

# Regulatory immune cells in regulation of intestinal inflammatory response to microbiota

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The intestinal lumen harbors nearly 100 trillion commensal bacteria that exert crucial function for health. An elaborate balance between immune responses and tolerance to intestinal microbiota is required to maintain intestinal homeostasis. This process depends on diverse regulatory mechanisms, including both innate and adaptive immunity. Dysregulation of the homeostasis between intestinal immune systems and microbiota has been shown to be associated with the development of inflammatory bowel diseases (IBD) in genetically susceptible populations. In this review, we discuss the recent progress reported in studies of distinct types of regulatory immune cells in the gut, including intestinal intraepithelial lymphocytes, Foxp3<sup>+</sup> regulatory T cells, regulatory B cells, alternatively activated macrophages, dendritic cells, and innate lymphoid cells, and how dysfunction of this immune regulatory system contributes to intestinal diseases such as IBD. Moreover, we discuss the manipulation of these regulatory immune cells as a potential therapeutic method for management of intestinal inflammatory disorders.

## INTRODUCTION

The gastrointestinal tract, considered as the largest immune interface with the environment, is continuously exposed to multiple antigens from diets, commensal bacteria, and pathogens. With such a high antigen load, a balance is required between inflammatory responses to harmful pathogens and tolerance to commensal flora or food antigens in maintaining immune homeostasis in the healthy gut.<sup>1</sup> Both innate and adaptive immunity have been suggested in maintaining intestinal homeostasis and various factors are involved in this process. The breakdown of this well-controlled balance results in intestinal disorders, such as inflammatory bowel diseases (IBD), food allergy, infectious diseases, diarrhea, and even cancer.

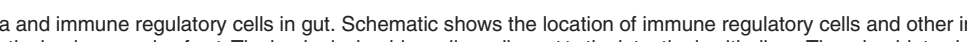
IBD comprises two distinct diseases, Crohn's disease (CD) and ulcerative colitis, and is a chronic relapsing disorder of the intestines caused by an exaggerated immune response to intestinal commensal bacteria in genetically susceptible individuals. However, the lesions and symptoms of IBD are disparate. CD can spread throughout the entire gastrointestinal tract and, thus, patients with CD may present with abdominal pain, diarrhea, weight loss, and fever, whereas ulcerative colitis

only affects the colon, with symptoms of rectal bleeding, frequent stool, tenesmus, and lower abdominal pain. Moreover, IBD patients have a high risk of developing extraintestinal autoimmunity (e.g., arthritis, psoriasis, and primary sclerosing cholangitis).<sup>2</sup> To date, the etiology and pathology of IBD have not been fully elucidated, and multiple factors are considered to play roles in the pathogenesis, including interactions among genetic factors, commensal microbiota, and host immune system.<sup>3</sup> IBD is characterized by the dysregulated immune responses to microbiota in intestinal mucosa, especially by CD4<sup>+</sup> T cell-mediated immune responses. For many years, T helper type 1 (Th1) cells, together with their proinflammatory cytokines (e.g., interferon- $\gamma$  (IFN- $\gamma$ ) and tumor necrosis factor (TNF)), were thought to be responsible for CD pathogenesis, whereas ulcerative colitis was considered to be associated with Th2-like responses, characterized by an enhanced production of interleukin (IL)-5 and IL-13.<sup>4</sup> However, studies on the IL-23/Th17 axis have demonstrated that a subtype of CD4<sup>+</sup> T cells that produce IL-17, called Th17 cells, is an essential player in the pathogenesis of IBD.<sup>5</sup> Recent discoveries have shown that ROR $\gamma$ t (retinoic acid-related orphan nuclear receptor  $\gamma$ t)-dependent innate lymphoid cells (ILCs) are another dominant

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In this review, we will discuss different regulatory cell subsets

