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OPEN Photosynthesis of subtropical forest species from different successional status in relation to foliar nutrients and phosphorus fractions

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The ecophysiological linkages of leaf nutrients to photosynthesis in subtropical forests along succession remain elusive. We measured photosynthetic parameters (A_{max}, V_{cmax}, J_{max}, PPUE), leaf phosphorus (P) and nitrogen (N), foliar P fractions and LMA from 24 species (pioneer, generalist, and climax). Amax was significantly related to N and P for the pooled data, while significant relationship between A_{max} and P was only found in climax species. The mixed-effect model including variables (N, P, and SLA or LMA) for predicting V_{cmax} and J_{max} best fitted but varied remarkably across succession. Climax species had higher N: P ratios, indicating an increasing P limitation at later succession stage; photosynthesis, however, did not show stronger P than N limitations across all species. Nevertheless, climax species appeared to increase nucleic acid P allocation and residual P utilization for growth, thereby reducing the overall demand for P. Our results indicate that the scaling of photosynthesis with other functional traits could not be uniform across succession, growth variables (e.g. photosynthesis) and species trait identity (e.g. successional strategy) should be considered in combination with N: P ratio when we investigate P limitation in subtropical forests, and variations in P allocation state further influencing photosynthetic rates and P-use efficiency.

Nutrient limitation to primary productivity is widespread in most terrestrial ecosystems globally, and low levels of nitrogen (N) and phosphorus (P) commonly limit or co-limit plant growth and rates of photosynthesis^{1,2}. P limitation generally occurs in lowland tropical forests, is particularly strong for sites in Panama and the Amazon basin³, while N limitation often occurs in temperate and boreal regions⁴⁻⁶. Leaf N:P ratio in terrestrial plants generally serves as a simple and useful indicator of nutrient limitation to primary productivity⁷⁻⁹. In forest ecosystems, leaf N:P ratios >16¹⁰ or 20⁸ usually indicate P limitation. However, P limitation has received comparatively less attention than N limitation, few studies have compared the limitations of leaf P and N concentration on photosynthesis of tropical forest species¹¹.

As we all know, the maximum net assimilation rate (A_{max}) is strongly affected by various leaf traits, for example, leaf thickness^{12,13}, leaf mass per area (LMA)¹⁴⁻¹⁶, and leaf nutrient concentration^{15,17,18}. A number of studies have explored how low leaf nutrient concentrations affect leaf photosynthetic capacity in the tropics, particularly for P6.19-22. Phosphorus limitation might be manifested in limiting ribulose-1,5-bisphosphate (RuBP) regeneration as the underlying control over A_{max} in leaves^{23,24}. Previous studies in nutrient-poor ecosystems have shown N limitation can induce larger proportion of leaf N allocated into cell walls, which increases LMA; an increase LMA reduces the photosynthetic N-use efficiency (PNUE) by decreasing $A_{max}^{25,26}$. However, the ecophysiological linkages between P and A_{max} and the underling mechanisms of P limitation are still poorly known²⁷.

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	N _a	P _a		LMA	A _{max,a}	V _{cmax,a}	J _{max,a}		PPUE
Group	(g m ⁻²)	$(g m^{-2})$	N:P ratio	$(g m^{-2})$	$(\mu mol m^{-2} s^{-1})$	$(\mu mol m^{-2} s^{-1})$	$(\mu mol m^{-2} s^{-1})$	$J_{\max,a}/V_{\max,a}$	$(\mu mol mol^{-1} s^{-1})$
Pioneer	$1.65\pm0.08~b$	$0.10 \pm 0.01 \ c$	18.28 ± 1.40 a	86.06 ± 8.73 b	12.43 ± 0.93 c	69.08 ± 4.86 b	$88.72 \pm 7.31 \text{ c}$	1.29 ± 0.05 a	$5222.20 \pm 520.82 \ b$
Generalist	$1.58\pm0.05~ab$	$0.08 \pm 0.01 \ b$	19.99 ± 0.69 a	$92.54 \pm 2.37 \ b$	$9.92 \pm 0.57 \text{ b}$	56.98 ± 3.11 a	$73.42 \pm 3.91 b$	1.31 ± 0.04 a	$5052.24 \pm 367.63 \ b$
Climax	1.46 ± 0.05 a	$0.06 \pm 0.01 \ a$	23.92 ± 0.88 b	70.50 ± 2.48 a	6.54 ± 0.28 a	49.51 ± 2.00 a	60.84 ± 2.40 a	1.25 ± 0.03 a	$3899.78 \pm 166.75 \ a$

Table 1. Area-based leaf traits for the subtropical forest species in three successional groups. Values shown are group averages (\pm SE). Means in a column followed by different letters are significantly different (P < 0.05). N_a, leaf nitrogen concentration; P_a, leaf phosphorus concentration; leaf N:P ratio, leaf nitrogen to phosphorus ratio; LMA, leaf mass per area; $A_{max,a}$, maximum photosynthesis assimilate rate; $V_{cmax,a}$, maximum carboxylation velocity; $J_{max,a}$, maximum electron transport rate; $J_{max;a}/V_{cmax;a}$, ratio of maximum carboxylation velocity over maximum rate of electron transport; PPUE, photosynthetic P-use efficiency.

