

Effect of p21^{Waf1} and p27^{Kip1} on centrosome replication and proliferation
of breast cancer cell

LI Fu-nian^{a*},JIANG Dan-dan^a, LIU Xiang-ping^b, GE Yin-lin^c, CAO Ming-zhi^a, WANG

Xin-gang^a

^a Department of Breast Surgery, The Affiliated Hospital of Qingdao University Medical
College, Qingdao 266003, China

^b Central laboratory, The Affiliated Hospital of Qingdao University Medical College,
Qingdao 266003, China

^c Department of Biochemistry and Molecular Biology, Qingdao University Medical
College, Qingdao 266021, China

Corresponding author: LI fu-nian, Department of Breast Surgery, The Affiliated
Hospital of Qingdao. Medical College, Qingdao 266003, China, Email:
drlifunian@yahoo.com.cn, telephone: 13583208308

LI Fu-nian made contributions to first concept and design. JIANG Dan-dan contributed
to design and write the manuscript. LIU Xiang-ping made contributions to technical
assistance and analysis of data. GE Yin-lin, CAO Ming-zhi and WANG Xin-gang

contributed to revise the manuscript critically for content.

Abstract

Aberrant centrosome numbers are detected in virtually all cancers increasing the risk for cell division errors and chromosomal instability.

Deregulation of the centrosome duplication cycle is considered as the major contributing factor for abnormal amplification of centrosomes. p21^{Waf1} and p27^{Kip1}, general CDK inhibitors by inhibiting cyclin-dependent kinase 2 (CDK2)/cyclin E and cyclin A complexes, controlled the initiation and progress of centrosome duplication .

We transfected p21^{Waf1}, p27^{Kip1} or p21^{Waf1}- p27^{Kip1} genes into MCF-7 cells by lipofection to explore the effect of the genes on centrosome duplication and proliferation of breast cancer cell. The result shows that the cell growth was obviously inhibited after being transfected, resulting in an accumulation of cells in G₁ and the proportion of cells which contained abnormal centrosomes was obviously decreased. Comparing with p21^{Waf1} or p27^{Kip1}, the effects of p21^{Waf1}- p27^{Kip1} genes are more significant. These results suggest that p21^{Waf1} and p27^{Kip1} genes could inhibit the growth of human breast cancer

cells and reverse abnormal duplication of centrosomes. p21^{Waf1} and p27^{Kip1} cooperate to regulate centrosome duplication and cell cycle progress, indicating p21^{Waf1}- p27^{Kip1} combined gene might be potential therapeutic agents of breast cancer which reveals suppressed p21^{Waf1} and p27^{Kip1} expression.

Key words: Centrosome; [Cell Proliferation](#); p21^{waf1} gene; p27^{kip1} gene; Breast neoplasms

Introduction

The centrosome is the major microtubule organizing center during interphase and mitosis in most animal and human cells [1]. Each centrosome consists of a pair of centrioles and a surrounding protein matrix referred to as pericentriolar materials [2].

During interphase, centrosomes organize the cytoplasmic microtubule network, which is involved in vesicle transport, proper distribution of small organelles, and establishment of cellular shape and polarity. During mitosis, duplicated centrosomes direct formation of bipolar spindles, which is critical for accurate segregation of chromosomes and cytokinesis [3]. The centrosome duplicates once every cell cycle, which starts during the G₁-S transition, coincident with the onset of DNA replication. Abrogation of regulatory mechanisms governing centrosome duplication leads to generation of amplified centrosomes (more than two centrosomes), which in turn leads to mitotic aberration (multipolar spindles) and unequal segregation of chromosomes[3]. Destabilization of chromosomes by centrosome amplification aids acquisition of further malignant phenotypes, hence promoting tumor progression. Recent studies have shown

