# Fetal Growth Restriction Is Associated With Prioritization of Umbilical Blood Flow to the Left Hepatic Lobe at the Expense of the Right Lobe

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ABSTRACT: Eighty to 85% of the venous perfusion to the fetal liver is from the umbilical vein, the rest from the portal vein. Umbilical venous flow to the liver is essential for intrauterine growth, and is impaired in placental insufficiency. We hypothesized that in growth-restricted fetuses portal blood flow compensates for insufficient umbilical blood flow to the liver. In 29 fetuses with fetal growth restriction (estimated fetal weight  $\leq$ 5th percentile), we used ultrasound to measure blood flows in the umbilical vein, ductus venosus, left portal vein, and main portal stem. Compared with normal fetuses, both absolute and normalized total venous liver blood flows were reduced in growth-restricted fetuses, related to the degree of placental compromise and equally affecting both liver lobes. However, portal replaced umbilical flow to the right lobe, in a manner graded according to placental vascular resistance; in extreme cases, the right lobe received no umbilical perfusion. In fetal growth restriction, the liver suffers from venous hypoperfusion, and portal blood partially replaces umbilical flow to the right lobe; this will result in right liver lobe hypoxemia. This striking prioritization in nutrient delivery of left over right lobes suggests an adaptive response to poor placental perfusion that may have functional consequences. (Pediatr Res 66: 113-117, 2009)

**P**lacental insufficiency is a major cause of impaired fetal growth and premature birth, which, in turn, is associated with an increased risk of chronic disease in later life (1). Decreased placental function forces the fetus to redistribute its circulation to maintain oxygen and nutrient supply to the heart and brain, but these responses that are beneficial in the short term may have deleterious long-term effects. One of these adaptive changes is increased shunting of nutrient-rich umbilical venous blood through the ductus venosus thereby decreasing the amount of umbilical venous blood perfusing the liver (2,3). Experimental data indicate a direct relationship between venous liver perfusion and fetal organ growth (4). Thus, the study of fetal liver flow may further elucidate the pathogenesis of fetal growth restriction (FGR).

The venous perfusion of the fetal liver differs from that after birth in that, venous blood flows to it both from the splanchnic bed *via* the portal vein and from the placenta *via* the umbilical

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vein (Fig. 4*A*). Although the latter has been studied extensively, the role of portal perfusion during placental compromise is unknown. Therefore, we developed a technique for blood flow measurement in the main portal stem and established reference ranges for this flow, showing that in uncompromised fetuses 15–20% of the venous liver perfusion is of splanchnic origin (5).

In this study, we have gone on to characterize hepatic venous perfusion in FGR and to test the hypothesis that portal blood flow may compensate for insufficient umbilical blood flow to the liver when the latter is diverted through the ductus venosus.

## MATERIALS AND METHODS

In a cross-sectional study, we examined 31 growth-restricted (*i.e.*, estimated fetal weight  $\leq$ 5th percentile) singleton fetuses without malformations or chromosomal abnormalities. Gestational age was determined by ultrasound in the second trimester (6). The study protocol was approved

 Table 1. Characterization of the study population, data given as number (N) or mean with SD in brackets

	Normal UA PI (N = $16$ )	Increased UA PI (N = 13)	
Preeclampsia (N)	3	6	
Hypertension (N)	0	2	
Gestational diabetes (N)	0	2	
Gestational age at			
Investigation (wks)	34.2 (3.5)	31.4 (4.4)	
Delivery (wks)	37.3 (3.5)	32.0 (4.5)	
Time span examination	22.1 (18.6)	4.3 (4.0)	
delivery (d)			
Delivery mode			
Vaginal delivery (N)	7	0	
Cesarean section (N)	9	13	
Birth weight (g)	2200 (591)	1263 (633)	
Cord acid base data			
Umbilical artery (N)	9	6	
pH	7.29 (0.06)	7.26 (0.05)	
Base deficit (mmol/L)	-0.38 (2.84)	1.18 (1.95)	
Po <sub>2</sub> (kPa)	1.83 (0.66)	1.24 (0.34)	
Umbilical vein (N)	10	8	
pH	7.34 (0.04)	7.30 (0.04)	
Base deficit (mmol/L)	-0.56 (3.14)	-0.20 (1.27)	