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In the photosynthesis model proposed by Farquhar *et al.*²⁸, 'the maximum carboxylation rate' (V_{cmax}) and 'the maximum electron transport rate' (J_{max}) were used to express photosynthetic capacity, which are generally positively related to N, P and specific leaf area in tree leaves^{11,21,29}. Various studies have shown that leaf photosynthetic characteristics correspond with successional status^{30–32}. The early successional (pioneer) species, which are usually fast-growing and light-demanding, make a greater fractional investment in leaf traits that maximize photosynthetic capacity than late successional (climax) species^{30,31,33}. In contrast, leaves of climax species are often have a longer lifespan, higher LMA, chlorophyll to N ratios and lower photosynthetic capacity than leaves of pioneer species. Unlike pioneer species, which occur early in succession, and climax species, which occur late in succession, generalist species can occur throughout succession. What is less clear is how P limitation, which is common in tropical forests, might affect the relationships of photosynthetic capacity to N and LMA during succession in subtropical forests.

Previous studies have shown that plants generally reduce their foliar P concentration in response to low P availability in tropical soils^{35,36}. Kedrowski³⁷ developed a successful fractionalization scheme based on differential solubility or hydrolysis for the extraction and analysis of various P-containing fractions from plant material. Using a trichloroacetic acid (TCA) extraction method, Close and Beadle³⁸ reported differences in the concentrations of insoluble P and inorganic P among plant species. More recently, Hidaka and Kitayama³⁹ divided foliar P into four fractions: structural P (lipid P, phospholipids of membranes), metabolic P (including Pi and easily soluble P-containing metabolites), nucleic acid P (RNA and DNA), and residual P (phosphoproteins and unidentified residue). It has been shown that tree species with high photosynthetic P-use efficiency (PPUE) on P-poor sites reduce the demand for foliar P by reducing concentrations of both metabolic P and nucleic acid P^{39,40}. However, how plants allocate P among foliar P fractions and how plants develop adaptive strategies to efficiently use P in subtropical region remain unclear.

Plants are probably P-limited in most soils of China, because of the low soil available P content⁴¹. Insufficient P has become the limiting factor of ecosystem primary productivity and other ecosystem processes in subtropical forests of China⁴²⁻⁴⁴; however, these studies mainly focused on plant N: P ratios, the real demand of leaf P for growth (e.g. photosynthesis) and its functional partitioning have not been concerned. To our knowledge, few data are available describing the photosynthetic parameters of the subtropical species. Here, we examined how leaf N, P, LMA (or SLA), and foliar P fractions affect photosynthetic performance along a subtropical forest succession. We tested the following hypotheses: (1) Average leaf trait values decrease with succession, pioneer species have higher mean P, N, and photosynthetic parameters (A_{max} , V_{cmax} , J_{max} and PPUE) than generalist and climax species; (2) Leaf P has a stronger influence over photosynthetic capacity (A_{max} , V_{cmax} , and J_{max}) than N, in particular at later succession stage; and (3) Foliar P fractions change substantially with succession in that climax species, unlike pioneer species, optimize the allocation of P among foliar P fractions in order to maintain their growth and to reduce the overall demand for P.

Results

Comparison of leaf traits among successional status. Leaf P concentration of the pioneer species was significantly higher than that of the generalist and climax species, on both an area and a mass basis (both P < 0.001; Table 1 and Supplementary Table S1). Pioneer species also exhibited the highest N_a values, although no significant differences in N_a were observed among successional groups (P = 0.08, Table 1). The pioneer and climax species leaves exhibited similar values for N_m; however, the climax species leaves had the highest N:P ratio (P < 0.001, Table 1). Given that a leaf N:P ratio >20 generally indicates P limitation as opposed to N limitation⁸, P limitation was evident in 21% of pioneer species, 38% of generalist species, and 66% of climax species (Fig. 1). On the basis of both area and mass, all photosynthetic parameters (A_{max} , V_{cmax} , J_{max} and PPUE) significantly differed among species with succession (P < 0.01; Table 1), i.e., these values were greater for pioneer species than for generalist and climax species. J_{max}/V_{cmax} ratio did not significantly differ among successional groups (P = 0.525, Table 1).

Bivariate relationships between photosynthesis and leaf traits. For all data pooled, leaf N_a and P_a were highly and positively correlated ($r^2 = 0.21$; P < 0.001, Fig. 1). Across all 24 species, N_a exhibited a weak, positive correlation with LMA (Supplementary Fig. S1a, $r^2 = 0.13$; P < 0.01). SMA tests for common slopes revealed a significant difference among the different successional status (Supplementary Table S2); the y-axis intercept



Figure 1. The relationship between area-based leaf P concentration (P_a) and leaf N concentration (N_a). Data points represent individual leaf values (19, 37, and 47 individuals from pioneer, generalist, and climax species, respectively). Dashed lines represent the N:P ratios of 10 and 20. Points above the N:P = 20 indicate P limitation; points below the N:P = 10 line indicate N limitation; and points between the lines indicate co-limitation.



