

SCIENTIFIC REPORTS



OPEN

Cytokine and Chemokine Profiling in Patients with Hand, Foot and Mouth Disease in Singapore and Malaysia

Fiona Mei Shan Teo¹, Min Nyo², Anng Anng Wong³, Natalie Woon Hui Tan³, Mia Tuang Koh⁴, Yoke Fun Chan⁵, Chia Yin Chong³ & Justin Jang Hann Chu^{1,2}

Hand, foot and mouth disease (HFMD) is a prevalent contagious childhood disease typically associated with fever, oral lesions and limb exanthema. While HFMD is caused by a plethora of serotypes of viruses under the genus *Enterovirus* within the *Picornaviridae* family, Coxsackievirus A16 (CV-A16) and Enterovirus 71 (EV-A71) are considered the main etiological agents. In recent years however, other viruses have also been isolated in considerable numbers from infected individuals in many regions, joining the legion commonly associated with HFMD. The present study investigated the cytokine and chemokine profiles of HFMD patients from Singapore and Malaysia for the first time. Comparative cohort studies of EV-A71-associated HFMD cases revealed that the Malaysia cohort had a distinct profile from the Singapore cohort, and this could be partly attributed by different EV-A71 genotypes. As the isolation of CV-A6, instead of CV-A16, had become prevalent in the Singapore cohort, it was also of particular interest to study the differential cytokine and chemokine profiles. Our data revealed that overlapping as well as unique profiles exist between the two major causative clinical isolates in the Singapore cohort. Having a better understanding of the respective immunological profiles could be useful for more accurate HFMD diagnosis, which is imperative for disease transmission control until multi-valent vaccines and/or broad-spectrum anti-viral drugs become available.

Hand, foot and mouth disease (HFMD) is a widespread transmissible infectious disease caused by a myriad of etiological agents under the genus *Enterovirus* within the *Picornaviridae* family, with Coxsackievirus A16 (CV-A16) and Enterovirus A71 (EV-A71) generally regarded as its major causative agents¹. CV-A16 is regarded as the most dominant serotype that causes asymptomatic or mild HFMD which resolves on its own most of the time. EV-A71, on the other hand, is a neurotropic virus associated with neurological complications and even deaths in infants and children in the Asia Pacific region²⁻⁷. Due to its virulence, EV-A71 causes a raising concern in the otherwise generally mild and self-limiting disease. Over the past two decades, tremendous research has been done to compare these two key players.

In addition to CV-A16 and EV-A71, other serotypes such as CV-A4, CV-A5, CV-A6, CV-A7, CV-A9, CV-A10, CV-A24, Coxsackievirus B2 (CV-B2), CV-B3, CV-B4, CV-B5, EV-G18, EV-D70 and Echovirus 7 (E-7) are also known to cause HFMD albeit in smaller numbers^{3,8}. Some of these serotypes, however, are becoming more prevalent in the recent years, being capable of existing as the main circulating virus of HFMD outbreaks in some regions.

In particular, CV-A6 is an emerging HFMD-causing virus strain capable of causing outbreaks in many regions⁹⁻¹⁸. In the recent years, the two predominant strains causing HFMD outbreaks in Singapore have also shifted from the conventional CV-A16 and EV-A71 to CV-A6 and EV-A71¹⁹⁻²³. Similar to CV-A16, CV-A6

¹Collaborative and Translation Unit for HFMD, Institute of Molecular and Cell Biology, Agency for Science, Technology and Research (A*STAR), Singapore, Singapore. ²Laboratory of Molecular RNA Virology and Antiviral Strategies, Department of Microbiology and Immunology, Yong Loo Lin School of Medicine, National University of Singapore, Singapore, Singapore. ³Infectious Disease Service, Department of Pediatrics, KK Women's and Children's Hospital, Singapore, Singapore. ⁴Department of Pediatrics, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia. ⁵Department of Medical Microbiology, Faculty of Medicine, University of Malaya, Kuala Lumpur, Malaysia. Correspondence and requests for materials should be addressed to J.J.H.C. (email: jhchu@imcb.a-star.edu.sg)

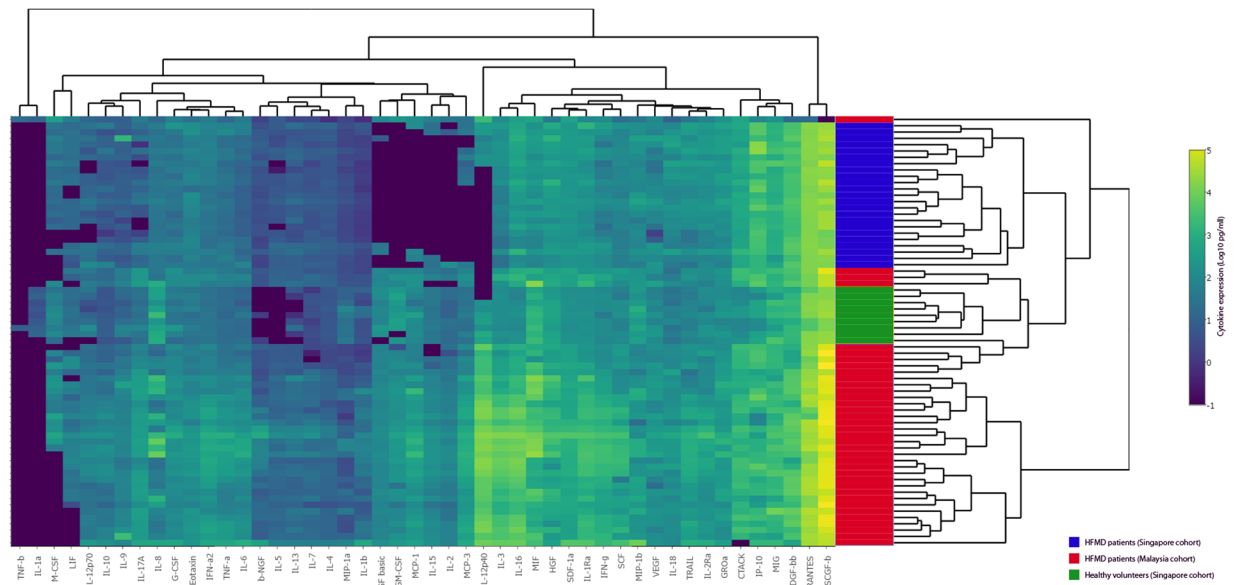


Figure 1. Heat map comparing the sera cytokine and chemokine expression profiles of HFMD patients revealed two distinctive clusters representing HFMD patients from Singapore and Malaysia respectively.

infections are usually self-limiting unlike neurotrophic EV-A71 infections. However, CV-A6 infections are reportedly associated with atypical clinical presentations including onychomadesis^{9–11,14,15}, eczema herpeticum¹⁷, skin rashes and/or eruptions at unusual sites^{14,18}, varicella-like skin eruptions¹⁸, allergic dermatitis-like rashes¹² and desquamation of palms and soles^{10,14,18}. As the paradigm changes, it is imperative that a better understanding of CV-A6, the emerging HFMD causative agent, is achieved.

This study was aimed at analysing the cytokine and chemokine profiles of HFMD patients from Singapore and Malaysia. Cytokines are a group of small secretory signalling molecules with diverse immune-related roles²⁴. Chemokines are a subclass of cytokines which have chemotactic properties²⁵. Several independent studies had previously shown an association between elevated inflammatory cytokines and HFMD pathogenesis and progression^{26–39}. However, none of these studies has examined HFMD cases from Singapore and Malaysia. In addition, previous studies have not analysed HFMD cases associated with CV-A6 infections, which was addressed in the present study.

