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Effect of serum concentrations of IL-6 and TNF-α on brain structure in anorexia nervosa: a combined cross-sectional and longitudinal study

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Previous studies of brain structure in anorexia nervosa (AN) have reported reduced gray matter in underweight patients, which largely normalizes upon weight gain. One underlying biological mechanism may be glial cell alterations related to low-grade inflammation. Here, we investigated relationships between brain structure as measured by magnetic resonance imaging and serum concentrations of two pro-inflammatory cytokines (interleukin-6 and tumor necrosis factor alpha) cross-sectionally in 82 underweight adolescent and young adult female patients (mean age 16.8 years; 59 of whom were observed longitudinally after short-term weight restoration; mean duration 2.8 months), 20 individuals long-term weight-recovered from AN (mean age 22.7 years) and 105 healthy control (HC) participants (mean age 17.2 years). We measured cortical thickness, subcortical volumes and local gyrification index, a measure of cortical folding. In contrast to most previous studies of cytokine concentrations in AN, we found no cross-sectional group differences (interleukin-6: p = 0.193, tumor necrosis factor alpha: p = 0.057) or longitudinal changes following weight restoration (interleukin-6: p = 0.201, tumor necrosis factor alpha: p = 0.772). As expected, widespread gray matter reductions (cortical thickness, subcortical volumes, cortical folding) were observed in underweight patients with AN compared to HC. However, we found no evidence of associations between cytokine concentrations and structural brain measures in any participant group. Furthermore, longitudinal changes in cytokine concentrations were unrelated to changes in gray matter. In conclusion, we did not identify any association between (sub-)inflammatory processes and structural brain changes in AN. Future studies are needed to elucidate which other factors besides nutritional status may contribute to brain morphological alterations.

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INTRODUCTION

The factors contributing to the development and maintenance of anorexia nervosa (AN), a life-threatening eating disorder with a typical onset in adolescence, are not fully understood. For instance, accumulating evidence points toward a low-grade inflammation with increased pro-inflammatory cytokines in patients with AN [1]. This might be an underlying mechanism of glial cell alterations in AN, which may in turn contribute to the well-documented substantial reduction of brain mass including cortical thickness (CT) in the acute state of the disorder. However, to date, no study has systematically investigated the connection between cytokines and CT reduction in AN.

Underlying mechanisms of low-grade inflammation found in AN are still largely unclear, but a few hypotheses have been suggested. Elevated cortisol concentrations in the underweight state, excessive physical activity and the altered gut microbiome [2–4] may modulate intestinal permeability and allow bacteria/ bacterial parts to traverse into peripheral circulation inducing low-grade inflammation [5-7]. In line with this, two large metaanalyses showed elevated blood concentrations of proinflammatory cytokines in acute AN, with the largest number of samples for interleukin-6 (IL-6) and tumor necrosis factor alpha (TNF- α) [1, 8]. Both IL-6 and TNF- α may facilitate a prolonged response of the pro-inflammatory nuclear factor kappa beta, and TNF-a may increase vascular permeability and thus contribute to the "leaky gut" in AN [9]. Furthermore, animal models have provided evidence for anorexigenic effects of IL-6 and TNF- α [8]. IL-6 interacts with the appetite-regulating hormone leptin and regulates multiple aspects of metabolism [10]. Additionally, preliminary evidence from human studies proposes a link between exercise-induced IL-6 release and the transient suppression of appetite and food intake [11, 12]. Also, TNF-a promotes the production of anorexigenic peptides [8] and may act as a modulator of energy storage in adipocytes [13].

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Data on longitudinal trajectories of cytokine concentrations during weight restoration are scarce [8, 14].

In the underweight state of AN, patients show widespread and substantial structural gray matter (GM) changes, such as a reduction of CT and cortical folding [15, 16]. After long-term weight recovery, previous studies have shown these GM changes to mostly normalize [17]. Our own findings point toward a rapid normalization of CT and cortical folding during short-term weight restoration therapy of ~3 months [15, 18, 19]. However, cortical folding has a different genetic origin [20] and pattern of alterations compared to CT [15]. Since cortical surface area alterations in AN seem smaller compared to CT (~1.6% vs. 6.4%) and the interregional pattern of cortical volume alterations largely overlaps that observed for CT [18], GM volume changes seem to be driven by CT changes. The underlying mechanisms of structural GM changes remain elusive, with hypothesized mechanisms including fluid shifts due to changes in oncotic pressure, the loss, damage or remodeling of glia and/or neurons and hormonal changes [17]. Furthermore, inflammatory processes may contribute to changes in brain structure. While evidence for this is still scarce in AN, previous studies in other psychiatric disorders support such a link [21-29]. For instance, findings in major depressive disorder, which is one of the most common comorbidities in AN, give rise to the hypothesis that alterations in brain structure may be due to the neurotoxic effects of increased inflammation [21]. In AN, using a virtual histology analysis, we found that brain regions with a high expression of genes specific to oligodendrocytes are most affected by the reductions [18]. Proinflammatory factors such as circulating cytokines could lead to an activation of astrocytes, of which a subgroup may promote toxicity for neurons and oligodendrocytes [30]. Consistent with this, blood concentrations of the neuronal damage marker neurofilament light (NF-L) and of glial fibrillary acidic protein (GFAP), which has been associated with astroglial injury, are elevated in AN [31, 32], and NF-L has been linked to cortical thinning in AN [33]. The loss or dysfunction of oligodendrocytes might result in demyelination and eventually neuronal death.

Therefore, the aim of the present study was to investigate the potential relationship between inflammatory processes and dynamic structural GM changes in AN. To this end, we assessed serum concentrations of TNF-a and IL-6 as well as CT, local gyrification index (IGI), a measure of cortical folding, and volumes of subcortical gray matter nuclei in (1) young patients with AN with a relatively short duration of illness, both when underweight and after short-term weight restoration (lasting ~3 months), (2) individuals long-term recovered from AN (for at least 6 months) and (3) healthy control (HC) participants. Based on previous research [1, 8], we expected IL-6 and TNF- α to be elevated in the acute state of AN as a sign of low-grade inflammation, and that altered cytokine concentrations would be linked to reduced CT, IGI and volumes of subcortical nuclei in underweight patients with AN. Furthermore, we hypothesized a decrease in IL-6 and TNF-α over the course of short-term weight restoration to predict normalization of structural GM measures. Long-term weightrecovered individuals were included to differentiate alterations related to undernutrition (state markers) from alterations that may confer vulnerability toward AN (trait markers) or long-term effects of the disorder. Furthermore, we explored associations of cytokine concentrations with markers of disorder severity, i.e., psychopathology, body mass index standard deviation scores (BMI-SDS) and plasma leptin concentrations. Plasma leptin concentrations served as a marker of undernutrition as they reflect energy stores and positively adipose tissue mass [34].