Abbreviation: FGR, fetal growth restriction

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**Table 2.** Liver flow parameters in FGR compared with reference population, expressed as z score

		95%				
	Mean	Std. error	Confidence	Interval	Ν	value
$Q_{\rm Liver}$						
Reference	0.00	0.04	-0.09	0.08	515	< 0.0001
FGR, normal UA Doppler	-1.48	0.25	-1.98	-0.99	16	
FGR, abnormal UA Doppler	-2.80	0.33	-3.46	-2.15	9	
$Q_{\text{Liver}}$ (kg)						
Reference	0.01	0.04	-0.08	0.10	518	< 0.0001
FGR, normal UA Doppler	-0.60	0.25	-1.09	-0.11	16	
FGR, abnormal UA Doppler	-1.49	0.33	-2.15	-0.84	9	
Q <sub>Left liver</sub>						
Reference	0.00	0.04	-0.09	0.09	498	< 0.0001
FGR, normal UA Doppler	-1.06	0.25	-1.54	-0.57	16	
FGR, abnormal UA Doppler	-1.47	0.33	-2.12	-0.81	10	
Q <sub>Right liver</sub>						
Reference	-0.01	0.04	-0.09	0.08	519	< 0.0001
FGR, normal UA Doppler	-1.61	0.25	-2.09	-1.12	16	
FGR, abnormal UA Doppler	-3.00	0.38	-3.74	-2.27	7	
$\frac{Q_{\text{Portal vein}}}{Q_{\text{portal vein}}}$ [%]	2.00					
21000000000000000000000000000000000000						
	0.00	0.04	0.16	0.01	517	<0.0001
Reference	-0.08	0.04	-0.16	0.01	517	< 0.0001
FGR, normal UA Doppler	0.60	0.25	0.11	1.09	16	
FGR, abnormal UA Doppler	2.54	0.38	1.80	3.28	7	
$\frac{Q_{\text{Left liver}}}{Q_{\text{Liver}}} [\%]$						
Reference	0.00	0.05	-0.09	0.09	487	0.97
FGR, normal UA Doppler	-0.06	0.25	-0.55	0.43	16	
FGR, abnormal UA Doppler	0.03	0.38	-0.71	0.77	7	
$\frac{Q_{\text{Right liver}}}{Q_{\text{Liver}}} [\%]$						
Reference	0.02	0.05	-0.06	0.11	487	0.95
FGR, normal UA Doppler	0.10	0.25	-0.39	0.59	16	0.00
FGR, abnormal UA Doppler	-0.01	0.38	-0.75	0.73	7	
$\frac{Q_{\text{Ductus venosus}}}{Q_{\text{Umbilical vein}}}$ [%]	5.01	0.00	5.75	5.75	,	
QUmbilical vein	0.00	0.04	0.08	0.00	524	<0.0001
Reference	0.00	0.04	-0.08	0.09	524	< 0.0001
FGR, normal UA Doppler	0.58	0.25	0.09	1.07	16	
FGR, abnormal UA Doppler	1.57	0.29	1.00	2.14	13	
Q <sub>Ductus venosus</sub>	0.00	0.04	0.00	0.00	522	-0.000
Reference	0.00	0.04	-0.08	0.09	533	< 0.0001
FGR, normal UA Doppler	-0.59	0.25	-1.08	-0.10	16	
FGR, abnormal UA Doppler	-1.07	0.29	-1.63	-0.50	13	
$Q_{\text{Ductus venosus}}(\text{kg})$	<i>~ ~~</i>	0.01	e			~ ~
Reference	0.00	0.04	-0.08	0.09	533	0.6
FGR, normal UA Doppler	0.16	0.25	-0.33	0.65	16	
FGR, abnormal UA Doppler	0.23	0.29	-0.34	0.79	13	

by the local research ethics committee (Regional Committee for Research Ethics, Western Norway, REK-Vest 04/3837) and participants were recruited after informed written consent from April 2005 until March 2007. At delivery, gender and birth weight were recorded. Cord acid-base data from both the umbilical artery and vein were obtained by clamping a cord segment immediately after delivery. The cord blood samples were analyzed by an ABL 735 Radiometer (Bergman Diagnostica, Oslo, Norway) calculating base deficit of the extracellular fluid according to Siggaard-Andersen algorithm (7).

