

biochemical and biophysical approaches support this notion. These include co-immunoprecipitation, various types of FRET, atomic force microscopy, covalent cross-linking, gel filtration, neutron scattering experiments, functional complementation, cell biology studies demonstrating cross-internalization and co-processing of GPCRs as well as binding studies showing positive and negative cooperativity. These approaches, their relative strengths and caveats, including methodological considerations and potential functional outcomes, have recently been reviewed^{16,17}. It is therefore premature to dismiss the GPCR oligomer hypothesis based on the interpretations of a single BRET study.

In conclusion, we believe that the results reported in the article by James *et al.* can be interpreted in different ways and that more controls would have been necessary to challenge the multidisciplinary work conducted on this topic by many groups over the past ten years. Clearly, BRET is gaining popularity in assessment of protein-protein interactions in living cells, and additional quantitative approaches will certainly be forthcoming. Maybe more importantly, additional studies performed in native tissues are needed to establish the generality of GPCR dimerization in physiologically relevant systems.

Michel Bouvier¹, Nikolaus Heveker², Ralf Jockers³, Stefano Marullo³ & Graeme Milligan⁴

¹Institute of Research in Immunology and Cancer, Department of Biochemistry, Université de Montréal, C.P. 6128 Succursale Centre-Ville, Montréal, Québec H3C 3J7, Canada. ²Ste-Justine Hospital Research Center, Department of Biochemistry, Université de Montréal, 3175 Chemin de la Côte Sainte-Catherine, Montréal, Québec H3T 1C5, Canada. ³Institut Cochin, Département de Biologie Cellulaire; INSERM, U567; Centre National de la Recherche Scientifique, Unité Mixte de Recherche 8104; and Université Paris 5, Faculté de Médecine René Descartes, UM 3, Paris, F-75014, France. ⁴Institute of Biomedical & Life Sciences, Division of Biochemistry and Molecular Biology, University of Glasgow, Wolfson Building, Glasgow, G12 8QQ, UK.
e-mail: michel.bouvier@umontreal.ca

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James and Davis reply: Bouvier *et al.* claim that several theoretical and/or technical limitations undermine our study¹ of class-A G protein-coupled receptor (GPCR) oligomerization using bioluminescence resonance energy transfer (BRET). First, they propose that in order to characterize the dependence of BRET efficiency (BRET_{eff}) on acceptor/donor ratio it is not strictly necessary to increase the acceptor/donor ratio by holding the combined amount of acceptor and donor constant and decreasing the amount of donor. They claim that it is equally valid to hold donor levels constant and increase the amount of acceptor. The reason why the acceptor/donor ratio should not be varied in this way is that BRET_{eff} will increase for both randomly interacting monomers and oligomers owing to the concomitant increase in overall protein density, preventing discrimination between the two types of interactions. Second, it is true that in the new method donor and acceptor densities each vary. The important point is that the acceptor density is effectively constant above a certain acceptor/donor ratio threshold, so that, in the case of randomly interacting donors and acceptors, each donor ‘experiences’ the same acceptor environment as the acceptor/donor ratio increases beyond this threshold. Third, they argue that we gave insufficient consideration to receptor expression level, but we actually show that GPCRs give similarly low acceptor/donor ratio-independent BRET_{eff} irrespective of overall expression from subphysiological amounts to levels 10–100-fold higher than that observed *in vivo*. Fourth, they claim that we were over-reliant on absolute BRET_{eff} levels when in fact we list four criteria that should be used to establish stoichiometry, including absolute BRET_{eff} levels. Fifth, they propose that in ‘type-2’ BRET experiments (our nomenclature) it is the slope of the line that is important, whereas our control experiments and theory indicate that the defining factor is whether or not the intercept is zero.

Bouvier *et al.* then raise the question of what is to be made of all the biochemical and biophysical data ostensibly supporting class-A GPCR dimerization. We have no direct experience of working with multipass membrane proteins ourselves, but we suspect that these extremely hydrophobic molecules would be prone to artefactual behavior and would exhibit, for example, a tendency to aggregate once extracted from their native membrane environments. The focus of our paper was on the proper implementation of BRET, which, in principle, allows the organizational properties of cell-surface receptors to be examined *in situ* in living cells, free from potential artefacts of this nature. We also note that the controversy surrounding GPCR oligomerization is hardly new^{2,3}.

John R James & Simon J Davis

Nuffield Department of Clinical Medicine and Medical Research Council Human Immunology Unit, Weatherall Institute of Molecular Medicine, The University of Oxford, John Radcliffe Hospital, Oxford, OX3 9DU, UK.
e-mail: simon.davis@clinical-medicine.oxford.ac.uk

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