Results and Discussion

The recruited HFMD study cohort was screened for its viral etiology and the individuals were grouped according to the causative etiological agents that they were infected with. There were a total of 2 CV-A16-infected patients, 11 EV-A71-infected patients, 10 CV-A6-infected patients, and 9 healthy volunteers enrolled for the Singapore cohort, as well as 1 CV-A16-infected patient, 34 EV-A71-infected patients, and 1 CV-A6-infected patient enrolled for the Malaysia cohort. Since this study involved children of young age, it was difficult to obtain parental consent and voluntary compliance in the collection of sera from the HFMD patients and healthy children in Singapore/Malaysia's context. Hence, we were unable to enrol a large cohort for our studies.

While EV-A71 remained as the major serotype isolated in both the Singapore and Malaysia cohorts, we observed a paradigmatic shift towards the CV-A6 serotype in the more recent recruitment from Singapore. On the contrary, the supposedly common CV-A16 serotype was rarely isolated in both our study cohorts. Several studies also reported similar epidemiological trends⁴⁰.

In this retrospective study, we focused on evaluating the cytokine and chemokine responses of EV-A71-associated HFMD cases between the Singapore and Malaysia cohort. In addition, we also explored the cytokine and chemokine responses of HFMD patients infected by EV-A71 and CV-A6 from Singapore. To our knowledge, cytokine and chemokine profiling has not been done in CV-A6-associated HFMD cases, which is of our interest to its recent predominance in Singapore. The fine-tuning of the levels of myriad cytokines and chemokines, which are crucial immune modulators, upon viral infection determines an anti-viral state advantageous to the hosts or a pro-viral state advantageous to the invading viruses. Hence, evaluation of the cytokines and chemokines would provide insights to the dynamics of HFMD disease development depending on the causative agents implicated. We profiled the HFMD and healthy control sera for an array of 48 cytokines and chemokines and compared the levels across the different serotypes (Supplementary Table S1). CV-A16-infected patients from both cohorts and CV-A6-infected patients from the Malaysia cohort were excluded from the comparative analyses as the extremely small sample size is not a true representative of the respective serotypes.

The heat map shown in Fig. 1 revealed two distinctive clusters representing HFMD patients from the two different cohorts, implying dissimilarities in the dysregulation of cytokines/chemokines between the Singapore and Malaysia cohorts.

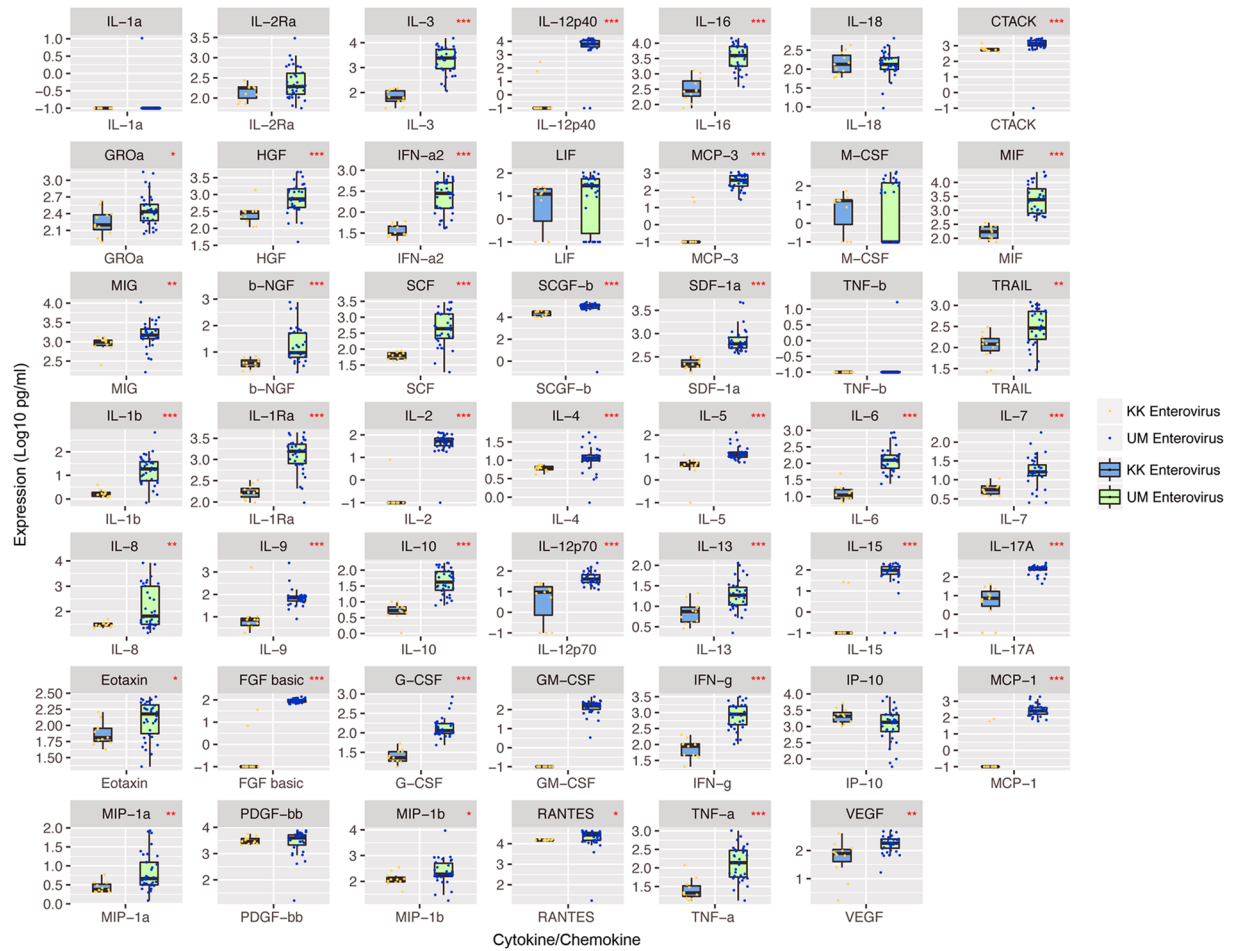


Figure 2. The expression profiles of EV-A71-infected patients from Singapore and Malaysia showed significant differences in 39 out of the panel of 48 cytokines and chemokines assayed. The statistical analyses were performed using GraphPad Prism version 4.0 (GraphPad software, USA), using Mann-Whitney non-parametric tests (without Gaussian distribution assumption). *p value < 0.05, **p value < 0.01, ***p value < 0.001. KK Enterovirus refers to EV-A71 cases from Singapore cohort, while UM Enterovirus refers to EV-A71 cases from Malaysia cohort.

EV-A71 serotypes were compared between the two cohorts, and Mann-Whitney non-parametric tests confirmed statistically significant differences between the two cohorts in 39 out of the panel of 48 cytokines and chemokines assayed (Fig. 2).

Since the Malaysia cohort was recruited in 2000 and 2012 while the Singapore cohort was recruited between 2013 and 2014, we speculated that the circulating EV-A71 (the major serotype isolated) genotype might be a contributing factor to the differences observed. To address this, comparisons were made between the EV-A71-infected patients recruited in 2000 and those recruited in 2012, where the main circulating EV-A71 genotypes were B4/C5 and B5 respectively⁴¹. Mann-Whitney non-parametric tests showed that 13 out of 48 cytokines were statistically different between the two periods of recruitment, where the HFMD patients from 2000 had higher sera levels of these cytokines/chemokines except for CTACK (Supplementary Figure S2). This might suggest that different EV-A71 genotypes could trigger host immune responses in different magnitudes, thereby emphasising a need to consider the circulating genotype when studying cytokine/chemokine dysregulation in EV-A71-associated HFMD.