METHODS AND MATERIALS

Participants

A total of 217 female participants took part in the study: (85 underweight patients with AN (acAN, 12–28 years old), 24 individuals long-term weight-

recovered from AN (recAN, 17–29 years old) and 108 healthy control participants (HC, 12–28 years old). All acAN participants were admitted to intensive treatment of specialized eating disorder programs at the child and adolescent psychiatry and psychosomatic medicine department of a university hospital. All acAN underwent the first assessments within 96 h of beginning nutritional rehabilitation (acAN-TP1). Sixty-three acAN were re-examined after partial weight restoration (at least 12% increase in body mass index (BMI) as inclusion criterion, range: 14.1–45.8%; acAN-TP2). After applying quality control procedures (see below), 266 scans were used in the analysis (82 acAN-TP1, 59 acAN-TP2, 20 recAN, 105 HC).

Diagnoses of eating disorders and other information regarding potential confounding variables (e.g., medication, comorbidities, smoking) were obtained using the expert form of the Structured Interview for Anorexia and Bulimia Nervosa (SIAB-EX) [35], supplemented with our own semistructured interview. Interviews were adapted to DSM-5 criteria and carried out by clinically experienced research assistants under the supervision of a child and adolescent psychiatrist. A diagnosis of AN required a BMI below the 10th age percentile (if <15.5 years old) or below 17.5 kg/m² (if >15.5 years old). RecAN participants had to have previously met diagnostic criteria for AN, maintained a BMI > 18.5 kg/m^2 (or the 10th age percentile if <18 years), had regular menstruation and not engaged in significant restrictive eating behavior (or binging/purging) for 6 months prior to study participation. HC participants had to have a normal BMI (18.5-29 kg/m² or >10th and <95th age percentile if <18 years), regular menstruation and have no history of psychiatric illness and were recruited through advertisement among middle/high school and university students. For exclusion criteria and comorbid diagnoses, see Supplementary 1.1. All protocols received ethical approval by the local Institutional Review Board, and all participants (or legal guardians) gave written informed consent.

Clinical measures

In addition to the evaluation with the SIAB-EX [35], we assessed eating disorder-related psychopathology with the Eating Disorder Inventory-2 (EDI-2) [36], depressive symptoms with the Beck Depression inventory-II (BDI-II) [37] and anxiety symptoms using the anxiety scale of the revised Symptom Checklist 90 (SCL-90-R) [38]. IQ was estimated with the Wechsler Adult Intelligence Scale (if age > 16 years) [39] or the Wechsler Intelligence Scale for Children (if age < 16 years) [40]. BMI-SDS [41] were computed to provide an age-corrected index.

Blood sample preparation and analysis of cytokines and leptin

Venous blood samples were collected between 7 and 9 a.m. after an overnight fast. To yield blood serum for IL-6 and TNF-a assessment, samples were left to clot for 30 min at 6–8 °C and then centrifuged $(2500 \times q, 15 \text{ min}, 15 \text{ min})$ 5 °C). To yield plasma for leptin assessment, aprotinin was added during blood sampling to prevent protein degradation by serine proteases and samples were immediately centrifuged ($2500 \times q$, 15 min, 5 °C). All samples were aliquoted immediately after centrifugation and stored at -80 °C. Serum IL-6 and TNF-a concentrations were measured in duplicate with commercially available high-sensitivity enzyme immunoassays (IL-6: IBL International GmbH, Hamburg, Germany; TNF-α: R&D Systems Inc., Minneapolis, MN, USA). Participants were excluded from analyses if the coefficient of variation between duplicate measurements exceeded 20% for either IL-6 or TNF-a (2 recAN, 2 HC). Only two IL-6 measurements (none for TNF-a) were below the limit of detection of 0.03 pg/ml and were imputed as $\frac{\text{limit of detection}}{\sqrt{2}}$. Plasma leptin concentrations were measured using a commercially available enzyme-linked immunosorbent assay (ELISA, BioVendor Research and Diagnostic Products, Brno, Czech Republic), For Jeptin, 20 acAN-TP1 measurements were below the limit of detection and were imputed using a quantile regression multiple imputation approach for left-censored missing data [42] (Supplementary 1.2).

Structural MRI acquisition and image data processing

All participants underwent MRI between 8 and 9 a.m. following an overnight fast. High-resolution three-dimensional T1-weighted structural scans were acquired on Siemens Magnetom Trio 3T Scanner with a magnetization-prepared rapid gradient-echo (MP-RAGE) sequence using the same parameters as in our previous studies [19, 43] (Supplementary 1.3). Reconstruction of the cerebral cortex was accomplished automatically with FreeSurfer 7.1 [44, 45] (Supplementary 1.4), followed by standardized quality control by trained raters (Supplementary 1.5). Scans with artefacts that exert a significant influence on parcellation (mainly dura inclusions in the pial surface) were excluded from the analysis (3 acAN-TP1, 4 acAN-TP2,

2 recAN and 1 HC). All images underwent further longitudinal preprocessing with the FreeSurfer longitudinal stream [46]. At each vertex on the resulting tessellated pial surface, CT [47] and IGI [48] were measured and smoothed using a Gaussian kernel with a full-width-at-half-maximum of 10 mm, and volumes of 8 subcortical gray-matter nuclei were measured (Supplementary 1.6).

Statistical analyses

Due to deviations from normality, IL-6, TNF- α and leptin were log_etransformed before parametric tests were applied. Spearman correlation coefficients were calculated to investigate possible associations between cytokine concentrations and clinical variables including BMI-SDS, leptin concentrations and psychopathology. Statistical significance was defined as p < 0.05.

