

Noninvasive measurement of intestinal epithelial damage at time of refeeding can predict clinical outcome after necrotizing enterocolitis

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BACKGROUND: Reintroduction of enteral nutrition in neonates with necrotizing enterocolitis (NEC) should take place when the gut is ready for its normal function. Too early a start of oral feeding might lead to disease relapse, whereas prolonged discontinuation of enteral nutrition is associated with impaired gut function and parenteral nutrition-related complications. This study evaluated whether noninvasive urinary measurement of intestinal fatty acid binding protein (I-FABP) at the time of refeeding can predict clinical outcome in neonates with NEC.

METHODS: Urinary I-FABP concentrations were measured in 21 infants with NEC just before reintroducing enteral nutrition. Poor outcome was defined as unsuccessful reintroduction of enteral feeding (EF), (re)operation for NEC, or death related to NEC after reintroduction of EF.

RESULTS: Median urinary I-FABP levels in neonates with poor outcome ($n = 5$) were significantly higher as compared with I-FABP levels in neonates with good outcome ($n = 16$) ($P < 0.01$). A clinically significant cutoff value of 963 pg/ml was found to discriminate between infants with poor outcome and those with good outcome (sensitivity 80%, specificity 94%).

CONCLUSION: Noninvasive urinary I-FABP measurement at time of refeeding differentiates neonates with poor outcome from neonates with good outcome in NEC. Urinary I-FABP measurement may therefore be helpful in the timing of EF in neonates with NEC.

Necrotizing enterocolitis (NEC) is a severe gastrointestinal disease with high morbidity and mortality (20–40%), affecting predominantly premature neonates (1). The treatment of choice for infants suffering from mild to moderate NEC includes cardiorespiratory support if necessary, broad-spectrum antibiotics, and discontinuation of enteral feeding (EF) (1–3). Surgical intervention is considered in patients with a perforation or advanced NEC that does not improve with conservative treatment (2,4).

After the initial treatment of NEC, there is no common opinion regarding the time when EF can be reintroduced. There are

no clinical, radiological, or laboratory parameters to determine when refeeding of the infant is safe. Refeeding at a very early stage has been suggested to lead to recurrence of the disease (5), whereas prolonged discontinuation of EF can worsen gut function (6) and impair growth. Moreover, prolonged use of parenteral nutrition is associated with catheter-related septicemia and longer duration of hospital stay (7).

In a recent study, we demonstrated that urinary intestinal fatty acid binding protein (I-FABP) is a suitable noninvasive marker to confirm the diagnosis of NEC in patients with early symptoms of NEC (8). I-FABP, a small (14–15 kDa) cytosolic protein, is solely present in mature epithelial cells of the small and large intestine (9). It is immediately released into the circulation upon loss of cell membrane integrity and passes the glomerular filter with a fractional renal excretion of 28% and a half-life of 11 min, which makes it detectable in the urine (10). Therefore, urinary I-FABP levels accurately reflect the actual extent of intestinal epithelial cell damage, which makes I-FABP an indicator of disease progression (8). Although initially elevated, urinary I-FABP levels decrease when disease severity diminishes (11). We therefore hypothesized that I-FABP levels may be useful to determine the ideal time for reintroducing EF.

In this study, we measured urinary I-FABP concentrations in infants who were previously treated for NEC at the time oral feeding was introduced and evaluated the outcome of these children.

RESULTS

Patients and Outcome

Twenty-one neonates were included in the study, of whom 12 were males. Patient characteristics are listed in **Table 1**. There were no significant differences between groups in gestational age, birth weight, sex, or in blood laboratory tests (C-reactive protein, leukocytes, and platelets) at time of refeeding (**Table 1**).

