

ARTICLE

Received 12 Oct 2016 | Accepted 15 Apr 2017 | Published 5 Jun 2017

DOI: 10.1038/ncomms15632

OPEN

IL-17-producing $\gamma\delta$ T cells switch migratory patterns between resting and activated states

Duncan R. McKenzie¹, Ervin E. Kara^{1,†}, Cameron R. Bastow¹, Timona S. Tyllis¹, Kevin A. Fenix¹, Carly E. Gregor¹, Jasmine J. Wilson¹, Rachelle Babb^{1,†}, James C. Paton^{1,2}, Axel Kallies^{3,4}, Stephen L. Nutt^{3,4}, Anne Brüstle⁵, Matthias Mack⁶, Iain Comerford^{1,*} & Shaun R. McColl^{1,7,*}

Interleukin 17-producing $\gamma\delta$ T ($\gamma\delta$ T17) cells have unconventional trafficking characteristics, residing in mucocutaneous tissues but also homing into inflamed tissues via circulation. Despite being fundamental to $\gamma\delta$ T17-driven early protective immunity and exacerbation of autoimmunity and cancer, migratory cues controlling $\gamma\delta$ T17 cell positioning in barrier tissues and recruitment to inflammatory sites are still unclear. Here we show that $\gamma\delta$ T17 cells constitutively express chemokine receptors CCR6 and CCR2. While CCR6 recruits resting $\gamma\delta$ T17 cells to the dermis, CCR2 drives rapid $\gamma\delta$ T17 cell recruitment to inflamed tissues during autoimmunity, cancer and infection. Downregulation of CCR6 by IRF4 and BATF upon $\gamma\delta$ T17 activation is required for optimal recruitment of $\gamma\delta$ T17 cells to inflamed tissue by preventing their sequestration into uninfamed dermis. These findings establish a lymphocyte trafficking model whereby a hierarchy of homing signals is prioritized by dynamic receptor expression to drive both tissue surveillance and rapid recruitment of $\gamma\delta$ T17 cells to inflammatory lesions.

¹Department of Molecular & Cellular Biology, University of Adelaide, Adelaide, South Australia 5005, Australia. ²Research Centre for Infectious Diseases, University of Adelaide, Adelaide, South Australia 5005, Australia. ³The Walter and Eliza Hall Institute of Medical Research, Parkville, Victoria 3052, Australia. ⁴Department of Medical Biology, University of Melbourne, Parkville, Victoria 3010, Australia. ⁵Department of Immunology and Infectious Diseases, John Curtin School of Medical Research, Australian National University, Canberra, Australian Capital Territory 2601, Australia. ⁶Department of Internal Medicine II, University Hospital Regensburg, Regensburg 93042, Germany. ⁷Centre for Molecular Pathology, University of Adelaide, Adelaide, South Australia 5005, Australia. * These authors contributed equally to this work. † Present address(es): Laboratory of Molecular Immunology, The Rockefeller University, New York, New York 10065, USA. (E.E.K.); Department of Medicine, Albert Einstein College of Medicine, New York, New York 10561, USA. (R.B.). Correspondence and requests for materials should be addressed to I.C. (email: iain.comerford@adelaide.edu.au) or to S.R.M. (email: shaun.mccoll@adelaide.edu.au).

Interleukin-17-producing $\gamma\delta$ T cells ($\gamma\delta$ T17 cells) are innate-like lymphocytes crucial for early defence against extracellular bacterial and fungal pathogens. $\gamma\delta$ T17 effector function is programmed in $V\gamma 4^+$ and $V\gamma 6^+$ cells during thymic development, resulting in their homeostatic localization to barrier tissues and ability to be rapidly activated by innate-derived cytokines^{1,2}. Production of interleukin 17A (IL-17A) and other inflammatory cytokines by $\gamma\delta$ T17 cells within hours of pathogen encounter orchestrates early neutrophil responses critical for mucocutaneous defence^{3–5}. However, dysregulated $\gamma\delta$ T17 cell responses contribute to pathogenesis associated with several models of autoimmunity and can enhance tumour growth and metastasis^{1,6–9}.

How $\gamma\delta$ T17 cells populate homeostatic barrier tissues and then infiltrate inflamed tissues from circulation is unclear. $\gamma\delta$ T17 cells seed dermis and mucosal tissues during perinatal life¹⁰. Although parabiosis experiments demonstrate that the majority of $V\gamma 4^+$ $\gamma\delta$ T17 cells in skin-draining lymph nodes (sLNs) are permanently resident¹¹, studies using photolabelling, adoptive transfers and receptor antagonism suggest that $\gamma\delta$ T17 cells constitutively circulate between dermis, sLNs and blood^{10,12–14}. Nevertheless, sLN $\gamma\delta$ T17 cells expand during autoimmune inflammation and infiltrate target tissues via circulation^{1,9}. Furthermore, dermal $V\gamma 4^+$ $\gamma\delta$ T17 cells home from skin to sLNs, proliferate, and repopulate inflamed and distal unaffected skin during psoriasis¹⁵. Thus despite a largely tissue-restricted distribution, $\gamma\delta$ T17 cells are motile and move between lymphoid and barrier tissues under homeostasis and experimental inflammatory conditions.

Chemokine receptor CCR6, involved in both homeostatic and inflammatory trafficking of leukocytes in barrier tissues, is expressed by both T helper 17 (Th17) and $\gamma\delta$ T17 cells^{16,17}. We reported a largely redundant function for CCR6 in recruitment of granulocyte-macrophage colony stimulating factor-producing encephalitogenic Th17 cells to the central nervous system (CNS) during experimental autoimmune encephalomyelitis (EAE). Instead, these cells display a $CCR6^- CCR2^+$ phenotype and infiltrate the CNS via CCR2, which is critical for T-cell-driven pathology¹⁸. In $\gamma\delta$ T17 cell biology, CCR6 has a debated function in regulating $V\gamma 4^+$ cell homeostasis, and is reported to direct $\gamma\delta$ T17 cell trafficking during inflammation^{10,11,19}. However, $V\gamma 4^+$ cells homing from inflamed skin to sLNs during psoriasis predominantly lack CCR6 expression¹⁴. By contrast, CCR2 is implicated in the migration of $\gamma\delta$ T17 cells to psoriatic skin and arthritic synovium^{15,20}, pointing to a clear interplay between CCR6 and CCR2 function in control of $\gamma\delta$ T17 cell homing. Nevertheless, a clear understanding of $\gamma\delta$ T17 cell trafficking mechanisms at rest and during inflammation is lacking.

Here, we find that CCR6 controls homeostatic $\gamma\delta$ T17 cell trafficking to the dermis, whereas constitutive CCR2 expression drives their rapid homing to inflammatory sites. In models of autoimmunity, cancer and infection, activation-induced downregulation of CCR6 releases $\gamma\delta$ T17 cells from their homeostatic immunosurveillance trafficking circuit through the skin and circulation, which then enhances their CCR2-dependent homing to inflamed tissue. Therefore, the dynamic interplay between CCR6 and CCR2 expression defines $\gamma\delta$ T17 cell trafficking patterns between resting and activated states.

Results

$\gamma\delta$ T17 cells downregulate CCR6 upon activation. We recently reported that Th17 cell development during EAE is coupled with a dynamic, temporally regulated switch from CCR6 to CCR2 expression as Th17 cells propagate their differentiation. Expression patterns of CCR6 and CCR2 define distinct effector

phenotypes of Th17 cells, with a $CCR6^- CCR2^+$ phenotype marking the encephalitogenic granulocyte-macrophage colony-stimulating factor/interferon- γ -producing population¹⁸. Unlike Th17 cells, $\gamma\delta$ T17 cell effector function is programmed during thymic development and these cells populate barrier tissues prior to inflammation^{2,21,22}. Thus, we initially examined CCR6 and CCR2 expression in sLN and dermis in unimmunized *Il17a^{Cre} × Rosa26^{eYFP}* mice, where *Il17a* expression drives permanent marking of cells with eYFP²³. $\gamma\delta$ T17 cells in these compartments constitutively co-expressed CCR2 and CCR6 (Fig. 1a and Supplementary Fig. 1a). Expression of CCR6 and CCR2 was restricted to $\gamma\delta$ T cells bearing a $CD27^- CD44^{hi}$ phenotype, characteristic of $\gamma\delta$ T17 cells (Supplementary Fig. 2a)²⁴. CCR6/CCR2 co-expression was similar between $V\gamma 4^+$ and $V\gamma 6^+$ $\gamma\delta$ T17 cell subsets as distinguished by both $V\gamma 4$ expression and CD3/T-cell receptor (TCR) expression level, as previously reported ('CD3^{bright} staining')²⁵ (Supplementary Fig. 1b,c), and both receptors were functional as determined by *ex vivo* chemotaxis (Fig. 1b). However, examination of $\gamma\delta$ T17 cells from diverse tissues revealed a heterogeneous pattern of CCR6 expression. While thymic and most lymphoid $\gamma\delta$ T17 cells uniformly expressed both CCR6 and CCR2, populations of $\gamma\delta$ T17 cells lacking CCR6 expression ($CCR6^- CCR2^+$) were prominent in lung and gut-associated tissues (Fig. 1c). As the gut is tonically immunologically active due to interactions with commensal microbiota, we hypothesized that $\gamma\delta$ T17 cells downregulate CCR6 expression during inflammation.

In support of this idea, activation of sLN and spleen $\gamma\delta$ T17 cells *in vivo* during EAE resulted in downregulation of CCR6 expression compared to unimmunized mice (Fig. 1d). CNS-infiltrating $\gamma\delta$ T17 cells were also largely $CCR6^-$. BrdU incorporation revealed that CCR6 expression was downregulated in proliferated $\gamma\delta$ T17 cells, while BrdU⁻ cells remained $CCR6^+$ (Fig. 1e). Other $\gamma\delta$ T-cell subsets did not express CCR2 or CCR6 at rest, and did not gain expression of these receptors over the course of EAE (Supplementary Fig. 2a). Unlike Th17 cells, $\gamma\delta$ T17 cells are predominantly activated by TCR-independent signals including IL-23 and IL-1 β ¹. *In vitro* stimulation of lymphocytes with a range of known stimuli including IL-23/IL-1 β , IL-23/IL-18 (ref. 26), IL-7 (ref. 27) and $\gamma\delta$ -TCR signalling²⁸ uniformly repressed CCR6 surface expression in $\gamma\delta$ T17 cells (Fig. 1f and Supplementary Fig. 2b). IL-12 did not impact CCR6 expression, consistent with a reported absence of IL-12R expression by $\gamma\delta$ T17 cells²⁹ (Supplementary Fig. 2b). Activation-induced CCR6 downregulation correlated with induction of activation markers CD69 and CD25 and increased CD44 expression, and occurred in both $V\gamma 4^+$ and $V\gamma 6^+$ $\gamma\delta$ T17 cells (Supplementary Fig. 2c,d). In all *in vivo* and *in vitro* systems, $\gamma\delta$ T17 cells maintained high levels of CCR2 following activation, and virtually all $\gamma\delta$ T17 cells were $CCR2^+$ (Fig. 1a,c,d,f). Therefore, $\gamma\delta$ T17 cells are programmed to co-express CCR6 and CCR2 during development, but lose CCR6 expression upon activation.