Statistical modeling of CT and IGI was based on a linear mixed effect model implemented using Freesurfer mixed effect tools [49]. We aimed to test whether changes in cytokine concentrations explain variance in GM changes not already accounted for by changes in BMI-SDS [15, 19]. Given the distinct developmental trajectories of the three investigated measures [50], age was included as a linear covariate. In sum, we modeled CT and IGI as:

 $CT, IGI = A + \Delta_{recAN} + \Delta_{acAN} + B_{acAN}(b_t - b_{TP2}) + D_{acAN}(d_t - d_{TP2}) + C \text{ age}$

where b_t and d_t represent BMI-SDS and the log_e-transformed serum concentrations of the cytokines IL-6 or TNF- α , respectively, at timepoint *t* (TP1 or TP2). For subcortical volumes, we included total intracranial volume as an additional covariate.

This approach allowed us to examine cross-sectional group comparisons (acAN-TP1 vs. HC, recAN vs. HC), and longitudinal changes associated with BMI-SDS and cytokine concentrations [19]. Compared to the use of repeated measures to examine longitudinal changes, this approach allowed us to include also participants with single point data, which lead to a better estimation of the variance. To assess the role of AN subtype and of comorbidities, we repeated the above analyses by excluding participants with AN of binge/purge subtype and with psychiatric and somatic comorbidities (including very mild/local infections), respectively. Furthermore, to control for possible effects of smoking on IL-6 and TNF- α [51], and non-linear age effects, we included the variables "current number of cigarettes per day" and "ever-smoking status", and age 2 as covariates in supplementary analyses [52]. To investigate relationships between cytokine levels and brain structure in each group, we used general linear models with cytokine levels and age as covariates. We corrected for multiple comparisons using a false discovery rate q < 0.05 for each hemisphere unless otherwise noted, and across cytokine type using the Bonferroni criterium. Further analysis details are provided in Supplementary 1.7, 1.8. Statistical analyses were performed using Matlab, R [53] and IBM SPSS Statistics for Windows, version 27.0 (IBM Corp., Armonk, NY).

RESULTS

Demographic and clinical characteristics

The demographic and clinical data are presented in Table 1. As expected, acAN-TP1 patients had lower BMI-SDS and leptin concentrations and higher symptom levels (EDI-2, BDI-II, SCL-90-R anxiety scale) than HC, which improved after short-term weight restoration (except for anxiety). Most notably, BMI increased by mean(SD) = 27.0(8.4)% during weight-restoration treatment (duration mean(SD) = 2.77(0.84) months, range 35–155 days), but was still under the age-appropriate average in acAN-TP2. RecAN were slightly older and still had some residual psychological symptoms. Furthermore, recAN consumed a higher number of cigarettes per day than acAN.

Cytokines

One-way ANOVAs revealed no significant differences between the acAN-TP1, recAN and HC groups in IL-6, F(2,204) = 1.66, p = 0.193 (Fig. 1), or TNF- α concentrations, F(2,204) = 2.91, p = 0.057 (Fig. 2). These results did not change when covarying for age (Supplementary 2.1) or smoking (Supplementary 2.2), or using non-parametric tests (Supplementary 2.3). Bayesian analysis provided anecdotal evidence in support for the null hypothesis (Supplementary 2.4).

In acAN, paired samples *t*-tests showed no change of serum concentrations over the course of short-term weight restoration for IL-6, t(58) = -1.30, p = 0.201 (Fig. 1), or TNF- α , t(58) = -0.29, p = 0.772 (Fig. 2), and Bayesian analysis provided moderate evidence in support for the null hypothesis (Supplementary 2.4). Both the cross-sectional and the longitudinal analyses yielded the same results when excluding participants with psychiatric and somatic comorbidities (Supplementary 2.5) or of the AN binge/ purge subtype (Supplementary 2.6). In the correlation analysis, no link between cytokine concentrations and BMI-SDS, leptin or measures of psychopathology (EDI-2, BDI-II total scores, SCL-90-R anxiety scale) emerged after correcting for multiple comparisons (Supplementary Table S1).

Structural MRI measures

Replicating our previous findings [15, 19], we observed a widespread reduction of CT and IGI in acAN at baseline (acAN-TP1) relative to HC, and longitudinal increases related to partial weight restoration (Figs. S1 and S2). No significant differences between recAN and HC were revealed, neither in CT nor in IGI. However, IGI was still reduced in some areas of the cortex in acAN-TP2 (Fig. S3). Contrary to our motivating hypothesis, neither CT, nor IGI were associated with IL-6 or TNF-α concentrations in any of the groups considered (acAN-TP1, acAN-TP2, recAN, HC, Figs. S4–S11). Furthermore, longitudinal changes in IL-6 (Fig. 3) or TNF- α (Fig. 4) were not associated with longitudinal changes in either CT, IGI, or subcortical volumes (Table S3). This was also the case when (1) excluding participants with psychiatric and somatic comorbidities (Figs. S12 and S13) or of the binge-purge subtype from the AN groups (Figs. S14 and S15). No effects of age² (Fig. S16) and "ever-smoking status" (Fig. S17) were evident, but the "current number of cigarettes per day" was associated with CT (but not IGI) alterations in the motor and pre-motor cortex (Fig. S18). However, even when accounting for these additional covariates, longitudinal changes in IL-6 (Fig. S19) or TNF-a (Fig. S20) were still not associated with longitudinal changes in either CT or IGI.

DISCUSSION

In the present study, we measured serum concentrations of the pro-inflammatory cytokines IL-6 and TNF- α in conjunction with sMRI scans cross-sectionally in a large sample of underweight adolescents and young adults with AN, individuals long-term weight-recovered from AN, and HC. Most patients with AN were re-examined after short-term weight restoration. To the best of our knowledge, our study was the first to investigate the potential relationship between peripheral inflammatory markers and structural brain changes in AN. Contrary to our hypothesis of a low-grade systemic inflammation in the acute state of the disorder, there were no significant group differences in cytokine concentrations between acAN-TP1, recAN, and HC in our adolescent and young adult sample. Also, the longitudinal analysis indicated no change in cytokine concentrations over the course of short-term weight restoration (mean duration 2.8 months). We replicated previous findings of widespread cortical thinning and reduced cortical folding (assessed by IGI) and reduced volumes of subcortical GM nuclei in acAN-TP1 compared to HC, which were largely reversed already after short-term weight restoration. However, no measure of GM structure was linked to cytokine concentrations in any of the groups. Furthermore, there were no associations between longitudinal changes in CT, IGI, or subcortical volumes and changes in cytokine concentrations. These findings were confirmed in supplementary analyses controlling for non-linear effects of age and smoking as well as excluding participants with the AN binge/purge subtype and with psychiatric and somatic comorbidities.