Previous profiling studies mainly explored HFMD or HFMD of different severities in EV-A71- and/or CV-A16- infected patients^{26–39}. Since CV-A6-associated HFMD cases were becoming more prevalent in the Singapore cohort, we were particularly interested in its differential cytokine/chemokine dysregulation in comparison to EV-A71-associated HFMD cases since this area had not been explored. A comparative analysis was performed between EV-A71- and CV-A6-associated HFMD cases in the Singapore cohort, using healthy control sera as a reference (Fig. 3).

Interestingly, EV-A71 and CV-A6 shared a large number of cytokines/chemokines dysregulation: EV-A71 and CV-A6-infected HFMD patients had significantly reduced levels of sera IL-1 α , MIF, IL-1 β , IL-2, IL-8, IL-15, FGF-basic, GM-CSF, MCP-1, MIP-1 α , and MIP-1 β as well as increased levels of sera IL-16, IL-18, CTACK, β -NGF, SDF-1 α , IL-5, IL-7, IL-13, IP-10, and PDGF- β β . In addition to the large set of commonly dysregulated

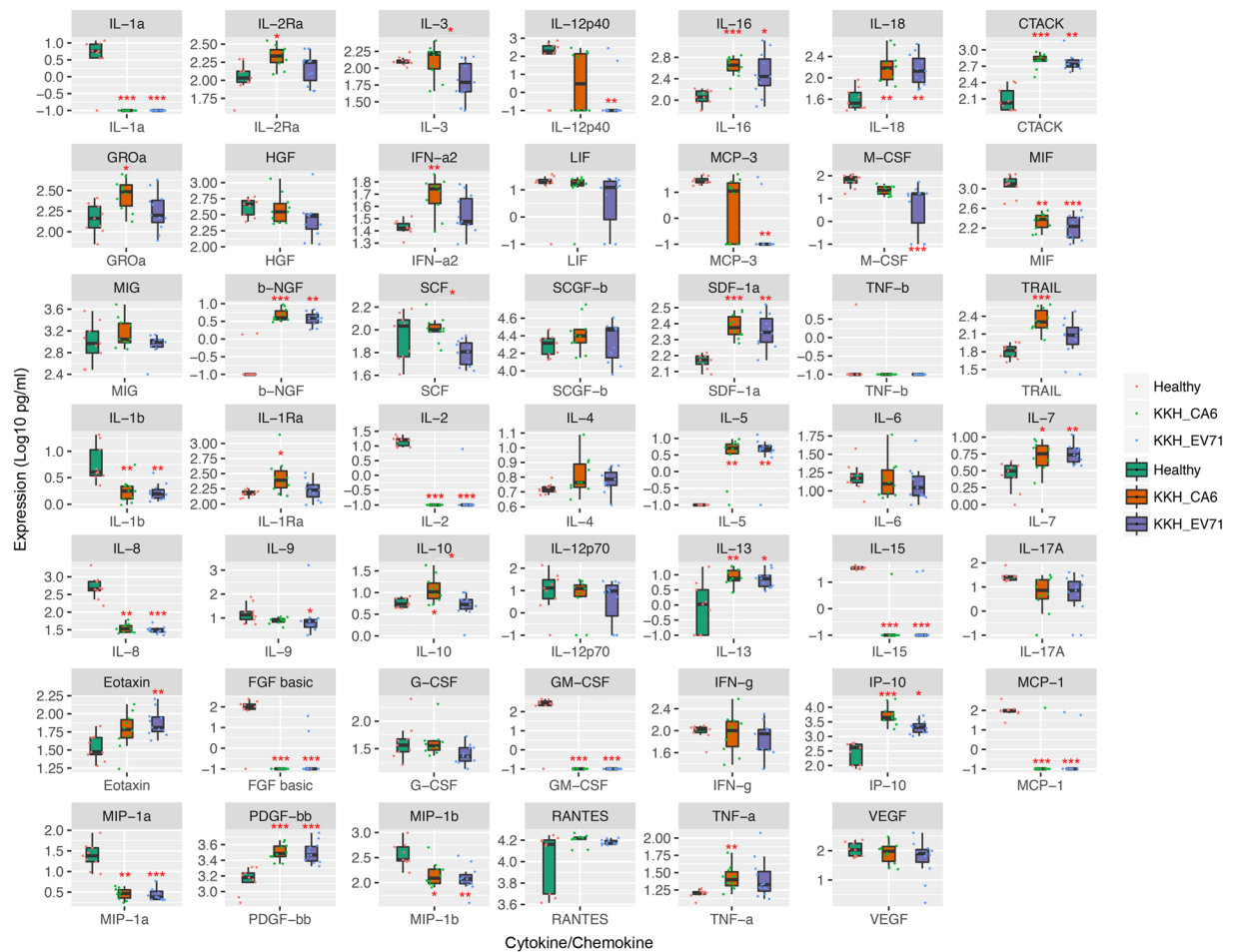


Figure 3. The cytokine and chemokine expression profiles of HFMD patients from Singapore showed significant differences between the two major etiological agents they were infected with. The statistical analyses were performed using GraphPad Prism version 4.0 (GraphPad software, USA), using Kruskal-Wallis non-parametric test (without Gaussian distribution assumption) coupled with Dunns post-test. All p-values were automatically adjusted by the program to account for false discovery rates associated with multiple comparisons. *p value < 0.05, **p value < 0.01, ***p value < 0.001. All p values stated were referenced to the healthy cohort, unless otherwise indicated by lines. Healthy refers to healthy volunteers, KKH_CA6 refers to CV-A6 cases, and KKH_EV71 refers to EV-A71 cases from Singapore cohort.

cytokines and chemokines, the two disease serotypes had their own subset of cytokine and chemokine dysregulation. EV-A71-associated HFMD cases displayed depressed expression of IL12p40, MCP-3, M-CSF, and IL-9 as well as augmented eotaxin. CV-A6-associated HFMD cases showed increased expression of IL-2R α , GRO- α , IFN- α 2, TRAIL, IL-1R α , IL-10, and TNF- α .

Indeed, previous studies also presented HFMD cases of decreased levels of MIF, IL-8, MIP-1 β , and M-CSF²⁹, as well as increased levels of IL-18⁴², IL-5³⁶, IP-10⁴²⁻⁴⁴, and eotaxin²⁹. In addition, EV-A71 cases of increased levels of IL-18³² and IL-13³³ have also been reported. Hence, these cytokines probably present generic cytokines implicated in HFMD cases. Some of the cytokines/chemokines including IL-8³², GM-CSF³⁰, MCP-1^{42,44}, IL-12p40²⁹, and MCP-3²⁹ were found to be dysregulated in opposite trends in other studies, thereby emphasising possible functional redundancies.

MIF, a pro-inflammatory cytokine of the immune system, is associated with disease severity and poor disease outcome⁴⁵. Since HFMD is generally a mild and self-limiting disease and the patients from the Singapore cohort suffered from uncomplicated HFMD with full recoveries, MIF was expectedly decreased in most of them. IL-7 has also been reported to inhibit immunosuppressive Socs3 protein responsible for chronic virus infections⁴⁶, possibly a key factor for a generally self-limiting disease. Cook *et al.* (1995) demonstrated that coxsackievirus-infected homozygous MIP-1 α mutant mice suffered from less severe pneumonitis and were protected from virus-induced myocarditis even though there was delayed virus clearance in comparison to infected wildtypes⁴⁷. Since MIP-1 β shared 67% sequence identity with MIP-1 α ⁴⁸, it was speculated that down-regulation of MIP-1 β was also a measure to avert disease progression. IP-10 augmentation was not surprising since it was shown to inhibit CB3 viral replication⁴⁹. MCP-3 has also been shown to inhibit viral replication as part of the anti-viral host defense

mechanism⁵⁰. M-CSF, shown to suppress CVB3 virus infection, reduce disease severity, and improve survival⁵¹, was reduced in EV-A71-associated HFMD cases.