Five of the 21 infants with NEC had a poor clinical outcome, and reintroduction of EF was unsuccessful (total parenteral nutrition was restarted). Four of these five received an operation

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Table 1. Baseline characteristics and conventional laboratory tests

	Poor outcome	Good outcome	Total	P
Gestational age (days)	207 (191–268)	224 (184–242)	224 (184–268)	0.55
Birth weight (g)	1,300 (1,185–2,960)	1,463 (860–2,106)	1,460 (860–2,960)	0.78
Sex	3M 2F	9M 7F	12M 9F	1.00
Surgery before refeeding	3/5 (60%)	6/16 (37.5%)	9/21 (42.9%)	0.61
CRP (mg/l)	1.8 (1.0–5.8)	40 (1.0–93)	5.8 (1.0–93)	0.11
Leukocytes (×10 ⁹ cells/l)	14.5 (10.3–16.5)	10.1 (8.0–20.1)	10.4 (8.0–20.1)	0.16
Platelets (×10 ⁹ cells/l)	309 (96–892)	120 (29–296)	191.5 (29–892)	0.11

Data are presented as median (range).
CRP, C-reactive protein; F, female; M, male.

or reoperation on NEC within 30 d after reintroduction of EF, and the fifth child died within 30 d after reintroduction of oral feeding (Table 2). Three of the four infants operated for NEC after refeeding had been operated on earlier, 0–2 d after the initial diagnosis. In all four cases, NEC was the indication for operation. This was confirmed by histopathology. The other 16 neonates recovered from NEC after EF was restarted, without the need for surgery (group with a good clinical outcome).

The median period of time between birth and NEC diagnosis was 9 (4–30) d in the good outcome group and 5 (2–18) d in the poor outcome group, *P* = 0.28. The median period of time after NEC diagnosis that infants received no feeding by mouth was 9 (6–19) d in the group with good outcome and 10 (6–14) d in the group with poor outcome, *P* = 0.50. In the poor outcome group, the period of refeeding lasted for 8 (3–34) d. These results are shown in Table 3. The median period of time until full EF was achieved was 9 d (no difference between groups).

High Urinary I-FABP Levels at Time of Refeeding Correlates With Poor Outcome

Median urinary I-FABP:creatinine ratio (I-FABP corrected for urine concentration) just before reintroduction of EF was significantly higher in neonates with poor outcome (6.00 (0.53–247.10) pg/nmol creatinine) as compared with neonates with good outcome (0.38 (0.03–1.41) pg/nmol creatinine), *P* < 0.01. To distinguish neonates with poor outcome from neonates with good outcome, an ideal cutoff value of 2.23 pg/nmol creatinine was found with a sensitivity of 80% and a specificity of 100%. This resulted in a positive likelihood ratio (LR) of >12.8 and a negative LR of 0.2.

Similar data were obtained when I-FABP was not corrected for urine concentration (Figure 1). This analysis led to an ideal cutoff value of 963 pg/ml, with a sensitivity of 80% and a specificity of 94%, and a positive LR of 12.8 and a negative LR of 0.21. The area under the curve for urinary I-FABP in

Table 2. Specification of patients with poor clinical outcome

	Discontinuation of enteral feeding after refeeding	Operation after refeeding	Death after refeeding
Patient A	Yes	Yes ^a	No
Patient B	Yes	No	Yes ^b
Patient C	Yes	Yes	Yes ^c
Patient D	Yes	Yes ^a	Yes ^d
Patient E	Yes	Yes ^a	No

Columns are ordered from left to right by increasing severity of adverse outcome. NEC, necrotizing enterocolitis.

^aSecond operation for NEC. Active NEC was found in all cases. ^bDied within 30 d after diagnosis of NEC. ^cDied 71 d after diagnosis of NEC; cause of death: persistent neonatal seizures. ^dDied 162 d after diagnosis of NEC; cause of death: treatment discontinued because of severe neurological damage.

Table 3. Time course

	Poor outcome	Good outcome	Total	P
Diagnosis at birth (days)	5 (2–18)	9 (4–30)	9 (2–30)	0.28
Diagnosis after refeeding (days)	10 (6–14)	9 (6–19)	9 (6–19)	0.50
Refeeding after the second discontinuation of enteral feeding (days)	8 (3–34)			

Data are presented as median (range).

differentiating neonates with poor outcome from neonates with good outcome was 0.93 (95% confidence interval 0.78–1.00) for I-FABP corrected for urine concentration and 0.95 (95% confidence interval 0.86–1.00) for I-FABP not corrected for urine concentration. Median urinary I-FABP for 20 premature neonates without gastrointestinal symptoms (matched controls) was 358 (174–924) pg/ml (Figure 1).