lable I. Demographic ar	ng ciinical gata.								
	Observation time	e point	Longitudinal change	Control samples		Cross-sec	tional ar	nalysis	
	acAN-TP1 (N = 82)	acAN-TP2 (<i>N</i> = 59)	acAN-TP2–acAN-TP1 (N = 59)	recAN (N=20)	HC (N = 105)	F/H	đ	d	post hoc tests
Age	16.8(3.2)	16.7(2.2)	0.23(0.07)***	22.7(3.4)	17.2(3.5)	25.47	204	<0.001	recAN > acAN-TP1*** recAN > HC***
BMI (kg/m ²)	15.0(1.2)	19.2(1.0)	4.0(1.0)***	21.6(1.5)	20.6(2.0)	279.71	204	<0.001	acAN-TP1 < recAN,HC***
BMI-SDS	-3.1(1.2)	-0.7(0.6)	2.3(0.7)***	-0.23(0.54)	-0.13(0.66)	257.57	204	<0.001	acAN-TP1 < recAN,HC***
Minimal lifetime BMI (kg/m²)	14.6(1.3)	n/a	n/a	14.6(1.6)	19.5(1.8)	233.65	192	<0.001	HC > acAN-TP1,recAN***
Q	111(12)	n/a	n/a	111.9(10.5)	111.68(9.07)	0.03	197	0.975	All not significant
BDI-II total score	24.1(10.8)	15.1(11.2)	7.8(10.5)***	8.1(8.0)	4.8(5.6)	129.19	202	<0.001	acAN-TP1 > recAN,HC***
EDI-2 total score	213.0(45.0)	195.1(52.7)	12.4(40.9)*	154.7(34.9)	138.1(27.2)	98.55	198	<0.001	acAN-TP1 > recAN,HC***
SCL-90-R anxiety scale	1.0 (0.8)	0.7 (0.7)	-0.1 (0.4)	0.4 (0.4)	0.3 (0.5)	37.03	202	<0.001	acAN-TP1 > recAN,HC***
Plasma leptin (μg/l)	1.46(1.75)	12.05(6.37)	10.56(5.80)***	9.37(5.01)	12.12(8.95)	136.54	202	<0.001	acAN-TP1 < recAN, HC***
Cigarettes per day	0.1(0.9)	0.2(1.3)	0.03(0.18)	2.1(4.6)	0.3(1.6)	9.92	2	0.007	acAN-TP1 < recAN**
Mean (standard deviation) (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.01$	are presented for eac 0.001), and longitudir	ch group in a separate c al differences between a	column. Group differences betwe acAN-TP1 and acAN-TP2 were tes	en acAN-TP1, recAN a ted with dependent se	and HC were tested amples t-tests. Due	d with a one to deviation	e-way AN	OVA follow ormality, lep	ed by Tukey's post hoc test tin concentrations and SCL-

process, process were loge-transformed before parametric tests were applied. Due to the right-skewed distribution of the variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) of the variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for constant) states scores were loge-transformed before parametric tests were applied. Due to the right-skewed distribution of the variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) and variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) and variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) and variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) and variable cigarettes per day, non-parametric tests were used (Kruskal–Wallis test for construction) and variable cigarettes or to having states. 11/20 recAN and 24/105 HC reported currently smoking cigarettes or to having smoked cigarettes in the past. The mean age of onset of AN in the acAN sample was 14.6(2.8) years and the mean duration of illness uses 14(20) months. 71 acAN were of the restrictive subtype (87%) and 11 (13%) were of the binge/purge subtype. 12 (60%) recAN were of the restrictive subtype (187%) and 11 (13%) were of the binge/purge subtype. In the acAN group, 80/82 identified as White and 2/82 identified as Asian. In the recAN group, 103/105 identified as the test of the binge/purge subtype. In the acAN group, 103/105 identified as the acAN were of the binge/purge subtype. In the acAN group, 103/105 identified as Asian. In the recAN group, 20/20 identified as White. In the HC group, 103/105 identified as Asian. as White, 1/105 identified as African and 1/105 identified as Asian.

ac4N-TP1 acutely underweight participants with AN at timepoint 1 (admission), ac4N-TP2 participants with AN at timepoint 2 (after short-term weight restoration), HC healthy control participants, recAN individuals long-term weight-recovered from AN, BDHI Beck Depression Inventory-II, BMI-SDS body mass index-standard deviation score, EDI-2 Eating Disorder Inventory-2, SCL-90R Symptom Checklist-90-Revised.



Fig. 1 Serum IL-6 concentrations. Violin plots showing the median, upper and lower quartile, outliers (depicted as circles, values deviating more than 1.5 times the interquartile range from the upper or lower quartile) and the kernel probability density of the data. Serum IL-6 concentrations (pg/ml): HC: Median = 0.62, interquartile range = 0.70, range = <0.03 - 10.44; acAN-T1: Median = 0.51, interquartile range = 0.61, range = <0.03 to 9.82; acAN-T2: Median = 0.52, interquartile range = 0.63, range = 0.08-7.13; recAN: Median = 0.63, interquartile range = 0.63, range = 0.27-6.45. The figure was created with JASP [65]. acAN-TP1 acutely underweight participants with AN at timepoint 1 (admission), acAN-TP2 participants with AN at timepoint 2 (after short-term weight restoration), HC healthy control participants, recAN individuals long-term weight-recovered from AN.