IL-2 and GM-CSF were reduced perhaps to subvert host immunity. A recent publication by Felix *et al.* (2016) proposed the concept of virus-induced molecular mimicry in antagonising host-induced anti-viral cytokines (such as IL-2 and GM-CSF)⁵². The phenomenon of IL-8 reduction was not surprising as Dodd *et al.* (2001) showed how poliovirus, which belongs to the same family as enteroviruses, could limit IL-8 secretion during infection⁵³. IL-15, a key regulator of anti-viral response during innate immunity⁵⁴, was suppressed probably in the favor of the viruses. IL-9, reported to inhibit viral replication in CVB3-induced myocarditis⁵⁵, was suppressed as well. IL-1R α enhancement was also shown to be a virus' evasion strategy^{56,57}. FGF basic, which was reported to induce local infection to promote the healing process⁵⁸, was significantly diminished. This might explain the higher probability of complications in EV-A71 infections²⁻⁷, as well as more persistency in the generally mild CV-A6 infections⁵⁹ causing consequences such as onychomadesis as late onset manifestations. IL-16 was demonstrated to retard virus spread and control virus propagation⁶⁰, and this could account for the late onset manifestations in the generally mild CV-A6 infections. TRAIL⁶¹ and IL-10⁶² has both pro- and anti-viral roles, hence it is unclear which role it serves in CV-A6-associated HFMD.

The suppression of IL-2, a Th1 cytokine, might possibly be a virus strategy to counter IL-2R α increment. Indeed, IL-12, which is responsible for the induction of Th1 cytokines, was also diminished. IL-12 induction of Th1 cytokines was reported to be vital for anti-viral immunity and virus clearance^{63,64}. These supported previous studies which reported implications of skewed Th2 cytokine response resembling the pathophysiology of allergic diseases in HFMD patients. The authors reasoned that the Th2-biased responses could be the fatal trigger for respiratory dysfunctions and failure³² or indicator of poor prognosis with higher fever and longer illness duration⁶⁵.

Indeed, many cytokines and chemokines involved in a Th2 response or implicated in allergic diseases were dysregulated. IL-18, an imperative contributor of the generation of cytotoxic T lymphocytes against invading viruses⁶⁶, was reported to be able to skew towards a Th2 response in the absence of IL-12 and IL-15⁶⁷. β -NGF, produced by effector cells of allergic diseases, was amplified in HFMD patients as in allergic diseases such as asthma^{68,69} and allergic dermatitis^{70,71}. SDF-1 α , another factor implicated in chronic inflammatory skin diseases⁷² and asthma⁷³, could also explain previous publications on HFMD similarity to allergic diseases. IL-5 is a pleiotropic Th2 cytokine commonly associated to asthma pathogenesis⁷⁴. IL-13, also a Th2 cytokine, was implicated as an important contributor to virus-induced acute airway hyper-responsiveness and inflammation leading to asthma⁷⁵. IL-7 was reported as a plausible contributor to airway inflammation in asthmatics⁷⁶. Eotaxin, an eosinophil chemoattractant, was implicated in virus-induced airway inflammatory processes and disease exacerbations⁷⁷⁻⁸⁰. MCP-1, however, was surprisingly decreased since its augmentation was also previously linked to Th2 polarisation⁸¹. C-TACK, a predominantly cutaneous T cell selective chemokine⁸², was up-regulated. A skin-specific immune response was anticipated since HFMD is enanthematous and exanthematous in nature.

IFN- α 2 and TNF- α have pyrogenic roles which may contribute to the febrile response commonly observed in patients with HFMD^{83,84}. However the supposedly most potent endogenous pyrogens – IL-1 α and IL-1 β ⁸⁵ – were oddly diminished. Dinarello (2004) reported that infection-associated fever could be induced independent of pyrogenic cytokines (such as IL-1 and TNF) via TLR⁸⁶. Another speculation was the virus' strategy to regulate host temperature changes in order to persist in the host without much deterrence. Indeed, virus-shedding studies by Li *et al.*⁸⁷ and Zhao *et al.*⁸⁸ revealed EV-A71 could persist in the body for weeks even after the disease symptoms had subsided.

PDGF-BB induces proliferation through MAPK, JAK/STAT and PI3K pathways⁸⁹. Wu *et al.* (2016) reported a positive correlation between cell proliferation and EV-A71 infection through a genome-wide RNAi screen, and revealed that many top hits were involved in cell proliferation or growth factor signalling pathways⁹⁰.

In summary, it should be stressed that interpreting the overall outcome of the dynamic fluctuations of cytokine/chemokine cascade, instead of monitoring specific cytokine/chemokine alterations, would be more insightful to understanding the disease due to the functional redundancies of cytokines and chemokines⁹¹. This could also explain the irreproducibility of cytokine/chemokine dysregulation implicated in HFMD across different publications²⁶⁻³⁹. Our study also showed that the different cytokine/chemokine dysregulation triggered by EV-A71- and CV-A6-associated HFMD seemed to result in a similar clinical outcome. These observations could perhaps reflect the virus' versatility in eliciting the same outcome through different channels. Hence, it is important to take into account the overall outcome(s) of the different combination of cytokine/chemokine dysregulation rather than a particular (set of) cytokine/chemokine dysregulation as disease biomarkers. From this study, it seemed that the viruses were striking an astute balance between exploitation of the host machinery for invasion and aversion of the host immunity for evasion.

First, we found that the dysregulation was seemingly triggered by the viruses in order to attain a balanced control of the host immune responses. In order for viruses to invade and propagate within the host, viruses need to modulate the host immune system such that they can take dominance over the host yet avoid over-stimulating the host immune system which will result in death. As such, viruses have to be able to strike a delicate balance to co-exist with and persist in the host, fighting off host anti-viral responses and restraining its invasion simultaneously. Next, a skewed Th2 response mirroring allergic diseases was presumably an attempt to avoid anti-viral immunity and virus clearance by Th1 cytokines^{63,64}. In addition, the enanthematous and exanthematous nature of HFMD would naturally beget a skin-specific immune response. Since HFMD is a febrile illness, pyrogenic cytokines were also anticipated to be implicated. However, it was not surprising that cytokine-independent pyretic factors could still result in fever⁸⁶. Lastly, an active cell mitotic status (G2/M phase of cell cycle) was postulated to promote viral replication in the proliferating cells. Moreover, mitotic progression could also initiate a switch from cap-dependent translation to internal-ribosome site-dependent translation necessary for enterovirus translation processes⁹⁰.

