Check for updates

CORRESPONDENCE OPEN (CEBPA bZIP in-frame mutations in acute myeloid leukemia: prognostic and therapeutic implications

© The Author(s) 2024

Blood Cancer Journal (2024)14:59; https://doi.org/ 10.1038/s41408-024-01042-6

TO THE EDITOR:

The transcription factor CCAAT/enhancer-binding proteinalpha(CEBPA) is a critical mediator of granulocytic differentiation. Mutations of the CEBPA gene (CEBPA^{mut}) occur in 5%–15% of adult AML patients and in-frame mutations within the bZIP domain of *CEBPA* (*CEBPA*^{bZIP-inf}) define a distinct entity associated with favorable prognosis in AML patients when treated by conventional chemotherapy [1, 2]. CEBPA mutations could activate the BCL2 P2 promoter and induce its expression via interaction with nuclear factor-kB (NF-kB) p50 in hematopoietic cell lines and display a markedly hypermethylated profile by multi-omics analysis in primary leukemia cells [3–5]. Meanwhile, venetoclax plus hypomethylating agents (VEN + HMA) were efficient in AML patients with specific molecular profiles (such as NPM1, IDH2, etc.) [6, 7]. However, relevant data related to the role of VEN + HMA regimens in $CEBPA^{bZIP-inf}$ AML patients is limited. Therefore, in the current study, we retrospectively analyzed the clinical features, co-mutational spectrum, and prognostic role of CEBPA mutations, particularly CEBPA^{bZIP-inf} mutations, in 996 newly diagnosed adult AML patients inducted with chemo-free regimens or chemotherapy between 2016–2022 at the First Affiliated Hospital of Soochow University in China.

A high frequency of *CEBPA* mutations (17.8%,177/996) and *CEBPA*^{bZIP-inf} cases (13.6%,135/996) were identified in our analysis (Fig. S1). *CEBPA*^{bZIP-inf} patients were diagnosed at a younger age, higher hemoglobin counts, lower platelet counts, more likely to have intermediate cytogenetics (normal karyotype) and less number of co-mutations compared with other *CEBPA* mutated (*CEBPA*^{other-mut}) and *CEBPA*^{wt} AML patients (Table S1 and Fig. S2). *CEBPA*^{bZIP-inf} mutations exhibited a higher complete remission/complete remission with incomplete cell recovery (CR/CRi) rate (87.6% vs. 64.1% vs. 47.3%, *P* < 0.001), favorable overall survival (OS) (3-year OS: 91.2% vs. 66.0% vs. 55.3%, *P* < 0.001) and relapse-free survival (RFS) (3-year RFS: 73.8% vs. 52.0% vs. 55.3%, *P* < 0.001) compared with *CEBPA*^{other-mut} and *CEBPA*^{wt} cases in the matched cohort (Fig. S3) (Table S2 and S3).

Among 130 *CEBPA*^{bZIP-inf} patients with available information, 116 cases were inducted by 7 + 3 chemotherapy while 14 patients were by VEN + HMA regimens and consolidated by chemotherapy or hematopoietic stem cell transplantation (HSCT). There was no difference in the baseline and genetic characteristics between the 7 + 3 cohort and the VEN + HMA cohort (Table 1). With a median follow-up time of 22 months, among 13 patients evaluable all attained CR/CRi, 46.2%(6/13) relapsed and 3 patients died in the VEN + HMA group (disease progression, n = 1; transplant-related complications, n = 1; pneumonia, n = 1); simultaneously, 113 of 116 patients attained CR/CRi, 22.1%(25/113) relapsed and 6 cases died in the 7 + 3group. Intriguingly, the VEN + HMA regimens seemed to show similar CR/CRi rates after one cycle of induction therapy (100.0% vs. 86.2%, P = 0.324), lower RFS (1-year RFS: 46.9% vs. 88.9%; P < 0.001) and OS (1-year OS: 84.6% vs. 99.1%; P < 0.001) than 7 + 3 regimens (Fig. 1A, B). When patients were censored at HSCT, worse 1-year RFS and 1-year OS were also observed in the VEN + HMA cohort (41.5% vs. 83.5%, P = 0.003 for RFS; 90.0% vs. 98.4%, P = 0.010 for OS) (Fig. 1C, D). In accordance with these findings, the multivariable analysis demonstrated adverse RFS (HR, 2.72; 95% CI: 1.01–7.30; P = 0.047) and a trend to dismal OS (HR, 5.14; 95% CI: 0.83-31.60; P = 0.078) in VEN + HMA cohort compared to 7 + 3 cohorts (Table S4). Totally, 126 CEBPA^{bZIP-inf} cases (96.9%) achieved CR/CRi post-