CCR2 drives $\gamma\delta$ T17 cell recruitment to inflamed tissues. Tissue-infiltrating $\gamma\delta$ T17 cells are best understood in the context of cancer and autoimmunity. $\gamma\delta$ T17 cells infiltrate B16 melanomas and promote tumour growth^{30,31}, and infiltrate the CNS at disease onset and exacerbate disease pathogenesis during EAE^{1,32}. How $\gamma\delta$ T17 cells infiltrate these inflammatory lesions is unknown. We thus used these models to investigate CCR6 and CCR2 function in control of $\gamma\delta$ T17 cell migration during inflammation. Consistent with the observation that activation induces downmodulation of CCR6 expression, *Ccr6*-deficiency did not affect $\gamma\delta$ T17 cell infiltration of B16 melanomas (Fig. 2a,b), nor recruitment to the CNS during EAE onset

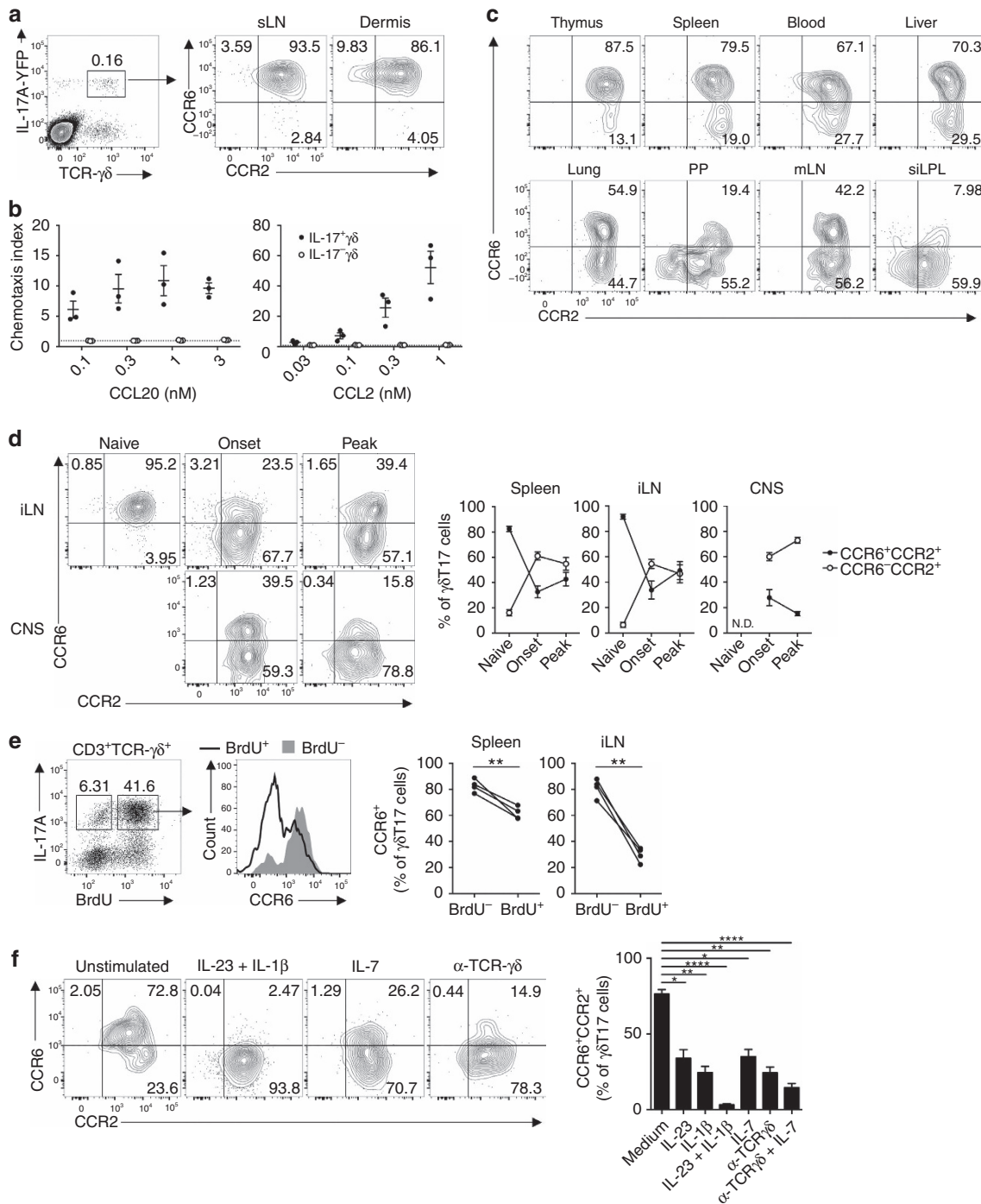


Figure 1 | $\gamma\delta$ T17 cells downregulate CCR6 upon activation. (a) Representative flow cytometry of CCR6 and CCR2 expression in skin-draining lymph nodes (sLN) and dermal CD3⁺TCR- $\gamma\delta$ ⁺IL-17A-YFP⁺ $\gamma\delta$ T17 cells from *Il17a*^{Cre} × *Rosa26*^{eYFP} mice (n = 3). (b) Ex vivo transwell chemotaxis of *Il17a*^{Cre} × *Rosa26*^{eYFP} splenic IL-17A^{+/-} $\gamma\delta$ T cells to CCL20 and CCL2 (n = 3). (c) Representative flow cytometry of CD45⁺ $\gamma\delta$ T17 cells from organs of naive *Il17a*^{Cre} × *Rosa26*^{eYFP} mice (n = 3). mLN, mesenteric lymph node; PP, Peyer’s patches; siLPL, small intestinal lamina propria lymphocytes. (d) Representative flow cytometry and quantitation of CCR6 and CCR2 expression by $\gamma\delta$ T17 cells from organs of *Il17a*^{Cre} × *Rosa26*^{eYFP} mice either naive (n = 6) or at experimental autoimmune encephalomyelitis (EAE) onset (n = 7) or peak (n = 5). CNS, central nervous system; iLN, inguinal lymph node; ND, not detected. (e) Representative flow cytometry and quantitation of CCR6 expression by $\gamma\delta$ T17 cells from wild type (WT) mice given BrdU at d3 post-immunization for EAE, and analysed at d8 (n = 4). (f) Representative flow cytometry and frequency of CCR6 and CCR2 expression by $\gamma\delta$ T17 cells from *Il17a*^{Cre} × *Rosa26*^{eYFP} lymphocytes cultured with indicated stimuli for 72 h (n = 5). See also Supplementary Figs 1 and 2. Mean ± s.e.m. (a–c) Representative of two experiments. (d,f) Pooled from two experiments. (e) Paired two-tailed Student’s t-test, (f) one-way paired ANOVA with Dunnett’s multiple comparisons test relative to unstimulated control. *P < 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001.

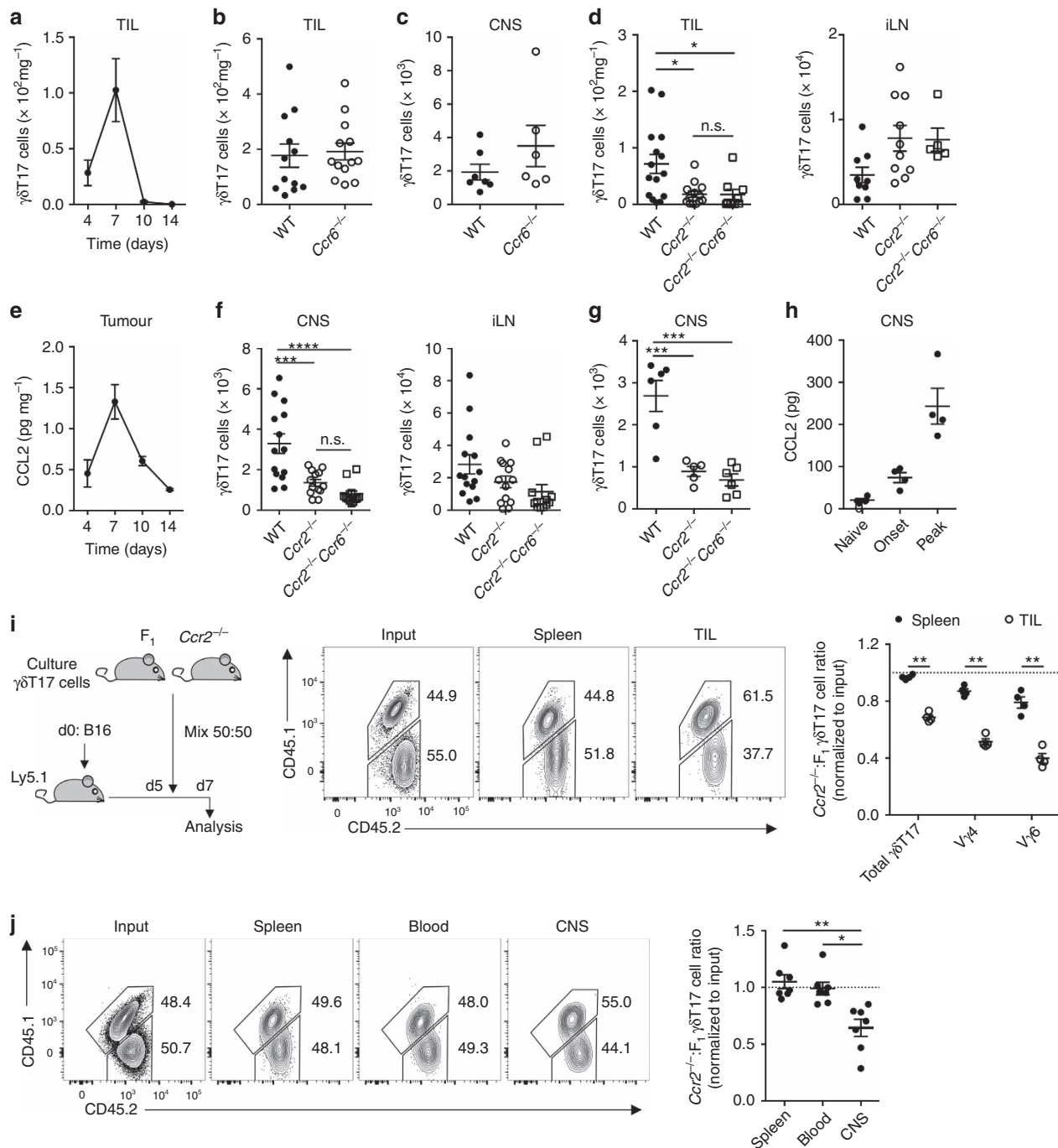


Figure 2 | CCR2 recruits $\gamma\delta T17$ cells to inflammatory sites. (a) $CD45^+ CD3^+ TCR\gamma\delta^+ IL-17A^+$ $\gamma\delta T17$ cell numbers in tumour-infiltrating lymphocytes (TIL) following B16 melanoma challenge ($n = 5/\text{time point}$). (b) $\gamma\delta T17$ cell numbers in TIL d7 post-challenge with B16 melanoma in wild type (WT) ($n = 12$) and $Ccr6^{-/-}$ mice ($n = 13$). (c) $\gamma\delta T17$ cell numbers in central nervous system (CNS) at experimental autoimmune encephalomyelitis (EAE) onset in WT ($n = 7$) and $Ccr6^{-/-}$ mice ($n = 6$). (d) $\gamma\delta T17$ cell numbers in TIL and inguinal lymph nodes (iLN) d7 post-challenge with B16 melanoma in WT ($n = 15$ (TIL), 9 (iLN)), $Ccr2^{-/-}$ ($n = 13$ (TIL), 10 (iLN)) and $Ccr2^{-/-}Ccr6^{-/-}$ mice ($n = 9$ (TIL), 5 (iLN)). (e) ELISA for CCL2 in tumour supernatant from WT mice challenged with B16 melanoma ($n = 5/\text{time point}$). (f) $\gamma\delta T17$ cell numbers in CNS and iLN at EAE onset in WT ($n = 14$), $Ccr2^{-/-}$ ($n = 13$) and $Ccr2^{-/-}Ccr6^{-/-}$ mice ($n = 12$). (g) $\gamma\delta T17$ cell numbers in CNS at peak disease in WT ($n = 6$), $Ccr2^{-/-}$ ($n = 5$) and $Ccr2^{-/-}Ccr6^{-/-}$ mice ($n = 6$). (h) ELISA for CCL2 in CNS of WT mice with EAE ($n = 4/\text{time point}$). (i) Ly5.1 mice ($n = 4$) at d5 post-challenge with B16 melanoma were transferred i.v. with *in vitro*-expanded $\gamma\delta T17$ cells from $Ccr2^{-/-}$ ($CD45.2^+$) and F_1 ($CD45.1^+ CD45.2^+$) mice. $Ccr2^{-/-}:F_1$ total, $V\gamma 4$ and $V\gamma 6$ $\gamma\delta T17$ cell ratios in spleen and tumours were normalized to input ratio. $V\gamma 4$ and $V\gamma 6$ $\gamma\delta T17$ cells were determined by $CD3^{\text{bright}}$ gating. Representative flow cytometry of $CD45.2^+ \gamma\delta T17$ cells at d7 or input. (j) Ly5.1 mice ($n = 7$) at EAE onset were transferred with F_1 and $Ccr2^{-/-}$ $\gamma\delta T17$ cells as in (i). Twenty-four hours later, ratios of $Ccr2^{-/-}:F_1 \gamma\delta T17$ cells in spleen, blood and CNS were normalized to input. Representative flow cytometry of $CD45.2^+ \gamma\delta T17$ cells 24 h later or input. See also Supplementary Figs 3 and 4. Mean \pm s.e.m. (a,c,e,i) Representative of two experiments. (b,d,f,j) Pooled from two experiments. (b,c) Unpaired two-tailed Student's *t*-test, (d,f-g,j) one-way ANOVA with Bonferroni's multiple comparisons test (paired in j), (i) paired two-tailed Student's *t*-test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.