While independent reports of EV-A71-induced ‘cytokine storm’, which describes a persistently over-stimulated immune response as a result of a positive feedback loop between cytokines and immune cells^{92,93}, have been documented previously^{37,94–96}, our study showed that the augmented cytokines in EV-A71-infected patients were also up-regulated in CV-A6-infected patients with the exception of eotaxin. Instead, EV-A71-infected patients had decreased sera levels of a subset of cytokines/chemokines not found to be significantly dysregulated in CV-A6-infected patients. In addition, CV-A6-infected patients had increased sera levels of another subset of cytokines/chemokines not found to be significantly dysregulated in EV-A71-infected patients. As previous studies have not explored CV-A6-associated cytokine/chemokine dysregulation, this pilot study showed for the first time the dynamics of this emerging HFMD causative agent in the host. Although CV-A6 usually causes self-limiting infections unlike neurotrophic EV-A71, its persistence causing consequences such as onychomadesis as late onset manifestations remained puzzling. These findings showed that CV-A6 is capable of eliciting greater (or at least the same) cytokine/chemokine dysregulation than (as) EV-A71, thereby highlighting a need to explore serotype-specific cytokine/chemokine dysregulation in HFMD patients to understand the clinical presentations associated with the respective etiological agents.

For example, onychomadesis was reported in young children who did not receive treatment from autosomal recessive loss-of-function mutations in *IL1RN* gene resulting in IL-1Ra deficiency. Interestingly, CV-A6-infected patients were found to have diminished levels of IL-1 α and IL-1 β as well as augmented levels of IL-1Ra⁹⁷. Since onychomadesis is a late onset manifestation observed in CV-A6-associated HFMD cases, it was speculated that the over-compensation of cytokine/chemokine dysregulation might have triggered this. Other than CV-A6-associated HFMD, patients recovering from toxic shock syndrome also experienced desquamation one to three weeks after onset as well as nail shedding and Beau’s lines after recovery⁹⁸. Cytokines such as TNF- α , IL-1, IL-6, IL-2, and IFN- γ were amplified in these patients. The expression of TNF- α was also increased in CV-A6-infected patients. Psoriasis lesions were found to have increased levels of cytokines including TNF- α and decreased levels of cytokines including IL-1 and IL-10. However, Saulite and colleagues found augmented IL-10 in psoriasis-affected nail beds, suggesting that IL-10 could specifically be linked to nail changes⁹⁹. Coincidentally, IL-10 was also exhibited at high levels in CV-A6-associated HFMD cases. It would be interesting to study the implications of differential cytokine/chemokine dysregulation between EV-A71 and CV-A6 by exploring the influence(s) of these cytokines and/or chemokines on the pathogenesis of the two viruses.

Despite a small sample size, this is the first study on the cytokine/chemokine profiles of the HFMD patients from the Singapore and Malaysia cohort. In particular, we showed that the Singapore and Malaysia cohorts had differential cytokine/chemokine dysregulation, even when infected with the same causative serotype (i.e. EV-A71). Further analyses showed that differences were observed even when comparison was made between different periods of recruitment within the same cohort. This showed that causative EV-A71 genotype played a contributing role in the differential dysregulation. Comparative analyses between the two major causative serotypes isolated from the Singapore cohort showed that causative serotype played a contributing role in the differential dysregulation as well. We showed for the first time the similarity and dissimilarity in the cytokine/chemokine dysregulation between EV-A71- and CV-A6-infected HFMD patients. As samples were obtained from consented patients upon hospitalisation, it was not possible to pre-determine the stage of infection the patients were in during sample collection. Hence, the differential profiles could also be partly influenced by the different stages of infection. Nevertheless, we hypothesised that the viruses were capable of flexible fine-tuning of the plethora of cytokines/chemokines towards a similar outcome. Further studies remain to explore and understand the intricate network of cytokines and chemokines triggered or attenuated by the virus or host to beget a particular clinical outcome.

Materials and Methods

Subject recruitment, sample collection and disease status verification. This study was approved by and carried out under the guidelines of the Ethics Committee of the SingHealth Centralised Institutional Review Board (CIRB; CIRB reference number: 2012/448/E) for the recruitment from KK Women’s and Children’s Hospital, Singapore, as well as the Medical Ethics Committee (reference number: 872.7) and the Medical Research and Ethics Committee of the Ministry of Health, Malaysia (reference number: NMRR-12-1038-13816) for the recruitment from the University of Malaya Medical Center, Malaysia. Informed consent was obtained from the parents/guardians of all the recruited subjects.

In this study, the patients were evaluated by clinicians using a clinical case definition of fever, oral ulcers, and skin lesions on the palms and soles for HFMD. The clinically-defined HFMD cases were confirmed using at least one of the laboratory tests, including viral genotyping, virus isolation and genotyping, as well as anti-EV-A71 IgM antibody detection. Only laboratory-confirmed cases were included in this study. Serum samples from patients diagnosed with HFMD and healthy volunteers were collected for cytokine and chemokine profiling.

Human Cytokine and Chemokine Bio-Plex Assays. The patient and healthy sera were assayed using the Bio-Plex Pro™ human cytokine 21-plex assay kit (Bio-Rad #MF0005KMII) and Bio-Plex Pro™ human cytokine 27-plex assay kit (Bio-Rad #M500KCAF0Y) according to the manufacturer’s instructions. The raw data was processed using Bio-Plex Manager software version 6.1 (Bio-Rad Laboratories, USA). Cytokines with out-of-range (OOR) readings that were below its detection limits were recorded as 0.1 pg/ml for subsequent statistical analyses which involved log transformation of data.

Statistical Analyses. All statistical analyses were performed using GraphPad Prism version 4.0 (GraphPad software, USA), using Mann-Whitney or Kruskal-Wallis non-parametric tests (without Gaussian distribution assumption). Kruskal-Wallis non-parametric tests were coupled with Dunns post-test to compare all pairs of groups. All p-values were automatically adjusted by the program to account for false discovery rates associated with multiple comparisons. (Appropriately-adjusted) p-values below 0.05 were considered to be statistically significant.