The luminal surface of

In the stimulation of bacteria or helminth, IECs can provide

including IL-1 family (e.g., IL-1 $\beta$ , IL-18, IL-33, IL-37), IL-6, IL-8, and some anti-inflammatory cytokines (e.g., IL-10, transforming growth factor- $\beta$  (TGF- $\beta$ ), IL-25). Studies have shown that IL-1 family members possess pleiotropic functions in different phase of inflammation. IL-1 $\beta$  participates in the inflammatory responses by augmenting infiltration of neutrophils, and by activating T cells and ILCs.<sup>13</sup> IL-18, also known as IFN- $\gamma$ -inducing factor, induces the production of proinflammatory cytokines and chemokines,<sup>14</sup> and may also exert a role in suppressing colitis and contributing to wound healing.<sup>15</sup> IL-33 induces Th2 response and drives production of IL-5 and IL-13 by Th2 cells and ILC2. In the chronic dextran sulfate sodium (DSS)-induced colitis model, IL-33 promotes the recruitment of neutrophils for the bacterial clearance<sup>16</sup> and induces regulatory T-cell differentiation in the TGF- $\beta$ -mediated pathway.<sup>17</sup> IL-37, a novel cytokine that is found to suppress innate immune response, could protect mice from colitis through decreasing the production of IL-1 $\beta$  and TNF and inducing expression of IL-10.<sup>18</sup> IL-8, a chemokine that stimulates neutrophil migration, could recruit neutrophils from the circulation to inflammatory sites to clear the pathogens. IEC-derived IL-6 is capable of mediating an inflammatory response through binding to the IL-6R $\alpha$  and gp130 molecules of target cells, such as T cells. IL-6 in synergy with TGF- $\beta$  can induce ROR $\gamma$ t expression and Th17 cell differentiation. However, in the presence of IL-6, TGF- $\beta$ -induced Foxp3 expression is inhibited.<sup>19</sup> Evidences have demonstrated that IL-6<sup>-/-</sup> mice exhibit a mild DSS-induced colitis,<sup>20</sup> and that blockage of gp130 signaling in IECs pronouncedly decreases the recruitment of neutrophils and macrophages to inflamed sites via reducing the production of chemokines or adhesion molecules and inhibiting the invading ability of myeloid cells.<sup>21</sup> Furthermore, IL-6 is also a crucial cytokine that could promote the differentiation of B cells and the secretion of IgA.<sup>22</sup> IL-25 is considered to promote Th2 immune response, and the reduced expression of IL-25 in inflamed mucosa of IBD patients exacerbates intestinal inflammation with the cause that IL-25 could inhibit CD4<sup>+</sup> T-cell activation.<sup>23</sup>

Besides the IECs, there are other specialized cells including goblet cells that can secrete a mucus inner layer to resist bacterial invasion, and Paneth cells that can secrete various antimicrobial proteins to hamper access and survival of bacteria adjacent to the epithelium. The function of mucus in inhibiting bacterial breach was emphasized by studies demonstrating that MUC2<sup>-/-</sup> mice have no bacteria-free zone and can develop intestinal inflammation spontaneously.<sup>24</sup> Moreover, recent study has also shown that mice with the nuclear factor- $\kappa$ B-inducible protein A20 conditional knockout in IECs and myeloid cells (A20<sup>IEC-KO</sup> mice) could develop severe colitis because of impaired goblet cells causing reduced mucus.<sup>25</sup> The IECs of A20<sup>IEC-KO</sup> mice were found to be sensitive to TNF-induced apoptosis. Thus, A20 possesses a dominant anti-apoptotic effect on IECs, although it could negatively regulate nuclear factor- $\kappa$ B signaling that also plays a role in protecting barrier integrity via inhibiting the apoptosis of IECs.<sup>26</sup>

### Intraepithelial lymphocytes

Beneath the intraepithelial lymphocytes (IELs), that is, in the basal layer of IECs, there is a population of dispersed immune cells called IELs. IELs, the exclusive T cells that become in direct contact with the enterocytes, are found to play an essential role in the maintenance of intestinal homeostasis.