(Fig. 2c). Thus, $\gamma\delta T17$ cell trafficking to inflamed tissues in these settings occurs independently of CCR6.

In contrast, deficiency of *Ccr2* abrogated $\gamma\delta T17$ cell infiltration of B16 melanomas but did not affect their expansion in draining lymph nodes (LNs) (Fig. 2d). CCR2-driven infiltration of $\gamma\delta T17$ cells was consistent with upregulation of its major ligand CCL2 in tumours (Fig. 2e). Similar results were found in EAE, where *Ccr2*-deficiency inhibited $\gamma\delta T17$ cell recruitment to the CNS at both onset and peak disease (Fig. 2f,g), time points at which CCL2 was induced in the CNS as reported³³ (Fig. 2h). CCR2 appeared to operate independently of CCR6 in regulation of $\gamma\delta T17$ cell trafficking, as compound deficiency of *Ccr6* and *Ccr2* (*Ccr6*^{-/-} *Ccr2*^{-/-}) did not further affect $\gamma\delta T17$ cell infiltration in either model (Fig. 2d,f,g).

Ccr2^{-/-} *Ccr6*^{-/-} mice exhibited enhanced tumour growth, while *Ccr2*^{-/-} and *Ccr2*^{-/-} *Ccr6*^{-/-} mice had decreased EAE severity (Supplementary Fig. 3). Therefore, to examine the cell-intrinsic requirements of CCR2 for $\gamma\delta T17$ cell migration, we developed a novel *in vitro* expansion protocol to generate large numbers of purified activated $\gamma\delta T17$ cells, which contained both $V\gamma 4^+$ and $V\gamma 6^+$ subsets and maintained functional CCR2 expression (Supplementary Fig. 4). Equal ratios of *in vitro*-expanded genetically marked wild type (WT) and *Ccr2*^{-/-} $\gamma\delta T17$ cells were co-transferred into B16 melanoma-bearing recipients. While donor $\gamma\delta T17$ cells recovered from spleen retained the input ratio, *Ccr2*^{-/-} $\gamma\delta T17$ cells did not migrate efficiently to tumours. This observation was true for both $V\gamma 4^+$ and $V\gamma 6^+$ subsets (Fig. 2i). Similar experiments using the EAE model revealed equivalent WT:*Ccr2*^{-/-} $\gamma\delta T17$ cell ratios in spleen and blood but reduced *Ccr2*^{-/-} $\gamma\delta T17$ cell recruitment to the CNS (Fig. 2j). Thus, CCR2, but not CCR6, drives activated $\gamma\delta T17$ cell migration to inflammatory sites during B16 melanoma and EAE.

CCR2 is essential for protective $\gamma\delta T17$ cell responses. The above models involve $\gamma\delta T17$ cell infiltration from circulation, as the CNS and tumours lack resident $\gamma\delta T17$ cell populations. However, many inflammatory scenarios implicate tissue-resident $\gamma\delta T17$ cells, which survey and rapidly defend against infection at barrier surfaces. The extent to which $\gamma\delta T17$ cell migration contributes to host defence during ongoing inflammation is unknown. To investigate whether CCR2 also directs tissue-infiltrating $\gamma\delta T17$ cells during infection, we used experimental *Streptococcus pneumoniae* infection, immunity to which requires $\gamma\delta T17$ cells⁴. Accordingly, *Tcrd*^{-/-} mice had higher bacterial burden and reduced neutrophils in the nasal wash (NW) than WT at 72h post-infection (Fig. 3a,b). *S. pneumoniae* infection induced $\gamma\delta T17$ cell expansion in draining LNs and nasal-associated lymphoid tissue (Fig. 3c). CCL2 was induced in the nasal passages (NPs) upon infection (Fig. 3d), and co-transfer of *in vitro*-expanded WT and *Ccr2*^{-/-} $\gamma\delta T17$ cells into infected mice revealed an intrinsic requirement of CCR2 for $\gamma\delta T17$ cell accumulation in NP (Fig. 3e). Thus, CCR2 drives circulating $\gamma\delta T17$ cell infiltration of mucosal tissue during *S. pneumoniae* infection.

To elucidate the ability of recruited $\gamma\delta T17$ cells to control infection, we transferred purified *in vitro*-expanded $\gamma\delta T17$ cells into *Tcrd*^{-/-} hosts prior to *S. pneumoniae* infection. In this model, tissue-infiltrating $\gamma\delta T17$ cells provide the only source of $\gamma\delta$ T-cell-driven protection. Transfer of WT $\gamma\delta T17$ cells reduced nasopharyngeal bacterial burden by ~10-fold, whereas *Ccr2*^{-/-} $\gamma\delta T17$ cells completely failed to control infection (Fig. 3f). Hence, CCR2 drives recruitment of protective $\gamma\delta T17$ cells to the nasal mucosa during *S. pneumoniae* infection. Collectively, we conclude that $\gamma\delta T17$ cell trafficking to diverse inflamed tissues is critically dependent on CCR2 signalling.

CCR6 regulates homeostatic positioning of $\gamma\delta T17$ cells.

Expression of CCR6 during $\gamma\delta T17$ cell thymic development followed by rapid downregulation upon activation suggests that CCR6 plays a more prominent function in regulation of $\gamma\delta T17$ cell homeostasis. While CCL20 is induced during inflammation, it is constitutively expressed in barrier tissues including skin, Peyer's patches and large intestine³⁴⁻³⁷. Both *Ccr6*^{-/-} and *Ccr2*^{-/-} *Ccr6*^{-/-} mice had markedly reduced number and frequency of $\gamma\delta$ T cells expressing intermediate amounts of CD3/TCR in the dermis ($\gamma\delta T10$), a population previously reported to produce IL-17 and distinct from TCR^{hi} dendritic epidermal T cells²² (Fig. 4a). We confirmed that $\gamma\delta T10$ cells were entirely marked by eYFP in *Il17a*^{Cre} × *Rosa26*^{eYFP} mice, despite negligible IL-17A production following *ex vivo* restimulation (Supplementary Fig. 5a). In contrast to a previous report¹⁰, *Ccr6*-deficiency reduced the number of both $V\gamma 4^+$ and $V\gamma 4^-$ ($V\gamma 6^+$) $\gamma\delta T10$ cells, although the ratio was skewed slightly towards $V\gamma 6^+$ cells (Fig. 4b). Examination of other organs revealed that deficiency in *Ccr2* had no effect on $\gamma\delta T17$ cell homeostasis, while *Ccr6*-deficiency increased $\gamma\delta T17$ cells in the peritoneal cavity (Supplementary Fig. 5b). We conclude that CCR6 regulates dermal $\gamma\delta T17$ cell residence.

To determine whether CCR6 drives recruitment of circulating $\gamma\delta T17$ cells into dermis, we transferred unstimulated WT or *Ccr6*^{-/-} lymphocytes into naive mice and tracked their accumulation in dermis. Transferred WT $\gamma\delta T17$ cells were substantially enriched in dermis, demonstrating that $\gamma\delta T17$ cells can constitutively populate the skin from circulation. In contrast, *Ccr6*^{-/-} $\gamma\delta T17$ cells were defective in infiltration of dermis and pooled in the blood (Fig. 4c). In support of earlier results, both $V\gamma 4^+$ and $V\gamma 4^-$ $\gamma\delta T17$ cells were recruited to the dermis, the ratio of which was unaltered by *Ccr6* deficiency, suggesting both populations are dependent on CCR6 for circulation-to-dermis trafficking (Fig. 4c). While constitutive expression of CCL20 in epidermis was reported, whether it is expressed in uninflamed dermis is unclear^{34,38}. We found that *Ccl20* was constitutively expressed in the dermis by an uncharacterized CD31⁻ CD90⁻ CD140 α ⁻ stromal population (Fig. 4d). Thus, CCR6 directs homeostatic recruitment of $\gamma\delta T17$ cells from circulation into dermis.

IRF4 and BATF regulate CCR6 expression in $\gamma\delta T17$ cells.

The downregulation of CCR6 upon $\gamma\delta T17$ cell activation is surprising, as T cells typically upregulate inflammatory chemokine receptors upon activation. Consequently we investigated the underlying mechanism regulating this process. *Ccr6* transcript levels were reduced by approx. fourfold in $\gamma\delta T17$ cells within 24h of stimulation, whereas *Ccr2* expression was maintained (Fig. 5a and Supplementary Fig. 6a). This indicated that CCR6 expression is transcriptionally regulated during $\gamma\delta T17$ cell activation. We thus examined expression of transcription factors previously implicated directly or indirectly in control of *Ccr6* expression, including ROR γ t¹⁷, IRF4 (ref. 39), IRF8 (refs 40,41), Blimp1 (refs 39,42), BATF⁴³ and T-bet and Eomes¹⁸. *Rorc* (ROR γ t) was highly expressed in resting $\gamma\delta T17$ cells but was downregulated by 24h of activation. *Batf* and *Prdm1* (Blimp1) were rapidly upregulated by 24h, while *Irf8* and *Irf4* were upregulated by 48h, although *Irf4* was already present in resting $\gamma\delta T17$ cells. Expression of *Eomes* and *Tbx21* (T-bet) at rest or following activation was minimal (Fig. 5b and Supplementary Fig. 6b). Therefore, we tested whether ROR γ t, IRF4, BATF, Blimp1 or IRF8 repressed *Ccr6* expression during $\gamma\delta T17$ cell activation.

The similar expression kinetics and known *Ccr6* regulatory activity of ROR γ t presented the possibility that its downregulation may result in loss of CCR6 expression. To test this,