References

- Li, L.-J. Review of hand, foot and mouth disease. *Front. Med. China* **4**, 139–146, <https://doi.org/10.1007/s11684-010-0033-6> (2010).
- Chow, K. C. *et al.* Congenital enterovirus 71 infection: a case study with virology and immunohistochemistry. *Clinical infectious diseases: an official publication of the Infectious Diseases Society of America* **31**, 509–512, <https://doi.org/10.1086/313975> (2000).
- Chan, K. P. *et al.* Epidemic hand, foot and mouth disease caused by human enterovirus 71, Singapore. *Emerg Infect Dis* **9**, 78–85, <https://doi.org/10.3201/eid0901.020112> (2003).
- He, S. J. *et al.* [Three cases of enterovirus 71 infection with pulmonary edema or pulmonary hemorrhage as the early clinical manifestation]. *Chinese journal of pediatrics* **46**, 513–516 (2008).
- Makonkawkeyoon, K., Sudjaritruk, T., Sirisanthana, V. & Silvilairat, S. Fulminant enterovirus 71 infection: case report. *Annals of tropical paediatrics* **30**, 245–248, <https://doi.org/10.1179/146532810x12703902516446> (2010).
- Zhang, Y. C. *et al.* Clinicopathologic features and molecular analysis of enterovirus 71 infection: report of an autopsy case from the epidemic of hand, foot and mouth disease in China. *Pathology international* **62**, 565–570, <https://doi.org/10.1111/j.1440-1827.2012.02837.x> (2012).
- Lee, D. S. *et al.* Massive pulmonary hemorrhage in enterovirus 71-infected hand, foot, and mouth disease. *Korean journal of pediatrics* **58**, 112–115, <https://doi.org/10.3345/kjp.2015.58.3.112> (2015).
- Repass, G. L., Palmer, W. C. & Stancampiano, F. F. Hand, foot, and mouth disease: identifying and managing an acute viral syndrome. *Cleveland Clinic journal of medicine* **81**, 537–543, <https://doi.org/10.3949/ccjm.81a.13132> (2014).
- Österback, R. *et al.* Coxsackievirus A6 and Hand, Foot, and Mouth Disease, Finland. *Emerging Infectious Diseases* **15**, 1485–1488, <https://doi.org/10.3201/eid1509.090438> (2009).
- Wei, S.-H. *et al.* An outbreak of coxsackievirus A6 hand, foot, and mouth disease associated with onychomadesis in Taiwan, 2010. *BMC Infectious Diseases* **11**, 346, <https://doi.org/10.1186/1471-2334-11-346> (2011).
- Tsuguto, F. *et al.* Hand, Foot, and Mouth Disease Caused by Coxsackievirus A6, Japan, 2011. *Emerging Infectious Disease journal* **18**, 337, <https://doi.org/10.3201/eid1802.111147> (2012).
- Mathes, E. F. *et al.* “Eczema coxsackium” and unusual cutaneous findings in an enterovirus outbreak. *Pediatrics* **132**, e149–157, <https://doi.org/10.1542/peds.2012-3175> (2013).
- Puenpa, J. *et al.* Hand, foot, and mouth disease caused by coxsackievirus A6, Thailand, 2012. *Emerg Infect Dis* **19**, 641–643, <https://doi.org/10.3201/eid1904.121666> (2013).
- Downing, C. *et al.* Coxsackievirus A6 associated hand, foot and mouth disease in adults: clinical presentation and review of the literature. *Journal of clinical virology: the official publication of the Pan American Society for Clinical Virology* **60**, 381–386, <https://doi.org/10.1016/j.jcv.2014.04.023> (2014).
- Fonseca, M. C. *et al.* Coxsackievirus A6 and enterovirus 71 causing hand, foot and mouth disease in Cuba, 2011–2013. *Archives of virology* **159**, 2451–2455, <https://doi.org/10.1007/s00705-014-2071-x> (2014).
- Hongyan, G. *et al.* Hand, foot and mouth disease caused by coxsackievirus A6, Beijing, 2013. *The Pediatric infectious disease journal* **33**, 1302–1303, <https://doi.org/10.1097/inf.0000000000000467> (2014).
- Sinclair, C. *et al.* Atypical hand, foot, and mouth disease associated with coxsackievirus A6 infection, Edinburgh, United Kingdom, January to February 2014. *Euro surveillance: bulletin European sur les maladies transmissibles = European communicable disease bulletin* **19**, 20745 (2014).
- Chatproedprai, S. *et al.* Unusual skin manifestation of hand, foot and mouth disease associated with coxsackievirus A6: cases report. *SpringerPlus* **4**, 362 (2015).
- Ang, L. *et al.* Epidemiology and control of hand, foot and mouth disease in Singapore, 2001–2007. *Ann Acad Med Singapore* **38**, 106–112 (2009).
- Lee, M.-S. *et al.* Challenges to Licensure of Enterovirus 71 Vaccines. *PLoS neglected tropical diseases* **6**, e1737, <https://doi.org/10.1371/journal.pntd.0001737> (2012).
- Ministry of Health, S. *Communicable Diseases Surveillance in Singapore*, https://www.moh.gov.sg/content/moh_web/home/Publications/Reports.html (2015).
- Pacific, W. H. O. R. O. f. t. W. *Emerging disease surveillance and response: Hand, Foot and Mouth Diseases*, http://www.wpro.who.int/emerging_diseases/HFMD.situation.updates.archive/en/ (2015).
- Wu, Y. *et al.* The largest outbreak of hand; foot and mouth disease in Singapore in 2008: the role of enterovirus 71 and coxsackievirus A strains. *Int J Infect Dis* 2010 Dec; 14(12), e1076–81, Epub 2010 Oct 16..
- Cameron, M. J. & Kelvin, D. J. (Landes Bioscience, Austin (Texas), 2000).
- Griffith, J. W., Sokol, C. L. & Luster, A. D. Chemokines and Chemokine Receptors: Positioning Cells for Host Defense and Immunity. *Annual Review of Immunology* **32**, 659–702, <https://doi.org/10.1146/annurev-immunol-032713-120145> (2014).
- Lin, T. *et al.* Different proinflammatory reactions in fatal and non-fatal enterovirus 71 infections: implications for early recognition and therapy. *Acta Paediatr* **91**, 632–635 (2002).
- Wang, S., Zhao, Y. & Zhang, X. Multiplex serum cytokine levels in patients with severe HFMD determined by Luminex suspension array. *Beijing Medical Journal* **34**, 185–188 (2012).
- Wang, W. *et al.* Interleukin-8 is elevated in severe hand, foot, and mouth disease. *J Infect Dev Ctries* **8**, 94–100 (2014).
- Zeng, M. *et al.* The cytokine and chemokine profiles in patients with hand, foot and mouth disease of different severities in Shanghai, China, 2010. *PLoS neglected tropical diseases* **7**, e2599, <https://doi.org/10.1371/journal.pntd.0002599> (2013).
- Zhang, Y. *et al.* Comparative study of the cytokine/chemokine response in children with differing disease severity in enterovirus 71-induced hand, foot, and mouth disease. *PLoS ONE* 2013 Jun 28; 8(6), e67430, Print 2013 (2013).
- Duan, G. *et al.* Serum Inflammatory Cytokine Levels Correlate with Hand-Foot-Mouth Disease Severity: A Nested Serial Case-Control Study. *PLoS ONE* **9**, e112676, <https://doi.org/10.1371/journal.pone.0112676> (2014).
- Han, J. *et al.* Serum cytokine profiles of children with human enterovirus 71-associated hand, foot, and mouth disease. *Journal of Medical Virology* **86**, 1377–1385, <https://doi.org/10.1002/jmv.23929> (2014).
- Chen, Z. *et al.* IL-6, IL-10 and IL-13 are associated with pathogenesis in children with Enterovirus 71 infection. *International journal of clinical and experimental medicine* **7**, 2718–2723 (2014).
- Li, W. *et al.* Study on Risk Factors for Severe Hand, Foot and Mouth Disease in China. *PLoS ONE* **9**, e87603, <https://doi.org/10.1371/journal.pone.0087603> (2014).
- Li, T., Jiang, T. & Liang, L. Changes in serum cytokine levels in hand-foot-and-mouth disease. *Biomedical Research - India* **26**, 594–598 (2015).
- Ye, N. *et al.* Cytokine responses and correlations thereof with clinical profiles in children with enterovirus 71 infections. *BMC Infectious Diseases* **15**, 225 (2015).
- Zhang, S. *et al.* Immunologic Characterization of Cytokine Responses to Enterovirus 71 and Coxsackievirus A16 Infection in Children. *Medicine* **94**, 000000000001137 (2015).
- Luo, Q., Peng, W. & Chen, L. I. Coxsackievirus A16 infection stimulates imbalances of T cells in children. *Experimental and Therapeutic Medicine* **9**, 2213–2218, <https://doi.org/10.3892/etm.2015.2405> (2015).
- Wang, J. *et al.* Clinical and Associated Immunological Manifestations of HFMD Caused by Different Viral Infections in Children. *Global Pediatric Health* **3**, 2333794X16643723, <https://doi.org/10.1177/2333794x16643723> (2016).
- Li, J. *et al.* Characterization of Coxsackievirus A6- and Enterovirus 71-Associated Hand Foot and Mouth Disease in Beijing, China, from 2013 to 2015. *Frontiers in Microbiology* **7**, 391, <https://doi.org/10.3389/fmicb.2016.00391> (2016).

