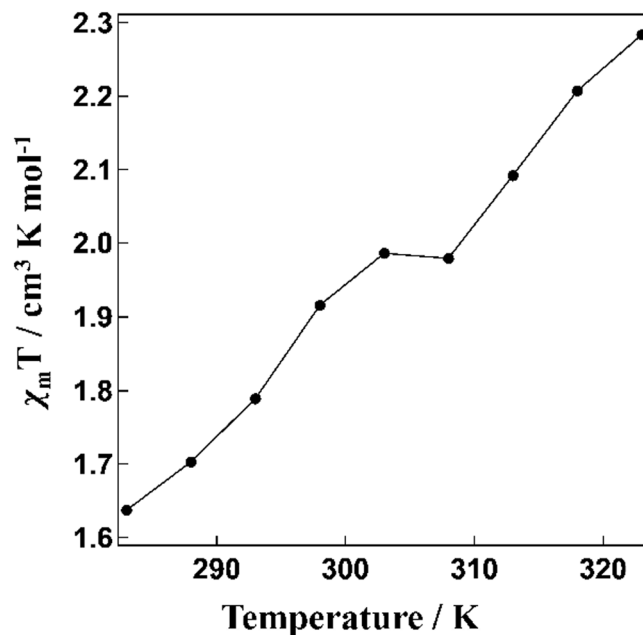
## Results

Of all the NPs investigated,  $[\text{Fe}(\text{Htrz})_{2.7}(\text{NH}_2\text{trz})_{0.3}](\text{BF}_4)_2$  (**1**) exhibited the steepest SCO behaviour (Fig. 2; that is, it showed the largest temperature dependence of  $\chi_m T$ ). Hence, we employed this NP derivative for use in our MRI investigation. The synthesized NPs **1** were dispersed in water and measured by transmission electron microscopy (TEM) to confirm that nanoparticulation had occurred. As estimated by TEM, the particle size was mainly around 20 nm (Fig. 3). The powder X-ray diffraction (PXRD) patterns for compound **1** and its corresponding NPs (NPs **1**) were determined at room temperature. The patterns for **1** and its NPs are almost the same, in accord with the NPs being successfully synthesized with the structure of **1** being retained intact (Fig. S2). NPs **1** were dispersed in water and investigated by NMR to obtain the  $T_1$  and  $T_2$  relaxation times (Figs 4 and 5). In the absence of NPs, water molecules give a  $T_{10}$  relaxation of 2.94 s at 20 °C. On the other hand, the  $T_1$  relaxation time was 2.03 s for the aqueous dispersion of NPs at 20 °C. NMR measurements were also carried out at higher temperatures: 50 °C for water and the aqueous dispersion of NPs. The  $T_{10}$  relaxation time for water was 4.70 s (50 °C), while for the aqueous dispersion of NPs it was  $T_1 = 3.20$  s (50 °C). The correlation between  $T_{10}$  and  $T_1$  relaxation times vs. temperature is shown in Fig. 4. We observed that the  $T_1$  relaxation time of water molecules with or without any contrast agent becomes longer with increasing temperature. The slope of the linear  $T_1$  vs  $T$  plot for water is 0.1215 (Fig. 4). However, in the case of the aqueous dispersion of NPs, the slope is only 0.0395. This difference between the slopes indicates that the presence of the NPs strongly shortens the  $T_1$  relaxation time at higher temperature (50 °C) compared to the lower temperature (20 °C). As such, these results demonstrate that a temperature-dependent shortening effect on the proton relaxation time occurs. The relaxivity ( $r$ ) of the MRI contrast agent was calculated using equations (2 and 3).

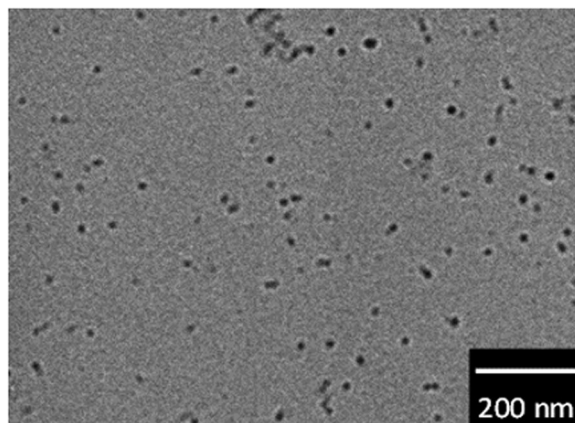
$$\frac{1}{T_{10}} = \frac{1}{T_1} - r_1 [\text{C}] \quad (2)$$