induction and 31 of them (24.6%) relapsed, especially in the VEN + HMA cohort with a CR/CRi rate of 100% and relapse rate of 46.2%, which may be different from primary refractory group or persistent remission group and could be classified into "remission but relapse" group. Thus, it's rather meaningful to explore high-risk indicators (such as co-mutations) for relapse and treatment regimens (such as consolidation) to reduce the relapse rate. With regard to consolidation therapy, 76 patients received cytarabine-based chemotherapy and 54 patients underwent transplantation in CR1 due to Measurable Residual Disease (MRD) positivity, more PTPN11, or FLT3-ITD mutations. HSCT at CR1 improved RFS (3-year RFS: 95.2% vs. 57.5%; P < 0.001) but not OS (3-year OS: 96.2% vs. 86.8%; P = 0.156) compared with chemotherapy alone in CEBPA^{bZIP-inf} AML patients (Fig. 1E, F), which is consistent with Cox Proportional Hazard Model (HR 0.04, 95% CI: 0.01-0.19; P < 0.001 for RFS in multivariable analysis; HR 0.34; 95% CI: 0.07-1.70; P = 0.184 for OS in univariable analysis) (Table S4).

On the other hand, overlapping gene mutations may affect the clinical outcome of *CEBPA*^{bZIP-inf} patients. The most frequently co-mutated genes within the *CEBPA*^{bZIP-inf} group were *WT1* (27.7%), *GATA2* (26.2%), *NRAS* (13.8%), *FLT3-ITD* (13.1%) and *RUNX1* (10.8%) (Fig. S2). Notably, *KIT* mutations had a significantly poor impact on RFS (P < 0.001) and OS (P = 0.002) compared to *KIT* wild type in *CEBPA*^{bZIP-inf} group (Fig. S4), and *CSF3R* mutation showed a worse RFS (P = 0.035) by performing landmark analysis after 10 months (crossover) (Fig. S4). Meanwhile, *WT1* and NRAS mutations revealed a tendency toward adverse RFS (P = 0.053, P = 0.067) and OS (P = 0.068, P = 0.069) (Fig. S4). Additionally, MDS-related gene mutations (MRs, including *ASXL1*, *BCOR*, *EZH2*, *SF3B1*, *SRSF2*, *STAG2*, *U2AF1*, *ZRSR2*) and *GATA2* aberrations had no effect on survival (Fig. S4). P-value