A Sonos 7500 ultrasound machine (Philips, Seattle, WA) with a 3.5 MHz (2–6 MHz) curved linear transducer including color Doppler (2.5 MHz) and pulsed Doppler (3 MHz) facilities was used for the study. The high-pass filter was set at 50 Hz. Vessel diameter and flow velocities [time-averaged maximum flow velocity (TAMXV)] were measured at the intra-abdominal portion of the umbilical vein, inlet of the ductus venosus, left portal vein, and main portal stem. Flow velocity was measured in an insonation along the vessel axis with the angle of insonation kept as low as possible and always lower than 30 degree. For diameter measurement, the vessel walls were insonated perpendicularly and the mean of the repeated measurements was used for flow calculation according to the formula:  $Q = \pi (D/2)^2 hTAMXV$  (*h*: velocity profile h = 0.7 for ductus venosus,

h = 0.5 for the umbilical and portal veins). Further details on measurement techniques and calculation of volume blood flow (*Q*) are described elsewhere (5,8,9). The pulsatility index of the umbilical artery (PI<sub>umb artery</sub>) was measured in a free cord loop.

The total venous blood supply of the liver ( $Q_{liver}$ ) was calculated as:  $Q_{liver} = Q_{untbilical vein} - Q_{ductus venous} + Q_{portal vein}$ . The venous blood flow to the left liver lobe ( $Q_{left liver}$ ) was calculated as:  $Q_{left liver} = Q_{untbilical vein} - (Q_{ductus venous} + Q_{left portal vein})$  and to the right lobe ( $Q_{right liver}$ ) as:  $Q_{right liver} = Q_{portal vein} + Q_{left portal vein}$ . The umbilical and portal fraction of the total venous liver supply was calculated as ( $Q_{umbilical vein} - Q_{ductus venous}$ )  $\times 100\%/Q_{liver}$  and  $Q_{portal vein} \times 100\%/Q_{liver}$ , respectively. Blood flow to the right and left lobes relative to the total venous liver flow was  $Q_{left liver} \times 100\%/Q_{liver}$  and  $Q_{right liver} \times 100\%/Q_{liver}$ , respectively. The ductus venosus shunt fraction was calculated as  $Q_{ductus venous}/Q_{umbilical vein} \times 100\%/Q_{liver}$  and  $Q_{right liver} \times 100\%/Q_{liver}$ , respectively. The ductus venosus shunt fraction was calculated as  $Q_{ductus venous}/Q_{umbilical vein} \times 100\%$ . The blood flow was normalized for estimated fetal weight, calculated by Combs formula. The corresponding percentile was obtained using gender-specific reference ranges (10). Blood flow data from FGR were compared with a reference population (11) using z scores (SDS). The relationship between continuous variables was assessed by linear regression analysis. Pl\_umb artery >97.5 percentile was considered representing a more severe degree of

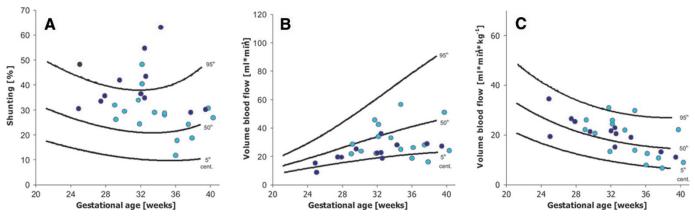
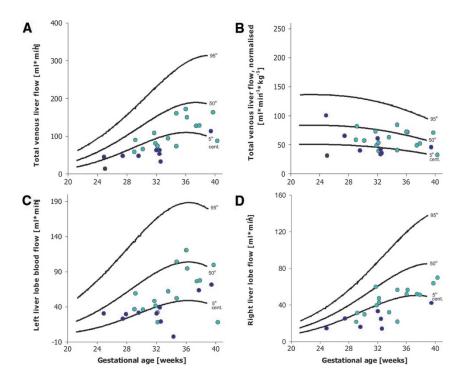


Figure 1. Ductus venosus shunt fraction (*A*), absolute (*B*), and normalized (*C*). Ductus venosus blood flow in growth-restricted fetuses with normal (*light blue bars*, n = 16) and increased (*dark blue bars*, n = 7) umbilical artery pulsatility. Data presented in relation to gestational age with mean, 5th and 95th percentile of the reference population.