that centrosome hyperamplification is commonly observed, and is the major contributing factor for chromosome instability in human tumors [4–7]. Moreover, a significant reduction of p21^{Waf1} and p27^{Kip1} expression, which occurs at a high frequency in human cancer [8-12], strongly correlates with the occurrence of centrosome hyperamplification. p21^{Waf1} and p27^{Kip1}, general CDK inhibitors, mainly inhibits cyclin/CDK2 complexes, thereby arresting the cell cycle[13,14]. Some reports have shown that the activation of CDK2/cyclinE (and cyclinA) is essential for the initiation of centrosome duplication[15]. It is not difficult to consider the role of p21^{Waf1} and p27^{Kip1} in centrosome homeostasis. In one of the first related studies, cells deficient of p21 and exposed to ionizing radiation were found to accumulate abnormal numbers of centrosomes following abortive mitoses[16]. Similar results were obtained in p27^{-/-}

and p27^{+/-} mouse embryonic fibroblasts and p27-silenced human cells [17]. Thus, both CDK inhibitors regulate centrosome duplication and cooperation of p21 and p27 might be important for accurate chromosome duplication. Furthermore, inhibitory effects of CDK inhibitors, p21^{Waf1} and p27^{Kip1}, on repeated centrosome reproduction were reported in frog embryos and *Xenopus* egg extracts[18-20]. But whether p21^{Waf1} and p27^{Kip1} can inhibit the abnormal duplication of centrosomes in breast cancer cells is not well known.

In this study, we transfected p21^{waf1}, p27^{kip1} and p21^{waf1}- p27^{kip1} genes into MCF-7 cells by lipofection to explore the role of p21^{Waf1} and p27^{Kip1} as a therapeutic target in breast cancer and the mechanism of interaction between p21^{Waf1} and p27^{Kip1}.

Materials and Methods

Reagents and antibodies.

pIRES-p21^{waf1} pIRES- p27^{kip1} and pIRES-p21^{waf1}- p27^{kip1} were generous gifts from

LIU Xiang-ping. A human breast carcinoma cell line MCF-7 was purchased from

Chinese Academy of Science. p21^{waf1} and p27^{Kip1} antibodies were purchased from Santa

Cruz Biotechnology (Santa Cruz, CA). Horseradish peroxidase (HRP)-conjugated anti-

mouse was obtained from Boster(Wuhan,China). The α -tubulin antibody and a FITC

conjugated secondary antibody were purchased from BD Pharmingen. All fine

chemicals were purchased from Sigma. The ECL detection system was obtained from

Amersham Biosciences (Piscataway, NJ). All cell culture products were purchased from

Jinuo(Hangzhou, China). Lipofectamine 2000 was purchased from Invitrogen (Grand

Island, NY).

Cell line and transfection.

The human breast carcinoma cell line MCF-7 was grown in Dulbecco's modified Eagle's medium (DMEM) containing 10% fetal bovine serum (FBS) at 37 °C in 5% CO₂ incubator. The recombinant plasmids pIRES-p21^{Waf1}, pIRES-p27^{Kip1} and pIRES-p21^{Waf1}-p27^{Kip1} were generously provided by Dr. Liu(center laboratory Qingdao, China). MCF-7 cells were transfected with these plasmids using lipofectamine.

Western blot analysis.

Cells were lysed in RIPA buffer [20 mM Tris-HCl, pH8.0, 1 mM EDTA, 1mM

EGTA, 150 mM NaCl, 0.1% (w/v)SDS, 1.0% (v/v) NP-40], which contained

Completek-Mini protease inhibitors. The amount of proteins was determined using a

Bradford protein assay kit (Bio-Rad, Hercules, CA). An aliquot of 50 g proteins from

a total lysate was electrophoresed on SDS-PAGE gel and then transferred to Immobilon-

P (Millipore, Bedford, MA). After blocking with 5% nonfat dry milk in Tris-buffered

saline (TBS), the membrane was incubated with a primary antibody at an appropriate

dilution overnight at 4 and then washed three times in TBST. The membrane was

then incubated with a secondary antibody at a dilution of 1:5,000 at room temperature

for 1h and again washed three times in TBST. The blots were visualized with ECL

(Amersham).

Flow cytometry.

The cells were fixed in 70 % ethanol for 1 hour at 4 °C. The fixed cells were incubated with PBS with 300 µg/mL RNase A at room temperature and stained with 50 µg/mL propidium iodide. Flow cytometric analysis was done with FACSC alibur and Cell Quest software (Becton Dickinson, San Jose, CA).

Indirect immunofluorescence.