VEN + HMA

Variables

	(<i>N</i> = 116)	(<i>N</i> = 14)			
Age in years, median [IQR]	34.5 [29.8,45.0]	40.5 [33.3,49.8]	0.167		
Sex (Male), n (%)	70 (60.3)	7 (50.0)	0.648		
WBC (median [IQR]) × 109/L	16.5 [8.6,68.2]	18.2 [8.3,42.7]	0.943		
Hemoglobin, (median [IQR])	102.0 [85.0,115.0]	103.5 [84.8,117.3]	0.834		
Platelet, (median [IQR]) × 109/L	26.0 [17.0,49.0]	32.0 [26.3,45.0]	0.554		
BM blast(median [IQR]) (%)	56.8 [37.8,70.0]	58.3 [40.7,80.1]	0.555		
Co-mutations, n (%)					
Activated signaling genes					
CSF3R	13 (11.2)	0 (0.0)	0.396		
FLT3-ITD	15 (12.9)	2 (14.3)	1.000		
KIT	4 (3.4)	1 (7.1)	1.000		
NRAS	14 (12.1)	4 (28.6)	0.201		
PTPN11	6 (5.2)	1 (7.1)	1.000		
Transcription factors genes					
GATA2	29 (25.0)	5 (35.7)	0.589		
RUNX1	11 (9.5)	3 (21.4)	0.365		
Chromatin modifiers genes					
ASXL1/2	4 (3.4)	0 (0.0)	1.000		
BCOR	2 (1.7)	0 (0.0)	1.000		
EZH2	12 (10.3)	0 (0.0)	0.439		
SETD2	6 (5.2)	0 (0.0)	0.844		
Tumor suppressors genes					
WT1	31 (26.7)	5 (35.7)	0.694		
DNA methylation genes					
DNMT3A	7 (6.0)	2 (14.3)	0.554		
IDH1/2	2 (1.7)	1 (7.1)	0.739		
TET2	12 (10.3)	2 (14.3)	1.000		

 Table 1. Baseline characteristics of patients with CEBPA^{bZIP-inf}
mutations in the 7 + 3 and VEN + HMA cohort.

7 + **3**

median [IQR]	[29.8,45.0]					
Sex (Male), <i>n</i> (%)	70 (60.3)	7 (50.0)	0.648			
WBC (median [IQR]) × 109/L	16.5 [8.6,68.2]	18.2 [8.3,42.7]	0.943			
Hemoglobin, (median [IQR])	102.0 [85.0,115.0]	103.5 [84.8,117.3]	0.834			
Platelet, (median [IQR]) × 109/L	26.0 [17.0,49.0]	32.0 [26.3,45.0]	0.554			
BM blast(median [IQR]) (%)	56.8 [37.8,70.0]	58.3 [40.7,80.1]	0.555			
Co-mutations, n (%)						
Activated signaling genes						
CSF3R	13 (11.2)	0 (0.0)	0.396			
FLT3-ITD	15 (12.9)	2 (14.3)	1.000			
KIT	4 (3.4)	1 (7.1)	1.000			
NRAS	14 (12.1)	4 (28.6)	0.201			
PTPN11	6 (5.2)	1 (7.1)	1.000			
Transcription factors	genes					
GATA2	29 (25.0)	5 (35.7)	0.589			
RUNX1	11 (9.5)	3 (21.4)	0.365			
Chromatin modifiers	genes					
ASXL1/2	4 (3.4)	0 (0.0)	1.000			
BCOR	2 (1.7)	0 (0.0)	1.000			
EZH2	12 (10.3)	0 (0.0)	0.439			
SETD2	6 (5.2)	0 (0.0)	0.844			
Tumor suppressors genes						
WT1	31 (26.7)	5 (35.7)	0.694			
DNA methylation genes						
DNMT3A	7 (6.0)	2 (14.3)	0.554			
IDH1/2	2 (1.7)	1 (7.1)	0.739			
TET2	12 (10.3)	2 (14.3)	1.000			
RNA splicing genes	RNA splicing genes					
SF3B1/U2AF1/ ZRSR2	2 (1.7)	1 (7.1)	0.739			
Cohesin complex gen	ies					
STAG2	4 (3.4)	0 (0.0)	1.000			
Cytogenetics, n (%)	116 (100)	14 (100)	0.739			
Favorable risk	0	0				
Intermediate risk	114 (98.3)	13 (92.9)				
Adverse risk	2 (1.7)	1 (7.1)				
Induction response, <i>n</i> (%)	116 (100%)	13 (92.9%)	0.311			
CR/CRi	100 (86.2)	13 (100.0)				
PR or NR	16 (13.8)	0 (0.0)				
HSCT in CR1, <i>n</i> (%)	51 (44.0)	3 (21.4)	0.184			

WBC white blood cell count, BM bone marrow, CR/CRi complete remission or CR with incomplete hematologic recovery, PR partial response, NR no response, HSCT hematopoietic stem cell transplantation, CR1 first complete remission.