Relative Difference in Urinary I-FABP Levels From Diagnosis to Time of Refeeding as a Predictor of Poor Outcome

Another approach to predicting the outcome of reintroduction of EF would be to measure the relative reduction of intestinal damage as reflected by the differences in I-FABP levels from the moment of diagnosis to the moment of oral refeeding. The relative difference (ΔRel) in urinary I-FABP from diagnosis (Diag) to reintroduction of EF was calculated as follows:

$$\Delta Rel_{Diag-EF} = ((I-FABP)_{EF} - (I-FABP)_{Diag}) / (I-FABP)_{Diag} \times 100\%$$

where (I-FABP) is the concentration of I-FABP (not corrected for urine concentration) (pg/ml). ΔRel_{Diag-EF} in neonates with good outcome (–100% (–100% to –22%)) was significantly lower than ΔRel_{Diag-EF} in neonates with poor outcome (–66% (–87% to +113%)), *P* < 0.05 (Figure 2). To distinguish neonates with poor outcome from neonates with good outcome, an ideal cutoff value of –90% was found with a sensitivity of 100% and a specificity of 75%. This resulted in a positive LR of 4.00 and a negative LR of 0.00. The area under the curve

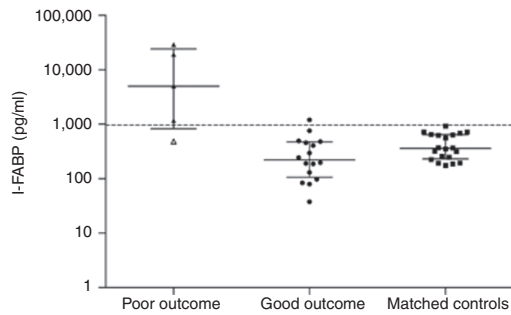


Figure 1. Urinary intestinal fatty acid binding protein (I-FABP) levels of infants on the day of reintroduction of enteral feeding, median 9 d after diagnosis of necrotizing enterocolitis. Data are expressed as median I-FABP levels with interquartile ranges. A cutoff value of 963 pg/ml is marked by the dotted line. I-FABP levels of controls matched for gestational age are depicted. Black triangles, infants operated on after refeeding; black circles, infants not operated on after refeeding; Δ symbol, died after refeeding; black squares, matched controls.

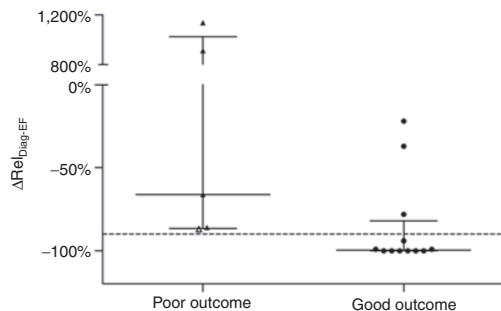


Figure 2. Relative differences with median and interquartile ranges of urinary intestinal fatty acid binding protein from diagnosis to reintroduction of enteral feeding ($\Delta\text{Rel}_{\text{Diag-EF}}$). A cutoff value of -90% is marked by the dotted line. Black triangles, infants operated on after refeeding; black circles, infants not operated on after refeeding; Δ symbol, died after refeeding.