Cells grown on slides were fixed with 4 % formalin for 20 min at 25 °C. The cells were permeabilized with 0.2 % Triton X-100 for 30 min, followed by incubation with blocking solution (10 % normal goat serum in PBS) for 1h. Cells were then probed with primary antibody for 1h and antibody-antigen complexes were detected with FITC-

conjugated goat secondary antibody by incubation for 1 h at 25 . The samples were washed three times with PBS after each incubation, and then counterstained with 4', 6-diamidino-2-phenylindole (DAPI).

Statistical analysis.

Differences between two groups were compared using the two-tailed unpaired Student's *t* test. $P < 0.05$ was considered statistically significant.

Results

Arrest of cell cycle and restoration of the normal centrosome duplication cycle by high expression of re-introduced p21^{Waf1}

Several reports have shown that breast cancer cells lacking p21^{Waf1} exhibit centrosome amplification after DNA damage. And after induced DNA damage, the level of p21^{Waf1} is gradually increases and centrosome amplification is suppressed. We decided to test whether re-introduction of p21^{Waf1} gene into MCF-7 can induce cell cycle arrest or restore the normal centrosome duplication cycle. The MCF-7 cells were transfected with a pIRES-p21^{Waf1}, generating a MCF-7- p21^{Waf1} cell line that constitutively expresses P21 protein. As a control, a pIRES vector was transfected into the MCF-7 cells (MCF-7-pIRES).

After transfected 48 hours, a statistical significant increased p21^{Waf1} expression was detected by immunoblot analysis (Fig 1). At the same time, cells were analysed by flow cytometry(Fig 2). MCF-7- p21^{Waf1} cells showed a different cell cycle phase distribution

from the control MCF-7-pIRES. MCF-7- p21^{Waf1} cell increased the number of cells in

G₁ phase P<0.01 and decreased the number of cells in S phase, indicating

that p21^{Waf1} overexpression can inhibit cell cycle progression resulting in an

accumulation of cells in G₁.

The centrosome in the MCF-7- p21^{Waf1} cell line was examined using antibody

against γ - tubulin, a major component of centrosomes (Figure 3). As the control, MCF-

7-pIRES cells were also immunostained. MCF-7-pIRES cells showed an expected

abnormal centrosome profile, 13% of the cells contained abnormally amplified

centrosomes (n = 3). In contrast, MCF-7- p21^{Waf1} cells showed a partial restoration of

centrosome profile; an increase in the number of cells with one or two centrosomes and

a decrease in the number of cells with n = 3 centrosomes(4%), demonstrating that re-

introduction of physiological high level of p21^{Waf1} re-established the normal centrosome profile.

These observations demonstrate that p21^{Waf1} is directly involved in the controlling the coordinated initiation of centrosome and DNA duplication. p21^{Waf1} overexpression can induce cell cycle arrest and restoration of the normal centrosome duplication cycle in breast cancer.

Arrest of cell cycle progression and restoration of the normal centrosome

duplication cycle by high expression of re-introduced p27^{Kip1}

p27^{Kip1}, as the related CDK inhibitor, also exhibited a significant reduction in most human aggressive tumors(16). We transfected pIRES-p27^{Kip1} into the MCF-7 cells to

explore effect of transfected p27^{Kip1} gene on the duplication of centrosomes and the proliferation of human breast carcinoma cells.

After transfected 48 hours, a statistical significant increase p27^{Kip1} expression was detected by immunoblot analysis (Fig 1). At the same time, cells were analysed by flow cytometry(Fig 2) MCF-7- p27^{Kip1} cells showed a different cell cycle phase distribution from the control MCF-7-pIRES. MCF-7- p27^{Kip1} cell increased he number of cells in G₁ phase P<0.01 and decreased the number of cells in S phase, indicating that p27^{Kip1} overexpression can inhibit cell cycle progression presumably during the G₁/S transition period, resulting in an accumulation of cells in G₁.

The centrosome in the MCF-7- p27^{Kip1} cell line was examined using antibody against

- tubulin (Fig3). Comparing with the controlled MCF-7-pIRES cells, MCF-7-p27^{Kip1}

cells showed a partial restoration of centrosome profile; an increase in the number of

cells with one or two centrosomes and a decrease in the number of cells with n = 3

centrosomes(6%), suggesting re-introduction of physiological high level of p27^{Kip1} re-

established the normal centrosome profile.