Vazguez previously found that all 4(100%) CEBPA-mutated AML patients achieved CR/CRi but 3(75%) relapsed with a shorter duration of remission (median: 4.65 months) in 19 unfit AML patients treated by venetoclax-based regimens [8]. Similarly, our cohort suggested that VEN + HMA induction exhibited a similar remission rate, worse RFS, and a tendency to poor OS in CEBPA^{bZIP-} cohort compared with 7 + 3 groups. It seems the treatment depth or clonal evolution of VEN + HMA needs to be further explored. On the side, as earlier studies reported [9, 10], we also demonstrated HSCT in CR1 significantly improved RFS but not OS compared to chemotherapy alone, possibly resulting from a high remission rate of re-induction by chemotherapy after relapse and high transplantation-related mortality. Therefore, HSCT is not recommended in consolidation therapy at first CR in CEBPA^{bZIP-inf} AML patients. In other words, conventional chemotherapy is preferred in both induction and consolidation courses among CEBPA^{bZIP-inf} AML cases.

The co-occurrence of other genetic mutations has had a controversial prognostic impact in patients with CEBPA^{mut} AML by previous studies, generally, mutations of WT1, CSF3R, KIT, NRAS, and CCS mutations (mutations in chromatic/DNA modifiers (C), cohesion complex (C), and splicing genes (S)) were associated with adverse prognosis while mutations of GATA2 were correlated with favorable outcome [9, 11–15]. Similar to these reports, KIT mutation was significantly related to inferior RFS and OS in Kaplan-Meier methods (RFS, P = 0.002; OS, P < 0.001) but not in multivariable analysis (HR 2.45, 95% CI: 0.63-9.55, P = 0.197 for RFS; HR 2.63, 95% CI: 0.30-23.04, P = 0.381 for OS), a possible explanation maybe the small sample size of KIT mutated patients in CEBPA^{bZIP-inf} patients. Inferior prognosis in terms of RFS was found in WT1(P = 0.053), CSF3R (P = 0.035), and NRAS (P = 0.069) mutated group while a trend of worse OS without statistical significance was observed (WT1, P = 0.068; NRAS, P = 0.067). However, no clinical significance of GATA2 mutation and MRs were found in the present cohort.

Recently, Georgi divided 1010 CEBPA^{mut} adult AML patients into 8 mutational subgroups and refined superior prognosis was associated with in-frame insertions/deletions within bZIP domain of CEBPA (CEBPA bZIP^{InDel}) rather than in-frame mutations within bZIP domain of CEBPA (CEBPA^{bZIP-inf}) by excluding missense mutations within bZIP domain (*CEBPA* bZIP^{ms}) [15]. The present *CEBPA*^{bZIP-inf} AML cohort was reanalyzed and 14 CEBPA bZIP^{ms} patients were excluded, the prognostic value of VEN + HMA induction and HSCT at CR1 consolidation was consistent with previous results (Fig. S5). Whether truncated mutations in the N-terminus and truncated sites influence the clinical outcome of CEBPA-mutated patients were analyzed. No statistical significance of OS (P = 0.490) and RFS (P = 0.412) were found in CEBPA transactivation domain (TAD) frameshift mutated patients and CEBPA other mutated patients (Fig. S6). No differences in OS (P = 0.240) and RFS (P = 0.571) were evaluated in N-terminal truncated mutations spanning the second start codon (Fig. S6).