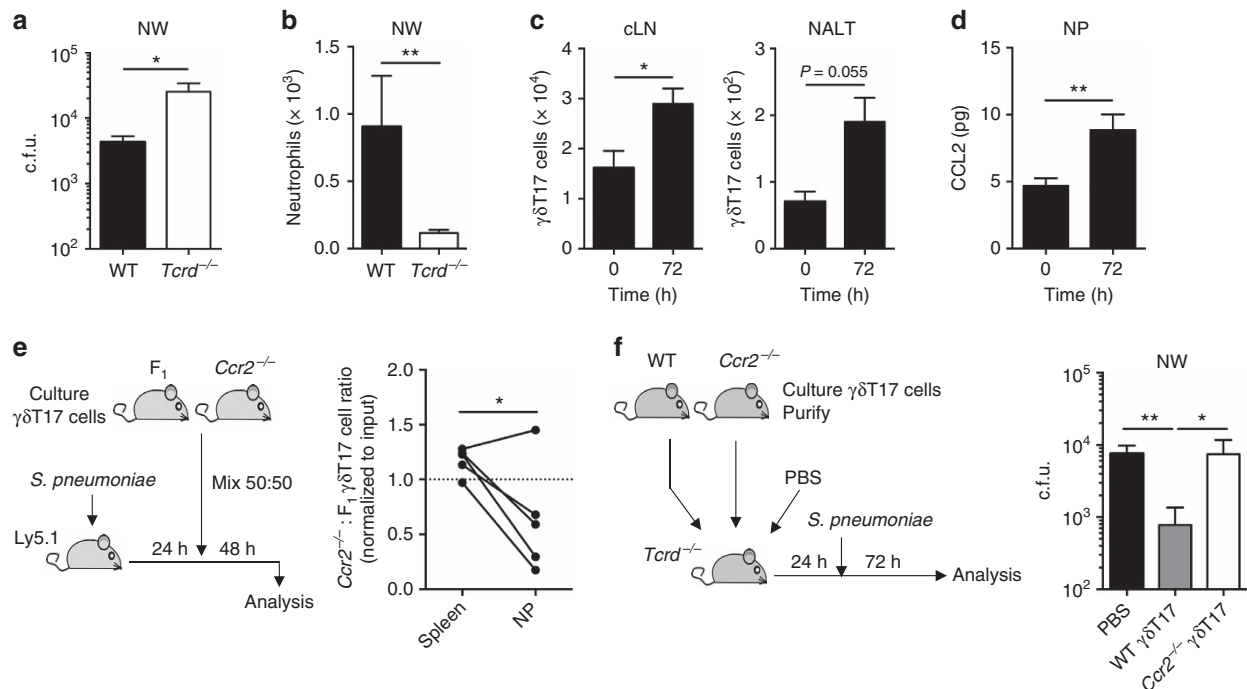


Figure 3 | CCR2 drives protective $\gamma\delta$ T17 cell responses. (a) Colony-forming units (c.f.u.) and (b) CD45⁺CD11b⁺Ly6G⁺ neutrophils recovered from nasal wash (NW) of wild type (WT) ($n=9$) and *Tcrd*^{-/-} mice ($n=10$) 72 h post-infection with *S. pneumoniae*. (c) $\gamma\delta$ T17 cell numbers in cervical lymph node (cLN) and nasal-associated lymphoid tissue (NALT) and (d) ELISA for CCL2 in digested nasal passage (NP) supernatant in unimmunized mice ($n=7$) and at 72 h post-*S. pneumoniae* infection ($n=13$). (e) Ly5.1 mice ($n=5$) 24 h post-*S. pneumoniae* infection were transferred i.v. with expanded $\gamma\delta$ T17 cells from *Ccr2*^{-/-} (CD45.2⁺) and *F*₁ (CD45.1⁺CD45.2⁺) mice. The *Ccr2*^{-/-}:*F*₁ $\gamma\delta$ T17 cell ratio in spleen and NP was normalized to input ratio. (f) Twenty-four hours prior to *S. pneumoniae* infection, *Tcrd*^{-/-} hosts received PBS ($n=8$) or expanded and purified $\gamma\delta$ T17 cells from WT ($n=9$) or *Ccr2*^{-/-} ($n=7$) mice. c.f.u. recovered from NW 72 h post-infection. Mean \pm s.e.m. (a–d) Pooled from two experiments. (a,b) Mann-Whitney test, (c,d) unpaired two-tailed Student's *t*-test, (e) paired two-tailed Student's *t*-test, (f) Kruskal-Wallis test with Dunn's multiple comparisons test. * $P<0.05$, ** $P<0.01$.

we retrovirally forced *Rorc* expression in *in vitro*-expanded $\gamma\delta$ T17 cells (Fig. 5c). However, this failed to alter CCR6 expression even in the highest GFP-expressing cells, suggesting that ROR γ t downregulation is not required for repression of CCR6 in activated $\gamma\delta$ T17 cells (Fig. 5d).

To determine whether IRF4, BATF, IRF8 or Blimp1 actively repress *Ccr6* expression, we cultured genetically-marked WT and transcription factor-deficient splenocytes with IL-23 and IL-1 β . $\gamma\delta$ T17 cells were present in all strains although at differing frequencies, and homeostatic CCR6 expression was comparable to WT (Supplementary Fig. 6c). IRF4- and BATF-deficient $\gamma\delta$ T17 cells were intrinsically defective in both proliferation and CCR6 downregulation upon stimulation (Fig. 5e). IRF8 and Blimp1 were not required for these processes, although Blimp1 appeared to moderately promote CCR2 expression in activated $\gamma\delta$ T17 cells (Supplementary Fig. 6d). *Irf4*^{-/-} and *Batf*^{-/-} $\gamma\delta$ T17 cells exhibited comparable surface expression of IL-23R and IL-1R1 to WT cells, indicating that maintained CCR6 expression was likely due to defective signalling downstream of IL-23 and IL-1 β stimulation (Supplementary Fig. 7a). Analysis of our and others' ChIP-Seq datasets in T cells^{43–45} revealed binding of IRF4 and BATF to a shared site in the *Ccr6* promoter (Supplementary Fig. 7b), suggesting that these factors cooperatively and directly repress *Ccr6* in $\gamma\delta$ T17 cells. To assess whether defective proliferation in absence of IRF4 or BATF was the cause of impaired CCR6 downregulation, dye-labelled WT splenocytes were pre-treated with proliferation inhibitor mitomycin C prior to stimulation. Although proliferation was effectively blocked, CCR6 downregulation still occurred upon mitomycin C treatment, suggesting that proliferation and CCR6

downregulation are coincident but independent (Fig. 5f). Together, these data indicate that activation-induced CCR6 downregulation in $\gamma\delta$ T17 cells is promoted by IRF4 and BATF, and is largely uncoupled from proliferation.

Loss of CCR6 promotes $\gamma\delta$ T17 cell homing to inflamed tissues.

Given the constitutive expression of CCL20 in mucocutaneous sites, we hypothesized that repression of CCR6 during activation enables homing of $\gamma\delta$ T17 cells toward inflammatory lesions by preventing their accumulation in uninfamed skin. To test this, we first compared the trafficking of *in vitro*-activated WT $\gamma\delta$ T17 cells with resting WT and *Ccr6*^{-/-} $\gamma\delta$ T17 cells upon transfer into unimmunized hosts. Activated WT $\gamma\delta$ T17 cells demonstrated the same defect in homing to the dermis as resting *Ccr6*^{-/-} $\gamma\delta$ T17 cells, and both pooled in blood compared to resting WT $\gamma\delta$ T17 cells (Fig. 6a). $\gamma\delta$ T17 cells lack CD62L and CCR7 expression, and traffic from skin to sLNs in a CCR7-independent manner¹². Thus $\gamma\delta$ T17 cell entry to sLNs following adoptive transfer likely occurs via afferent lymph draining from dermis. In keeping with this idea, resting *Ccr6*^{-/-} or *in vitro*-activated WT $\gamma\delta$ T17 cells, impaired in their ability to home to uninfamed skin, also accumulated less than resting WT $\gamma\delta$ T17 cells in sLNs (Fig. 6a). These data are consistent with the notion that activation switches off $\gamma\delta$ T17 cell homeostatic circulation patterns, enabling directed migration toward inflammatory cues.

To investigate this proposal directly, we studied the migratory patterns of *in vitro*-activated $\gamma\delta$ T17 cells retrovirally forced to maintain CCR6 expression. Infection with *Ccr6*^{tg} virus restored CCR6 expression in activated $\gamma\delta$ T17 cells, which regained the

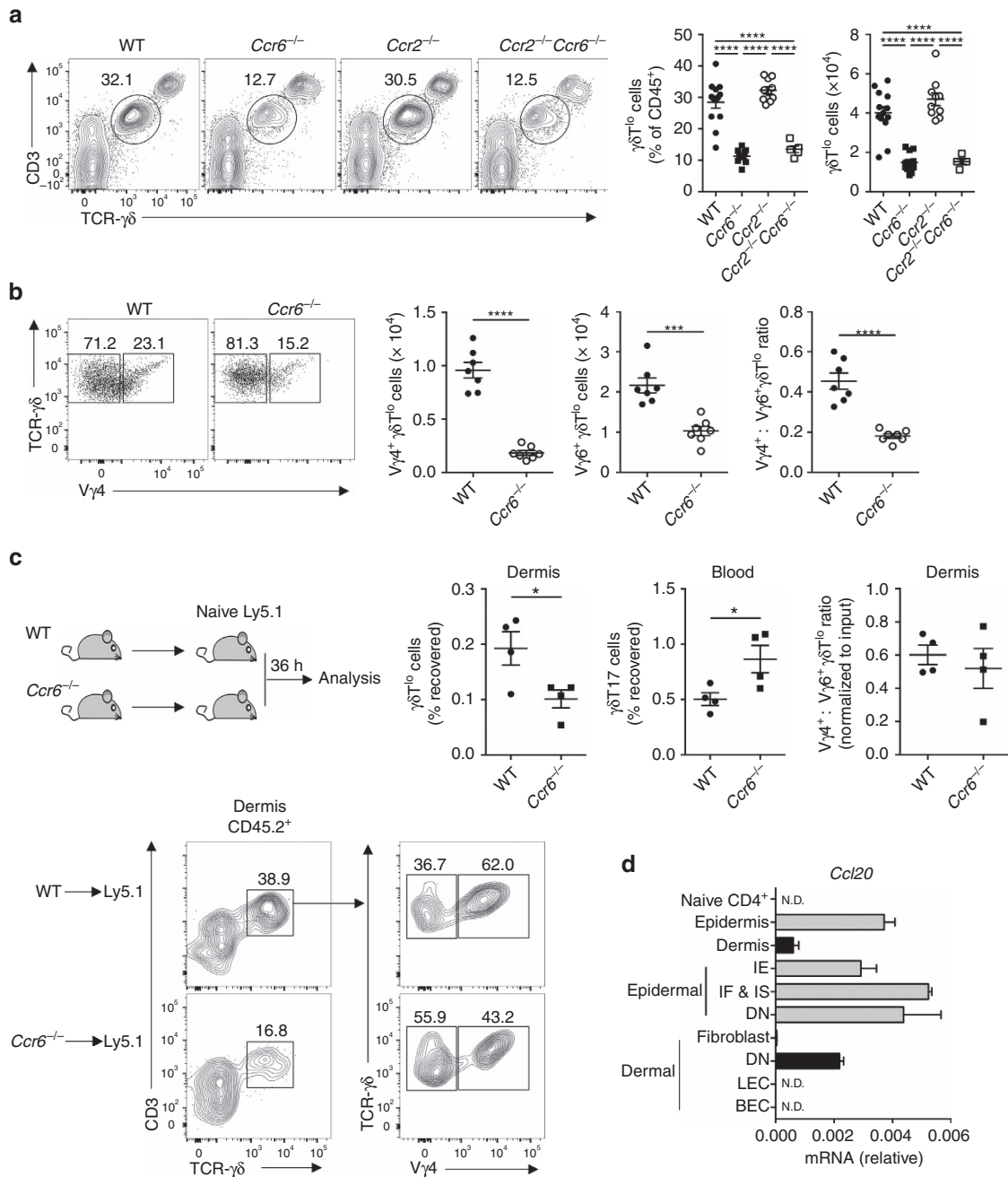


Figure 4 | CCR6 regulates homeostatic $\gamma\delta T17$ cell recruitment to dermis. (a) Representative flow cytometry and quantitation of CD45⁺ CD3^{lo}TCR- $\gamma\delta^{lo}$ ($\gamma\delta T^{10}$) cells from ear skin dermis of naïve wild type (WT) ($n = 13$), $Ccr6^{-/-}$ ($n = 11$), $Ccr2^{-/-}$ ($n = 10$) and $Ccr2^{-/-} Ccr6^{-/-}$ mice ($n = 5$). (b) Representative flow cytometry of V $\gamma 4$ expression by dermal $\gamma\delta T^{10}$ cells and quantitation of V $\gamma 4^+$ and V $\gamma 4^-$ $\gamma\delta T^{10}$ cells in dermis of WT and $Ccr6^{-/-}$ mice ($n = 7$ /group). (c) WT or $Ccr6^{-/-}$ lymphocytes were transferred i.v. into naïve Ly5.1 mice ($n = 4$ /group). After 36 h, number of CD45.2⁺ $\gamma\delta T^{10}$ / $\gamma\delta T^{17}$ cells recovered was expressed as % of number transferred. Representative flow cytometry of dermal CD45.2⁺ cells and quantitation of $\gamma\delta T^{17}$ cell recovery and V $\gamma 4^+$: V $\gamma 4^-$ ratio (normalized to input). (d) *Ccl20* mRNA from whole tissues or sorted CD45⁻ epidermal keratinocytes (Sca-1⁺Ep-CAM^{lo} interfollicular epidermis (IE), Sca-1⁺Ep-CAM⁺ infundibulum and isthmus (IF & IS), Sca-1^{lo}Ep-CAM^{lo} double negative (DN)) or CD45⁻ dermal populations (CD31⁻CD90⁺CD140 α ⁺ fibroblast, gp38⁺CD31⁺ lymphatic endothelial cells (LEC), gp38^{lo}CD31⁺ blood endothelial cells (BEC), CD31⁻CD90⁻CD140 α ⁻ double negative (DN)) from naïve WT mice (pooled from 5 mice/experiment). ND, not detected. See also Supplementary Fig. 5. Mean \pm s.e.m. (a,d) Pooled from three experiments, (c) representative of two similar experiments. (a) One-way ANOVA with Bonferroni's multiple comparisons test, (b,c) unpaired two-tailed Student's *t*-test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.