**The effect of cooperation of p21^{Waf1} and p27^{Kip1} on the duplication of centrosome
and the proliferation of human breast cancer cell**

The above results demonstrated that both p21^{Waf1} and p27^{Kip1} can inhibit cell cycle

transition and restore centrosome duplication. As the related CDK inhibitors, the

cooperation of both might be important for the progression of cell cycle and centrosome

duplication. Therefore we test whether p21^{Waf1}- p27^{Kip1} combination get more effective

inhibit effects.

The recombinant pIRES-p21^{Waf1}- p27^{Kip1} was constructed and transfected into the

MCF-7 to generate a MCF-7- p21^{Waf1}- p27^{Kip1} cell line that constitutively expresses P21

and P27 proteins.

After transfected 48 hours, a similar significant increased P21 and P27 proteins

expression were detected as the same as the expression in MCF-7- p21^{Waf1} cells and

MCF-7- p27^{Kip1} cells(Fig 1). By flow cytometry, MCF-7- p21^{Waf1}- p27^{Kip1} cells showed a

different cell cycle phase distribution from the controls, MCF-7- p21^{Waf1} and MCF-7-

p27^{Kip1}. MCF-7- p21^{Waf1}- p27^{Kip1} cells exhibited an increase in the number of cells in G₁

phase P<0.01 and a decrease in the number of cells in S phase (Fig 2),

indicating that p21^{Waf1} and p27^{Kip1} combination exhibited more evident inhibit effects on

cell cycle progression than p21^{Waf1} or p27^{Kip1} single gene.

As anticipation, in contrast to the controls, MCF-7- p21^{Waf1}- p27^{Kip1} cells showed a

statistical increase in the number of cells with one or two centrosomes and a decrease in

the number of cells with n = 3 centrosomes(2%) (Fig 3), further demonstrating that

p21^{Waf1}-p27^{Kip1} combination is a more effective inhibitor than p21^{Waf1} or p27^{Kip1} single

gene.

Several studies have suggested that both p21^{Waf1} and p27^{Kip1}, general inhibitors of

cyclin-dependent kinases in G₁ and S phase, can interact with many different cyclin-

CDK complexes. This interaction is mediated by a homologous domain. Our results

indicated the association between p21^{Waf1} and p27^{Kip1} and suggested that the association

of p21^{Waf1} and p27^{Kip1} plays a crucial role to assure the accurate cell cycle transition and

centrosome duplication. But the specific mechanism is still poorly understood.

Discussion

Defect in the fidelity of chromosome segregation is a common characteristic of cancer

cells and are likely to be important in the progression of a cancerous phenotype. Studies

of cultured cells and tumor tissues have shown that tumorigenesis associated with

suppressed p21^{Waf1} and p27^{Kip1} is primarily attributed to the deregulated centrosome

duplication cycle and the consequential centrosome hyperamplification.

CDKs, a family of serine/threonine kinases, control the onset of the major cell

cycle events such as DNA synthesis and mitosis, and also the accurate duplication of

centrosome. The activity of CDKs is in part regulated by association of different cyclins, which are temporally expressed at specific cell cycle stages. CDK inhibitors p21^{Waf1} and p27^{Kip1} interact with these complexes and thereby inhibit CDKs activity [19].

Labaer[21] and Nomura[22] found the differences between p21^{Waf1} and p27^{Kip1}, when they study the function of CDK inhibitors. p21^{Waf1} inhibits the assembly of cyclinD/CDK4 complex which is the mainly active kinase complexes in mid-earlier period of G₁, and is considered as the major inhibitor in earlier G₁. p27^{Kip1} firstly binds to cyclinD/CDK4 in earlier G₁. After cyclinE/CDK4 complex formation, p27^{Kip1} is visible to bind to cyclinE/CDK4 complex in later G₁ and inhibits the cell cycle transition. The accurate regulation of p21^{Waf1} and p27^{Kip1} is the major contributing factor for cell normal growth and development.