In conclusion, CEBPA^{bZIP-inf} patients exhibited higher CR rates, improved OS and RFS, and benefited from traditional chemotherapy compared with VEN + HMA induction and transplantation consolidation at CR1 in our preliminary study. Furthermore, CEBPA^{bZIP-inf} patients with WT1 and NRAS mutation adversely affected the RFS in multivariable analysis. Taken the retrospective nature and limited sample size into consideration, future research efforts aimed at validating our results and elucidating the potential molecular mechanisms are warranted to improve therapeutic strategy in specific types of CEBPA^{mut} AML.



20 60 80 40 Time (Months) Number at risk 69 44 7 0 0 0 0 1 20 40 Time (Months) 60 80 VEN+HMA 7+3 P = 0.00320 80 60 40 Time (Months) Number at risk 30 18 2 0 0 0 0 1 20 40 Time (Months) 60 80 -HSCT in CR1 Others P< 0.001 80 20 60 40 Time (Months) Number at risk 31 18 2 0 39 26 5 0 20 80 40 Time (Months) 60

P < 0.001

Fig. 1 Kaplan-Meier survival curves for overall survival (OS) and relapse-free survival (RFS) in CEBPA^{bZIP-inf} patients. A, B compared survival outcomes among patients inducted with standard 7 + 3 and VEN + HMA regimens; C, D showed survival outcomes in the standard 7 + 3 and VEN + HMA cohorts with censoring at the time of HSCT; E, F illustrated survival by consolidation treatment (HSCT in CR1 or not). HSCT hematopoietic stem cell transplantation, CR1 first complete remission.

Fenghong Zhang $1^{2,3}$, Zhen Shen^{1,2,3}, Jundan Xie^{1,2}, Jingren Zhang^{1,2}, Qian Wu^{1,2}, Rui Jiang^{1,2}, Xiangyu Zhao^{1,2}, Xiaofei Yang^{1,2 \bowtie} and Suning Chen ^{1,2 \limsup}

¹National Clinical Research Center for Hematologic Diseases, Jiangsu Institute of Hematology, The First Affiliated Hospital of Soochow University, Suzhou, People's Republic of China. ²Institute of Blood and Marrow Transplantation, Collaborative Innovation Center of Hematology, Soochow University, Suzhou, China. ³These authors contributed equally: Fenghong Zhang, Zhen Shen. ⊠email: yangxiaofei1977@163.com; chensuning@sina.com

DATA AVAILABILITY

The data used for this study are not publicly available. Molecular and clinical characteristics are available upon request from the corresponding author.

REFERENCES

- Taube F, Georgi JA, Kramer M, Stasik S, Middeke JM, Röllig C, et al. CEBPA mutations in 4708 patients with acute myeloid leukemia: differential impact of bZIP and TAD mutations on outcome. Blood. 2022;139:87–103.
- Taskesen E, Bullinger L, Corbacioglu A, Sanders MA, Erpelinck CA, Wouters BJ, et al. Prognostic impact, concurrent genetic mutations, and gene expression features of AML with CEBPA mutations in a cohort of 1182 cytogenetically normal AML patients: further evidence for CEBPA double mutant AML as a distinctive disease entity. Blood. 2011;117:2469–75.
- Paz-Priel I, Cai DH, Wang D, Kowalski J, Blackford A, Liu H, et al. CCAAT/enhancer binding protein alpha (C/EBPalpha) and C/EBPalpha myeloid oncoproteins induce bcl-2 via interaction of their basic regions with nuclear factor-kappaB p50. Mol Cancer Res. 2005;3:585–96.
- Mo Q, Yun S, Sallman DA, Vincelette ND, Peng G, Zhang L, et al. Integrative molecular subtypes of acute myeloid leukemia. Blood Cancer J. 2023;13:71.
- Figueroa ME, Lugthart S, Li Y, Erpelinck-Verschueren C, Deng X, Christos PJ, et al. DNA methylation signatures identify biologically distinct subtypes in acute myeloid leukemia. Cancer Cell. 2010;17:13–27.