Our null finding in the cross-sectional analysis of cytokines, i.e., no differences in concentrations between the groups, is partially consistent with the findings of prior studies. While two large metaanalyses indicated that patients with AN had elevated blood concentrations of IL-6 and TNF- α [1, 8], more recent studies presented heterogenous results [54–57]. For instance, a study on a large adult sample reported normal IL-6 concentrations in patients with AN compared to HC [54]. Keeler et al. reported normal TNF- α and reduced IL-6 concentrations in AN patients, while carefully controlling for potentially confounding variables, including, as in the current study age, smoking status and use of psychopharmacological medications [55]. Of note, one meta-analysis [1] highlighted the moderate to high levels of heterogeneity among previous studies, suggesting that variation in clinical characteristics such as age at onset, illness duration, or stage of treatment may be contributing factors. Additionally, they identified age as a factor influencing IL-6 concentrations [1]. This may be especially important because the participants in our study were younger than those included in the meta-analysis (mean age: our study sample: 16.8 years, Solmi et al. [8]: 21.7 years, Dalton et al. [1]: 21.4 years). Also, it has been speculated that in the early stages of AN, higher endogenous opioid release, which may positively reinforce fasting, could lead to a lower inflammatory response or even an immunosuppressive state [58]. This would be consistent with research suggesting that intermittent fasting leads to antiinflammatory effects as well as to a reduction of anxiety and depressive symptoms [59-61]. Our data suggests that young patients with a relatively short duration of illness (mean: 14 months) 5



Fig. 2 Serum TNF-a concentrations. Violin plots showing the median, upper and lower quartile, outliers (depicted as circles, values deviating more than 1.5 times the interquartile range from the upper or lower quartile) and the kernel probability density of the data. Serum TNF- α concentrations (pg/ml): HC: Median = 0.69, interquartile range = 0.23, range = 0.35–2.18; acAN-T1: Median = 0.63, interquartile range = 0.22, range = 0.29–2.75; acAN-T2: Median = 0.62, interquartile range = 0.28, range = 0.30–1.79; recAN: Median = 0.62, interquartile range = 0.32, range = 0.33–1.30. The figure was created with JASP [65]. acAN-T1 acutely underweight participants with AN at timepoint 1 (admission), acAN-T2 participants with AN at timepoint 2 (after short-term weight restoration), HC healthy control participants, recAN individuals long-term weight-recovered from AN.

may not show significant signs of systemic inflammation as assessed by serum TNF- α and IL-6. This is partly in line with a study in adolescents with AN, which found elevated TNF- α but reduced IL-6 concentrations [56]. However, a study in adolescents (mean age 14.9 years) with short illness duration (mean: 14 months) reported elevated TNF- α plasma concentrations [62]. In another young transdiagnostic sample including male and female patients with AN and other eating disorders (mean age 17.7 years for female participants), patients showed normal serum TNF- α levels and lower concentrations of IL-6 and the pro-inflammatory cytokine IL-1 beta [58].

Furthermore, recAN showed no group differences in cytokines when compared to HC, which is consistent with the findings from the largest study to date that included recovered patients [54], while another study reported reduced TNF- α in recovered individuals in comparison to HC but no group difference for IL-6 [55]. Our finding of no longitudinal change in cytokine concentrations over the course of short-term weight restoration is partly in line with previous findings: The meta-analysis from Solmi et al. [8] as well as a more recent study [14] found no significant difference in IL-6 concentrations before and after weight gain, while for TNF- α no difference [8] or a decrease from baseline to reassessment [14] were reported.

Additionally, our exploratory correlational analyses did not show any correlation between cytokine concentrations and indicators of undernutrition (BMI-SDS, leptin concentrations) or psychopathology for any of the groups. The heterogeneity of the published results emphasizes the complexity of possible inflammatory processes in AN and the need for further large-scale studies to identify potential patient subgroups prone to systemic inflammation [54].



Fig. 3 No significant associations between longitudinal changes in cortical structure (acAN-TP2-acAN-TP1) and changes in serum concentrations of the IL-6 cytokine. Left: Associations with CT changes. Right: Associations with IGI changes. Uncorrected statistical maps (p < 0.05) plotted on the inflated surface of the standard average subject and displaying regions in which differences in CT or IGI between acAN-TP2 and acAN-TP1 were associated with changes in IL-6. The color scale shows p values expressed as $-\log_{10}(p)$. Warm colors indicate a positive correlation between changes in IL-6 concentrations and CT or IGI. After FDR-correcting for multiple comparisons (q < 0.05) across each hemisphere and Bonferroni correcting across cytokine type, these associations were not significant. LH left hemisphere, RH right hemisphere. Colored outlines correspond to anatomical labels of the Desikan-Killiany atlas [44].



Fig. 4 No significant associations between longitudinal changes in cortical structure (acAN-TP2-acAN-TP1) and changes in serum concentrations of the TNF-a cytokine. Left: Associations with CT changes. Right: Associations with IGI changes. Uncorrected statistical maps (p < 0.05) plotted on the inflated surface of the standard average subject and displaying regions in which differences in CT or IGI between acAN-TP2 and acAN-TP1 were associated with changes in TNF- α . The color scale shows p values expressed as $-\log_{10}(p)$. Warm colors indicate a positive correlation between changes in TNF- α and CT or IGI. After FDR-correcting for multiple comparisons (q < 0.05) across each hemisphere and Bonferroni correcting across cytokine type, these associations were not significant. LH left hemisphere, RH right hemisphere. Colored outlines correspond to anatomical labels of the Desikan-Killiany atlas [44].

The absence of any association between measures of gray matter (GM) and serum concentrations of IL-6 or TNF- α in any of the groups may be attributed to the lack of peripheral inflammation in the sample. None of the study participants exhibited cytokine concentrations indicative of systemic inflammation, as all values were below established clinical thresholds. Therefore, it seems unlikely that the marked and widespread CT reductions observed in underweight adolescent and young females with AN are due to (sub)-inflammatory processes. Recently, higher serum concentrations of the neuronal damage marker NF-L were found to be associated with lower CT in several brain regions in a study sample of acAN-TP1 similar to ours, which implies axonal damage as a potential underlying mechanism of cortical thinning [33]. However, an exploratory analysis in part of this sample did not find a correlation between serum concentrations of NF-L and IL-6 or TNF-a (Supplementary Table S2), again rendering it unlikely that (sub)-inflammatory processes that can be captured with IL-6 or TNF-α explain GM decreases in acute AN. While various other mechanisms of CT reduction have been hypothesized (e.g., the loss, damage or remodeling of glia and/or neurons, see Introduction), a recent study using a virtual histology approach highlighted that the most vulnerable regions are those with a higher density of energetically demanding cells in adolescents and young female patients [18]. This further supports longitudinal findings showing a strong association between GM changes in AN and nutritional status. However, these findings were mostly obtained in adolescents and young female participants, and the recovery was slower in older participants [18]. Since also the heterogeneity of findings regarding cytokine concentrations in different eating disorder patient populations seems to be related to age effects [1], future studies should investigate whether sub-inflammatory processes might play a role in the long-term maintenance or worsening of CT reductions in (chronic) adult patients.