p21^{Waf1}, a general inhibitor of cyclin-dependent kinases in G₁ and S phase, can bind to many different cyclin-CDK complexes, inhibit activity of nuclear cyclin-CDK complexes. Recent reports suggested that p21^{Waf1} controlled the initiation and progress of centrosome duplication by regulating cyclin/CDK2 activity[15]. One of the first related investigation was that cells lacking p21^{Waf1} were prone to centrosome amplification following a prolonged S phase arrest [23]. Additional support for a role of p21^{Waf1} in centrosome duplication came from studies that The human papillomavirus type 16 E7 (HPV-16 E7) oncoprotein, which inactivates p21^{Waf1} [24,25] was found to stimulate centrosome overduplication[26]. In keeping with these findings, depletion of p21^{Waf1} in human hematopoietic cells was found to cause abnormal centrosome numbers together with a deformed nuclear architecture and polyploidy[27]. In our studies, we

found the abnormal duplication of centrosome in MCF-7 breast cancer, following a suppressed expression of p21^{Waf1}. This is consistent with previous research result that centrosome aberrations in primary invasive breast cancer are associated with nodal status and hormone receptor expression[28]. Furthermore, after transfected exogenous p21^{Waf1}, MCF-7 breast cancer cells with p21^{Waf1} overexpression, exhibited the arrest of cell cycle progression and restoration of the normal centrosome duplication cycle.

These results are consistent with our prior studies and supported by a report that p21^{Waf1} overexpression can inhibit centrosome overduplication of cells treated with hydroxyurea for a long time[15,29].

p27^{Kip1}, similar with p21^{Waf1}, was initially identified as a negative regulator of G₁ progression in growth-arrested cells [30-32] and required to ensure the coordinated

progression of centrosome duplication and cell cycle progress. Some reports have suggested that abnormal mitotic cells with amplified centrosomes were frequently observed in p27-silenced cells [17]. Furthermore, silencing of Skp2 by siRNA leads to P27 accumulation, reduction of CDK2 activity and suppression of centrosome amplification in lung cancer cells [33]. We observed that MCF-7 breast cancer cells transfected with p27^{Kip1} presented cell cycle inhibition and abnormal centrosome amplification suppression, suggesting that p27^{Kip1} could suppress cell cycle transition and restore centrosome amplification in breast cancer cells.

We show here that both p21^{Waf1} and p27^{Kip1} contribute to suppress CDK2 activity, arresting cell cycle progress and centrosome amplification. Either defection of p21^{Waf1} or p27^{Kip1} can induce cell cycle disorder and centrosome amplification. Thus, there

remained a possibility that p21^{Waf1} and p27^{Kip1} cooperate to regulate cell cycle progress and centrosome duplication depending on the differences between p21^{Waf1} and p27^{Kip1} in molecular structure, control methods, pathway activity and mode of action. Our study found that p21^{Waf1}- p27^{Kip1} combination had more significant inhibition of cell cycle progress and centrosome amplification than p21^{Waf1} or p27^{Kip1} single gene. These results lend support to the hypothesis that the cooperation of p21^{Waf1} and p27^{Kip1} play a synergistic role in regulating the cell cycle progress and centrosome duplication.

The tumorigenesis and progression of breast cancer refer to a variety of oncogenes, tumor suppressor genes forming a network-control. Therefore study of a single gene is difficult to see the whole picture, and that of multiple genes and their linkages is closer to the body the actual situation. Our findings suggest that p21^{Waf1} and

p27^{Kip1} overexpression can inhibit cell proliferation through G₁ cell cycle arrest and restore centrosome duplication through centrioles separation suppression in breast cancer. Possibly because of the differences in molecular structure, control methods, pathway activity and mode of action, p21^{Waf1} and p27^{Kip1} cooperate to inhibit cell proliferation and centrosome duplication. Then p21^{Waf1}- p27^{Kip1} combination might have potential clinical significance as therapeutic agents of breast cancer with suppressed p21^{Waf1} and p27^{Kip1} expression. Multi-gene therapy may provide a new idea for breast cancer gene therapy, but the mechanism of the synergy between p21^{Waf1} and p27^{Kip1} remains to be further explored.