IELs are specific T cells that consist of antigen-experienced T-cell subsets bearing T-cell receptor- $\gamma\delta$  (TCR $\gamma\delta$ ) and TCR $\alpha\beta$ , and can be divided into two populations, including natural and induced IELs. Natural IELs that have undergone alternative selection with self-antigens and have differentiated into double-negative TCR $\gamma\delta$ <sup>+</sup> or TCR $\alpha\beta$ <sup>+</sup> cells from triple-positive (CD4<sup>+</sup>CD8 $\alpha\beta$ <sup>+</sup>CD8 $\alpha\alpha$ <sup>+</sup>) cells in thymus directly migrate to the intestinal epithelium.<sup>27</sup> On the other hand, induced IELs that have gone through the conventional selection in thymus and differentiated into naive CD4<sup>+</sup> or CD8 $\alpha\beta$ <sup>+</sup> T cells are activated at mesenteric lymph nodes or Peyer's patches when they accept major histocompatibility complex (MHC) class I peptides and MHC class II peptides, respectively, and then migrate to the intestinal epithelia. The induced IELs include CD4<sup>+</sup>, CD8 $\alpha\beta$ <sup>+</sup>, and CD8 $\alpha\alpha$ <sup>+</sup> TCR $\alpha\beta$ <sup>+</sup> cells.<sup>27</sup> Furthermore, CD8 $\alpha\alpha$  homodimer, a hallmark of T-cell activation, is expressed on the majority of IELs and mainly function as a TCR corepressor.<sup>28</sup> Because of the specific location of IELs in intestinal mucosa, they are involved in resisting to the invasion of foreign pathogens and preserving immune regulation. In response to infection, TCR $\gamma\delta$ <sup>+</sup> IELs can secrete IFN- $\gamma$  and TNF to defend the intestine against pathogens, and importantly to drive proinflammatory effects.<sup>29</sup> Fascinatingly, challenging the conventional view that TCR $\gamma\delta$ <sup>+</sup> IELs are stationary in intestinal epithelium, Edelblum *et al.*<sup>30</sup> have found that TCR $\gamma\delta$ <sup>+</sup> IELs could migrate within the epithelia and into lamina propria dynamically through occludin-occludin interactions between IECs and TCR $\gamma\delta$ <sup>+</sup> IELs in the sites of TCR $\gamma\delta$ <sup>+</sup> IEL/IEC contact. Recently, they also found that TCR $\gamma\delta$ -deficient mice that were infected with *Toxoplasma gondii* had upregulated parasite translocation into the lamina propria in comparison with wild-type mice, suggesting that the migration of TCR $\gamma\delta$ <sup>+</sup> IELs into the lateral intercellular space plays a critical role in host defense response to infection.<sup>31</sup> Therefore, the migration of TCR $\gamma\delta$ <sup>+</sup> IELs exerts a key role in innate immune surveillance and could prevent the translocation of bacteria and pathogens in intestinal epithelia. For intestinal barrier protection, TCR $\gamma\delta$ <sup>+</sup> natural IELs could secrete keratinocyte growth factor-1 to repair the impaired intestinal epithelium.<sup>32</sup> Studies demonstrated that TCR $\gamma\delta$ <sup>+</sup> cell-deficient mice suffered from even more serious DSS-induced colitis because of keratinocyte growth factor deficiencies causing tissue repair delay.<sup>33</sup> In addition, TCR $\gamma\delta$ <sup>+</sup> T cell-deficient mice showed an increase in susceptibility to DSS-induced colitis because of reduced mucin expression and impaired goblet cell function.<sup>34</sup> For immune regulation of intestinal mucosa, TCR $\gamma\delta$ <sup>+</sup> T cells could increase the production of TGF- $\beta$  and IL-10 to perform their suppressive function in intestinal inflammation.<sup>35,36</sup> CD8 $\alpha\alpha$ <sup>+</sup>TCR $\alpha\beta$ <sup>+</sup> IELs have also been reported that provide the protective effects



for intestinal mucosa by expressing some immune regulatory molecules, such as TGF- $\beta$ , lymphocyte activation gene 3, and killer cell immunoglobulin-like receptors.<sup>37</sup> In addition, there is evidence that another subset of IELs, CD8 $\alpha^+\beta^-$  TCR $\alpha\beta^+$  cells, could prevent inflammation in a naive CD4 $^+$ CD45RB<sup>high</sup> T-cell transfer model of colitis in an IL-10-dependent manner.<sup>38</sup>

Collectively, IELs are a critical part of the intestinal immune system, simultaneously participating in the innate and adaptive immune regulation.

## REGULATORY T CELLS

Tregs are a subset of CD4 $^+$  T cells that play an important role in maintaining peripheral tolerance and are involved in the pathogenesis of various clinical disorders. In the gut, regulatory T cells (Tregs) have also been observed to be a primary mediator in maintaining the immune homeostasis. Conventionally, Tregs are known to be generated in the thymus through IL-2 signaling, referred to as thymus-derived Tregs. However, another defined subset of Tregs has been discovered to develop from Foxp3 $^-$ CD25 $^-$ CD4 $^+$  cells in peripheral organs/tissues in the presence of TGF- $\beta$  and all-*trans* retinoic acid, and are identified as peripherally derived Tregs.<sup>39</sup> The surface marker to define Tregs was long unknown until researchers identified the high-affinity IL-2R $\alpha$  chain (CD25) that constitutively expresses in Tregs. However, CD25 was not found to be a unique marker for Tregs, because it is also expressed in activated T cells. The finding of a forkhead box P3 (Foxp3) transcription factor, which is specifically expressed in CD4 $^+$ CD25 $^+$  Tregs, provides a unique marker for Tregs. Foxp3 is not only the marker to define the Treg lineage, but also the key regulatory gene for the development and function of Tregs.<sup>40,41</sup> The level of Foxp3 protein expression in Tregs is critical for their suppressive function, and deficiency of the Foxp3 gene leads to autoimmune diseases, e.g., polyendocrinopathy, enteropathy, and X-linked autoimmunity allergic dysregulation syndrome.<sup>42</sup> Several studies have also suggested that when CD4 $^+$ CD25 $^-$  T cells were transfected with Foxp3 gene, they would possess the behavior like Tregs. Foxp3-transduced T cells have been shown to display suppressive activities and inhibit the development of IBD and autoimmune gastritis in severe combined immunodeficient (SCID) mice.<sup>41</sup> Consistent with these notions, in the presence of TGF- $\beta$ , nonregulatory T cells can be induced to express Foxp3, endowing these cells with the capacity of suppression.<sup>43</sup>