41. NikNadia, N. M. N. *et al.* Cyclical Patterns of Hand, Foot and Mouth Disease Caused by Enterovirus A71 in Malaysia. *PLoS neglected tropical diseases* **10**, e0004562, <https://doi.org/10.1371/journal.pntd.0004562> (2016).
42. Shang, W., Qian, S., Fang, L., Han, Y. & Zheng, C. Association study of inflammatory cytokine and chemokine expression in hand foot and mouth disease. *Oncotarget* **8**, 79425–79432, <https://doi.org/10.18632/oncotarget.18341> (2017).
43. Hua, Y., Wang, Y. & Gong, W. Inflammatory cytokine profiles of serum and cerebrospinal fluid in Chinese children with hand, foot and mouth disease. *Int J Clin Exp Pathol* **10**, 11022–11029 (2017).
44. Shao, P. *et al.* Clinical significance of inflammatory cytokine and chemokine expression in hand, foot and mouth disease. *Molecular medicine reports* **15**, 2859–2866, <https://doi.org/10.3892/mmr.2017.6324> (2017).
45. Calandra, T. & Roger, T. Macrophage migration inhibitory factor: a regulator of innate immunity. *Nat Rev Immunol* **3**, 791–800 (2003).
46. Pellegrini, M. *et al.* IL-7 engages multiple mechanisms to overcome chronic viral infection and limit organ pathology. *Cell* **144**, 601–613, <https://doi.org/10.1016/j.cell.2011.01.011> (2011).
47. Cook, D. N. *et al.* Requirement of MIP-1 alpha for an inflammatory response to viral infection. *Science (New York, N.Y.)* **269**, 1583–1585 (1995).
48. Ren, M. *et al.* Polymerization of MIP-1 chemokine (CCL3 and CCL4) and clearance of MIP-1 by insulin-degrading enzyme. *The EMBO Journal* **29**, 3952–3966, <https://doi.org/10.1038/emboj.2010.256> (2010).
49. Yuan, J. *et al.* CXCL10 inhibits viral replication through recruitment of natural killer cells in coxsackievirus B3-induced myocarditis. *Circulation research* **104**, 628–638, <https://doi.org/10.1161/circresaha.108.192179> (2009).
50. Schols, D., Proost, P., Van Damme, J. & De Clercq, E. RANTES and MCP-3 inhibit the replication of T-cell-tropic human immunodeficiency virus type 1 strains (SF-2, MN, and HE). *Journal of Virology* **71**, 7300–7304 (1997).
51. Hiraoka, Y., Kishimoto, C., Takada, H., Suzaki, N. & Shiraki, K. Colony-stimulating factors and coxsackievirus B3 myocarditis in mice: macrophage colony-stimulating factor suppresses acute myocarditis with increasing interferon-alpha. *Am Heart J* **130**, 1259–1264 (1995).
52. Felix, J. *et al.* Structural basis of GM-CSF and IL-2 sequestration by the viral decoy receptor GIF. *Nature Communications* **7**, 13228, <https://doi.org/10.1038/ncomms13228> (2016).
53. Dodd, D. A., Giddings, T. H. & Kirkegaard, K. Poliovirus 3A Protein Limits Interleukin-6 (IL-6), IL-8, and Beta Interferon Secretion during Viral Infection. *Journal of Virology* **75**, 8158–8165, <https://doi.org/10.1128/JVI.75.17.8158-8165.2001> (2001).
54. Rodrigues, L. & Bonorino, C. Role of IL-15 and IL-21 in viral immunity: applications for vaccines and therapies. *Expert Rev Vaccines* **8**, 167–177 (2009).
55. Yu, M. *et al.* IL-9 Inhibits Viral Replication in Coxsackievirus B3-Induced Myocarditis. *Frontiers in Immunology* **7**, 409, <https://doi.org/10.3389/fimmu.2016.00409> (2016).
56. Roberge, C. J., Poubelle, P. E., Beaulieu, A. D., Heitz, D. & Gosselin, J. The IL-1 and IL-1 receptor antagonist (IL-1Ra) response of human neutrophils to EBV stimulation. Preponderance of IL-1Ra detection. *Journal of immunology (Baltimore, Md.: 1950)* **156**, 4884–4891 (1996).
57. Hill-Batorski, L. *et al.* Loss of Interleukin 1 Receptor Antagonist Enhances Susceptibility to Ebola Virus Infection. *J Infect Dis* **212**(Suppl 2), S329–335, <https://doi.org/10.1093/infdis/jiv335> (2015).
58. Kalicke, T. *et al.* Local application of basic fibroblast growth factor increases the risk of local infection after trauma: an *in-vitro* and *in-vivo* study in rats. *Acta Orthop* **78**, 63–73 (2007).
59. Huang, Y. *et al.* Characterization of severe hand, foot, and mouth disease in Shenzhen, China, 2009–2013. *J Med Virol* **87**, 1471–1479 (2015).
60. Archin, N. M., van den Boom, L., Perelygina, L., Hilliard, J. M. & Atherton, S. S. Delayed spread and reduction in virus titer after anterior chamber inoculation of a recombinant of HSV-1 expressing IL-16. *Invest Ophthalmol Vis Sci* **44**, 3066–3076 (2003).
61. Cummins, N. & Badley, A. The TRAIL to Viral Pathogenesis: The Good, the Bad and the Ugly. *Current molecular medicine* **9**, 495–505 (2009).
62. Rojas, J. M., Avia, M., Martin, V. & Sevilla, N. IL-10: A Multifunctional Cytokine in Viral Infections. *J Immunol Res* **2017**, 6104054, <https://doi.org/10.1155/2017/6104054> (2017).
63. Rossol, S. *et al.* Interleukin-12 induction of Th1 cytokines is important for viral clearance in chronic hepatitis B. *Journal of Clinical Investigation* **99**, 3025–3033 (1997).
64. Tagawa, T. *et al.* Epstein-Barr viral miRNAs inhibit antiviral CD4+T cell responses targeting IL-12 and peptide processing. *The Journal of Experimental Medicine* <https://doi.org/10.1084/jem.20160248> (2016).
65. Wei, R. *et al.* Elevated antigen-specific Th2 type response is associated with the poor prognosis of hand, foot and mouth disease. *Virus Research* **177**, 62–65, <https://doi.org/10.1016/j.virusres.2013.07.009> (2013).
66. Dinarello, C. A. Interleukin-1 beta, interleukin-18, and the interleukin-1 beta converting enzyme. *Annals of the New York Academy of Sciences* **856**, 1–11 (1998).
67. Nakanishi, K., Yoshimoto, T., Tsutsui, H. & Okamura, H. Interleukin-18 is a unique cytokine that stimulates both Th1 and Th2 responses depending on its cytokine milieu. *Cytokine and Growth Factor Reviews* **12**, 53–72, [https://doi.org/10.1016/S1359-6101\(00\)00015-0](https://doi.org/10.1016/S1359-6101(00)00015-0).
68. Braun, A. *et al.* Role of nerve growth factor in a mouse model of allergic airway inflammation and asthma. *Eur J Immunol* **28**, 3240–3251, [https://doi.org/10.1002/\(sici\)1521-4141\(199810\)28:10<3240::aid-immu3240#62;3.0.co;2-u](https://doi.org/10.1002/(sici)1521-4141(199810)28:10<3240::aid-immu3240#62;3.0.co;2-u) (1998).
69. Bonini, S. *et al.* Nerve growth factor and asthma. *Allergy* **57**(Suppl 72), 13–15 (2002).
70. Hodeib, A. *et al.* Nerve growth factor, neuropeptides and cutaneous nerves in atopic dermatitis. *Indian Journal of Dermatology* **55**, 135–139, <https://doi.org/10.4103/0019-5154.62735> (2010).
71. Yamaguchi, J., Aihara, M., Kobayashi, Y., Kambara, T. & Ikezawa, Z. Quantitative analysis of nerve growth factor (NGF) in the atopic dermatitis and psoriasis horny layer and effect of treatment on NGF in atopic dermatitis. *J Dermatol Sci* **53**, 48–54, <https://doi.org/10.1016/j.jdermsci.2008.08.011> (2009).
72. Zraggen, S., Huggenberger, R., Kerl, K. & Detmar, M. An important role of the SDF-1/CXCR4 axis in chronic skin inflammation. *PLoS One* **9**, e93665, <https://doi.org/10.1371/journal.pone.0093665> (2014).
73. Mo, B. *et al.* The role of SDF-1/CXCR4 on airway inflammation and airway remodeling in a rat asthma model. *Chinese journal of tuberculosis and respiratory diseases* **38**, 39–44 (2015).
74. Takatsu, K. Interleukin-5 and IL-5 receptor in health and diseases. *Proceedings of the Japan Academy. Series B, Physical and Biological Sciences* **87**, 463–485, <https://doi.org/10.2183/pjab.87.463> (2011).
75. Cormier, S. A. & Kolls, J. K. Innate IL-13 in virus-induced asthma? *Nat Immunol* **12**, 587–588 (2011).
76. Kelly, E. A. B. *et al.* Potential Contribution of IL-7 to Allergen-Induced Eosinophilic Airway Inflammation in Asthma. *Journal of immunology (Baltimore, Md.: 1950)* **182**, 1404–1410 (2009).
77. Conroy, D. M. & Williams, T. J. Eotaxin and the attraction of eosinophils to the asthmatic lung. *Respiratory Research* **2**, 150, <https://doi.org/10.1186/rr52> (2001).
78. Kawaguchi, M. *et al.* Expression of eotaxin by normal airway epithelial cells after influenza virus A infection. *Int Arch Allergy Immunol* **122** Suppl 1, 44–49, doi:53632 (2000).
79. Kawaguchi, M. *et al.* Influenza virus A stimulates expression of eotaxin by nasal epithelial cells. *Clin Exp Allergy* **31**, 873–880 (2001).
80. Papadopoulos, N. G. *et al.* Rhinovirus infection up-regulates eotaxin and eotaxin-2 expression in bronchial epithelial cells. *Clin Exp Allergy* **31**, 1060–1066 (2001).