Acknowledgments

We thank Yang Kun, Sui Ai-hua and Li Jing-hua for their technical help. We also thank the strongly support from Department of Breast Surgery in the Affiliated Hospital of Qingdao University Medical College.

Conflict of interest statement

None declared.

Funding

Project funded by grants from Natural Science Foundation of Shandong Province

(y2007C134) and Natural Science Foundation of Qingdao Municipalit (06-2-2-5-nsh-1).

References

1. Bornens M. Centrosome composition and microtubule anchoring mechanisms. *Curr Opin Cell Biol.* 2002;14(1):25–34.
2. Fukasawa K. Introduction. *Centrosome: Oncogene* 2002;21(40):6140–5.
3. Meraldi P, Nigg EA. The centrosome cycle. *FEBS Lett* 2002;521(1-3):9–13.
4. Lingle WL, Lutz WH, Ingle JN, Maihle NJ, Salisbury JL. Centrosome hypertrophy in human breast tumors: implications for genomic stability and cell polarity. *Proc Natl Acad Sci U S A* 1998;95(6):2950–5.
5. Pihan GA, Purohit A, Wallace J, Knecht H, Woda B, Quesenberry P, Doxsey SJ. Centrosome defects and genetic instability in malignant tumors. *Cancer Res* 1998;58(17):3974–85.
6. Carroll PE, Okuda M, Horn HF, Biddinger P, Stam Brook PJ, Gleich LL, Li YQ, Tarapore P, Fukasawa K. Centrosome hyperamplification in human cancer: chromosome instability induced by p53 mutation and/or Mdm2 overexpression. *Oncogene* 1999;18(11):1935–44.
7. Sato N, Mizumoto K, Nakamura M, Nakamura K, Kusumoto M, Niiyama H, Ogawa T, Tanaka M. Centrosome abnormalities in pancreatic ductal carcinoma. *Clin Cancer Res* 1999;5(5):963–70.
8. Tsihlias J, Kapusta L, Slingerland J. The prognostic significance of altered cyclin-dependent kinase inhibitors in human cancer. *Annu Rev Med.* 1999;50:401-23.
9. Skomedal H, Kristensen GB, Lie AK, Holm R. Aberrant expression of the cell cycle associated proteins TP53, MDM2, p21, p27, cdk4, cyclin D1, RB, and EGFR in cervical carcinomas. *Gynecol Oncol.* 1999;73(2):223-8.

10. Lu X, Toki T, Konishi I, Nikaido T, Fujii S. Expression of p21^{WAF1/CIP1} in adenocarcinoma of the uterine cervix: a possible immunohistochemical marker of a favorable prognosis. Cancer. 1998;82(12):2409-17.
11. Huang LW, Chao SL, Hwang JL, Chou YY. Down-regulation of p27 is associated with malignant transformation and aggressive phenotype of cervical neoplasms. Gynecol Oncol. 2002;85(3):524-8.
12. Kataoka TR, Tsukamoto Y, Matsumura M, Miyake A, Kamiura S, Ishiguro S, Nishizawa Y. Expression of p21Cip1/Waf1 and p27Kip1 in small cell neuroendocrine carcinoma of the uterine cervix. Int J Surg Pathol. 2008;16(1):11-5.
13. Xiong Y, Hannon GJ, Zhang H, Casso D, Kobayashi R, Beach D. p21 is a universal inhibitor of cyclin kinases. *Nature*. 1993;366:701-4.
14. Toyoshima H, Hunter T. p27, a novel inhibitor of G₁ cyclin-Cdk protein kinase activity, is related to p21. *Cell*. 1994;78:67-74.
15. Matsumoto Y, Hayashi K, Nishida E. Cyclin-dependent kinase 2 (Cdk2) is required for centrosome duplication in mammalian cells. Curr Biol. 1999 Apr 22;9(8):429-32.
16. Bunz F, Dutriaux A, Lengauer C, Waldman T, Zhou S, Brown JP, Sedivy JM, Kinzler KW, Vogelstein B. Requirement for p53 and p21 to sustain G2 arrest after DNA damage. *Science*. 1998;282:1497-501.
17. Eiji Sugihara, Masayuki Kanai, Soichiro Saito, Takayuki Nitta, Hideo Toyoshima, Keiko Nakayama, et al. Suppression of Centrosome Amplification after DNA Damage depends on p27 Accumulation. *Cancer Research*,2006;66:4020-9.
18. Meraldi P, Lukas J, Fry AM, Bartek J, Nigg EA. Centrosome duplication in mammalian