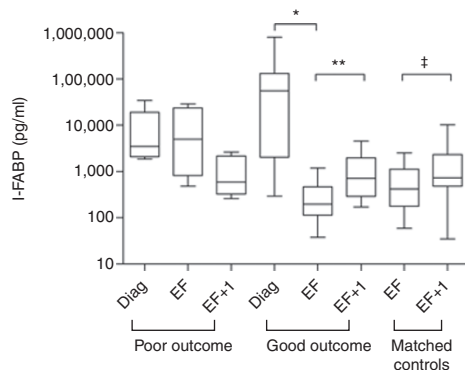


Figure 3. Urinary intestinal fatty acid binding protein levels on the day of diagnosis (Diag), the day of reintroduction of enteral feeding (EF), and 1 d after reintroduction of EF. * $P = 0.002$; ** $P = 0.04$; † $P = 0.03$. I-FABP, intestinal fatty acid binding protein.

for $\Delta\text{Rel}_{\text{Diag-EF}}$ in differentiating neonates with poor outcome from neonates with good outcome was 0.87 (95% confidence interval 0.69–1.00) (receiver operating characteristic curve not shown). Actual urinary I-FABP levels at the time of diagnosis are depicted in **Figure 3**. There was no significant difference

in urinary I-FABP levels at the time of diagnosis between the groups. In conclusion, the former analysis based on actual values at the moment of refeeding produced better data.

Kinetics of I-FABP Following Reintroduction of EF

In the analyses, both I-FABP corrected and uncorrected for urine concentration were used. Next, we analyzed whether differences could be detected between neonates with good clinical outcome and those with poor clinical outcome early during feeding. To this end, we compared data at EF with data on EF+1. Analysis of the data revealed a significant increase in urinary I-FABP from EF (0.38 (0.03–1.41) pg/nmol creatinine) to EF+1 (1.41 (0.53–2.74) pg/nmol creatinine, $P < 0.05$) in the neonates with good outcome. In contrast, neonates with poor outcome showed no increase from EF to EF+1.

Similar data were obtained in the case in which I-FABP was not corrected for urine concentration; a significant increase in urinary I-FABP from EF (197 (37.7–1,196) pg/ml) to EF+1 (712 (170–4,537) pg/ml, $P = 0.04$) was observed in the group with good outcome (**Figure 3**). This increase (EF, 421 (59–2,553); EF+1, 733 (35–10,278), $P = 0.03$, **Figure 3**) was also observed in the control group of nine infants without intestinal injury in whom feeding was discontinued and reintroduced, indicating a physiological rather than a pathological response.

DISCUSSION

One of the challenges in the treatment of the infant with NEC is to choose the best moment to reintroduce EF. Although no benefit has been shown for delayed (12) or slow (13) introduction of EF in the prevention of NEC, evidence is lacking on the length of the period between the treatment of NEC and the moment at which EF can be restarted safely. Although a period of 7–10 d of discontinuation of EF has been recommended (14), faster reintroduction after NEC seems feasible in some cases. In a study by Bohnhorst *et al.* (7), EF was restarted as early as 4 d after diagnosis of NEC when portal vein gas was absent on ultrasound. These infants showed less catheter-related sepsis and a shorter hospital stay as compared with historical controls in which EF was reinitiated on day 10 (7). Moreover, return to EF may reduce the development of villus atrophy (15). We consider that daily measurement of urinary I-FABP could help to identify infants who can be safely returned to oral feeding.

The results of this study show that urinary I-FABP is a suitable noninvasive marker to determine which infants with NEC in whom EF is restarted are at risk for a poor outcome, in terms of mortality or reoperation. In addition to clinical assessment, urinary I-FABP measurement can therefore be a helpful tool for the neonatologist in deciding which infants can be refed safely in the course of NEC, especially when clinical improvement is doubted. At the moment, no laboratory markers exist to confirm this decision. In the population studied, a cutoff value at the time of oral refeeding of 963 pg/ml could discriminate neonates with poor outcome from those with good outcome, with a sensitivity of 80% and a specificity of 94%. Whether clinical deterioration

could have been avoided by delaying reintroduction of EF, or whether clinical deterioration can directly be caused by onset of refeeding cannot be determined by these data. Further study in a large number of patients is needed to evaluate the cutoff value of 963 pg/ml and its clinical usefulness.