Symbols: pioneer species (solid); generalist species (grey); climax species (open).

Figure 2. The log-log relationships between photosynthesis capacity (the maximum photosynthesis assimilate rate, A_{max}) and leaf traits including nitrogen concentration (N), phosphorus concentration (P), and leaf mass per area (LMA). Note that A_{max} values are based on measurements of mass in the upper panels and on area in the lower panels. Data points represent individual leaf values (19, 37, and 47 individuals from pioneer, generalist, and climax species, respectively). Standardized major axis (SMA) regressions are presented in Table S3. Symbols are the same as in Fig. 1.

of the relationship was higher for climax species, indicating that the climax species might have a higher N_a for a given LMA than pioneer species. Compared to N_a, P_a showed a stronger relation with LMA (Supplementary Fig. S1b, $r^2 = 0.34$; P < 0.001). Additionally, P_a for a given LMA was lower for climax species than pioneer species (Supplementary Table S2).

Bivariate relationships of A_{max} on N and P (either area-based or mass-based) were highly significant for the pooled data (Fig. 2, P < 0.01). When the area-based data were pooled, variations in $A_{max,a}$ were slightly explained by variations in P_a (P < 0.001; $r^2 = 0.19$) and N_a (P = 0.001; $r^2 = 0.10$). However, Supplementary Table S3 shows



Figure 3. The relationship between area-based maximum carboxylation velocity ($V_{cmax,a}$) and area-based maximum electron transport rate ($J_{max,a}$). Data points represent individual leaf values (19, 37, and 47 individuals from pioneer, generalist, and climax species, respectively). Standardized major axis (SMA) regressions are given in Table S4. Symbols are the same as in Fig. 1.

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that when the data were grouped by successional status, area-based relationships were significant between $A_{\max,a}$ and N_a for the pioneer species (P=0.012; $r^2=0.32$) and between $A_{\max,a}$ and P_a for the climax species (P=0.001; $r^2=0.31$). For all data pooled, LMA was weakly positively related to $A_{\max,a}$ (P<0.05; $r^2=0.04$), but this relationship was not significant for any of the three successional status (Supplementary Table S3). When the mass-based data were pooled, $A_{\max,m}$ was significantly and positively correlated with P_m ($r^2=0.26$), N_m ($r^2=0.21$), and negatively with LMA ($r^2=0.15$) (all P<0.001). Still, $A_{\max,m}$ was significantly correlated with N_m and LMA for all successional groups, while $A_{\max,m}$ and P_m were poorly related for pioneer or generalist species (Supplementary Table S3).

The regression of $J_{\max,a}$ on $V_{\max,a}$ using data from all species suggested a very tight co-ordination between the two parameters (P < 0.001, $r^2 = 0.74$, Fig. 3). Variations in $J_{\max,a}$ were strongly correlated with $V_{\max,a}$ all for the three successional groups (Supplementary Table S4). Across all species, $V_{\max,a}$ and $J_{\max,a}$ were correlated with leaf traits, and the bivariate relationships were stronger with P_a (P < 0.001; $r^2 = 0.17$ for $V_{\max,a}$; $r^2 = 0.23$ for J_{\max}) than with N_a (P < 0.01; $r^2 = 0.13$ for $V_{\max,x}$; $r^2 = 0.18$ for $J_{\max,a}$) (Fig. 4). No significant relationship was found between $V_{\max,a}$ (or $J_{\max,a}$) and N:P ratio (Fig. 4). Both $V_{\max,a}$ and $J_{\max,a}$ were marginally correlated with LMA (P < 0.01; $r^2 = 0.07$ and $r^2 = 0.06$, respectively) (Fig. 3). Within the pioneer group, significant relationship of $V_{\max,a}$ was observed only with LMA (P = 0.021, $r^2 = 0.27$), no significant relations with N_a , P_a , or N:P ratios (Supplementary Table S4). Within the generalist and climax group, $V_{\max,a}$ was positively related to N_a (P = 0.002, $r^2 = 0.24$ for generalist species; P = 0.050, $r^2 = 0.08$ for climax species) but not with LMA, P_a , or N:P ratio (Supplementary Table S5). Similar patterns were observed for $J_{\max,a}$ (Supplementary Table S4).

Modelling variations in photosynthetic capacity from leaf traits. We used linear mixed-effect to model variations in photosynthetic parameters (V_{cmax} and J_{max}) (Table 2 and Supplementary Table S5). The regression coefficients (r^2) ranged from 0.30 to 0.53 and were substantially higher when the variables were expressed on a mass basis than on an area basis. The model's random variance indicated that species accounted for less than 3% of the unexplained variance for models.