Some limitations should be considered. First, the cytokines IL-6 or TNF- α are the most thoroughly studied in AN, but other cytokines (e.g., IL-17A, IL-1 β , IL-10) may also play an important role in inflammatory processes in AN and should be included in future studies as well as brain cell damage markers such as NF-L and

GFAP. Furthermore, we only measured markers of peripheral inflammation and not central inflammatory markers, which would more accurately reflect neuroinflammation. Similarly, we have tested for associations with structural brain measures, but rapid inflammatory changes could impact neural function in ways that would change macroscopic brain structure. Second, our findings may be specific to young and non-chronic patients with AN. However, a sub-inflammatory state in adult chronic patients may also be related to deteriorating living conditions due to socioeconomic decline and increasing health burden rather than specifically to AN [63]. Multiple influencing factors on cytokine concentrations, such as smoking, age, psychiatric and somatic comorbidities and immunomodulating medication, were addressed in our inclusion criteria and additional sub-analyses. Furthermore, psychosocial stress, which we did not measure directly, may influence cytokine concentrations. Third, the relatively small number of patients with AN of the binge/purge subtype did not enable us to investigate the effect of AN subtype. Nonetheless, since our results did not change when excluding these participants, the impact of subtype seemed negligible in the current study. Finally, as all participants were all female and 98% identified as White, our findings cannot be generalized to all individuals with AN.

In conclusion, the present study provides no evidence for a link between measurable (sub-)inflammatory states (measured in peripheral blood) and the well-established widespread reductions of CT and gyrification in young patients with AN. Recent findings that brain regions with a higher density of energetically demanding cells have been found to be most affected by CT reduction [18] implicate nutritional status as an important factor in structural brain changes in acute AN. However, the present study cannot outrule neuroinflammation as a potential influence factor for structural brain changes. Future large-scale studies are needed to elucidate which other metabolic [18], hormonal [64] or cell-damaging processes [33] may also contribute to CT reduction in AN and should include further relevant parameters, such as MRI measures of neuroinflammation, measures of permeability of the blood-brain barrier or glymphatic measures. They should also explore differences between young individuals with a short duration of illness and adults with severe and enduring AN and include multiple longitudinal timepoints.

DATA AVAILABILITY

The datasets generated during and/or analyzed during the current study are available from the corresponding author on reasonable request.

REFERENCES

- Dalton B, Bartholdy S, Robinson L, Solmi M, Ibrahim MAA, Breen G, et al. A metaanalysis of cytokine concentrations in eating disorders. J Psychiatr Res. 2018; 103:252–64.
- Ehrlich S, Burghardt R, Schneider N, Broecker-Preuss M, Weiss D, Merle JV, et al. The role of leptin and cortisol in hyperactivity in patients with acute and weightrecovered anorexia nervosa. Prog Neuropsychopharmacol Biol Psychiatry. 2009;33:658–62.
- Hebebrand J, Exner C, Hebebrand K, Holtkamp C, Casper RC, Remschmidt H, et al. Hyperactivity in patients with anorexia nervosa and in semistarved rats: evidence for a pivotal role of hypoleptinemia. Physiol Behav. 2003;79:25–37.
- Dhopatkar N, Keeler JL, Mutwalli H, Whelan K, Treasure J, Himmerich H. Gastrointestinal symptoms, gut microbiome, probiotics and prebiotics in anorexia nervosa: a review of mechanistic rationale and clinical evidence. Psychoneuroendocrinology. 2023;147:105959.
- Jésus P, Ouelaa W, François M, Riachy L, Guérin C, Aziz M, et al. Alteration of intestinal barrier function during activity-based anorexia in mice. Clin Nutr Edinb Scotl. 2014;33:1046–53.
- 6. Herpertz-Dahlmann B, Seitz J, Baines J. Food matters: how the microbiome and gut-brain interaction might impact the development and course of anorexia nervosa. Eur Child Adolesc Psychiatry. 2017;26:1031–41.
- Seitz J, Dahmen B, Keller L, Herpertz-Dahlmann B. Gut feelings: how microbiota might impact the development and course of anorexia nervosa. Nutrients. 2020;12:E3295.