-VEN+HMA

- 7+3

- Gangat N, Karrar O, Iftikhar M, McCullough K, Johnson IM, Abdelmagid M, et al. Venetoclax and hypomethylating agent combination therapy in newly diagnosed acute myeloid leukemia: genotype signatures for response and survival among 301 consecutive patients. Am J Hematol. 2024;99:193–202.
- DiNardo CD, Tiong IS, Quaglieri A, MacRaild S, Loghavi S, Brown FC, et al. Molecular patterns of response and treatment failure after frontline venetoclax combinations in older patients with AML. Blood. 2020;135:791–803.
- Vazquez R, Breal C, Zalmai L, Friedrich C, Almire C, Contejean A, et al. Venetoclax combination therapy induces deep AML remission with eradication of leukemic stem cells and remodeling of clonal haematopoiesis. Blood Cancer J. 2021;11:62.
- Ahn SY, Kim T, Kim M, Song GY, Jung SH, Yang DH, et al. Clinical significance of bZIP in-frame CEBPA-mutated normal karyotype acute myeloid leukemia. Cancer Res Treat. 2023;55:1011–22.
- Schlenk RF, Taskesen E, van Norden Y, Krauter J, Ganser A, Bullinger L, et al. The value of allogeneic and autologous hematopoietic stem cell transplantation in prognostically favorable acute myeloid leukemia with double mutant CEBPA. Blood. 2013;122:1576–82.
- Chen X, Abuduaini D, Zhang Y, Long J, Lin X, Zhu H, et al. Characteristic immunophenotype and gene co-mutational status orchestrate to optimize the prognosis of CEBPA mutant acute myeloid leukemia. Blood Cancer J. 2023;13:70.
- Wei H, Zhou C, Liu B, Lin D, Li Y, Wei S, et al. The prognostic factors in acute myeloid leukaemia with double-mutated CCAAT/enhancer-binding protein alpha (CEBPAdm). Br J Haematol. 2022;197:442–51.
- Fasan A, Eder C, Haferlach C, Grossmann V, Kohlmann A, Dicker F, et al. GATA2 mutations are frequent in intermediate-risk karyotype AML with biallelic CEBPA mutations and are associated with favorable prognosis. Leukemia. 2013;27:482–5.
- Zhang Y, Wang F, Chen X, Zhang Y, Wang M, Liu H, et al. Companion gene mutations and their clinical significance in AML with double mutant CEBPA. Cancer Gene Ther. 2020;27:599–606.
- Georgi JA, Stasik S, Kramer M, Meggendorfer M, Röllig C, Haferlach T, et al. Prognostic impact of CEBPA mutational subgroups in adult AML. Leukemia. 2024;38:281–90.

ACKNOWLEDGEMENTS

The authors thank all the patients who participated in the current study. The authors appreciate the samples from the Hematological Biobank, Jiangsu Biobank of Clinical Resources. This study was supported by a grant from the National Key R&D Program of China(2022YFC2502701), and the National Natural Science Foundation of China (82170158, 81970142).

AUTHOR CONTRIBUTIONS

FHZ and ZS contributed to the study design. JDX, JRZ, and QW were responsible for mutation analysis and interpretation; RJ and XYZ collected data for the study. FHZ and ZS wrote the manuscript. All authors reviewed the manuscript, and XFY and SNC approved the final version, and supported this publication.

COMPETING INTERESTS

The authors declare no competing interests.

ADDITIONAL INFORMATION

Supplementary information The online version contains supplementary material available at https://doi.org/10.1038/s41408-024-01042-6.

Correspondence and requests for materials should be addressed to Xiaofei Yang or Suning Chen.

Reprints and permission information is available at http://www.nature.com/ reprints

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creativecommons.org/licenses/by/4.0/.

© The Author(s) 2024