Based on leaf mass, a combination of leaf N, P and SLA accounted for 47% of the variation in $V_{cmax,m}$. Similar to $V_{cmax,m}$, variations in $J_{max,m}$ were largely explained by a combination of N, P and SLA; the best model explained 48% of the variation in $J_{max,m}$. When these analyses were repeated using area-based data, relationships were similar to those described for mass-based measurements. For the pioneer species, the species with the highest N and P, SLA was the important fixed effect for explaining V_{cmax} and J_{max} . For the generalist and climax species, SLA (or LMA) was important and the importance of N and P varied depending on whether data were expressed on a mass or an area basis. Importantly, for both the mass- and area-based mixed-effect models, both N and P are important predictors of V_{cmax} and J_{max} , when data from all species were combined.

Variation in foliar P fractions. The overall average concentration of each foliar P fraction was significantly higher in pioneer species than in the other two groups (all P < 0.05) (Table 3), although the mean concentration of both structural P and nucleic acid P were not different between generalist species and climax species. For each group of species, concentrations tended to be higher for nucleic acid P than for the other fractions (all P < 0.05) (Table 3). The mean percentage of P represented by structural P did not differ among the three successional groups (P = 0.766). The metabolic P percentage was significantly lower in generalist species than in the other groups (P < 0.001). Conversely, the residual P percentage was significantly lower in climax species than in the other groups (P < 0.001).

Across all 24 species, the concentrations of each foliar P fraction were positively related to P_m (Table 4). For regressions of concentrations of P fractions on P_m , SMA slopes did not significantly differed (P = 0.107), while the intercept was significantly higher for the concentration of nucleic acid P than for the concentrations of other



Figure 4. The log-log relationships between area-based photosynthesis capacity and leaf traits including nitrogen concentration (N_a) (**a**,**e**), phosphorus concentration (P_a) (**b**,**f**), leaf N:P (**c**,**g**), and leaf mass per area (LMA) (**d**,**h**). (**a**–**d**) Show the maximum carboxylation velocity ($V_{cmax,a}$), while e-h show the maximum electron transport rate ($J_{max,a}$). Data points represent individual leaf values (19, 37, and 47 individuals from pioneer, generalist, and climax species, respectively). Standardized major axis (SMA) regressions are given in Table S4. Symbols are the same as in Fig. 1.

	V _{cmax}	J _{max}								
	Fixed effect			Random variance (%)		Fixed effect			Random variance (%)	
	Best model	r ²	Р	Species	Residual	Best model	r ²	Р	Species	Residual
(a) Mass-based model										
All	$P_m + N_m * SLA$	0.47	< 0.01	0.19	99.81	$P_m + N_m * SLA$	0.48	< 0.01	0.00	100.00
Pioneer	SLA	0.39	< 0.01	0.00	100.00	N _m * SLA	0.32	< 0.01	0.00	100.00
Generalist	N _m * SLA	0.41	< 0.01	0.00	100.00	N _m * SLA	0.50	< 0.01	0.00	100.00
Climax	$P_m + N_m * SLA$	0.53	< 0.01	0.00	100.00	N _m * SLA	0.49	< 0.01	0.00	100.00
(b) Area-based model										
All	$N_a + P_a * LMA$	0.23	< 0.01	0.47	99.53	$N_a + N_a * P_a$	0.36	< 0.01	0.20	99.80
Pioneer	LMA	0.19	< 0.05	0.00	100.00	LMA	0.27	< 0.05	0.00	100.00
Generalist	P _a * LMA	0.13	< 0.05	0.00	100.00	LMA + N _a * LMA	0.31	< 0.01	0.00	100.00
Climax	$N_a + N_a * LMA$	0.16	< 0.01	2.56	97.44	N _a	0.17	< 0.01	0.00	100.00

Table 2. The linear mixed-effect models, with mass-based (a) and area-based (b) leaf photosynthetic capacity $(V_{cmax} \text{ and } J_{max})$ as the response variables, each showing fixed and random effects. For the best models, explanatory variables are: leaf nitrogen (N) and phosphorus (P) concentrations; leaf area per mass (SLA) and leaf mass per area (LMA). Species was used as a random component of the model. The best model for V_{cmax} and J_{max} for all species (All), and for the pioneer species (Pioneer), the generalist species (Generalist) and the climax species (Climax) are shown. The coefficient of variation (r²) and significance (P) for the linear regression of the modelled vs measured data are also shown and the contribution of random effect to the variance with the dataset.

fractions (P=0.001, Table 4). But the values for both nucleic acid P and metabolic P were followed by a lowercase c, indicating that the slopes were not different. The concentration of each foliar P fraction had significant positive relationship with N_m, and negative relationship with N:P ratio (Table 4). Moreover, except residual P, LMA significantly increased with decreasing P fractions concentration, and the intercept of nucleic acid P for LMA was larger than those of the other two fractions (P<0.001, Table 4). The SMA slopes of regressions of concentration of each foliar P fraction on N_m, N:P ratio, and LMA did not significantly differ among foliar fraction types (all P > 0.1).