In intestinal lamina propria, Tregs maintain intestine homeostasis through negative regulation of effector T cells and play a critical role in suppressing intestinal inflammation by producing IL-10 or TGF- $\beta$  and highly expressing cytotoxic T lymphocyte antigen (CTLA-4 or CD152) (Figure 1).<sup>44,45</sup> However, the abnormal frequency and function of Tregs have been suggested in the pathogenesis of IBD.<sup>46–48</sup> Tregs have also been reported to remarkably increase in peripheral blood from inactive IBD patients, but decrease in active disease,<sup>46</sup> and the suppressive role of Tregs in autologous CD25 $^-$ CD4 $^+$  T-cell proliferation is impaired in active IBD,<sup>48</sup> consistent with other

discoveries about impaired Treg function in autoimmune conditions such as rheumatoid arthritis.<sup>47</sup> However, it has also been observed that Tregs from both peripheral blood and mucosal lymphoid tissues of IBD patients still retain suppressive activities.<sup>46</sup> Thus, whether the suppressive roles of Tregs are impaired in IBD is still elusive.

Several lines of evidence have shown that the altered cytokine milieu of the intestine contribute to the altered frequency and function of Tregs in IBD. Moreover, an imbalance of Tregs and Th17 exists in IBD. In an environment rich in TGF- $\beta$ , CD4 $^+$ CD25 $^+$ Foxp3 $^+$  Tregs could not only induce CD4 $^+$ CD25 $^-$  T cells into IL-17-producing cells, but also facilitate themselves into Th17 cells. A recent study has also demonstrated that IL-17 $^+$ Foxp3 $^+$ CD4 $^+$  T cells are present in the intestinal mucosa of IBD patients.<sup>48</sup> CD patients had a markedly higher frequency of IL-17 $^+$ Foxp3 $^+$  T cells in inflamed intestinal mucosa and circulating blood compared with the levels in healthy controls. Moreover, CD25 $^+$  regulatory T cells from IBD patients have been demonstrated to have an impaired suppressive function compared with healthy controls that is associated with the coexpression of ROR $\gamma$ t and Foxp3 in IL-17 $^+$ Foxp3 $^+$  T cells. Our recent work has also demonstrated that psychological stress-derived prolactin is able to induce DCs to secrete IL-6 and IL-23 and facilitate Tregs to differentiate into Foxp3 $^+$ TNF $^+$ IL-17 $^+$  T cells, further aggravating intestinal inflammation. In addition, this study highlights the influence of a cytokine environment on immune regulation in gut mucosa.<sup>49</sup>

IBD are associated with upregulation of TNF and, accordingly, anti-TNF therapy is now considered an effective therapy for IBD. Accumulating evidence indicates that anti-TNF therapy has an effect on Tregs. The frequency of peripheral blood Tregs in IBD patients was increased after anti-TNF therapy, and this treatment was able to enhance the number of Tregs in the peripheral blood from chronic IBD cases by reducing the apoptosis of these cells.<sup>50</sup> In addition, *in vitro* experiments have indicated that the suppressive role of Tregs in regulating effector T cells was enhanced after an anti-TNF injection, and Tregs from IBD patients after anti-TNF therapy also displayed an enhanced ability to suppress effector CD4 $^+$  T cells *in vivo*.<sup>51</sup>

With respect to the role of bacteria, the commensal bacteria play a crucial role in the maintenance of gut homeostasis and, moreover, abnormal bacteria composition is considered to be one of the pathogenic factors in IBD. Intestinal microbiota regulate the development of the immune system and play an critical role in the induction of Tregs.<sup>52</sup> *Bacteroides fragilis* and *Clostridia* species have been found to suppress an inflammatory response in the intestine by promoting Tregs, and the strain of *Clostridium butyricum* is also found to induce IL-10-producing macrophage in intestine and prevent DSS-induced acute colitis in mice,<sup>53</sup> whereas segmented filamentous bacteria could enhance the Th17 response to protect the intestines from infection (e.g., *Citrobacter rodentium*) in mice.<sup>54</sup> Multiple mechanisms have been found to be involved in microbiota regulation of Treg differentiation and function. For example, polysaccharide A of *B. fragilis* could promote the suppressive