ability to migrate toward CCL20 (Fig. 6b,c). Genetically marked control- and $Ccr6^{tg}$ -transduced $\gamma\delta T17$ cells were mixed 50:50 and transferred into B16 melanoma-bearing recipients. While the input ratio of transferred GFP⁻ cells was maintained in all examined organs as expected, among GFP⁺ cells, $Ccr6^{tg}$ $\gamma\delta T17$

cells were enriched in the dermis but deficient in tumours (Fig. 6d). Similar results were observed during *S. pneumoniae* infection: $Ccr6^{tg}$ $\gamma\delta T17$ cells were selectively deficient at homing to NP, but accumulated to a greater extent than control-transduced cells in uninflamed dermis (Fig. 6e). $Ccr6^{tg}$ $\gamma\delta T17$

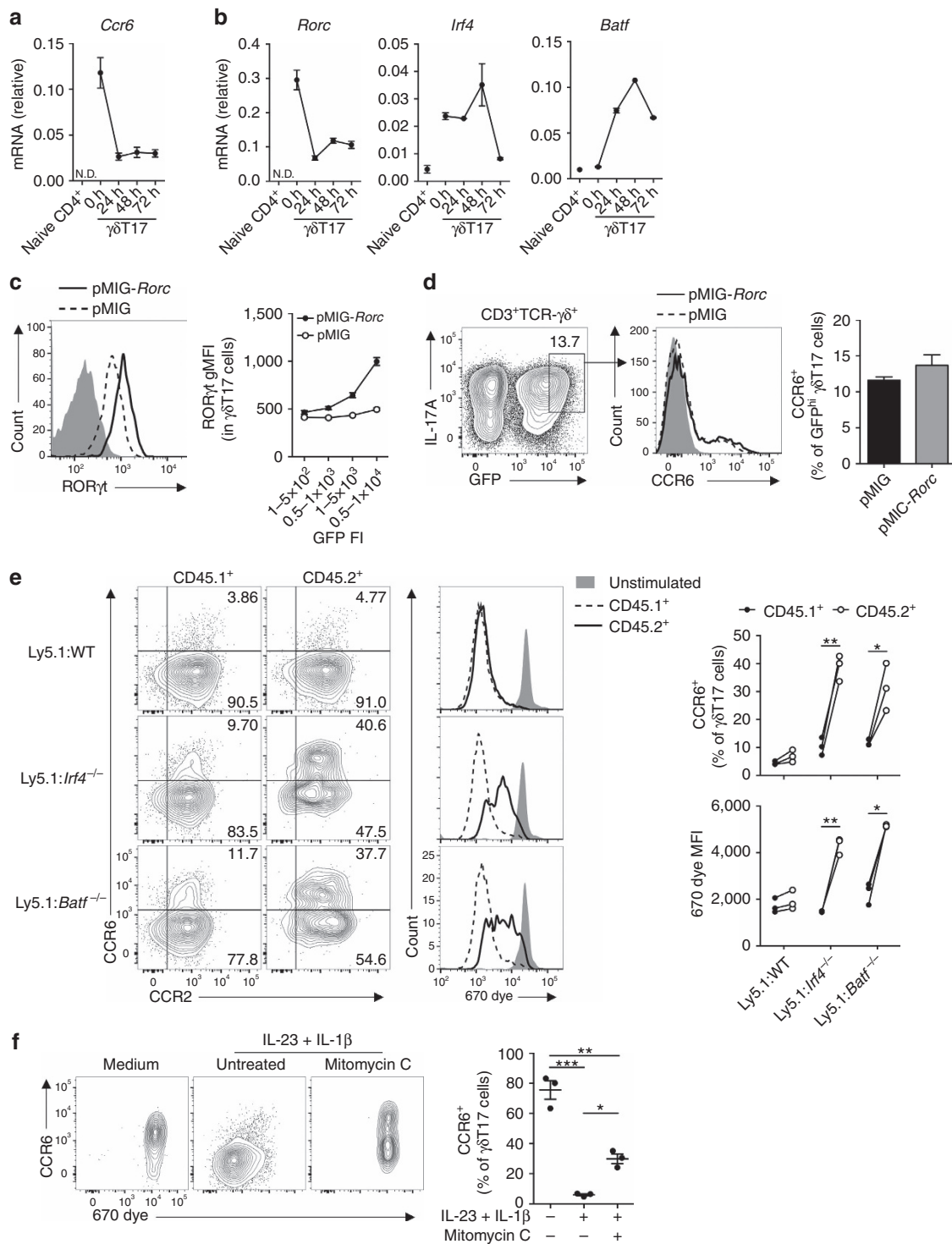


Figure 5 | IRF4 and BATF promote CCR6 downregulation in $\gamma\delta$ T17 cells. (a) *Ccr6* and (b) transcription factor mRNA in sorted $\gamma\delta$ T17 cells from *Irf4*^{Cre} \times *Rosa26*^{eYFP} lymphocytes ex vivo or cultured with IL-23/IL-1 β for indicated times (pooled from 5 to 7 mice). ND, not detected. (c,d) Expanded $\gamma\delta$ T17 cells ($n = 3$) were transduced with empty pMIG or pMIG-*Rorc* retrovirus. (c) Representative flow cytometry of ROR γ t expression in GFP^{hi} $\gamma\delta$ T17 cells (gated as in d), relative to isotype (grey) and geometric mean fluorescence intensity (gMFI) relative to GFP fluorescence intensity (FI). (d) Representative flow cytometry of CCR6 expression and quantitation in GFP^{hi} $\gamma\delta$ T17 cells. (e) Splenocytes from Ly5.1 and either wild type (WT), *Irf4*^{-/-} or *Batf*^{-/-} mice were 670 dye-labelled, mixed 50:50 and stimulated with IL-23/IL-1 β for 72 h. Representative flow cytometry and quantitation of CCR6 expression and proliferation in CD45.1⁺ or CD45.2⁺ $\gamma\delta$ T17 cells ($n = 3$ /group). (f) Representative flow cytometry and quantitation of CCR6 expression by 670 dye-labelled $\gamma\delta$ T17 cells from WT splenocytes cultured with IL-23/IL-1 β for 72 h with/without mitomycin C pre-treatment ($n = 3$). See also Supplementary Figs 6 and 7. (a,b) Mean \pm s.d., (c-f) Mean \pm s.e.m. (a-f) Representative of two similar experiments. (d,e) Paired two-tailed Student's *t*-test, (f) one-way paired ANOVA with Bonferroni's multiple comparisons test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$.

cells also homed less efficiently to the CNS during EAE, although subcutaneous complete Freund's adjuvant immunization precluded analysis of homing to uninfamed skin in this model

(Fig. 6f). Together, these experiments demonstrated that activated $\gamma\delta T17$ cells with forced CCR6 expression were recruited to uninfamed dermis at the expense of homing to inflamed tissue.

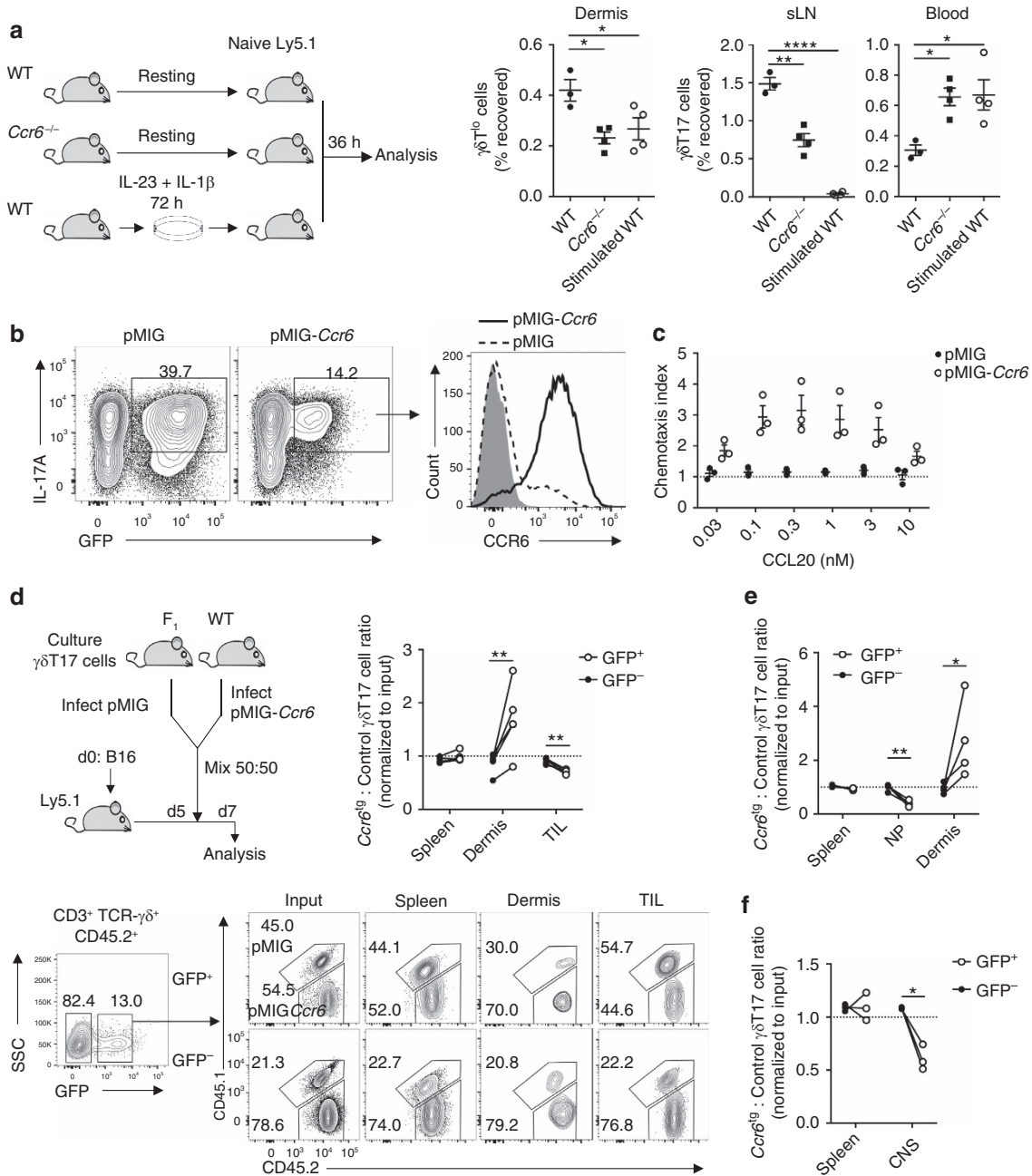


Figure 6 | CCR6 downregulation by $\gamma\delta T17$ cells enhances migration to inflamed tissue. (a) Resting lymphocytes from wild type (WT) ($n = 3$) or *Ccr6*^{-/-} ($n = 4$) mice, or WT lymphocytes stimulated with IL-23/IL-1 β for 72 h ($n = 4$) were transferred i.v. into separate naive Ly5.1 hosts. After 36 h, number of CD45.2⁺ $\gamma\delta T17$ cells recovered was expressed as % of number transferred. sLN, skin-draining lymph node. (b) Representative flow cytometry for CCR6 expression by GFP⁺ *in vitro*-expanded $\gamma\delta T17$ cells transduced with empty pMIG or pMIG-*Ccr6*, relative to isotype (grey) ($n = 3$). (c) Chemotaxis of GFP⁺ $\gamma\delta T17$ cells transduced as in (b) to CCL20 ($n = 3$). (d) *In vitro*-expanded $\gamma\delta T17$ cells from F₁ (CD45.1⁺CD45.2⁺) or WT (CD45.2⁺) mice were transduced with empty pMIG or pMIG-*Ccr6*, respectively. Equal numbers of mixed GFP⁺ cells were transferred i.v. into Ly5.1 mice challenged with B16 melanoma 5 days prior and analysed at d7 ($n = 5$). Representative flow cytometry and ratio of recovered F₁ to WT $\gamma\delta T17$ cells within transduced (GFP⁺) and untransduced (GFP⁻) populations. Recovered values were normalized to input values. TIL, tumour-infiltrating lymphocytes. (e,f) *In vitro*-expanded $\gamma\delta T17$ cells from WT or F₁ mice were transduced with empty pMIG or pMIG-*Ccr6*, respectively. Equal numbers of mixed GFP⁺ cells were transferred i.v. into Ly5.1 mice either (e) 24 h post-infection with *S. pneumoniae* ($n = 4$) or (f) at experimental autoimmune encephalomyelitis (EAE) onset ($n = 3$) and organs were analysed 48 h later. Ratio of recovered WT to F₁ $\gamma\delta T17$ cells within transduced (GFP⁺) and untransduced (GFP⁻) populations, normalized to input values. CNS, central nervous system; NP, nasal passage. Mean \pm s.e.m. (a) Representative of three similar experiments, (b,d) representative of two experiments. (a) One-way ANOVA with Dunnett's multiple comparisons test relative to resting WT $\gamma\delta T17$ cells, (d-f) paired two-tailed Student's *t*-test. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.