Relationships between foliar P fractions and photosynthetic rates and P-use efficiency. Among all 24 species, $A_{\text{max,m}}$ was positively correlated with the concentration of foliar P fractions (Fig. 5). The coefficients

P fraction	Pioneer		Generalist		Climax		P value			
Structural P										
Concentration	0.248 ± 0.007	Ba	0.204 ± 0.009	BCb	0.206 ± 0.009	СЪ	P=0.009			
Percentage	ercentage 21.88±1.27		22.63 ± 0.45	a	22.49 ± 0.51	a	P=0.766			
Metabolic P										
Concentration	0.358 ± 0.022	A a	0.226 ± 0.010	Вc	0.271 ± 0.009	Вb	P<0.001			
Percentage	29.98 ± 0.94	a	25.12 ± 0.44	b	30.15 ± 0.77	a	P<0.001			
Nucleic acid P										
Concentration	0.349 ± 0.017	A a	0.288 ± 0.009	A b	0.309 ± 0.013	A b	P=0.021			
Percentage	29.47 ± 0.66	b	32.66 ± 0.57	a	33.26 ± 0.48	a	P<0.001			
Residual P										
Concentration 0.227 ± 0.020		Ba	0.183 ± 0.013	СЪ	0.129 ± 0.007	Dc	P<0.001			
Percentage	18.76 ± 0.94	a	19.58 ± 0.68	a	14.07 ± 0.57	b	P<0.001			

Table 3. Concentrations (mg g⁻¹) and percentages (%) of foliar P fractions of subtropical forest species in different successional groups. Values are means \pm SE. Within each P fraction, concentrations followed by different lowercase letters are significantly different at P < 0.05 among successional groups. Within each P fraction, percentages followed by different by different lowercase letters are significantly different at P < 0.05 among successional groups. Within each successional group, concentrations followed by different uppercase letters are significantly different at P < 0.05 among successional groups. Within each successional group, concentrations followed by different uppercase letters are significantly different at P < 0.05 among P fractions.

Leaf trait Structural P Metabolic P Nucleic acid P Residual P P-value Pm 0.675*** r^2 0.630*** 0.820*** 0.568*** Slope 1.051 (0.810, 1.370) 1.133 (0.828, 1.472) 1.191 (0.980, 1.437) 1.771 (1.346, 2.326) 0.107 Intercept -0.656 b -0.546 c -0.485 c -0.770 a 0.001 Nm 0.381*** 0.285*** 12 0 124*** 0.036* 1.764 (1.090, 2.847) Slope 1.044 (0.636, 1.727) 0.858 (0.640, 1.185) 0.937 (0.656, 1.427) 0.311 -2.017 b -3.070 a < 0.001 Intercept -1.669 c -1.722 c N:P ratio 0.286*** 12 0 145*** 0.149*** 0.034 Slope -1.065(-0.681, -1.813)-1.263(-0.831, -1.843)-1.067(-0.703, -1.746)-1.950(-1.287, -3.038)0.430 0.713 a 0.872 b 1.741 d < 0.001 Intercept 1.081 c LMA 0.319*** 0.325*** 0.400*** r 0.016 Slope -1.116(-0.791, -1.671)-1.160(-0.766, -1.789)-1.244(-0.908, -1.741)-1.987(-1.246, -3.323)0.370 1.438 a 1.631 ab 1.838 b 2.962 c < 0.001 Intercept

Table 4. Standardized major axis (SMA) relationships between log-log transformed foliar phosphorus (P) fractions and total leaf P concentration (P_m), leaf mass per area (LMA), leaf nitrogen concentration (N_m), and N:P ratio across the three successional groups. Analysis undertaken using species replicates. For each leaf trait, intercepts followed by different letters indicate significant differences among P fractions. *, **, and *** indicate significance at P < 0.05, < 0.01, and < 0.001, respectively.

of determination between $A_{\text{max,m}}$ and P fraction concentrations (r² = 0.26 for structural P and 0.27 for nucleic acid P) were similar to that between $A_{\text{max,m}}$ and P_{m} (r² = 0.26). The correlation coefficients between $A_{\text{max,m}}$ and metabolic P (r² = 0.11) and residual P (r² = 0.10) were low and positive. PPUE was positively correlated with the percentage of structural P (r² = 0.07, P < 0.01; Fig. 6a) but was negatively correlated with the percentage of metabolic P (r² = 0.03, P < 0.05; Fig. 6b). PPUE was not correlated with the percentage of nucleic acid P or residual P (Fig. 6c,d).

Discussion

The first hypothesis was supported, and overall average area-based $N_{av} P_{av}$, and photosynthetic parameters (A_{max} , V_{cmax} , J_{max} , PPUE) were higher in pioneer species than in generalist and climax species (Table 1), indicating the photosynthesis capacity with accompanying P use efficiency generally declined with succession proceeding. These results were consistent with what is known about photosynthetic physiology, such as, photosynthetic capacity decreases along the successional axis^{30,31,33,34}. Rijkers *et al.*⁴⁵ found that early-successional species had higher SLA than that of late-successional species in a lowland tropical area of French Guiana. However, no clearly consistent pattern of LMA variations along with succession was found in the present study, i.e. mean LMA values were





higher for the generalist species than for the pioneer and climax species (Table 1). This result might be due to the large variations in LMA among species within the same successional group.