activity of inducible Foxp3<sup>+</sup> Tregs via Toll-like receptor 2 signaling and protect the TNBS (2,4,6-trinitrobenzenesulfonic acid)-induced colitis in mice.<sup>55</sup> Different bacterial species could regulate the cytokine milieu to affect T-cell differentiation and activation. IL-6, IL-21, IL-23, and IL-27 could prevent Treg development, but facilitate the induction of proinflammatory Th1/Th17 cells.<sup>56,57</sup> Of note, the same commensal bacteria in a different context are also able to induce varying immune responses. *B. fragilis*, unbalanced *Clostridia* flora, and altered *Schaedler* flora species tend to exert a suppressive function by inducing Tregs in normal conditions of the intestine, whereas they could enhance Th1/Th17 cell development under the context of an absence of Tregs.<sup>54,58</sup> Furthermore, recent work has also indicated that the bacterial metabolites are involved in regulating the gut immune response. Butyrate, a colon microbial fermentation product, could induce the differentiation of Tregs in the colonic lamina propria and ameliorate T cell-dependent experimental colitis in mice.<sup>59</sup> Butyrate could also downregulate the production of proinflammatory mediators (e.g., IL-6, IL-12, nitric oxide) of lamina propria macrophages via inhibiting histone deacetylases, furthering the importance of butyrate in maintenance of tolerance to intestinal microbiota.<sup>60</sup> In addition, previous work has demonstrated that butyrate could impart anti-inflammatory properties on colonic macrophages and DCs to induce differentiation of Tregs and IL-10-producing T cells through activation of Gpr109a signaling.<sup>61</sup>

In addition to Foxp3<sup>+</sup> Tregs, other distinct sets of Tregs have also been discovered to be involved in immune regulation in gut mucosa. The type 1 regulatory T (Tr1) cells, induced by the presence of IL-10, are a different class of CD4<sup>+</sup> Tregs from that of Foxp3<sup>+</sup> Tregs. Tr1 cells display their suppressive function via the production of IL-10 and TGF- $\beta$ , whereas the lack of Foxp3 expression is observed in this population.<sup>62</sup> Previous work has demonstrated that the IL-10-producing Tr1 cells could suppress the antigen-specific proliferation of naive CD4<sup>+</sup> T cells and cotransfer of Tr1 T-cell clones could prevent experimental colitis in SCID mice induced by naive CD4<sup>+</sup> CD45RB<sup>high</sup> splenic T cells or microbiota antigen-specific Th1 cells.<sup>63,64</sup> As an anti-inflammatory cytokine, IL-10 is able to directly inhibit cytokine production (e.g., TNF), to indirectly suppress the proliferative responses of antigen-specific CD4<sup>+</sup> T effector cells, and maintain the persistence of Foxp3 expression in Tregs.<sup>65</sup> In addition, IL-10 can limit inflammation by indirectly suppressing the antigen-presenting ability of distinct types of professional antigen-presenting cells (APCs). Recently, a phase I/IIa clinical trial has been performed, in which the efficacy and safety of ovalbumin-specific Tr1 cells were assessed in the treatment of patients with refractory CD, showing that injections of these cells significantly reduced the Crohn's Disease Activity Index of CD patients.<sup>66</sup>

Th3 cells are a unique subset of CD4<sup>+</sup> regulatory cells, by which TGF- $\beta$  is required for their biologic activity and regulatory functions, and which are determined during the course of investigating mechanisms of oral tolerance. Like Tr1 and Foxp3<sup>+</sup> Tregs, Th3 cells secrete a large amount of TGF- $\beta$ ,

low level of IL-4 and IL-10, but no IFN- $\gamma$  or IL-2.<sup>67</sup> Evidences have been reported that antigen-specific TGF- $\beta$ -producing Th3 cells could maintain peripheral tolerance by promoting the differentiation of antigen-specific Foxp3<sup>+</sup> regulatory cells, and adoptive transfer of myelin oligodendrocyte glycoprotein TCR-Tg  $\times$  TgTGF- $\beta$  T cells into wild-type mice induced autoreactive Th3 cells to ameliorate experimental autoimmune encephalomyelitis (EAE) in mice.<sup>68</sup>