$$\frac{1}{T_{20}} = \frac{1}{T_2} - r_2 [\text{C}] \quad (3)$$

- $T_{10}$ :  $T_1$  relaxation time for original sample (s).  
 $T_1$ :  $T_1$  relaxation time shortened by contrast agent (s).  
 $r_1$ : Relaxivity for  $T_1$  relaxation time ( $\text{mM}^{-1} \text{s}^{-1}$ ).  
 $T_{20}$ :  $T_2$  relaxation time for original sample (s).  
 $T_2$ :  $T_2$  relaxation time shortened by contrast agent (s).  
 $r_2$ : Relaxivity for  $T_2$  relaxation time ( $\text{mM}^{-1} \text{s}^{-1}$ ).  
 $C$ : Concentration of contrast agent (mM).



**Figure 2.**  $\chi_m T$  vs  $T$  plot for NPs 1 in water.

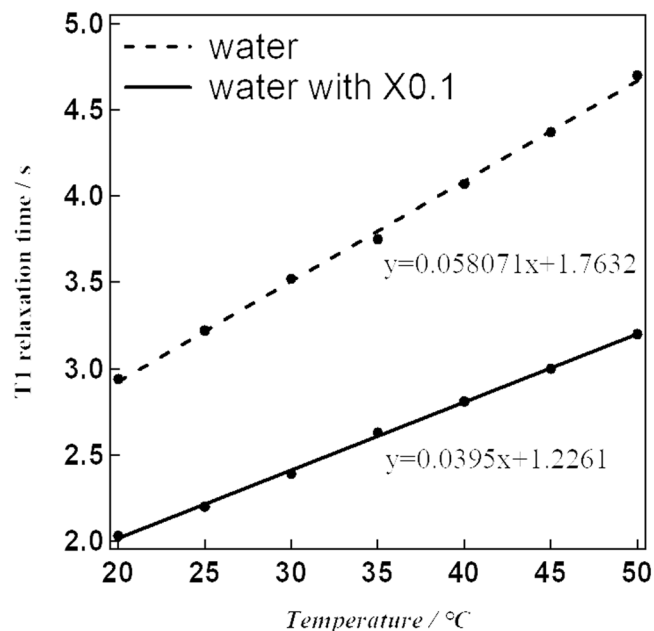


**Figure 3.** TEM image of NPs 1.

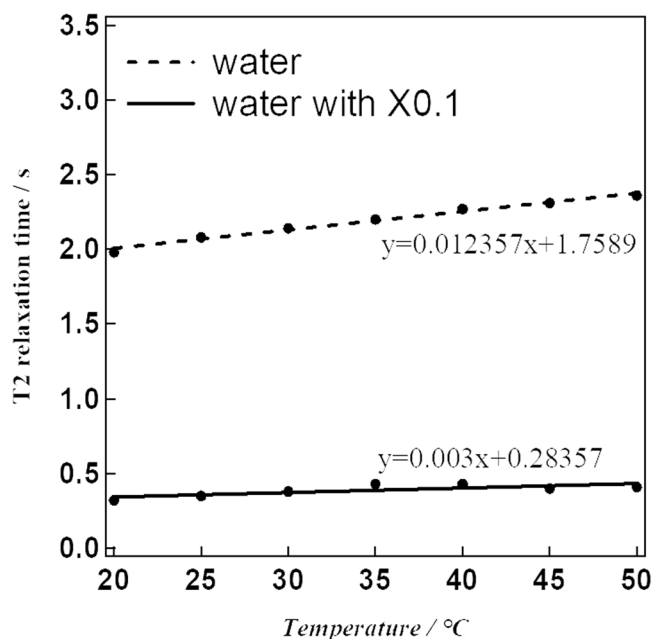
## Discussion

The  $T_1$  relaxivity ( $r_1$ ) values were  $0.16 \text{ mM}^{-1} \text{ s}^{-1}$  at  $20^\circ\text{C}$  and  $0.10 \text{ mM}^{-1} \text{ s}^{-1}$  at  $50^\circ\text{C}$ . The corresponding proton relaxation time of water at  $50^\circ\text{C}$  was calculated using the linear equation given in Fig. 4; the calculation parameters are listed in Table 1. The calculated  $r_1$  value is far from that for the commonly used MRI contrast agent, Gd-DTPA<sup>20</sup>.  $T_2$  relaxation times were also obtained by NMR measurements (Fig. 5). At  $20^\circ\text{C}$ , the values were 1.98 for water and 0.32 s for the aqueous dispersion of NPs. In higher temperature experiments, the  $T_2$  relaxation times were 2.36 s for water at  $50^\circ\text{C}$  and 0.41 s for the aqueous dispersion of NPs at  $50^\circ\text{C}$ . The slope of the linear plot for the  $T_2$  relaxation time of water was 0.003. While for the aqueous dispersion of NPs, the slope was effectively zero. As for the  $T_1$  relaxation times, this result shows that the presence of the NPs strongly shortens the  $T_2$  proton relaxation time at the higher temperature ( $50^\circ\text{C}$ ). The calculated  $T_2$  relaxivities ( $r_2$ ) were  $2.44 \text{ mM}^{-1} \text{ s}^{-1}$  at  $20^\circ\text{C}$  and  $1.91 \text{ mM}^{-1} \text{ s}^{-1}$  at  $50^\circ\text{C}$ , respectively. The calculated parameters are shown in Table 1. The ratio of transverse/longitudinal relaxivity,  $r_2/r_1$ , was found to be 19.1 at  $50^\circ\text{C}$ , which seems an effective value for the use as a negative contrast agent.