- Solmi M, Veronese N, Favaro A, Santonastaso P, Manzato E, Sergi G, et al. Inflammatory cytokines and anorexia nervosa: a meta-analysis of cross-sectional and longitudinal studies. Psychoneuroendocrinology. 2015;51:237–52.
- Brooks SJ. Targeting cytokines in the 5-Lox pro-inflammatory pathway for treatment-resistant anorexia nervosa. J Mol Genet Med Int J Biomed Res. 2018;1:4 https://doi.org/10.4172/1747-0862.1000376.
- Han MS, White A, Perry RJ, Camporez J-P, Hidalgo J, Shulman GI, et al. Regulation of adipose tissue inflammation by interleukin 6. Proc Natl Acad Sci USA. 2020;117:2751–60.
- 11. Almada C, Cataldo LR, Smalley SV, Diaz E, Serrano A, Hodgson MI, et al. Plasma levels of interleukin-6 and interleukin-18 after an acute physical exercise: relation with post-exercise energy intake in twins. J Physiol Biochem. 2013;69:85–95.
- Hunschede S, Schwartz A, Kubant R, Thomas SG, Anderson GH. The role of IL-6 in exercise-induced anorexia in normal-weight boys. Appl Physiol Nutr Metab. 2018;43:979–87.
- 13. de Baat A, Trinh B, Ellingsgaard H, Donath MY. Physiological role of cytokines in the regulation of mammalian metabolism. Trends Immunol. 2023;44:613–27.
- Dalton B, Leppanen J, Campbell IC, Chung R, Breen G, Schmidt U, et al. A longitudinal analysis of cytokines in anorexia nervosa. Brain Behav Immun. 2020;85:88–95.
- Bernardoni F, King JA, Geisler D, Birkenstock J, Tam Fl, Weidner K, et al. Nutritional status affects cortical folding: lessons learned from anorexia nervosa. Biol Psychiatry. 2018;84:692–701.
- Seitz J, Herpertz-Dahlmann B, Konrad K. Brain morphological changes in adolescent and adult patients with anorexia nervosa. J Neural Transm. 2016;123:949–59.
- 17. King JA, Frank GKW, Thompson PM, Ehrlich S. Structural neuroimaging of anorexia nervosa: future directions in the quest for mechanisms underlying dynamic alterations. Biol Psychiatry. 2018;83:224–34.
- Bahnsen K, Bernardoni F, King JA, Geisler D, Weidner K, Roessner V, et al. Dynamic structural brain changes in anorexia nervosa: a replication study, mega-analysis, and virtual histology approach. J Am Acad Child Adolesc Psychiatry. 2022;61:1168–81.
- Bernardoni F, King JA, Geisler D, Stein E, Jaite C, Nätsch D, et al. Weight restoration therapy rapidly reverses cortical thinning in anorexia nervosa: a longitudinal study. NeuroImage. 2016;130:214–22.
- Zilles K, Palomero-Gallagher N, Amunts K. Development of cortical folding during evolution and ontogeny. Trends Neurosci. 2013;36:275–84.
- Kim Y-K, Won E. The influence of stress on neuroinflammation and alterations in brain structure and function in major depressive disorder. Behav Brain Res. 2017;329:6–11.
- Tu P-C, Li C-T, Lin W-C, Chen M-H, Su T-P, Bai Y-M. Structural and functional correlates of serum soluble IL-6 receptor level in patients with bipolar disorder. J Affect Disord. 2017;219:172–7.
- Williams JA, Burgess S, Suckling J, Lalousis PA, Batool F, Griffiths SL, et al. Inflammation and brain structure in schizophrenia and other neuropsychiatric disorders: a Mendelian randomization study. JAMA Psychiatry. 2022;79:498–507.
- 24. Bai Y-M, Chen M-H, Hsu J-W, Huang K-L, Tu P-C, Chang W-C, et al. A comparison study of metabolic profiles, immunity, and brain gray matter volumes between patients with bipolar disorder and depressive disorder. J Neuroinflammation. 2020;17:42.
- Laskaris L, Mancuso S, Shannon Weickert C, Zalesky A, Chana G, Wannan C, et al. Brain morphology is differentially impacted by peripheral cytokines in schizophreniaspectrum disorder. Brain Behav Immun. 2021;95:299–309.
- Chen M-H, Kao Z-K, Chang W-C, Tu P-C, Hsu J-W, Huang K-L, et al. Increased proinflammatory cytokines, executive dysfunction, and reduced gray matter volumes in first-episode bipolar disorder and major depressive disorder. J Affect Disord. 2020;274:825–31.
- Licinio J, Wong ML. Pathways and mechanisms for cytokine signaling of the central nervous system. J Clin Invest. 1997;100:2941–7.
- Harrison NA, Cooper E, Dowell NG, Keramida G, Voon V, Critchley HD, et al. Quantitative magnetization transfer imaging as a biomarker for effects of systemic inflammation on the brain. Biol Psychiatry. 2015;78:49–57.
- Lesh TA, Careaga M, Rose DR, McAllister AK, Van de Water J, Carter CS, et al. Cytokine alterations in first-episode schizophrenia and bipolar disorder: relationships to brain structure and symptoms. J Neuroinflammation. 2018;15:165.
- Seitz J, Trinh S, Kogel V, Beyer C. Brain volume loss, astrocyte reduction, and inflammation in anorexia nervosa. Adv Neurobiol. 2021;26:283–313.
- Hellerhoff I, King JA, Tam FI, Pauligk S, Seidel M, Geisler D, et al. Differential longitudinal changes of neuronal and glial damage markers in anorexia nervosa after partial weight restoration. Transl Psychiatry. 2021;11:86.
- Nilsson IAK, Millischer V, Karrenbauer VD, Juréus A, Salehi AM, Norring C, et al. Plasma neurofilament light chain concentration is increased in anorexia nervosa. Transl Psychiatry. 2019;9:180.