## REGULATORY B CELLS

B cells are typically characterized by their capacity to produce antibodies. In addition, they release cytokines and serve as secondary APCs. As with T cells, B cells possess distinct subpopulations that perform pathogenic and regulatory functions, respectively. In 2002, Bhan and Mizoguchi used the term "Bregs" to describe the IL-10-producing B cells, a subpopulation of B cells capable of maintaining immune homeostasis and performing the suppressive function chiefly depending on their secretion of IL-10 (**Figure 1**).<sup>69</sup> Regulatory B cells (Bregs) not only regulate the balance of Th1, Th17, and Tregs, and subsequently inhibit inflammation, but also can convert effector T cells into Tr1 cells. Accumulating evidence has demonstrated that Bregs play a regulatory role in certain autoimmune diseases. In the myelin oligodendrocyte glycoprotein-induced EAE mouse model, B cell-deficient mice failed to enter the remission phase, whereas transfer of IL-10<sup>+</sup> B cells could inhibit EAE.<sup>69</sup> IL-10-producing B cells in mesenteric lymph nodes regulate the progression of chronic colitis of TCR $\alpha$  knockout mice by directly downregulating inflammatory cascades associated with IL-1 $\beta$  production and signal transducer and activator of transcription 3 activation.<sup>8</sup> The intact gut microbiota could also facilitate IL-10-producing B-cell differentiation in the spleen and mesenteric lymph nodes by increasing IL-1 $\beta$  and IL-6 production in mice.<sup>70</sup> In humans, Bregs have been studied primarily in systemic lupus erythematosus and multiple sclerosis, and it was reported that these cells could regulate autoimmune responses.<sup>71</sup> With regard to IBD, evidence has shown that the frequency of CD19<sup>high</sup>CD1d<sup>high</sup> IL-10-producing B cells was reduced in patients with CD, and that this subset of cells could reduce intestinal inflammation in colitis in a Treg-independent manner.<sup>72</sup> Most recently, IL-33 was found to induce a subset of IL-10-producing B cells (Breg<sup>IL-33</sup>) in mice, and adoptive transfer of these Breg<sup>IL-33</sup> into IL-10<sup>-/-</sup> mice effectively attenuated mucosal inflammatory responses in the intestines.<sup>73</sup> Accordingly, evidence has also been generated that certain Breg subsets in performing their suppressive function are dependent on factors other than IL-10. In addition, TGF- $\beta$ -producing B cells were observed to have regulatory function in allergic diseases.<sup>74</sup> CD19<sup>+</sup>CD5<sup>+</sup>Foxp3<sup>+</sup> B cells, a newly discovered type of Bregs, also express Foxp3, but their immunologic function remains unclear.<sup>75</sup> Furthermore, a novel IL-10-independent immunoregulatory role of B cells has also been found in suppressing autoimmunity, such as EAE, by promoting the expansion of Tregs through GITRL (glucocorticoid-induced TNF ligand).<sup>76</sup> Moreover, B cells are found to regulate the immunity and promote

the recovery of EAE by secreting IL-35. Importantly, CD138<sup>+</sup> plasma cells, a subset of activated B cells, produce IL-35 and could prevent intestinal inflammation after salmonella infection. Therefore, these regulatory B cells or plasma cells may provide a novel therapy for IBD.<sup>77</sup>

### INNATE LYMPHOID CELLS

ILCs are recently characterized members of hematopoietic cells of the innate immune system that have been unappreciated for a long period, partly owing to the lack of surface markers distinct from other immune cell lineages. As more and more research work is done in this field, ILCs are emerging to be a brand new star. According to their cytokine profile, ILCs have been classified into three subsets.<sup>78</sup> The group 1 ILC population, comprising ILC1 and natural killer cells, express T-bet, IFN- $\gamma$ , and TNF. The group 2 ILC, similar to Th2 cells, expresses ROR $\alpha$  and GATA3 and could produce Th2 cell-associated cytokines. ILC2 cells consist of natural helper cells, nuocyte, innate helper 2 cells, and multipotent progenitor type 2 cells. In response to IL-25 and IL-33 that are produced by IECs subjected to the infection of helminth, ILC2 could secrete IL-5 and IL-13 to resist the parasite infection.<sup>6,79</sup> Moreover, studies have shown that lung ILC2 could produce amphiregulin to promote the lung barrier integrity and tissue remodeling after influenza virus-induced damage.<sup>80</sup> Analogous to Th17 cells, the group 3 ILC population requires the transcription factor ROR $\gamma$ t, and they produce IL-22, IL-17A, and IL-17F.<sup>78</sup> This group of ILC comprises ILC3 and lymphoid tissue-inducer cells. The lymphoid tissue-inducer cells are essential in secondary lymphoid organ formation during fetal development. A recent report<sup>81</sup> on dietary effects on the developing immune system has revealed that mouse fetal ILC3 are controlled by both maternal retinoid intake and retinoic acid signaling in a cell-autonomous manner. Although ILCs, particularly ILC1, have been described to participate in the regulation of immune responses in the intestine, the underlying mechanism remains unclear. ROR $\gamma$ t<sup>+</sup> ILC3 have been recently demonstrated as possibly serving as APCs, owing to the expression of MHC II, but unlike the conventional APCs that do not induce T-cell proliferation. Interestingly, the deletion of MHC II on ROR $\gamma$ t<sup>+</sup> ILC3 led to commensal bacteria-dependent intestinal inflammation, suggesting that ILC3 restrain intestinal CD4<sup>+</sup> T cell responses to commensal bacteria in an MHC II-dependent manner.<sup>82</sup> IL-22, important to maintaining intestinal homeostasis, directly induces epithelial defense through the expression of antimicrobial peptides.<sup>83</sup> As the main source of intestinal IL-22, ROR $\gamma$ t<sup>+</sup> ILC3 are implicated in the protection and recovery from DSS-mediated epithelial damage and colitis.<sup>84</sup> Furthermore, ILC3 can also work as potent innate immune effector cells and facilitate resistance to intestinal pathogens (Figure 1).<sup>85</sup> IL-22-producing ROR $\gamma$ t<sup>+</sup> ILC3 show the ability to enhance resistance to Gram-negative extracellular bacterial infection at the surfaces of the lungs and intestines. Collectively, identification of the previously unappreciated family of ILCs, which displays various functions in innate and adaptive immunity, might

provide new insight into the regulation of intestinal immune responses.