Remarkably, I-FABP levels increased early during feeding in the group with good outcome and in a control group without NEC or other severe intestinal injury. This may be a result of a physiological response of the gut when challenged by EF. In a piglet model, rapid enterocyte turnover and mucosal structural changes were observed 8 h after introduction of EF (16). Unexpectedly, I-FABP levels seemed to decrease early during refeeding in the group with poor outcome; however, the difference was not statistically significant. This could be a consequence of the reduction of enterocyte mass as described earlier in severe mucositis induced by myeloablative conditioning (17). However, the size of this group is relatively small, which is an inevitable problem, given that many of the children who die of NEC die shortly after NEC diagnosis without the possibility to restart EF.

Conclusion

In conclusion, when I-FABP levels were >963 pg/ml following clinical progression that resulted in reintroducing EF, there was a high chance of clinical deterioration in this NEC population. Urinary I-FABP measurement may therefore be helpful in the timing of EF in neonates with NEC. However, this hypothesis has to be validated in a prospective trial, with different treatment strategies determined on the basis of urinary I-FABP measurement that serves as an addition to clinical judgment.

METHODS

Patients, Feeding Protocol, and Sample Collection

Twenty-one consecutive infants, in whom EF was reintroduced after treatment for NEC in the neonatal intensive care units at Maastricht University Medical Centre and Wilhelmina Children's Hospital in Utrecht, between July 2005 and August 2010, were included in the study. NEC was diagnosed with the current gold standard of abdominal X-ray showing pneumatosis intestinalis (Bell stage II or higher) (2). Twenty consecutive premature infants without gastrointestinal symptoms (matched with NEC infants for gestational age at birth) admitted to the neonatal intensive care unit at Maastricht University Medical Centre were also included for the evaluation of control values. Indication for admission into the neonatal intensive care unit was small for gestational age or prematurity in all cases. In these premature neonates serving as controls, a urine sample was collected. A second control group of nine neonates without intestinal injury was included, in whom EF was stopped and restarted. Indications for cessation of EFs were suspicion of NEC (eventually diagnosed as sepsis) or laparotomy for malformation or atresia without significant intestinal epithelial cell damage. Written consent was obtained from both parents, and the study was conducted with the approval of the local medical ethical committees and according to the revised version of the Declaration of Helsinki (October 2008, Seoul). The principles of good clinical practice were followed during this study.

Feeding was restarted in patients who improved clinically: spontaneously breathing patients without intestinal pneumatosis on abdominal X-ray, without abdominal distention, and with peristalsis and a soft abdomen. This decision was made by the attending neonatologist, who was independent of the investigators. EF consisted of formula feeding, breast milk, or a combination of both and was increased with a maximum of 20 ml/kg/day.

Urine was obtained daily by placing a dental cotton roll (Henry Schein, Almere, The Netherlands) in the diaper of the neonate. Once the roll was filled with urine, it was placed into a sterile 5-ml syringe (Becton Dickinson, Oxford, UK), and the urine was pressed into Micronic tubes (Micronic B.V., Lelystad, The Netherlands) and stored at -20°C until analysis. Urine samples were collected at the time of diagnosis (Diag), just before reintroduction of EF, and the day after reintroduction of EF (EF+1).

All patient materials were collected in a prospective manner; however, the analyses were performed by one person who was blinded to clinical diagnoses after completion of patient inclusion. Attending clinicians were therefore not aware of I-FABP levels when deciding to restart EF.

Urinary I-FABP Measurement

Urinary I-FABP was measured using an in-house enzyme-linked immunosorbent assay that selectively detects human I-FABP (standard: 10–1,280 pg/ml). Values are expressed as concentration (pg/ml) and ratio (pg/nmol creatinine) of I-FABP (pg/ml) to creatinine ($\mu\text{mol/l}$), to compensate for variations in urine concentration.