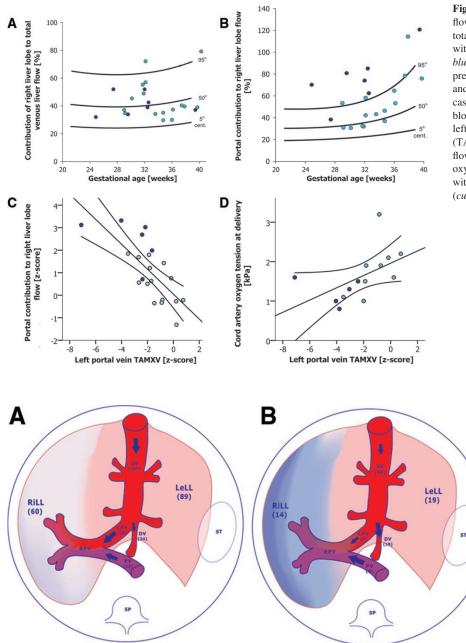
**Figure 2.** Total venous liver flow in absolute terms (*A*) and normalized for fetal weight (*B*); venous blood flow to the left (*C*) and right (*D*) liver lobe in growth-restricted fetuses with normal (*light blue bars*; n = 16) and increased (*dark blue bars*; n = 9 [*A* and *B*]; n = 10 [*C*]; n = 7 [*D*]) umbilical artery pulsatility. Data presented in relation to gestational age with mean, 5th and 95th percentile of the reference population.

hemodynamic compromise than those below and the group was divided accordingly for subanalyses. Statistical analysis was performed using SPSS (Statistical Package for the Social Sciences, SPSS Inc, Chicago, IL).

# RESULTS

Of the 31 fetuses with estimated fetal weight ≤5th percentile, 2 had a birth weight on the 25th percentile, giving 29 fetuses eligible for analysis. Sixteen of 29 fetuses had normal and 13 of 29 increased  $PI_{umb artery}$  (>97.5 percentile). Further characteristics of the study population are shown in Table 1. Missing measurements (Table 2) were due to unfavorable fetal position, fetal movements, or time constraints. FGR fetuses shunted an increased fraction of umbilical blood through the ductus venosus (Table 2, Fig. 1A). Although absolute ductus venosus flow is decreased (Table 2, Fig. 1B), normalized flow is maintained compared with the reference population (Table 2, Fig. 1C). FGR fetuses had relatively low total venous liver flow (Table 2, Fig. 2A), even when normalized for fetal weight (Table 2, Fig. 2B). The low total venous blood flow was related to the degree of placental compromise and equally affected the left and right liver lobes (Table 2, Fig. 2C and D), thus maintaining the fractional distribution between left (60%) vs. right liver lobe (40%) as in the reference population (Table 2, Fig. 3A). However, growthrestricted fetuses substituted umbilical with portal blood to the right liver lobe depending on the degree of placental compromise, and in extreme cases the right lobe was exclusively perfused by portal blood (Table 2, Fig. 3B). There was a strong inverse relationship between the portal contribution to the right liver lobe flow and the time-averaged maximum blood velocity in the left portal vein (r = 0.73; p < 0.0001) (Fig. 3C). Furthermore, our data suggest a direct relationship between time-averaged maximum blood velocity in the left portal vein and the oxygen partial pressure in the umbilical artery sampled at birth (Fig. 3D).

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**Figure 3.** The portal contribution to right liver lobe flow (*A*) and the contribution of right liver lobe flow to total venous liver flow (*B*) in growth-restricted fetuses with normal (*light blue bars*, n = 16) and increased (*dark blue bars*, n = 7) umbilical artery pulsatility. Data presented in relation to gestational age with mean, 5th and 95th percentile of the reference population. The two cases with a portal contribution >100% had reversed blood flow in the left portal vein. Relationship between left portal vein time-averaged maximum flow velocity (TAMXV) and the portal contribution to right liver lobe flow (r = 0.73, p < 0.0001) (*C*), and the cord artery oxygen tension at delivery (r = 0.52, p = 0.059) (*D*) with 95% confidence interval for the regression lines (*curved lines*).