Thus, CCR6 downregulation promotes $\gamma\delta$ T17 cell migration to inflammatory sites.

Discussion

In the present study we define the molecular regulation of $\gamma\delta$ T17 cell trafficking between resting and activated states. Our data are consistent with a model in which $\gamma\delta$ T17 cells are imprinted with expression of both CCR6 and CCR2 during thymic development. CCR6 coordinates steady-state recruitment of circulating $\gamma\delta$ T17 cells into the dermis, where CCL20 is constitutively expressed, thus orchestrating their homeostatic recirculation. Upon activation, $\gamma\delta$ T17 cells rapidly downregulate CCR6 in an IRF4- and BATF-dependent manner. CCR2 expression is maintained and drives homing of activated $\gamma\delta$ T17 cells to inflamed tissue during autoimmunity, cancer and infection. CCR6 downregulation is required to promote optimal recruitment of activated $\gamma\delta$ T17 cells to inflamed tissue, by preventing their diversion and sequestration into uninfamed skin. These data identify a novel mode of lymphocyte trafficking that facilitates both $\gamma\delta$ T17 cell innate-like surveillance of host tissues and their rapid recruitment to distal sites of ongoing inflammation.

Our data demonstrate that CCR6 coordinates homeostatic recirculation of $V\gamma 4^+$ and $V\gamma 6^+$ $\gamma\delta$ T17 cells through blood, skin and sLNs. Given that $\gamma\delta$ T17 cells likely populate sLNs via afferent lymph^{12,14}, our data suggest that entry is contingent on first trafficking through the dermis, as *Ccr6*-deficient $\gamma\delta$ T17 cells fail to localize to both organs. Thus, in addition to driving infiltration of skin, CCR6 ultimately regulates $\gamma\delta$ T17 cell homing to sLNs. These findings complement a recent report that CCR6 positions sLN $V\gamma 4^+$ $\gamma\delta$ T17 cells in the subcapsular sinus¹¹. Our findings are also in agreement with previous reports of steady-state $\gamma\delta$ T17 cell circulation, although we show that in addition to $V\gamma 4^+$, $V\gamma 6^+$ $\gamma\delta$ T17 cells also undergo constitutive trafficking^{10,13,14}. While most $V\gamma 4^+$ $\gamma\delta$ T17 cells in sLNs do not recirculate within a 2-week period, approximately 25% undergo extensive circulation¹¹. The non-recirculating population of sLN $\gamma\delta$ T17 cells is likely those positioned in interfollicular zones and subcapsular sinus, which were proposed to scan for sLN-invading microbes^{11,46}. However, the function of the recirculating population of $\gamma\delta$ T17 cells is unclear. As $\gamma\delta$ T17 cells do not need to scan sLNs for antigen like conventional T cells, why they adopt a constitutive CCR6-dependent circulation loop between tissues and sLNs remains to be resolved.

We found that programmed expression of CCR2 equips $\gamma\delta$ T17 cells with the ability to rapidly home from circulation into diverse inflammatory environments. Our description of CCR2-driven $\gamma\delta$ T17 cell infiltration of the autoimmune CNS reflects other reports during psoriasis and arthritis^{15,20}. Additionally, CCR2-mediated infiltration of both $V\gamma 4^+$ and $V\gamma 6^+$ $\gamma\delta$ T17 cells into B16 melanomas suggests that this axis is a universal inflammatory homing signal for $\gamma\delta$ T17 cells. Our data reveal a redundant function for CCR6 in these processes, due to its immediate downregulation upon $\gamma\delta$ T17 cell activation. However, these data are inconsistent with reports of CCR6-driven migration of $\gamma\delta$ T17 cells during skin and liver inflammation^{19,47}. Tissue-specific signals directing maintained CCR6 expression in activated $\gamma\delta$ T17 cells in these particular scenarios could possibly explain discrepancies between these reports and our data, although this requires further investigation. The function of tissue-infiltrating $\gamma\delta$ T17 cells during inflammation in barrier tissues, where a resident $\gamma\delta$ T17 cell population already exists, is unclear. This phenomenon was previously reported during psoriasis, although did not appear to affect disease^{14,48}. Here, we show that $\gamma\delta$ T17 cells expand in draining LNs and infiltrate nasal mucosa via CCR2 during bacterial infection. In absence of endogenous $\gamma\delta$ T

cells, transferred circulating $\gamma\delta$ T17 cells are able to control infection in a CCR2-dependent manner. Thus, we propose that LN-expanded $\gamma\delta$ T17 cells home to lesions via CCR2 to supplement the local $\gamma\delta$ T17 cell pool during ongoing tissue inflammation.

Along with our previous work, this study identifies a shared chemokine receptor program used by IL-17-producing cells. We show that like Th17 cells¹⁸, $\gamma\delta$ T17 cells express CCR6 early during their effector program, but lose CCR6 expression via IL-23 signalling during ongoing inflammation. Instead, CCR2 is the defining IL-17-program inflammatory homing signal. Whether CCR6 repression in Th17 cells occurs to disrupt barrier tissue-homing and promote recruitment to inflamed tissue, as we show in $\gamma\delta$ T17 cells, is unclear. Despite $\gamma\delta$ T17 cell development occurring independently of IRF4 and BATF^{49,50}, we report that these factors suppress *Ccr6* expression in $\gamma\delta$ T17 cells upon IL-23- and IL-1 β -driven activation. This is likely to be cooperative, as has been shown during Th17 cell development⁴³. Moreover, we identified a common binding site for IRF4 and BATF in the *Ccr6* promoter, suggesting that repression of *Ccr6* expression is directly mediated by these factors. It remains to be seen if a similar mechanism operates in Th17 cells. Therefore, while $\gamma\delta$ T17 and Th17 cells both lose CCR6 expression during inflammation, the mechanism and function of this process may differ between these populations.

Investigating the trafficking of human $\gamma\delta$ T17 cells is of clinical relevance as they are increasingly implicated in autoimmunity and cancer, and it will be important to determine whether the model we have established here for murine $\gamma\delta$ T17 cells applies to humans. Of interest, CCR6 is known to be expressed by both $V\delta 1^+$ and $V\delta 2^+$ $\gamma\delta$ T17 cells in humans, as well as by circulating $V\delta 2^+$ cells with a skin-homing CLA⁺ phenotype^{51–53}. CCR2 expression has been identified in human $\gamma\delta$ T cells, but to our knowledge this has not been examined specifically in IL-17⁺ cells^{54,55}. Whether human $\gamma\delta$ T17 cells also undergo activation-induced CCR6 downregulation to enhance inflammatory homing has yet to be determined. While the human dermal $\gamma\delta$ T17 cell compartment is relatively small compared to the murine system, the relative abundance of circulating skin-homing $V\delta 2^+$ cells suggests that similar recirculation mechanisms may also operate in humans^{8,52}. These issues await further experimental resolution.

Our description of $\gamma\delta$ T17 cell trafficking between homeostasis and inflammation presents a novel mode of lymphocyte migration. Conventional T cells downregulate the homeostatic recirculation signal CCR7 and induce expression of inflammatory chemokine receptors upon differentiation into effector subsets, a slow process. In contrast, we show that $\gamma\delta$ T17 cells constitutively co-express homeostatic and inflammatory homing receptors. The switch in $\gamma\delta$ T17 cell trafficking from homeostatic to inflammatory programs is solely driven by downregulation of the homeostatic receptor CCR6, rather than induction of additional inflammatory homing receptors. This model likely facilitates immediate homing to inflammatory sites in addition to homeostatic scanning behaviour, consistent with the 'activated-but-resting' phenotype of $\gamma\delta$ T17 cells. Use of CCR6 expression to distinguish resting and activated states may facilitate future investigation of $\gamma\delta$ T17 cell biology. We conclude that $\gamma\delta$ T17 cells exhibit a unique bi-phasic trafficking program driven by programmed changes in homing receptor expression to facilitate tissue sentinel responses and rapid homing to distal inflammatory sites.

Methods

Mice. C57Bl/6 (WT) and Ly5.1 mice were purchased from Animal Resource Centre (WA, Australia) or bred at the University of Adelaide animal facility. *Il17a^{Cre} × Rosa26^{eYFP}*, *Ccr6^{-/-}*, *Ccr2^{-/-}*, *Ccr2^{-/-} Ccr6^{-/-}*, *Tcrd^{-/-}* and

C57Bl/6 \times Ly5.1 (F₁) mice were bred at the University of Adelaide animal facility. *Irf4*^{-/-}, *Irf8*^{-/-}, *Batf*^{-/-} and *Lck*^{Cre}*Prdm11*^{fl/fl} mice were bred at the WEHI animal facility. Mice were age- and gender-matched and used at 6–14 weeks of age. Experiments were conducted with approval of the University of Adelaide Animal Ethics Committee.

Disease models. Mice were immunized for chronic EAE by subcutaneous injection of 100 μ g of MOG_{35–55} (GL Biochem) in phosphate-buffered saline (PBS) emulsified 1:1 in complete Freund's adjuvant, coupled with i.p. injection of 300 ng Pertussis toxin (Sapphire Bioscience) on days 0 and 2. Mice were analysed at clinical scores of 0.5 (onset) and 2–3 (peak) in wild type (WT) mice, where scoring criteria were: 0.5 tremor, 1 partially limp tail, 2 fully limp tail, 2.25 unable to right, 2.5 sprawled hindlimbs, 2.75 one hindlimb paralysed, 3 both hindlimbs paralysed, 3.5 one forelimb paralysed. B16.F10 melanoma cells (provided by Prof. Mark Smyth, QIMR Berghofer, mycoplasma free and verified by short tandem repeat) were cultured in RPMI 1640 containing 10% fetal calf serum (FCS) and 5×10^4 cells were injected subcutaneously into mice at four sites. *S. pneumoniae* EF3030 was grown to a D₆₀₀ of 0.18 in nutrient broth with 10% horse serum at 37 °C 5% CO₂ and stored at -80 °C. Stocks were defrosted and 1×10^6 colony-forming units were delivered intranasally. Bacterial load was determined by serial dilution of concentrated NW onto blood agar with 5 μ g ml⁻¹ gentamicin (Sigma).