81. Deshmane, S. L., Kremlev, S., Amini, S. & Sawaya, B. E. Monocyte Chemoattractant Protein-1 (MCP-1): An Overview. *Journal of Interferon & Cytokine Research* **29**, 313–326, <https://doi.org/10.1089/jir.2008.0027> (2009).
82. Morales, J. *et al.* CTACK, a skin-associated chemokine that preferentially attracts skin-homing memory T cells. *Proceedings of the National Academy of Sciences of the United States of America* **96**, 14470–14475 (1999).
83. Dinarello, C. A. Cytokines as Endogenous Pyrogens. *Journal of Infectious Diseases* **179**, S294–S304, <https://doi.org/10.1086/513856> (1999).
84. Vybiral, S., Bárczayová, L., Pešanová, Z. & Janský, L. Pyrogenic effects of cytokines (IL-1 β , IL-6, TNF- α) and their mode of action on thermoregulatory centers and functions. *Journal of Thermal Biology* **30**, 19–28, <https://doi.org/10.1016/j.jtherbio.2004.06.003> (2005).
85. Dinarello, C. A. The history of fever, leukocytic pyrogen and interleukin-1. *Temperature: Multidisciplinary Biomedical Journal* **2**, 8–16, <https://doi.org/10.1080/23328940.2015.1017086> (2015).
86. Dinarello, C. Infection, fever, and exogenous and endogenous pyrogens: some concepts have changed. *J Endotoxin Res* **10**, 201–222 (2004).
87. Li, J. *et al.* Excretion of enterovirus 71 in persons infected with hand, foot and mouth disease. *Virology Journal* **10**, 31, <https://doi.org/10.1186/1743-422x-10-31> (2013).
88. Zhao Shi-yong, L. X.-Y. *et al.* Survey of virus shedding and spread mechanism in children infected with EV71. *Disease Surveillance* **30**, 23–26, <https://doi.org/10.3784/j.issn.1003-9961.2015.01.007> (2015).
89. van Roeyen, C. R. C. *et al.* Biological responses to PDGF-BB versus PDGF-DD in human mesangial cells. *Kidney International* **69**, 1393–1402, <https://doi.org/10.1038/sj.ki.5000332> (2006).
90. Wu, K. X. *et al.* Human genome-wide RNAi screen reveals host factors required for enterovirus 71 replication. *Nature Communications* **7**, 13150, <https://doi.org/10.1038/ncomms13150> (2016).
91. Ozaki, K. & Leonard, W. J. Cytokine and Cytokine Receptor Pleiotropy and Redundancy. *Journal of Biological Chemistry* **277**, 29355–29358, <https://doi.org/10.1074/jbc.R200003200> (2002).
92. Clark, I. A. The advent of the cytokine storm. *Immunol Cell Biol* **85**, 271–273 (0000).
93. Tisoncik, J. R. *et al.* Into the Eye of the Cytokine Storm. *Microbiology and Molecular Biology Reviews: MMBR* **76**, 16–32, <https://doi.org/10.1128/mmb.05015-11> (2012).
94. Chen, L.-C. & Yeh, T.-M. Enterovirus 71 Infection of Human Immune Cells Induces the Production of Proinflammatory Cytokines. *Journal of Biomedical and Laboratory Sciences* **21**, 82–90 (2009).
95. Gong, X. *et al.* Excessive proinflammatory cytokine and chemokine responses of human monocyte-derived macrophages to enterovirus 71 infection. *BMC Infectious Diseases* **12**, 224, <https://doi.org/10.1186/1471-2334-12-224> (2012).
96. Zhang, Y. *et al.* The Gene Expression Profile of Peripheral Blood Mononuclear Cells from EV71-Infected Rhesus Infants and the Significance in Viral Pathogenesis. *PLoS ONE* **9**, e83766, <https://doi.org/10.1371/journal.pone.0083766> (2014).
97. De Jesus, A. A. & Goldbach-Mansky, R. Monogenic Autoinflammatory Diseases: Concept And Clinical Manifestations. *Clinical Immunology* **147**, 155–174, <https://doi.org/10.1016/j.clim.2013.03.016> (2013).
98. Bologna, J. L., Jorizzo, J. L. & Schaffer, J. V. In *Toxic Shock Syndrome* Vol. 1 (Elsevier Saunders, USA, 2012).
99. Ventura, A. *et al.* New insight into the pathogenesis of nail psoriasis and overview of treatment strategies. *Drug Design, Development and Therapy* **11**, 2527–2535, <https://doi.org/10.2147/DDDT.S136986> (2017).

Acknowledgements

The authors would like to thank Biomedical Research Council, Agency for Science, Technology and Research and National Medical Research Council, Ministry of Health, Singapore for financially supporting this study.

Author Contributions

F.M.S.T. – designed and conducted the experiment, analysed the data, co-wrote the manuscript; M.N. – analysed the data; A.A.W., M.T.K., Y.F.C. – recruited the subjects and collected samples; N.W.H.T., C.Y.C. – designed the experiment; J.J.H.C. – designed the experiment, analysed the data, and co-wrote the manuscript.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-018-22379-6>.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018