### ALTERNATIVELY ACTIVATED MACROPHAGES

The innate immune cells that are considered to exert the function of bacterial killing, antigen presentation, and proinflammatory cytokine secretion also possess regulatory functions, such as regulatory macrophages and DCs. Macrophages display plasticity in their activation profile under different cytokine conditions. Classically activated macrophages (or M1) induced by IFN- $\gamma$  and microbial products are characterized by microbicidal activity and proinflammatory properties. However, alternatively activated macrophages (AAM; or M2), induced by IL-4 or IL-13, display an anti-inflammatory phenotype.<sup>86</sup> In humans, AAMs are characterized by a different set of cell surface receptors such as CD163, CD206, DECTIN-1, chemokines (CCL16, CCL17, CCL18, CCL22, CCL24), and chemokine receptors (CCR2, CXCR1, CXCR2). In mice, AAMs also express some specific effector molecules that are induced upon exposure to IL-4/IL-13, such as arginase-1, Fizz1, Ym1, and Ym2.<sup>87,88</sup> In addition to IL-4 and IL-13, several other cytokines are also found to be involved in AAM induction indirectly through Th2 cells. IL-25 has been found to increase the gene expression of AAMs, and transfer of IL-25-induced AAMs could ameliorate TNBS colitis in mice.<sup>89</sup> In addition, IL-33 could bind to ST2L to increase Th2 cytokine production and AAM induction.<sup>90</sup> Similar to Th2 cells, ILC2 could also respond to epithelium-derived IL-33 and facilitate AAM polarization by producing IL-13 and IL-5.<sup>91</sup>

AAMs have been found to inhibit the proliferation of T cells and suppress protozoan infection via cell-to-cell contact or production of regulatory cytokines (e.g., IL-10 and TGF- $\beta$ ).<sup>87</sup> Therefore, AAMs mainly participate in Th2 cell responses (e.g., chronic obstructive pulmonary disease, asthma) and play a key role in immune regulation (Figure 1).<sup>92</sup> Accumulating evidence has implied that AAMs could regulate intestinal inflammation. The initial data related to the suppressive function of AAMs in intestinal inflammation come from an interesting phenomenon by which helminth worm infections ameliorate the severity of colitis in mice.<sup>93</sup> In line with this, a clinical trial has demonstrated the efficacy of the intestinal helminth *Trichuris suis* in the treatment of active CD patients.<sup>94</sup> Moreover, injection of AAM significantly ameliorated the severity of dinitrobenzene sulfonic acid-induced colitis in mice by increasing IL-10 production,<sup>10</sup> indicating that AAMs could maintain the intestinal homeostasis and resist inflammation.

Recent work has shown that active CD patients had low amount of CD68<sup>+</sup>CD206<sup>+</sup> macrophages (namely AAMs), whereas colonic biopsies from inactive CD patients had increased numbers of these cells.<sup>10</sup> In addition, evidence has shown that anti-TNF therapy could induce CD14<sup>+</sup> monocytes into AAMs *in vitro* when the CD14<sup>+</sup> monocytes were cocultured with CD4<sup>+</sup> T cells, and further that these AAMs could inhibit the activity of CD4<sup>+</sup> T cells.<sup>95</sup> Consistent with this, anti-TNF therapy was found to promote AAM infiltrate in intestinal mucosa of active CD patients.<sup>95</sup> The later



finding certainly implies that AAMs may be involved in the process of IBD, indicating that adoptive transfer of AAMs from patients' blood could be a novel approach in the treatment of IBD.<sup>10</sup> Our recent data implicated that monocyte could also develop anti-inflammatory properties with producing IL-10 by the induction of insulin-like growth factor-1, and this insulin-like growth factor-1-primed monocyte has the capacity of ameliorating mouse colitis. However, there is no evidence to show that this group of cells possesses the phenotypes like AAMs.<sup>96</sup>

## REGULATORY DENDRITIC CELLS

It has been long established that DCs are one of the most important components of immune system. As a specialized type of APCs, DCs are considered to be the most effective APCs to initiate primary immune responses and be able to link innate and adaptive immunity. After "meeting" foreign intruders, DCs take up antigens, present antigens to T cells, activate them, and further influence the type of immune response.