Cell preparation. Single-cell suspensions were prepared from lymphoid organs by pressing through 70 μ m filters. Peripheral blood was collected into heparinized Vacutainer tubes (BD). Red blood cells were lysed as required. CNS from PBS-perfused mice was pressed through 70 μ m filters and then separated over a 30/70% Percoll (GE) gradient at 500g. The following digestions were performed at 37 °C with 30 U ml⁻¹ DNase (Sigma). Epidermis and dermis from ears or shaved trunk skin were separated by incubation in 0.375% trypsin for 2 h at 37 °C, and then digested separately with 85 μ g ml⁻¹ Liberase TM (Roche) for 1 h. Tumours and perfused lungs were digested in 2 mg ml⁻¹ collagenase (Sigma) for 1 h. NP tissue between the nose tip and eyes was dissected following removal of nasal-associated lymphoid tissue, digested with 2 mg ml⁻¹ collagenase for 1.5 h and separated by a 40/80% Percoll gradient at 600g. Livers were pressed through 70 μ m filters and then lymphocytes were isolated by a 37.5% Percoll gradient at 850g. Flushed and longitudinally opened small intestine free of Peyer's patches was washed in PBS then incubated in 5 mM EDTA for 40 min at 37 °C to remove epithelium, before remaining lamina propria was digested with 0.5 mg ml⁻¹ collagenase for 1.5 h at 37 °C and separated over a 40/80% Percoll gradient at 1,000g to isolate lamina propria lymphocytes. Peritoneal exudate cells were collected by 3×1 ml PBS washes.

Flow cytometry. Single-cell suspensions were stained in 96-well round- or v-bottom plates (Corning) at 2×10^6 lymphocytes per well using antibodies and other reagents detailed in Supplementary Table 1. For intracellular cytokine staining, cells were first incubated in complete IMDM with 20 μ g ml⁻¹ PMA (Life Technologies), 1 nM ionomycin (Life Technologies) and 1/1,500 GolgiStop (BD) for 4 h at 37 °C. Cells were washed in PBS and stained with Near Infrared fixable dye diluted 1/1,000 (Life Technologies) for 15 min at room temperature. Cells were then washed with FACS buffer (PBS 1% bovine serum albumin 0.04% azide) and blocked with mouse γ -globulin (m γ g) (200 μ g ml⁻¹) for 5 min at room temperature. All subsequent steps were incubated at 4 °C. For purified antibodies, cells were stained with purified antibody for 20–60 min, washed in FACS buffer, stained with secondary antibody in m γ g (200 μ g ml⁻¹) and normal mouse serum (NMS) (1%) for 20 min, washed in FACS buffer and blocked with rat γ -globulin for 15 min. Cells were stained with directly conjugated and biotinylated antibodies for 20 min. In the case of biotinylated antibodies, cells were then washed in FACS buffer and stained with streptavidin for 15 min. Cells were then washed in PBS 0.04% azide. For intracellular cytokine staining, cells were incubated in Cytofix/Cytoperm (BD) for 20 min, washed in Permwash (BD) and stained with intracellular directly conjugated antibodies for 20 min. For transcription factor staining, cells were incubated in Foxp3 kit perm buffer (eBioscience) for 30 min to overnight, and then washed in Foxp3 kit permwash (eBioscience). Cells were then stained with directly conjugated α -transcription factor antibodies in NMS (2%) and normal rat serum (2%), and then washed in PBS 0.04% azide. All stains were washed in PBS 0.04% azide, resuspended in PBS 1% paraformaldehyde and stored at 4 °C in the dark.

Specificity of α -CCR2 was confirmed by negative staining on *Ccr2*^{-/-} γ δ T17 cells. CCR2 and CCR6 gating was determined by relevant isotype controls. For measurement of fluorescence intensity, relevant isotype control geometric mean fluorescence intensity was subtracted from raw geometric mean fluorescence intensity value. For *in vivo* proliferation analysis, mice were given 2 mg BrdU i.p. and then drinking water with 0.8 mg ml⁻¹ BrdU 2% glucose. Following restimulation and surface staining, cells were permeabilized, DNase-treated and stained with α -BrdU (BD) according to the manufacturer's instructions. For *in vitro* proliferation analysis, cells were labelled with Cell Proliferation Dye (eBioscience) according to the manufacturer's instructions. Flow cytometry was

acquired on a BD LSR II or FACSAria and analysed with FlowJo (Treestar). Gating strategies are detailed in Supplementary Fig. 8.

γ δ T17 cell expansion culture and retroviral transduction. Pooled spleen and LN cells were cultured at 1×10^6 cells per ml in RPMI 1640 containing 10% FCS, antibiotics, 1 \times Glutamax (Gibco), 10 mM HEPES (SA Pathology), 1 mM sodium pyruvate, 54 μ M β -mercaptoethanol and 1 \times non-essential amino acids (Gibco) with 5 ng ml⁻¹ recombinant (r)IL-23 (eBioscience), 5 ng ml⁻¹ rIL-1 β (Miltenyi Biotec) and 10 μ g ml⁻¹ α -IFN- γ (BioXCell) in 96-well round-bottom plates coated with 1 μ g ml⁻¹ α -TCR- γ δ (clone GL3; Biologend) for 3 days. Cells were washed and re-seeded on fresh plastic at 1×10^6 cells per ml for a further 3 days as above without TCR- γ δ stimulation. Cells were then washed and re-seeded in 20 ng ml⁻¹ rIL-7 (Peprotech) and 10 μ g ml⁻¹ α -IFN- γ for a further 3 days. pMIG, pMIG-*Rorc* and pMIG-*Ccr6* (cloned from mouse *Ccr6* cDNA) were transfected into EcoPack 2 293 cells (Clontech; mycoplasma free) with Lipofectamine 2000 (ThermoFisher), and supernatant collected after 48 h. γ δ T17 cells at days 4 and 5 of culture were centrifuged at 2,500 r.p.m. (30 °C for 1.5 h) in supernatant with 8 μ g ml⁻¹ polybrene (Sigma) in flat-bottom 96 well trays before being returned to culture.

Adoptive transfers. For *Ccr2*^{-/-} and *Ccr6*^{fl/fl} trafficking experiments, 1–2 $\times 10^6$ each of *in vitro*-expanded F₁ (CD45.1⁺CD45.2⁺) and *Ccr2*^{-/-} (CD45.2⁺), or transduced control and *Ccr6*^{fl/fl} γ δ T17 cells (F₁ or WT), were mixed and transferred i.v. into Ly5.1 (CD45.1⁺) recipient mice d5 post-challenge with B16 melanoma, d8–10 post EAE induction or 24 h post-*S. pneumoniae* infection. γ δ T17 cell infiltration of target organs was analysed 24–48 h post-transfer, and CD45 congenic ratios were normalized to input sample. For *S. pneumoniae* *Tcrd*^{-/-} reconstitution, *in vitro*-expanded WT and *Ccr2*^{-/-} γ δ T17 cells were further purified by MACS (Miltenyi Biotec) before 3×10^6 cells were transferred into separate *Tcrd*^{-/-} hosts 24 h prior to infection. For naive dermis trafficking experiments, 5–10 $\times 10^7$ fresh WT or *Ccr6*^{-/-} lymphocytes or 3×10^7 72 h IL-23/IL-1 β -stimulated WT lymphocytes were transferred into separate unimmunized Ly5.1 mice and analysed 36 h later. Number of recovered cells was normalized to number of γ δ T17 cells transferred.

ELISA. Tumour and NP supernatants from digested samples and supernatants from filtered CNS were supplemented with protease inhibitors (Sigma) and stored at -80 °C. Mouse CCL2 DuoSet ELISA (R&D) was conducted according to the manufacturer's instructions.

qPCR. γ δ T cells from *Il17a*^{Cre} \times *Rose26*^{YFP} mice were enriched by MACS using mouse TCR γ δ ⁺ isolation kit (Miltenyi Biotec, # 130-092-125), and then sorted using a BD FACSAria. Naive CD4⁺ T cells were sorted from WT splenocytes, and skin stromal populations were sorted from digested epidermal and dermal suspensions from WT mice. Sorting strategies are detailed in Supplementary Figure 9. RNA was extracted from sorted cells using Qiagen RNeasy Micro kit (# 74004). For epidermis and dermis, tissues were snap frozen in liquid nitrogen, crushed with mortar and pestle and RNA purified using Qiagen RNeasy Mini kit (# 74104) according to the manufacturer's instructions. cDNA was generated with the Roche Transcriptor First Strand cDNA synthesis kit (# 04896866001). qPCR was performed with Roche LightCycler 480 SYBR Green 1 master mix (# 04887352001) using primer sequences in Supplementary Table 2 on a LightCycler 480 instrument (Roche). Relative gene expression was calculated as $2^{-(\Delta\Delta CT)}$ where reference was *Rplp0*.

Chemotaxis. Splenocytes were rested in complete RPMI for 3–4 h at 37 °C, washed and suspended in chemotaxis buffer (RPMI 0.5% bovine serum albumin 20 mM HEPES). γ δ T17 cells from culture were washed and suspended in chemotaxis buffer. CCL2 or CCL20 (from the late Prof. Ian Clark-Lewis) were diluted in chemotaxis buffer and loaded into the lower chambers of 96-well 5 μ m pore transwell plates (Corning). 2×10^6 splenocytes or 2×10^5 *in vitro*-expanded γ δ T17 cells were loaded into the upper chambers and plates were incubated at 37 °C for 3 h. Lower wells were harvested and stained for flow cytometry. CountBrite beads (Invitrogen) were added to samples prior to acquisition to normalize event counts. Chemotaxis index was calculated as number of gated events divided by number in 0 chemokine control.

***In vitro* stimulation.** Splenocytes were cultured in complete IMDM (10% FCS, pen/strep, l-glutamine, β -mercaptoethanol) at 2.5×10^6 cells per ml with 10 ng ml⁻¹ rIL-23 (eBioscience), 10 ng ml⁻¹ rIL-1 β (Miltenyi Biotec), 20 ng ml⁻¹ rIL-7 (Peprotech), 10 ng ml⁻¹ rIL-12 (R&D), 10 ng ml⁻¹ rIL-18 (R&D) and/or with pre-coating in 1 μ g ml⁻¹ α -TCR- γ δ (Biologend) for up to 72 h at 37 °C. For mitomycin C pre-treatment, cells were first incubated with 10 μ g ml⁻¹ mitomycin C (Sigma) in complete IMDM at 2×10^7 cells per ml for 2 h at 37 °C before extensive washing.

ChIP-seq analysis. ChIP-seq data for IRF4 in CD8⁺ T cells⁴⁵, BATF in CD8⁺ T cells⁴⁴ and IRF4/BATF in Th17 cells⁴³ were previously published and were

obtained from NCBI database using accession codes GSE49930, GSE54191 and GSE40918, respectively. BAM files were loaded and displayed using the IGB genome browser.

Statistics. Data were analysed with GraphPad Prism 6. Appropriate statistical tests were two-sided and used as indicated in figure legends. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$. All replicates are biological except in qPCR experiments, where technical replicates are denoted. Sample sizes were determined empirically to ensure adequate power. *Tcrd*^{-/-} host mice were randomly assigned to groups before receiving adoptive transfers in Fig. 3f. No blinding was utilized. Minimal variance was generally observed between groups; Welch's correction was used in *t*-tests where standard deviations were significantly different. Most data sets were normally distributed; Mann-Whitney and Kruskal-Wallis tests were utilized where data were not normally distributed.

Data availability. Relevant data are available from authors upon reasonable request.