Our results showed that leaf N:P ratios ranged from 10.5 to 37.6 across the 24 species, and 48% of the species measured were limited by P and the remainder were co-limited by N and P (Fig. 1). The relatively high values for N:P ratios reported here agree with our prior knowledge that these systems are P limited^{42,44}. Moreover, the climax species had the highest N:P ratio (Table 1), showing that P limitation is more pronounced in climax species. However, some studies reported that leaf N:P ratios are not definitive indicators of N or P limitation^{46,47}. In a study in Australia, for example, median N:P ratios were relatively high (>20) at all sites, but evidence did not indicate that photosynthesis was limited by P for either forest or savanna trees⁴⁸. These inconsistencies between N:P ratios and P limitation may result from the great variability of N:P ratios throughout biomes and among and within species, and although analogous biogeochemical constraints may exist for individual plant organs, differences can vary by up to an order of magnitude^{8,49-51}. For this reason, we also explore how leaf N and P concentration in these subtropical systems may affect photosynthetic biochemistry.

Several studies have reported that positive relationships between leaf N (or P) and photosynthetic rates^{18,19,30,52}. Indeed, A_{max} was positively related to N and P, and these relationships were evident whether measurements were mass-based or area-based (Fig. 2). The slopes of the relationships between A_{max} and key leaf traits (N, P, and LMA) differed depending on the successional status, more importantly, significant relationship between A_{max} and P was only found in the climax species (Supplementary Table S3). The climax species had both the lowest P and the lowest A_{max} values. This result suggests that a reduction in the total P is a main factor limiting photosynthesis of climax species, in agreement with studies related low A_{max} in leaves with P limitation²⁴. The relationship between A_{max} and N may be constrained by low P in P-limited ecosystems^{18,27}, out results highlight that the slope of A_{max} on N was shallower for the climax species (with low P values) than for the pioneer species (with high P values).





Further, A_{max} was negatively correlated with LMA when A_{max} was based on mass measurements (Fig. 2f). This is in line with an increase in LMA leading to an increase in resistance to CO₂ diffusion within the leaf and eventually to a decrease in $A_{\text{max}}^{53,54}$.

In this study, J_{max} and V_{cmax} were closely related, and the linear regression had a slope of 1.15 ± 0.07 (Fig. 3). A larger common slope of J_{max} and V_{cmax} was 1.27 when the SMA test was used. The slope in our study was less than those from global analyses (1.64 in Wullschleger⁵⁵, 1.67 in Medlyn *et al.*⁵⁶). Furthermore, the slope of the J_{max} - V_{cmax} relationship was steeper for pioneer species than for generalist and climax species in the current study (Supplementary Table S4). This discrepancy may be explained by the relative high light conditions experienced by the pioneer species.

The photosynthetic capacity (V_{cmax} and J_{max}) of trees is generally positively related to leaf P concentration, especially under conditions of P limitation^{6,20,21,48,57}. In this study, we demonstrated that across all 24 species, $V_{cmax,a}$ and $J_{max,a}$ were significantly and positively related to N_a and P_a and weakly and negatively related to the N:P ratio (Fig. 4). When data from all 24 species were modelled, the line mixed-effect modeling analysis performed here demonstrated that, both N and P were important as variables for predicting V_{cmax} and J_{max} , whether measurements were area-based or mass-based (Table 2). Additionally, leaf P and the interaction between N and SLA have proven to be significant explanatory variables in mixed-effect models of variations in photosynthetic capacity, explaining about 50% of the observed variations in $V_{cmax,m}$ and $J_{max,m}$ (Table 2a). The best combination of variables (N, P, and SLA or LMA) for modelling V_{cmax} and J_{max} changed with succession. We also found that species played a very weak role in modelling photosynthetic capacity, and therefore that limitations to photosynthetic capacity were likely to be results of environmental factors. In addition, these differences in the best mixed-effect model were in line with the observed variation in the relationships of photosynthetic capacity (V_{cmax} and J_{max}) with leaf traits (N, P, and LMA) from SMA regression analysis (Supplementary Table S4). Collectively, these results show that N, P, and LMA or SLA, and especially N and P, help explain the variations in A_{max} , V_{cmax} and J_{max} in this study and that the degree of influence for different trait of three successional groups are different.

Taken together, the second hypothesis that P has a stronger influence over photosynthetic capacity than N, was only supported in the climax species, but was not supported in the other two species groups; leaf N and P are the best predictors of photosynthetic capacity. These results highlight the importance for the consideration of not solely N:P ratio but also growth variables (e.g. photosynthesis) and species identity (e.g. successional strategy) into nutrient limitation determination in the highly species-rich and diversified subtropical forest ecosystems.