In conclusion, we have synthesized the iron(II) triazole NPs (1). The NPs showed gradual spin transition around 300 K in aqueous dispersion and temperature-dependent shortened proton relaxation times. The calculated  $r_2/r_1$  value indicates that NPs is a candidate for use as a thermally-responsive  $T_2$  shortening MRI contrast agent.



**Figure 4.**  $T_1$  relaxation time vs temperature plot.



**Figure 5.**  $T_2$  relaxation time vs temperature plot.

Temperature	$T_{1_0}$	$T_1$	$r_1$	$T_{2_0}$	$T_2$	$r_2$
20°C	2.94	2.03	0.16	1.98	0.32	2.44
50°C	4.7	3.2	0.10	2.36	0.41	1.91

**Table 1.** Calculation parameters for  $T_1$  relaxivity ( $r_1$ ) and  $T_2$  relaxivity ( $r_2$ ). Concentration;  $1.0 \times 10^{-3}$  M.

## Methods

**Physical measurements.** The magnetic susceptibility in the solution was carried out with a JEOL (500-ECX) instrument operating at 500 MHz by the Evens method using  $D_2O$ . The inner tube was filled with 1% *t*-BuOH in  $D_2O$  solution. The NPs were dispersed in  $D_2O$  solution containing 2% *t*-BuOH, and the dispersion liquid was added to the outer tube. Transmission electron microscopy (TEM) was performed to determine particle

sizes (JEOL, 2000FX, 200 kV). NPs were dispersed in water and deposited on a holey carbon film. Powder X-ray diffractograms were recorded on a Rigaku X-ray diffractometer (RAD-2A with a 2.0 kW CuK $\alpha$  X-ray). The  $T_1$  and  $T_2$  relaxation times were performed on  $^1\text{H}$  NMR spectroscopy. The experiments were carried out with a Bruker AvanceIII at 500 MHz. The water sample was measured with the aid of double-walled NMR tube. The water sample was prepared by  $\text{H}_2\text{O}$  was put in the inner tube and 99.8% of  $\text{D}_2\text{O}$  was added to the outer tube. The NPs were dissolved in mixed solution 97%  $\text{H}_2\text{O}$  and 3%  $\text{D}_2\text{O}$  at a concentration of 1 mM and measured with single NMR tube. The magnetic susceptibilities in the solid state for NPs and bulk samples were measured between 100 K and 400 K with a superconducting quantum interference device (SQUID) magnetometer (Quantum Design MPMS-XL). Atomic absorption analysis was performed to measure the ratio of SCO complex in the NPs with an atomic absorption spectrometer (PerkinElmer, PinAAcle 500). Each NP sample (10 mg) was dissolved in 100 mL of 0.1 M nitric acid aqueous solution before measurement (Fig. S1 and Table S1).