References

- Sutton, C. E. *et al.* Interleukin-1 and IL-23 induce innate IL-17 production from gammadelta T cells, amplifying Th17 responses and autoimmunity. *Immunity* **31**, 331–341 (2009).
- Jensen, K. D. *et al.* Thymic selection determines gammadelta T cell effector fate: antigen-naïve cells make interleukin-17 and antigen-experienced cells make interferon gamma. *Immunity* **29**, 90–100 (2008).
- Kashem, S. W. *et al.* Nociceptive sensory fibers drive interleukin-23 production from CD301b+ dermal dendritic cells and drive protective cutaneous immunity. *Immunity* **43**, 515–526 (2015).
- Cao, J. *et al.* Activation of IL-27 signalling promotes development of postinfluenza pneumococcal pneumonia. *EMBO Mol. Med.* **6**, 120–140 (2014).
- Cho, J. S. *et al.* IL-17 is essential for host defense against cutaneous *Staphylococcus aureus* infection in mice. *J. Clin. Invest.* **120**, 1762–1773 (2010).
- Coffelt, S. B. *et al.* IL-17-producing gammadelta T cells and neutrophils conspire to promote breast cancer metastasis. *Nature* **522**, 345–348 (2015).
- Rei, M. *et al.* Murine CD27(-) Vgamma6(+) gammadelta T cells producing IL-17A promote ovarian cancer growth via mobilization of protumor small peritoneal macrophages. *Proc. Natl Acad. Sci. USA* **111**, E3562–E3570 (2014).
- Cai, Y. *et al.* Pivotal role of dermal IL-17-producing gammadelta T cells in skin inflammation. *Immunity* **35**, 596–610 (2011).
- Roark, C. L. *et al.* Exacerbation of collagen-induced arthritis by oligoclonal, IL-17-producing gamma delta T cells. *J. Immunol.* **179**, 5576–5583 (2007).
- Cai, Y. *et al.* Differential developmental requirement and peripheral regulation for dermal Vgamma4 and Vgamma6T17 cells in health and inflammation. *Nat. Commun.* **5**, 3986 (2014).
- Zhang, Y. *et al.* Migratory and adhesive cues controlling innate-like lymphocyte surveillance of the pathogen-exposed surface of the lymph node. *Elife* **5**, E18156 (2016).
- Nakamizo, S. *et al.* Dermal Vgamma4(+) gammadelta T cells possess a migratory potency to the draining lymph nodes and modulate CD8(+) T-cell activity through TNF-alpha production. *J. Invest. Dermatol.* **135**, 1007–1015 (2015).
- Maeda, Y. *et al.* IL-17-producing Vgamma4+ gammadelta T cells require sphingosine 1-phosphate receptor 1 for Their Egress from the lymph nodes under homeostatic and inflammatory conditions. *J. Immunol.* **195**, 1408–1416 (2015).
- Gray, E. E. *et al.* Deficiency in IL-17-committed Vgamma4(+) gammadelta T cells in a spontaneous Sox13-mutant CD45.1(+) congenic mouse substrain provides protection from dermatitis. *Nat. Immunol.* **14**, 584–592 (2013).
- Ramirez-Valle, F., Gray, E. E. & Cyster, J. G. Inflammation induces dermal Vgamma4+ gammadeltaT17 memory-like cells that travel to distant skin and accelerate secondary IL-17-driven responses. *Proc. Natl Acad. Sci. USA* **112**, 8046–8051 (2015).
- Haas, J. D. *et al.* CCR6 and NK1.1 distinguish between IL-17A and IFN-gamma-producing gammadelta effector T cells. *Eur. J. Immunol.* **39**, 3488–3497 (2009).
- Hirota, K. *et al.* Preferential recruitment of CCR6-expressing Th17 cells to inflamed joints via CCL20 in rheumatoid arthritis and its animal model. *J. Exp. Med.* **204**, 2803–2812 (2007).
- Kara, E. E. *et al.* CCR2 defines in vivo development and homing of IL-23-driven GM-CSF-producing Th17 cells. *Nat. Commun.* **6**, 8644 (2015).
- Mabuchi, T. *et al.* CCR6 is required for epidermal trafficking of gammadelta-T cells in an IL-23-induced model of psoriasiform dermatitis. *J. Invest. Dermatol.* **133**, 164–171 (2013).
- Akitsu, A. *et al.* IL-1 receptor antagonist-deficient mice develop autoimmune arthritis due to intrinsic activation of IL-17-producing CCR2(+)Vgamma6(+)gammadelta T cells. *Nat. Commun.* **6**, 7464 (2015).
- Sumaria, N. *et al.* Cutaneous immunosurveillance by self-renewing dermal gammadelta T cells. *J. Exp. Med.* **208**, 505–518 (2011).
- Gray, E. E., Suzuki, K. & Cyster, J. G. Cutting edge: identification of a motile IL-17-producing gammadelta T cell population in the dermis. *J. Immunol.* **186**, 6091–6095 (2011).
- Hirota, K. *et al.* Fate mapping of IL-17-producing T cells in inflammatory responses. *Nat. Immunol.* **12**, 255–263 (2011).
- Ribot, J. C. *et al.* CD27 is a thymic determinant of the balance between interferon-gamma- and interleukin 17-producing gammadelta T cell subsets. *Nat. Immunol.* **10**, 427–436 (2009).
- Paget, C. *et al.* CD3bright signals on gammadelta T cells identify IL-17A-producing Vgamma6Vdelta1+ T cells. *Immunol. Cell Biol.* **93**, 198–212 (2015).
- Lalor, S. J. *et al.* Caspase-1-processed cytokines IL-1beta and IL-18 promote IL-17 production by gammadelta and CD4 T cells that mediate autoimmunity. *J. Immunol.* **186**, 5738–5748 (2011).
- Michel, M. L. *et al.* Interleukin 7 (IL-7) selectively promotes mouse and human IL-17-producing gammadelta cells. *Proc. Natl Acad. Sci. USA* **109**, 17549–17554 (2012).
- Blink, S. E. *et al.* gammadelta T cell subsets play opposing roles in regulating experimental autoimmune encephalomyelitis. *Cell Immunol.* **290**, 39–51 (2014).
- Kulig, P. *et al.* IL-12 protects from psoriasiform skin inflammation. *Nat. Commun.* **7**, 13466 (2016).
- Li, J., Liu, J., Mao, X., Tang, Q. & Lu, H. IL-7 receptor blockade inhibits IL-17-producing gammadelta cells and suppresses melanoma development. *Inflammation* **37**, 1444–1452 (2014).
- Tang, Q. *et al.* Hmgb1-IL-23-IL-17-IL-6-Stat3 axis promotes tumor growth in murine models of melanoma. *Mediators Inflamm.* **2013**, 713859 (2013).
- Petermann, F. *et al.* gammadelta T cells enhance autoimmunity by restraining regulatory T cell responses via an interleukin-23-dependent mechanism. *Immunity* **33**, 351–363 (2010).
- Fife, B. T., Huffnagle, G. B., Kuziel, W. A. & Karpus, W. J. CC chemokine receptor 2 is critical for induction of experimental autoimmune encephalomyelitis. *J. Exp. Med.* **192**, 899–905 (2000).
- Nagao, K. *et al.* Stress-induced production of chemokines by hair follicles regulates the trafficking of dendritic cells in skin. *Nat. Immunol.* **13**, 744–752 (2012).
- Zhao, X. *et al.* CCL9 is secreted by the follicle-associated epithelium and recruits dome region Peyer's patch CD11b+ dendritic cells. *J. Immunol.* **171**, 2797–2803 (2003).
- Varona, R. *et al.* Molecular cloning, functional characterization and mRNA expression analysis of the murine chemokine receptor CCR6 and its specific ligand MIP-3alpha. *FEBS Lett.* **440**, 188–194 (1998).
- Comerford, I. *et al.* An immune paradox: how can the same chemokine axis regulate both immune tolerance and activation?: CCR6/CCL20: a chemokine axis balancing immunological tolerance and inflammation in autoimmune disease. *Bioessays* **32**, 1067–1076 (2010).
- Charbonnier, A. S. *et al.* Macrophage inflammatory protein 3alpha is involved in the constitutive trafficking of epidermal langerhans cells. *J. Exp. Med.* **190**, 1755–1768 (1999).
- Cretney, E. *et al.* The transcription factors Blimp-1 and IRF4 jointly control the differentiation and function of effector regulatory T cells. *Nat. Immunol.* **12**, 304–311 (2011).
- Newman, D. M., Leung, P. S., Putoczki, T. L., Nutt, S. L. & Cretney, E. Th17 cell differentiation proceeds independently of IRF8. *Immunol. Cell Biol.* **94**, 796–801 (2016).
- Ouyang, X. *et al.* Transcription factor IRF8 directs a silencing programme for TH17 cell differentiation. *Nat. Commun.* **2**, 314 (2011).
- Jain, R. *et al.* Interleukin-23-induced transcription factor Blimp-1 promotes pathogenicity of T helper 17 cells. *Immunity* **44**, 131–142 (2016).
- Ciofani, M. *et al.* A validated regulatory network for Th17 cell specification. *Cell* **151**, 289–303 (2012).
- Kurachi, M. *et al.* The transcription factor BATF operates as an essential differentiation checkpoint in early effector CD8+ T cells. *Nat. Immunol.* **15**, 373–383 (2014).
- Man, K. *et al.* The transcription factor IRF4 is essential for TCR affinity-mediated metabolic programming and clonal expansion of T cells. *Nat. Immunol.* **14**, 1155–1165 (2013).
- Gerner, M. Y., Torabi-Parizi, P. & Germain, R. N. Strategically localized dendritic cells promote rapid T cell responses to lymph-borne particulate antigens. *Immunity* **42**, 172–185 (2015).
- Hammerich, L. *et al.* Chemokine receptor CCR6-dependent accumulation of gammadelta T cells in injured liver restricts hepatic inflammation and fibrosis. *Hepatology* **59**, 630–642 (2014).
- Riol-Blanco, L. *et al.* Nociceptive sensory neurons drive interleukin-23-mediated psoriasiform skin inflammation. *Nature* **510**, 157–161 (2014).
- Barros-Martins, J. *et al.* Effector gammadelta T cell differentiation relies on Master but not auxiliary Th cell transcription factors. *J. Immunol.* **196**, 3642–3652 (2016).

50. Raifer, H. *et al.* Unlike alphabeta T cells, gammadelta T cells, LTi cells and NKT cells do not require IRF4 for the production of IL-17A and IL-22. *Eur. J. Immunol.* **42**, 3189–3201 (2012).
51. Wu, P. *et al.* gammadeltaT17 cells promote the accumulation and expansion of myeloid-derived suppressor cells in human colorectal cancer. *Immunity* **40**, 785–800 (2014).
52. Laggner, U. *et al.* Identification of a novel proinflammatory human skin-homing Vgamma9Vdelta2 T cell subset with a potential role in psoriasis. *J. Immunol.* **187**, 2783–2793 (2011).
53. Caccamo, N. *et al.* Differentiation, phenotype, and function of interleukin-17-producing human Vgamma9Vdelta2 T cells. *Blood* **118**, 129–138 (2011).
54. Lanca, T. *et al.* Protective role of the inflammatory CCR2/CCL2 chemokine pathway through recruitment of type 1 cytotoxic gammadelta T lymphocytes to tumor beds. *J. Immunol.* **190**, 6673–6680 (2013).
55. Brandes, M. *et al.* Flexible migration program regulates gamma delta T-cell involvement in humoral immunity. *Blood* **102**, 3693–3701 (2003).

Acknowledgements

We thank Prof. Ian Frazer (UQ Diamantina Institute, Australia), Prof. Brigitta Stockinger (Francis Crick Institute, UK) and Prof. Christian Engwerda (QIMR Berghofer, Australia) for mice and Prof. Dan Littman (New York University, USA) for pMIG-*Rorc* (Addgene #24069). We also thank the late Prof. Ian Clark-Lewis for chemokine ligands and staff at Laboratory Animal Services from University of Adelaide for animal husbandry. This work was supported by National Health and Medical Research Council project grants 1066781 and 1054925. A.K. is supported by the Sylvia and Charles Viertel foundation.

Author contributions

D.R.M., E.E.K., I.C. and S.R.M. designed experiments, analysed data and wrote the manuscript; D.R.M. performed most experiments; E.E.K., C.R.B., T.S.T., K.A.F., C.E.G. and J.J.W. performed experiments; R.B. and J.C.P. contributed to *S. pneumoniae*

experiments; A.K. and S.L.N. contributed to transcription factor experiments; A.B. contributed to retrovirus experiments; M.M. provided key reagents; J.C.P., A.K., S.L.N., A.B. and M.M. edited the manuscript; I.C. and S.R.M. supervised the study and attained funding.

Additional information

Supplementary Information accompanies this paper at <http://www.nature.com/naturecommunications>

Competing interests: The authors declare no competing financial interests.

Reprints and permission information is available online at <http://npg.nature.com/reprintsandpermissions/>

How to cite this article: McKenzie, D. R. *et al.* IL-17-producing $\gamma\delta$ T cells switch migratory patterns between resting and activated states. *Nat. Commun.* **8**, 15632 doi: 10.1038/ncomms15632 (2017).

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>

© The Author(s) 2017