In our study, the concentrations and the percentages of total P represented by functional P fractions were higher for nucleic acid P than for other foliar P fractions except for metabolic P in the pioneer species (Table 3). These results are in line with past studies showing that nucleic acid P is the largest pool throughout the growing season^{39,58}. A previous study suggested that a reduction in foliar P concentration is strongly correlated with a reduction in the concentrations of both metabolic P and nucleic acid P with decreasing soil P availability³⁹. Similarly, our results indicate that reductions in the concentrations of both nucleic acid P and metabolic P explain most of the reduction in leaf P concentration among the three successional groups (Table 4). Another previous study showed that the concentration of structural P is greater in fast-growing plants with small LMA than in slow-growing plants with large LMA⁵⁹, which was partial in line with the results of the present study. The pioneer species and climax species exhibited similar values for LMA, however, pioneer species had higher structural P concentration (Table 4). Moreover, our results show that the concentration of each foliar P fraction decreases along the successional axis, and that the climax species have a greater nucleic acid P percentage and a lower residual P percentage than pioneer and generalist species (Table 4). Overall, these findings suggest that there are trade-offs in the P allocation among nucleic acid P and metabolic P with residue P.

 $A_{\max,m}$ positively correlated with P_m and the concentrations of each foliar P fraction (Figs 2b and 5). Mean $A_{\max,m}$ values and mean concentrations of P_m and of each foliar P fraction were lower in climax species than in pioneer species (Table 3 and Supplementary Table S1). As a result, climax species had lower mean PPUE values than pioneer species in this study, which agrees with previous studies of tropical rainforest trees³¹. PPUE was positively correlated with $A_{\max,m}$ ($r^2 = 0.68$, P < 0.001) and weakly and negatively correlated with P_m and LMA (data not shown). Hidaka and Kitayama⁴⁰ found that PPUE was negatively correlated with the proportion of structural P but tended to be positively correlated with the proportion of metabolic P. In contrast, we found that PPUE was weakly and positively correlated with the structural P percentage and negatively correlated with the metabolic P percentage (Fig. 6). Hence, the lower PPUE values in climax species can be explained by reduced photosynthetic activity. Finally, the third hypothesis was supported by our results - the climax species may increase their nucleic acid P percentage to maintain their growth and decrease residual P percentage to reduce their demand for P (i.e., reducing total leaf P concentration) as adaptation strategies to soil available P impoverishment at later succession stage.

Previous studies were restricted to Alaskan tree species on P-rich soils^{58,60} or to Mount Kinabalu plant species on P-poor sites^{39,40}. Our studies on foliar P fractions and the relationships between foliar P fractions and leaf traits in species representing successional strategies can improve our understanding of what the magnitude species are adapted to low soil P availability and how forests can maintain their high productivity in the highly weathered and acidified soils in the subtropics. We conclude that predicting future dynamics of forest ecosystems in response to global change requires a better understanding of the variations of nutrient limitation, not solely on the base of plant N: P ratios but also by incorporating growth variables (e.g. photosynthesis), and in particular the P adaptation strategies created by ecosystem succession of the subtropical forests.

Materials and Methods

Study site and plant materials. This study took place at the Dinghushan Biosphere Reserve (DBR) in central Guangdong Province, southern China $(21^{\circ}09'21''-21^{\circ}11'30'' \text{ N}, 112^{\circ}30'39''-112^{\circ}33'41''\text{E})$. The region is characterized by a typical subtropical monsoon climate, with mean annual temperature is 21.4 °C, and annual average precipitation is 1927 mm with 80% occurring during the wet season (April to September). The soils are classified as Ultisol and Udult according to the USDA soil classification. A total of 24 species across successional stages were sampled in this study (Supplementary Table S6). Three to five individuals were selected for each species. The studied species are common and typical in each stage of the succession according to the long-term forest community studies of the reserve^{61,62}. We used long-reach pruner to collect middle canopy branches (supporting leaves considered to be typically exposed to full sunlight for much of the day) from tall plant species. The detached branches had been recut under water immediately after harvesting to preserve xylem water continuity, prior to subsequent leaf gas exchange measurements⁶³⁻⁶⁵.

Leaf gas exchange measurements. Measurements of leaf gas exchange were made on the most recently fully expanded leaves during August to September 2015, between 9:00 and 12:00 h using the LI-6400 portable photosynthesis system (Li-Cor, Lincoln, Nebraska, USA). Maximum photosynthetic rate per unit area ($A_{max,a}$) was determined for each species by measuring a light response curve (A-PPFD curves) at ambient 400 µmol mol⁻¹ CO₂, leaf temperature of 28–30 °C, and relative humidity of 40–60%, with the photosynthetic photon flux density (PPFD) order was 1500, 1200, 1000, 800, 500, 300, 200, 120, 100, 80, 50, 20,0 µmol m⁻² s⁻¹. For the photosynthesis measurement of masson pine needles, a bunch of needles were measured side by side, and then the average was calculated. Photosynthetic P-use efficiency (PPUE) was calculated as A_{max} divided by total P concentration (µmol CO₂ mol P⁻¹ s⁻¹). $V_{cmax,a}$ and $J_{max,a}$ on an area basis were estimated from relationships between photosynthetic rate (A) and sub-stomatal CO₂ mole fraction (C_i)²⁸ at fixed PPFD (1200 µmol photons m⁻² s⁻¹) following the CO₂ order 400, 300, 200, 100, 500, 700, 900, 1000, 1200, 1500 µmol mol⁻¹.