In recent years, the central role of DCs in inducing inflammatory responses against pathogens and tolerance to commensal microflora in the gut has been established with studies of DC function. There are several DC subsets located at intestinal mucosa that play a crucial role in the maintenance of the gut homeostasis. According to the expression of the surface marker CD103, intestinal DCs can be divided into two predominant subsets: CD11c<sup>high</sup>CD103<sup>+</sup> and CD11c<sup>high</sup>CD103<sup>-</sup> DCs.<sup>97</sup> Mesenteric lymph node CD11c<sup>high</sup>CD103<sup>-</sup> DCs have been reported to increase in murine colitis and exhibit proinflammatory properties. Moreover, Opn-expressing CD11c<sup>high</sup>CD103<sup>-</sup> DCs possess the ability to induce Th1 and Th17 cell development.<sup>97-99</sup> In contrast, one of the major functions of DCs is to help generate Tregs in the gut from naive T cells. CD11c<sup>high</sup>CD103<sup>+</sup> DCs have been known for their regulatory functions in the intestine. CD103<sup>+</sup> DCs derived from the lamina propria of the small intestine have been found to facilitate the differentiation of Tregs in a retinoic acid- and TGF- $\beta$ -dependent manner.<sup>100</sup> However, recently discovered intestinal CX3CR<sup>high</sup>CD11b<sup>+</sup>CD11c<sup>+</sup> cells, a subset of CX3CR<sup>+</sup>CD11b<sup>+</sup>CD11c<sup>+</sup> cells that mediate the inflammatory responses, are found to inhibit the proliferation of CD4<sup>+</sup> T cells and prevent intestinal inflammation.<sup>101</sup> Furthermore, a type of Tr1-like cells, which exert suppressive functions mainly by secreting IL-10, has been reported to be promoted by CD8 $\alpha$ <sup>+</sup> DCs with a CD11c<sup>low</sup>B220<sup>+</sup> phenotype, another subpopulation of DCs (Figure 1).<sup>102</sup>

Although the mechanism whereby intestinal DC maintains intestinal homeostasis remains unclear, great progress has been made over the past decade. On one hand, studies have shown that DCs could induce the generation of Tregs directly in the presence of retinoic acid or, as recently reported, CD31-conditioned DC could contribute to the development of CD25<sup>+</sup>Foxp3<sup>+</sup> T cells dependent on the signaling of coinhibitory receptor CD31.<sup>103</sup> On the other hand, DCs could combine with other factors to jointly maintain gut homeostasis. When DCs interact with commensal flora, they amplify their immune regulatory functions. Recently, MUC2 secreted by

goblet cells has been found to imprint DCs with anti-inflammatory abilities.<sup>104</sup> It augments DC transcription and secretion of TGF- $\beta$ 1 and retinaldehyde dehydrogenase (RALDH or ALDH1), an enzyme that converts vitamin A into RA and helps CD103<sup>+</sup> DCs to induce Tregs. Moreover, recent studies have highlighted the interaction of DCs and subsets of ILCs in regulating intestinal homeostasis.<sup>105,106</sup> In the steady state of the gut, after sensing and uptaking microbiota or their products, macrophages secrete IL-1 $\beta$  to stimulate ROR $\gamma$ t<sup>+</sup> ILC3 to produce colony-stimulating factor 2 that further promotes DCs to maintain Tregs by releasing RA.<sup>107</sup> Commensal microbiota, tissue-resident mononuclear phagocytes, and ILCs thus form an axis to regulate intestinal homeostasis. CD11b<sup>+</sup> classical DCs as a source of IL-23 are required for ILC3 activation, that are rendered as a type of immune regulatory cells characterized by producing IL-22, and inhibiting commensal bacteria-specific CD4<sup>+</sup> T-cell proliferation via MHC II-mediated antigen presentation.<sup>106</sup>

## CONCLUSION

Innate and adaptive immune systems maintain intestinal homeostasis, and regulatory immune cells, including IELs, Tregs, B cells, macrophages, DCs, and ILCs, play a key role in preventing the inappropriate immune responses. However, evidence has been indicated that the existence of an imbalance of proinflammatory and immunosuppressive cells is present in IBD. Although the cytokine milieu is considered to regulate the differentiation of immune cells, the exact mechanism that leads to the dysfunction of regulatory cells in IBD remains unclear. Currently, the main therapies for IBD rely on reducing proinflammatory stimuli, such as anti-TNF treatment. However, restoring the intestinal homeostasis by increasing the frequency and function of immune regulatory cells should be a novel therapeutic approach in the management of IBD. Many studies have demonstrated that the adoptive transfer of these regulatory cells to animal models efficiently alleviates experimental colitis. Therefore, therapy of human IBD with regulatory cells seems warranted.

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## DISCLOSURE

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