- Hellerhoff I, Bernardoni F, Bahnsen K, King JA, Doose A, Pauligk S, et al. Serum neurofilament light concentrations are associated with cortical thinning in anorexia nervosa. Psychol Med. 2023;53:7053–61.
- 34. Smitka K, Papezova H, Vondra K, Hill M, Hainer V, Nedvidkova J. The role of "mixed" orexigenic and anorexigenic signals and autoantibodies reacting with appetite-regulating neuropeptides and peptides of the adipose tissue-gut-brain axis: relevance to food intake and nutritional status in patients with anorexia nervosa and bulimia nervosa. Int J Endocrinol. 2013;2013:1–21.
- Fichter M, Quadflieg N. Strukturiertes Inventar f
 ür anorektische und bulimische Esstörungen (SIAB); Fragebogen (SIAB-S) und Interview (SIAB-EX) nach DSM-IV und ICD-10; Handanweisung. Göttingen, Germany: Hogrefe; 1999.
- Thiel A, Jacobi C, Horstmann S, Paul T, Nutzinger DO, Schüssler G. Eine deutschsprachige Version des Eating Disorder Inventory EDI-2 [A German version of the Eating DisorderInventory EDI-2]. Psychother Psychosom Med Psychol. 1997;47:365–76.
- 37. Hautzinger M, Kühner C, Keller F. Beck Depressions-Inventar (BDI-II). Frankfurt, Germany: Pearson Assessment and Information GmbH; 2009.
- Franke G, Derogatis L. Symptom-Checkliste von LR Derogatis: SCL-90-R; Deutsche Version. Göttingen, Germany: Beltz Test; 2002.
- von Aster M, Neubauer A, Horn R. WIE—Wechsler Intelligenztest f
 ür Erwachsene. Bern, Switzerland: Huber; 2006.
- Petermann F, Petermann U. HAWIK IV. Hamburg-Wechsler-Intelligenztest f
 ür Kinder IV.
 Übersetzung und Adaptation der WISC-IV von David Wechsler. G
 öttingen, Germany: Hogrefe; 2010.
- Kromeyer-Hauschild K, Wabitsch M, Kunze D, Geller F, Geiß HC, Hesse V, et al. Perzentile f
 ür den Body-mass-Index f
 ür das Kindes- und Jugendalter unter Heranziehung verschiedener deutscher Stichproben. Monatsschr Kinderheilkd. 2001;149:807–18.
- Wei R, Wang J, Jia E, Chen T, Ni Y, Jia W. GSimp: a Gibbs sampler based leftcensored missing value imputation approach for metabolomics studies. PLoS Comput Biol. 2018;14:e1005973.
- King JA, Geisler D, Ritschel F, Boehm I, Seidel M, Roschinski B, et al. Global cortical thinning in acute anorexia nervosa normalizes following long-term weight restoration. Biol Psychiatry. 2015;77:624–32.
- 44. Desikan RS, Ségonne F, Fischl B, Quinn BT, Dickerson BC, Blacker D, et al. An automated labeling system for subdividing the human cerebral cortex on MRI scans into gyral based regions of interest. NeuroImage. 2006;31:968–80.
- 45. Fischl B, Salat DH, Busa E, Albert M, Dieterich M, Haselgrove C, et al. Whole brain segmentation. Neuron. 2002;33:341–55.
- 46. Reuter M, Schmansky NJ, Rosas HD, Fischl B. Within-subject template estimation for unbiased longitudinal image analysis. NeuroImage. 2012;61:1402–18.
- 47. Fischl B, Dale AM. Measuring the thickness of the human cerebral cortex from magnetic resonance images. Proc Natl Acad Sci USA. 2000;97:11050–5.
- Schaer M, Cuadra MB, Tamarit L, Lazeyras F, Eliez S, Thiran J-P. A surface-based approach to quantify local cortical gyrification. IEEE Trans Med Imaging. 2008;27:161–70.
- Bernal-Rusiel JL, Greve DN, Reuter M, Fischl B, Sabuncu MR. Statistical analysis of longitudinal neuroimage data with linear mixed effects models. NeuroImage. 2013;66:249–60.
- Tamnes CK, Herting MM, Goddings A-L, Meuwese R, Blakemore S-J, Dahl RE, et al. Development of the cerebral cortex across adolescence: a multisample study of inter-related longitudinal changes in cortical volume, surface area, and thickness. J Neurosci. 2017;37:3402–12.
- Wang H, Chen H, Fu Y, Liu M, Zhang J, Han S, et al. Effects of smoking on inflammatory-related cytokine levels in human serum. Molecules. 2022;27:3715.
- 52. He H, Wang W, Crits-Christoph P, Gallop R, Tang W, Chen D-G, et al. On the implication of structural zeros as independent variables in regression analysis: applications to alcohol research. J Data Sci. 2014;12:439–60.
- 53. R Core Team. R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing; 2021.
- Nilsson IAK, Millischer V, Göteson A, Hübel C, Thornton LM, Bulik CM, et al. Aberrant inflammatory profile in acute but not recovered anorexia nervosa. Brain Behav Immun. 2020;88:718–24.
- 55. Keeler JL, Patsalos O, Chung R, Schmidt U, Breen G, Treasure J, et al. Reduced MIP- 1β as a trait marker and reduced IL-7 and IL-12 as state markers of anorexia nervosa. J Pers Med. 2021;11:814.
- 56. Specht HE, Mannig N, Belheouane M, Andreani NA, Tenbrock K, Biemann R, et al. Lower serum levels of IL-1 β and IL-6 cytokines in adolescents with anorexia nervosa and their association with gut microbiota in a longitudinal study. Front Psychiatry. 2022;13:920665.
- 57. Breithaupt L, Holsen LM, Ji C, Hu J, Petterway F, Rosa-Caldwell M, et al. Identification of state markers in anorexia nervosa: replication and extension of inflammation associated biomarkers using multiplex profiling in anorexia nervosa and atypical anorexia nervosa. bioRxiv. 2023. https://doi.org/10.1101/2023.06.30.547289.

- 58. Ruiz Guerrero F, González Gómez J, Benito Gonzalez P, García García J, Berja Miguel A, Calcedo Giraldo G, et al. Low levels of proinflammatory cytokines in a transdiagnostic sample of young male and female early onset eating disorders without any previous treatment: a case control study. Psychiatry Res. 2022;310:114449.
- Malinowski B, Zalewska K, Węsierska A, Sokołowska MM, Socha M, Liczner G, et al. Intermittent fasting in cardiovascular disorders—an overview. Nutrients. 2019;11:673.
- Dwaib HS, AlZaim I, Eid AH, Obeid O, El-Yazbi AF. Modulatory effect of intermittent fasting on adipose tissue inflammation: amelioration of cardiovascular dysfunction in early metabolic impairment. Front Pharm. 2021;12:626313.
- Berthelot E, Etchecopar-Etchart D, Thellier D, Lancon C, Boyer L, Fond G. Fasting interventions for stress, anxiety and depressive symptoms: a systematic review and meta-analysis. Nutrients. 2021;13:3947.
- Caso JR, Graell M, Navalón A, MacDowell KS, Gutiérrez S, Soto M, et al. Dysfunction of inflammatory pathways in adolescent female patients with anorexia nervosa. Prog Neuropsychopharmacol Biol Psychiatry. 2020;96:109727.
- 63. Friedman E, Shorey C. Inflammation in multimorbidity and disability: an integrative review. Health Psychol. 2019;38:791–801.
- 64. Wronski M-L, Geisler D, Bernardoni F, Seidel M, Bahnsen K, Doose A, et al. Differential alterations of amygdala nuclei volumes in acutely ill patients with anorexia nervosa and their associations with leptin levels. Psychol Med. 2023;53:6288–6303.
- 65. JASP Team (2019). JASP (Version 0.10) [Computer software].

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AUTHOR CONTRIBUTIONS

Conceptualization: FT, DP, and SE. Methodology: FB, FT, and SE. Investigation: FB, FT, IH, DA, DG, FL, and JK. Visualization: FB and FT. Funding acquisition: FB, FT, and SE. Project administration: SE. Supervision: KW, CP, VR, JK, and SE. Writing—original draft: FB, FT, JK, and SE. Writing—review and editing: FB, FT, DP, IH, DA, DG, FL, JK, KW, CP, VR, JK, and SE. Software: FB, DA, and DG. Formal analysis: FB and FT. Resources: KW, VR, and SE. Data curation: FB, DA, DG, and SE.

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