Statistical Analyses

Normality was tested by the Kolmogorov–Smirnov test. The Mann–Whitney *U* test was used for between-groups comparison for continuous data. Dichotomous variables were compared using Fisher's exact test. All data are presented as median and range. Outcome was defined as poor clinical outcome (unsuccessful introduction of EF (i.e., restarting total parenteral nutrition), (re)operation, or death within 30 d after diagnosis of NEC) or good clinical outcome. To find a cutoff point of urinary I-FABP levels at the time of refeeding that most accurately predicts outcome, a receiver operating characteristic curve was drawn by plotting sensitivity against 1-specificity for all possible thresholds. Overall accuracy of urinary I-FABP to predict outcome was represented by the area under the curve. The best cutoff point was defined as the cutoff point with the maximum sum of sensitivity and specificity, after taking the range of normal values into account. Statistical analyses were performed with Prism 5.0 for Windows (GraphPad Software, San Diego, CA). STARD statement for reporting studies of diagnostic accuracy was used in this study (18).

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REFERENCES

1. Lin PW, Stoll BJ. Necrotizing enterocolitis. *Lancet* 2006;368:1271–83.
2. Bell MJ, Ternberg JL, Feigin RD, et al. Neonatal necrotizing enterocolitis. Therapeutic decisions based upon clinical staging. *Ann Surg* 1978;187:1–7.
3. Neu J. Neonatal necrotizing enterocolitis: an update. *Acta Paediatr Suppl* 2005;94:100–5.
4. Lee JS, Polin RA. Treatment and prevention of necrotizing enterocolitis. *Semin Neonatol* 2003;8:449–59.
5. Berseth CL. Feeding strategies and necrotizing enterocolitis. *Curr Opin Pediatr* 2005;17:170–3.
6. Lin PW, Nasr TR, Stoll BJ. Necrotizing enterocolitis: recent scientific advances in pathophysiology and prevention. *Semin Perinatol* 2008;32:70–82.
7. Bohnhorst B, Müller S, Dördelmann M, Peter CS, Petersen C, Poets CF. Early feeding after necrotizing enterocolitis in preterm infants. *J Pediatr* 2003;143:484–7.
8. Thuijls G, Derikx JP, van Wijck K, et al. Non-invasive markers for early diagnosis and determination of the severity of necrotizing enterocolitis. *Ann Surg* 2010;251:1174–80.
9. Veerkamp JH, Paulussen RJ, Peeters RA, Maatman RG, van Moerkerk HT, van Kuppevelt TH. Detection, tissue distribution and (sub)cellular localization of fatty acid-binding protein types. *Mol Cell Biochem* 1990;98:11–8.
10. van de Poll MC, Derikx JP, Buurman WA, et al. Liver manipulation causes hepatocyte injury and precedes systemic inflammation in patients undergoing liver resection. *World J Surg* 2007;31:2033–8.

11. Evennett NJ, Hall NJ, Pierro A, Eaton S. Urinary intestinal fatty acid-binding protein concentration predicts extent of disease in necrotizing enterocolitis. *J Pediatr Surg* 2010;45:735–40.
12. Morgan J, Young L, McGuire W. Delayed introduction of progressive enteral feeds to prevent necrotising enterocolitis in very low birth weight infants. *Cochrane Database Syst Rev* 2011:CD001970.
13. Morgan J, Young L, McGuire W. Slow advancement of enteral feed volumes to prevent necrotising enterocolitis in very low birth weight infants. *Cochrane Database Syst Rev* 2011:CD001241.
14. Neu J, Walker WA. Necrotizing enterocolitis. *N Engl J Med* 2011;364:255–64.
15. Kliegman RM, Walker WA, Yolken RH. Necrotizing enterocolitis: research agenda for a disease of unknown etiology and pathogenesis. *Pediatr Res* 1993;34:701–8.
16. Oste M, Van Haver E, Thymann T, Sangild P, Weyns A, Van Ginneken CJ. Formula induces intestinal apoptosis in preterm pigs within a few hours of feeding. *JPEN J Parenter Enteral Nutr* 2010;34:271–9.
17. Derikx JP, Blijlevens NM, Donnelly JP, et al. Loss of enterocyte mass is accompanied by diminished turnover of enterocytes after myeloablative therapy in haematopoietic stem-cell transplant recipients. *Ann Oncol* 2009;20:337–42.
18. Bossuyt PM, Reitsma JB, Bruns DE, et al. The STARD statement for reporting studies of diagnostic accuracy: explanation and elaboration. *Clin Chem* 2003;49:7–18.