## References

- Huang, Y. *et al.* Chitosan oligosaccharide based Gd-DTPA complex as a potential bimodal magnetic resonance imaging contrast agent. *Magnetic Resonance Imaging* **34**, 1–7 (2016).
- Basti, H. *et al.* Catechol derivatives-coated  $\text{Fe}_3\text{O}_4$  and  $\gamma\text{-Fe}_2\text{O}_3$  nanoparticles as potential MRI contrast agents. *J. Colloid. Interface. Sci* **341**, 248–254 (2010).
- Thapa, B., Diaz-Diestra, D., Beltran-Huarac, J., Weiner, B. R. & Morell, G. Enhanced MRI  $T_2$  Relaxivity in Contrast-Probed Anchor-Free PEGylated Iron Oxide Nanoparticles. *Nano. Res. Lett* **12**, 312–324 (2017).
- Abujudeh, H. H. *et al.* Nephrogenic systemic fibrosis after gadopentetate dimeglumine exposure: case series of 36 patients. *Radiology* **1**, 81–89 (2009).
- Hellman, R. N. Gadolinium-induced nephrogenic systemic fibrosis. *Semin. Nephrol.* **3**, 310–316 (2011).
- Abu-Alfa, A. K. Nephrogenic systemic fibrosis and gadolinium-based contrast agents. *Adv. Chronic Kidney Dis.* **3**, 188–198 (2011).
- Jeon, I.-R., Park, J. G., Haney, C. R. & Harris, T. D. Spin crossover iron(II) complexes as PARACEST MRI thermometers. *Chem. Sci.* **5**, 2461–2465 (2014).
- Venkataramani, S. *et al.* Magnetic bistability of molecules in homogeneous solution at room temperature. *Science* **331**, 445–448 (2011).
- Sato, O., Iyoda, T., Fujishima, S. & Hashimoto, K. Photoinduced Magnetization of a Cobalt-Iron Cyanide. *Science* **272**, 704–705 (1996).
- Chastanet, G., Gaspar, A. B., Real, J. A. & Létard, J.-F. Photo-switching spin pairs—synergy between LIESST effect and magnetic interaction in an iron(II) binuclear spin-crossover compound. *Chem. Commun* **0**, 819–820 (2001).
- Hayami, S. *et al.* Photo-Induced Spin Transition of Iron(III) Compounds with  $\pi$ - $\pi$  Intermolecular Interactions. *Chem. Eur. J.* **15**, 3497–3508 (2009).
- Lawson, R. N. & Chughtai, M. S. Breast Cancer and Body Temperature. *Canad. Med. Ass. J.* **88**, 68–70 (1963).
- Hayashi, K. *et al.* Magnetically Responsive Smart Nanoparticles for Cancer Treatment with a Combination of Magnetic Hyperthermia and Remote-Control Drug Release. *Theranostics* **4**, 834–844 (2014).
- Krober, J., AudiBre, J.-P., Claude, R., Codjovi, E. & Kahn, O. Spin Transitions and Thermal Hysteresis in the Molecular-Based Materials  $[\text{Fe}(\text{Htrz})_2(\text{trz})](\text{BF}_4)$  and  $[\text{Fe}(\text{Htrz})_3](\text{BF}_4)_2 \cdot \text{H}_2\text{O}$  (Htrz = 1,2,4-4H-triazole; trz = 1,2,4-triazolato). *Chem. Mater.* **6**, 1404–1412 (1994).
- Kahn, O., Krober, J. & Jay, C. Spin Transition Molecular Materials for displays and data recording. *Adv. Mater* **4**, 718–728 (1992).
- Kahn, O. & Martinez, C. J. Spin-Transition Polymers: From Molecular Materials Toward Memory Devices. *Science* **279**, 44–48 (1998).
- Schubert, E. M. Utilizing the Evans Method with a Superconducting NMR Spectrometer in the Undergraduate Laboratory. *J. Chem. Educ* **69**, 62 (1992).
- Sur, S. K. Measurement of magnetic susceptibility and magnetic moment of paramagnetic molecules in solution by high-field fourier transform NMR spectroscopy. *J. Magn. Reson* **82**, 169–173 (1989).
- Bain, G. A. & Berry, J. F. Diamagnetic Corrections and Pascal's Constants. *J. Chem. Educ* **85**, 532–536 (2008).
- Zhou, X. *et al.* Dopamine-containing gadolinium complex as magnetic resonance imaging contrast agent. *J. Rare Earths* **30**, 884–889 (2012).

## Acknowledgements

This work was supported by KAKENHI Grant—in—Aid for Scientific Research (A) JP17H01200.

## Author Contributions

S.H. conceived and designed the project; A.T., K.S.M., H.K., H.T., S.Y. and M.T. performed the experiments and analysed the data; A.T., K.S.M., H.K., H.T., S.Y., M.T., L.F.L. and S.H. co-wrote the paper.

## Additional Information

**Supplementary information** accompanies this paper at <https://doi.org/10.1038/s41598-018-33362-6>.

**Competing Interests:** The authors declare no competing interests.

**Publisher's note:** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



**Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018