- somatic cells requires E2F and Cdk2- cyclin A. *Nat Cell Biol.* 1999;1:88–93.
19. Lacey KR, Jackson PK, Stearns T. Cyclin-dependent kinase control of centrosome duplication. *Proc Natl Acad Sci U S A.* 1999;96:2817–22.
 20. Hinchcliffe EH, Li C, Thompson EA, Maller JL, Sluder G. Requirement of Cdk2-cyclin E activity for repeated centrosome reproduction in *Xenopus* egg extracts. *Science.* 1999;283:851–4.
 21. Labaer J, Garrett MD, Stevenson LF, et al. New functional activities for the p21 family of CDK inhibitor. *Genes Dev* 1997;11(4):847–862.
 22. Nomura H, Sawada Y, Fujinaga K, et al. Cloning and characterization of rat p27^{kip1} a cyclin—dependent kinase inhibitor. *Gene* 1997;191(2):211–218.
 23. Tarapore P, Horn HF, Tokuyama Y, Fukasawa K. Direct regulation of the centrosome duplication cycle by the p53- p21Waf1/Cip1 pathway. *Oncogene.* 2001;20:3173–84.
 24. Jones DL, Alani RM, Munger K. The human papillomavirus E7 oncoprotein can uncouple cellular differentiation and proliferation in human keratinocytes by abrogating p21Cip1-mediated inhibition of cdk2. *Genes Dev.* 1997;11:2101–11.
 25. Funk JO, Waga S, Harry JB, Espling E, Stillman B, Galloway DA. Inhibition of CDK activity and PCNA-dependent DNA replication by p21 is blocked by interaction with the HPV-16 E7 oncoprotein. *Genes Dev.* 1997;11:2090–100.
 26. Duensing S, Lee LY, Duensing A, Basile J, Piboonniyom S, Gonzalez S, Crum CP, Munger K. The human papillomavirus type 16 E6 and E7 oncoproteins cooperate to induce mitotic defects and genomic instability by uncoupling centrosome duplication from the cell division cycle. *Proc Natl Acad Sci U S A.* 2000;97:10002–7.

27. Mantel C, Braun SE, Reid S, Henegariu O, Liu L, Hango G, Broxmeyer HE. p21^{cip1}/waf-1 deficiency causes deformed nuclear architecture, centriole overduplication, polyploidy, and relaxed microtubule damage checkpoints in human hematopoietic cells. *Blood*. 1999;93:1390–8.
28. Schneeweiss A, Sinn HP, Ehemann V, et al. Centrosome aberrations in primary invasive breast cancer are associated with nodal status and hormone receptor expression. *Int J Cancer* 2003;107(3):346–352.
29. Chong RF, Li FN, Liu XP, et al. Effects of transfected p21^{Waf1} gene on the duplication of centrosomes and the proliferation of human breast carcinoma cells. *Chin J Exp Surg*, 2008; 25(1):12-14.
30. Polyak K, Lee MH, Erdjument-Bromage H, et al. Cloning of p27^{Kip1}, a cyclin-dependent kinase inhibitor and a potential mediator of extracellular antimitogenic signals. *Cell* 1994;78:59–66.
31. Toyoshima H, Hunter T. p27, a novel inhibitor of G₁ cyclin-Cdk protein kinase activity, is related to p21. *Cell* 1994;78:67–74.
32. Hengst L, Dulic V, Slingerland JM, et al. A cell cycle-regulated inhibitor of cyclin-dependent kinases. *Proc Natl Acad Sci U S A* 1994;91:5291–5.
33. Jiang F, Caraway NP, Li R, et al. RNA silencing of S-phase kinase-interacting protein 2 inhibits proliferation and centrosome amplification in lung cancer cells. *Oncogene* 2005;24:3409–18.