Leaf structure and nutrients. After photosynthetic measures, leaves with the same position as used for gas-exchange measurements were collected. Some of the leaves were immediately snap-frozen in liquid N and transferred on dry ice to laboratory. The samples were freeze-dried and stored at $-80 \,^{\circ}$ C until they were used for determination of foliar P fractions. The remaining leaves were oven-dried at 70 $^{\circ}$ C and then ground and homogenized for subsequent analyses. Total N concentration in dried leaves was determined using the Kjeldahl method. An additional sample of ten leaves was scanned to determine leaf area (LA) by LI-3000A portable system, dried at 70 $^{\circ}$ C for 72 h to a constant weight and measured for oven-dried mass (DM). Since the needles of masson pine trees don't have flat leaf area, we record needle length and cross-section width and needle leaf area was estimated as: LA = $\pi^*L^*D/2$ (where LA is leaf area; L is needle length and D is needle width at the hale needle length). LMA, calculated as DM·LA⁻¹ (g m⁻²), was used to calculate area-based nutrient concentrations (N_a, P_a; g m⁻²) from mass-based concentrations (N_m, P_m; mg g⁻¹), and to shift area-based photosynthetic parameters ($A_{max,a}$, $V_{cmax,a}$, and $J_{max,a}$; µmol CO₂ m⁻² s⁻¹) to ($A_{max,m}$, $V_{cmax,m}$, and $J_{max,m}$; µmol CO₂ g⁻¹ s⁻¹). To compare LMA with mass-based leaf nutrient concentrations, we converted LMA into its reciprocal, leaf area per mass (SLA, cm² g⁻¹).

Foliar P fractions. Foliar P fractions were extracted following methods outlined in Hidaka and Kitayama³⁷. Each freeze-dried and ground sample (0.5 g DW) was homogenized and extracted twice with a total of 15 mL of 12:6:1 CMF (chloroform, methanol, and formic acid; v/v/v) and twice with a total of 19 mL of 1:2:0.8 CMW (chloroform, methanol, and water; v/v/v). Extracts were combined in a 50-mL centrifuge tube, and 9.5 mL of water was added to each tube. Thereafter, the extract was separated into an aqueous upper layer and a lipid-rich organic bottom layer. A subsample of the lipid layer was digested to give structural P. The residue was re-extracted for 1 h with 5 mL of 85% (w/v) methanol. The supernatant was added to the tube containing the aqueous layer. A 20 ml volume of the aqueous layer was added to another 50-ml graduated tube. The remaining aqueous layer was decanted into the tube containing the residue. The volume in the tube was increased to 20 ml with deionized water. After the preparation was cooled to 4°C, 1 ml of cold 100% (w/v) trichloroacetic acid (TCA) was added to make a 5% (w/v) cold TCA 4°C. A subsample of the supernatant was taken for the analysis of metabolic P. The residue was then re-extracted twice in a total of 35 mL of 2.5% (w/v) TCA at 95 °C for 1 h. A subsample of the supernatant was taken for the analysis of nucleic acid P. The residue was digested to obtain residue P.

Each subsample was evaporated to dryness at 70 °C (organic solutions) or 100 °C (aqueous solutions) and digested for P determination as described above. The concentration of P in the digest was determined at 700 nm in a UV-Vis spectrophotometer (UV1800, Shimadzu, Japan) after a standard molybdate reaction. Total P concentration was the sum of foliar P fractions.

Data analysis. To compare overall average leaf trait and foliar P fraction (concentration and percentage) values of the three successional groups using ANOVAs, and multiple comparisons were conducted using Duncan's multiple range test. Bivariate regression was used to explore relationships between photosynthetic parameters and other leaf traits (N, P and LMA) among the three successional groups. Standardized major axis (SMA) regression was used to examine for variations in the slope and the y-axis intercept of bivariate leaf trait relationships, using SMATR version 2.0. software. When we tested the relationships among leaf traits, values were log_{10} -transformed when it was necessary to normalize distributions. The significance of SMA regression was determined at the 0.05 level. We also used a linear mixed effects model combining fixed and random effects to account for variability in V_{cmax} and J_{max} on both area- and mass-bases. The model's fixed effect included N, P and LMA (or SLA), and species as random effects. Model specification and validation were conducted in 'R' (version 3.4.0; R Development Core Team, 2011), using the nlme package. All of our data for the mixed-effect modelling analysis were log transformed. To select the best model, Akaike's information criterion (AIC) was used. Statistical analysis was performed using SPSS 21.0 (IBM SPSS, USA), unless otherwise indicated.

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Author Contributions

D.Z.W. and G.H.Z. conceived the study; G.H.Z., L.L.Z. and D.Z.W. performed the experiment; G.H.Z. drafted the manuscript.

Additional Information

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