#### DISCUSSION

This study demonstrates that in FGR the venous perfusion of the liver is reduced. To minimize the volume discrepancy, portal blood (with low oxygenation) increasingly substitutes oxygen- and nutrient-rich umbilical flow to the right liver lobe, graded according to umbilical artery compromise, worsening oxygenation of this part of the liver (Fig. 4).

In agreement with previous human studies (2,3), we found an increased ductus venosus shunt fraction in FGR as part of the circulatory adaptation to placental insufficiency.

Our findings also confirm microsphere blood flow measurements in fetal sheep (12) and a Doppler study in human FGR (3) showing a greater drop in umbilical blood flow to the right compared with the left lobe of the liver during placental compromise. The fraction of cardiac output directed to the

Figure 4. Typical pattern of venous blood supply of the fetal liver with level of oxygenation, flow direction (blue arrows), and flow amount (arrow size) in the uncompromised (A) and growth-restricted fetus (B). Red, high oxygen saturation; purple, low oxygen saturation. UV, umbilical vein; DV, ductus venosus; LPV, left portal vein; RPV, right portal vein; PV, main stem of the portal vein; LeLL, left liver lobe; RiLL, right liver lobe; ST, stomach; SP, spine. Volume blood flow (mL/min) in brackets: Panel A, corresponding to the mean for 32 gestational wks in uncompromised pregnancies, panel B, one FGR case at 32 + 4 wk.

placenta for reoxygenation is decreased in growth-restricted fetuses (13) with a correspondingly increased fraction recirculating within the fetal body. In fetal lambs, portal venous blood is largely oxygen depleted with a saturation of 30% (14). The relative increase in portal blood directed to the liver observed in this study thus constitutes part of this recirculation although it mainly affects the right lobe. This is reflected in the blood velocity in the left portal vein (Fig. 3C). A reduced or reversed blood velocity in this vessel has been shown to be a direct reflection of the altered distribution between umbilical and portal blood supply to the liver, the umbilicoportal watershed (Fig. 4) (15). The shift in this watershed toward the left liver lobe was related to the degree of hypoxemia at birth (Fig. 3D). The decreased splenic artery pulsatility reported in FGR with low cord  $Po_2$  (16) provides further support for the concept that splanchnic vessels contribute to increased recirculation during hypoxemia.

It is likely that a reduction in well-oxygenated umbilical venous flow will affect liver growth and function. Indeed, a direct relationship between venous liver perfusion and tissue proliferation was demonstrated in fetal sheep (4). Our findings suggest that this will affect predominantly the right lobe and, interestingly, fetal lambs exposed to reduced uterine blood flow selectively show reduced growth of the right lobe of the liver (17). Selective right liver lobe damage was also reported in human fetuses that have suffered intrauterine asphyxia (18). Furthermore, exposure of the liver to hypoxia *per se* may also induce reduced fetal growth: human hepatocytes cultured in hypoxic conditions increase their expression of IGF binding proteins, which inhibits the IGF action (19). Impaired liver growth has been previously demonstrated in human FGR (20,21) although differences between the lobes were not addressed.

Even in apparently uncompromised pregnancies differences in microarchitecture, enzyme function (22), and gene expression (23) between the fetal right and left liver lobes have been found and mainly ascribed to fetal venous perfusion patterns. Because differences between the liver lobes seem to persist into adulthood (24,25), the circulatory changes within the liver of growth-restricted fetuses may have long-term implications on liver function after birth. Growth-restricted neonates showed a decreased hepatic biotransformation compared with appropriately grown babies (26). Moreover, a recent casecontrol study of children with nonalcoholic fatty liver disease showed a 39% prevalence of being born small for gestational age, compared with 7% among controls (27).

FGR fetuses with normal umbilical artery pulsatility index are commonly considered to be at minor risk for perinatal complications. Interestingly, this study demonstrated that such fetuses not only may show brain sparing (28,29) with potential consequences for postnatal development (30), but also an impaired and distorted liver perfusion.

In conclusion, both mild and severe placental insufficiency cause venous hypoperfusion of the fetal liver and an increased contribution of poorly oxygenated blood from the portal vein. This predominantly affects the right liver lobe (Fig. 4). The reduced oxygenation of the right lobe may have longer-term consequences, ranging from the right liver necrosis occasionally described in the perinatal period to alterations in metabolic processes, which may predispose to later disease.

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